Station B: cleanNA Clean Viral DNA & RNA kit

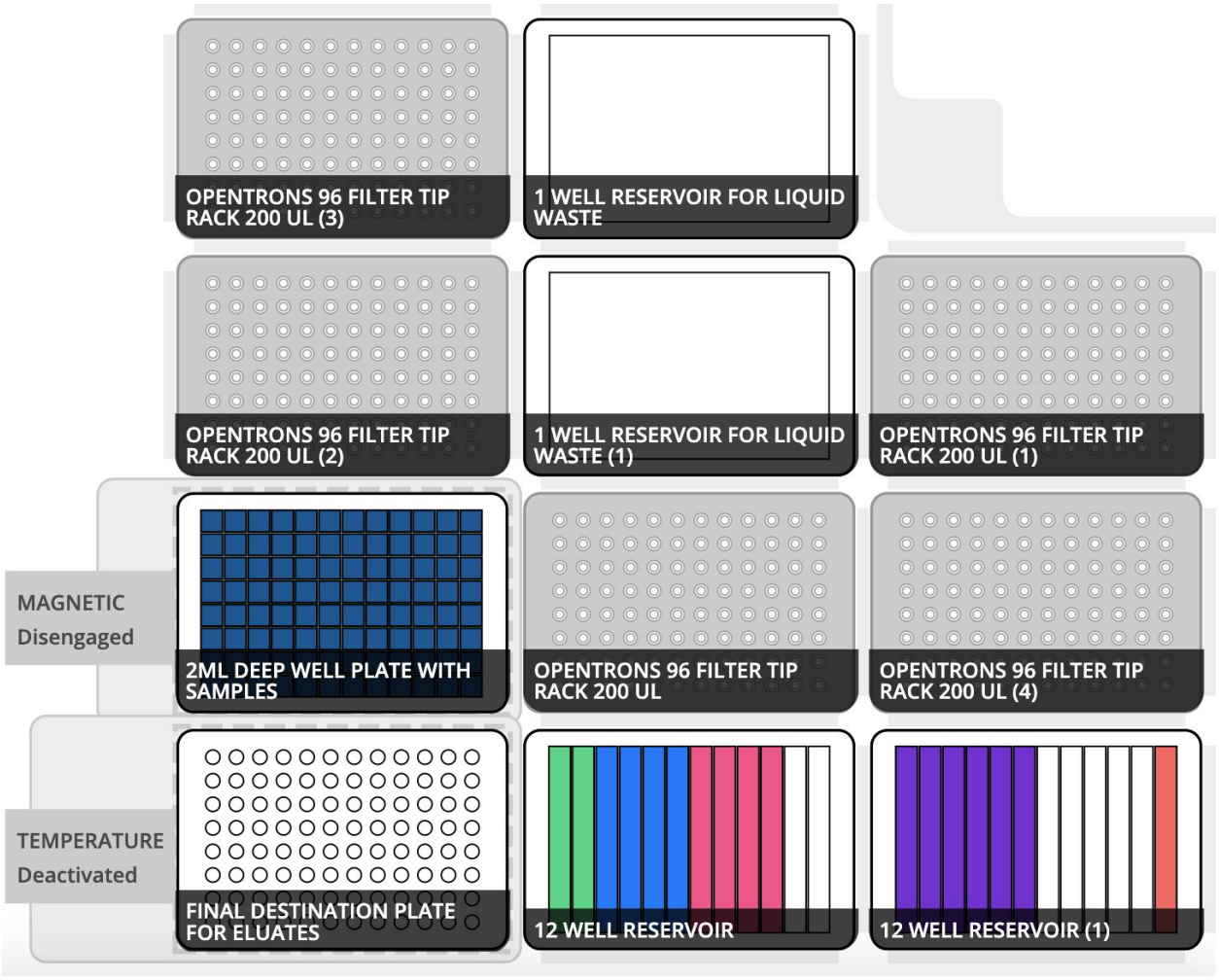
Code parameters:

- Change the sample number on line 11 (default is 8, max is 94)
- Change the elution volume on line 13 (default is 50µl)
- Change the starting volume on line 14 (default is 200µl)

Pipettes:

- P300 multichannel on the left mount

Deck layout:



Labware and module requirements:

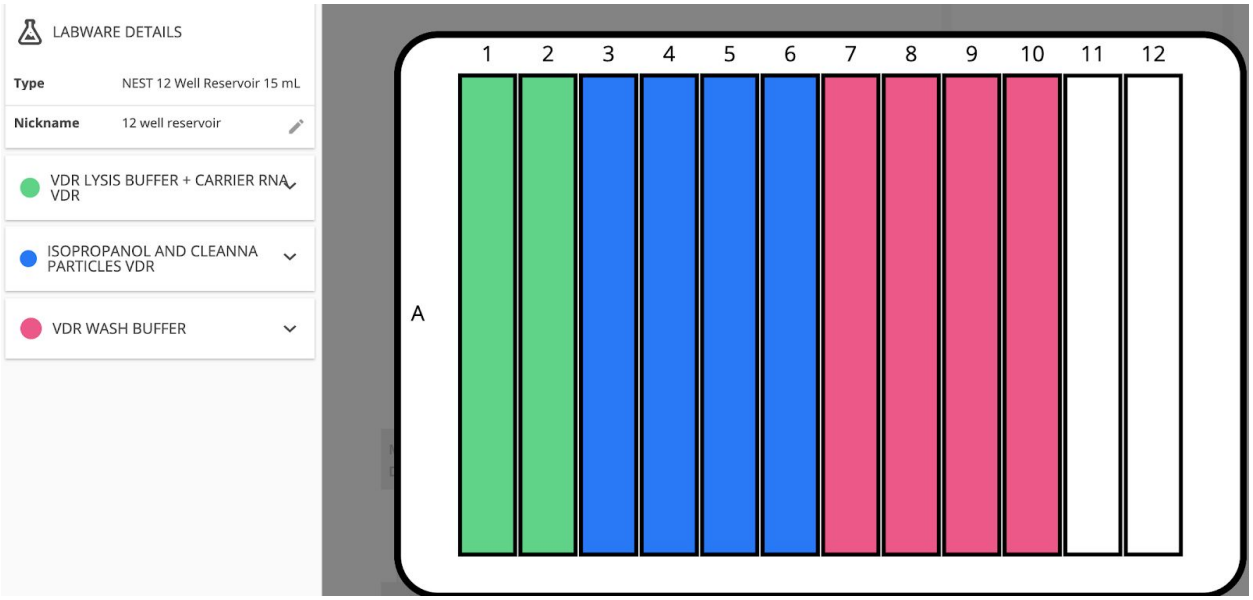
- 1 x magnetic module
- 1 x temperature module
- 1 x 2mL deep well plate **[input with samples]**
- 10 x 200µl filter tipracks

- 2 x 12 well reservoirs
- 2 x 12 well reservoir **[left empty for liquid waste]**
- 1 x 96 well aluminum block loaded on top of the temperature module in slot 1
- 1 x 96 well PCR plate OR PCR strip tubes to match the number of samples **[output with eluates/extractions]**

