

Station B: cleanNA Clean Viral DNA & RNA kit

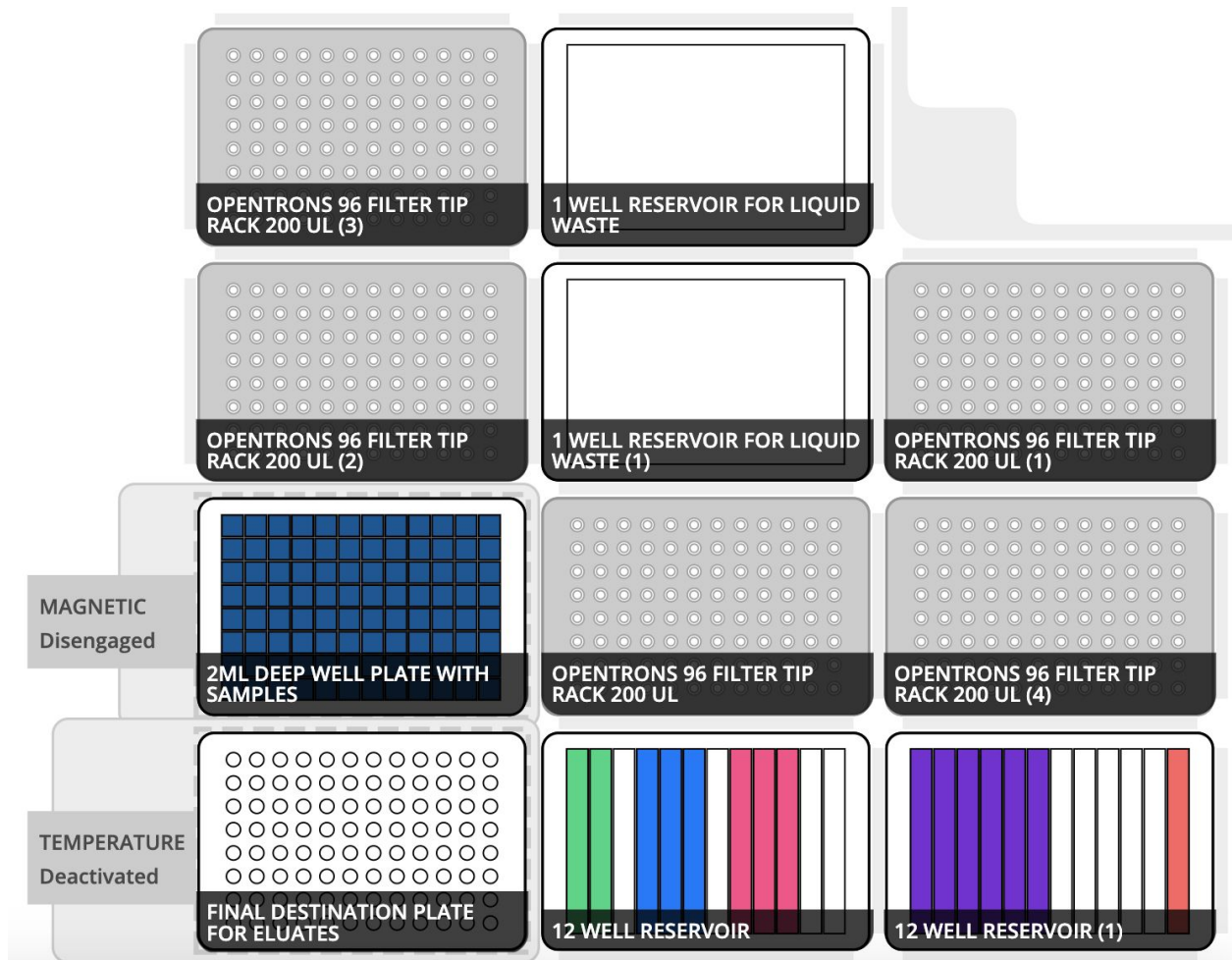
Code parameters:

- Change the sample number on line 11 (default is 8, max is 94)
- Change the elution volume on line 13 (default is 50µl)
- Change the starting volume on line 14 (default is 200µl)

Pipettes:

- P300 multichannel on the left mount

Deck layout:



Labware and module requirements:

- 1 x magnetic module
- 1 x temperature module
- 1 x 2mL deep well plate [input with samples]
- 10 x 200µl filter tipracks


- 2 x 12 well reservoirs
- 2 x 12 well reservoir **[left empty for liquid waste]**
- 1 x 96 well aluminum block loaded on top of the temperature module in slot 1
- 1 x 96 well PCR plate OR PCR strip tubes to match the number of samples **[output with eluates/extractions]**

Volume requirements:


Note: the below volumes account for a 10% overage - the dead volume can be adjusted depending on the calibration of the pipette to the labware, but it is recommended to have an overage of at least 10%


| Reagents | Volume per sample (µl) | Volume for 8 samples (µL) | Volume for 48 samples (mL) | Volume for 96 samples (mL) |
|--|------------------------|---------------------------|----------------------------|----------------------------|
| VDR Lysis buffer + Carrier RNA VDR | 240uL + 1uL | 2,880 + 12uL | 12.4mL + 52uL | 26.6mL + 105uL |
| Isopropanol and CleanNA Particles VDR | 280uL + 10uL | 3,360uL + 120uL | 24.5mL + 520uL | 28mL + 1mL |
| VDR Wash buffer | 350 | 4,200 | 18.2 | 35 |
| 80% Ethanol | 700 | 8,400 | 36.4 | 70 |
| Nuclease-free Water | 50 | 600 | 2.6 | 5 |


Slot 2 - 12 well reservoir


 LABWARE DETAILS

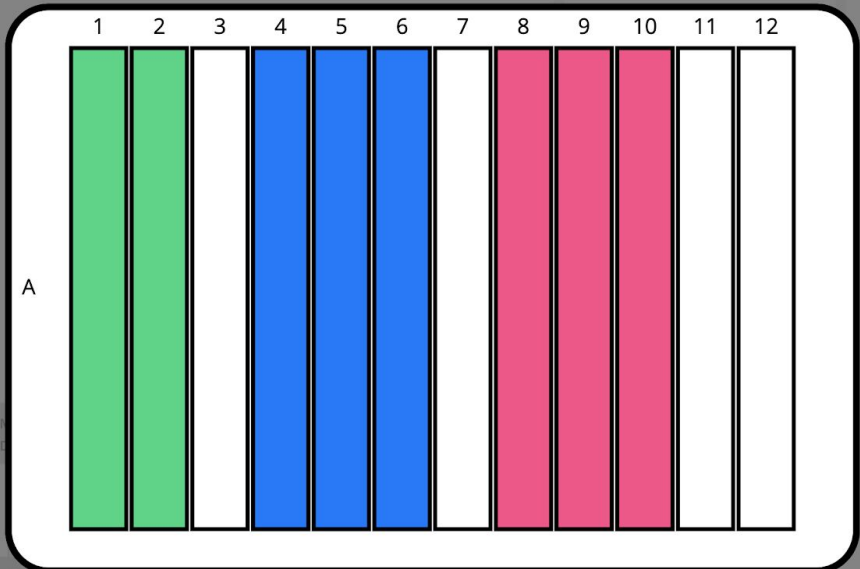
Type NEST 12 Well Reservoir 15 mL

Nickname 12 well reservoir 


 VDR LYSIS BUFFER + CARRIER RNA
VDR

 ISOPROPANOL AND CLEANNA
PARTICLES VDR


 VDR WASH BUFFER





Slot 3 - 12 well reservoir

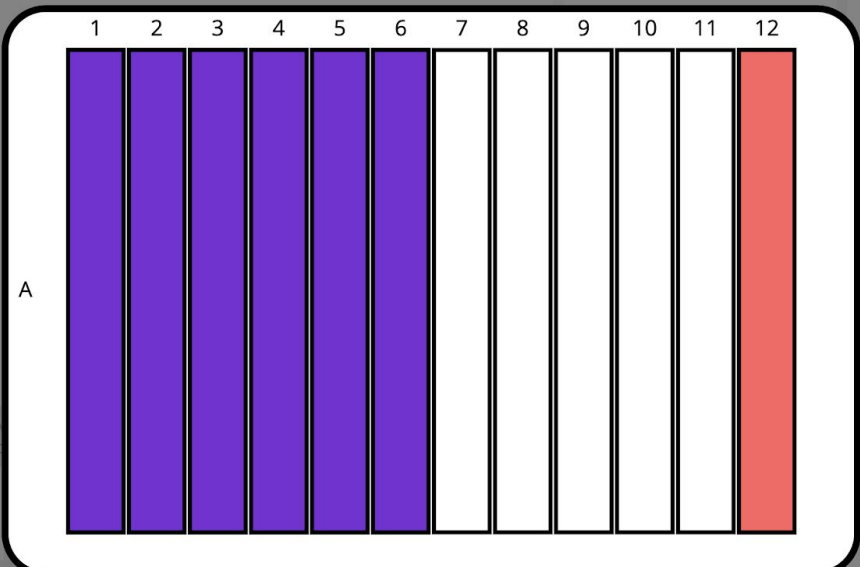
 LABWARE DETAILS

Type NEST 12 Well Reservoir 15 mL

Nickname 12 well reservoir (1) 

 80% ETHANOL

 NUCLEASE-FREE WATER



Before you begin:

1. Pre-cool the Temperature Module in the Opentrons App to 4°C
2. Add the buffers to the appropriate wells in the 12 well reservoirs
3. Place the deep well plate of samples from Station A to on top of the magnetic module in slot 4.
4. Add a 96 well aluminum block and the 96 well PCR plate or PCR strip tubes on top of the temperature module

The final plate of eluates/extractions will be found on top of the temperature module in slot 1. Once the run is complete, please proceed to Station C for RT-qPCR set up.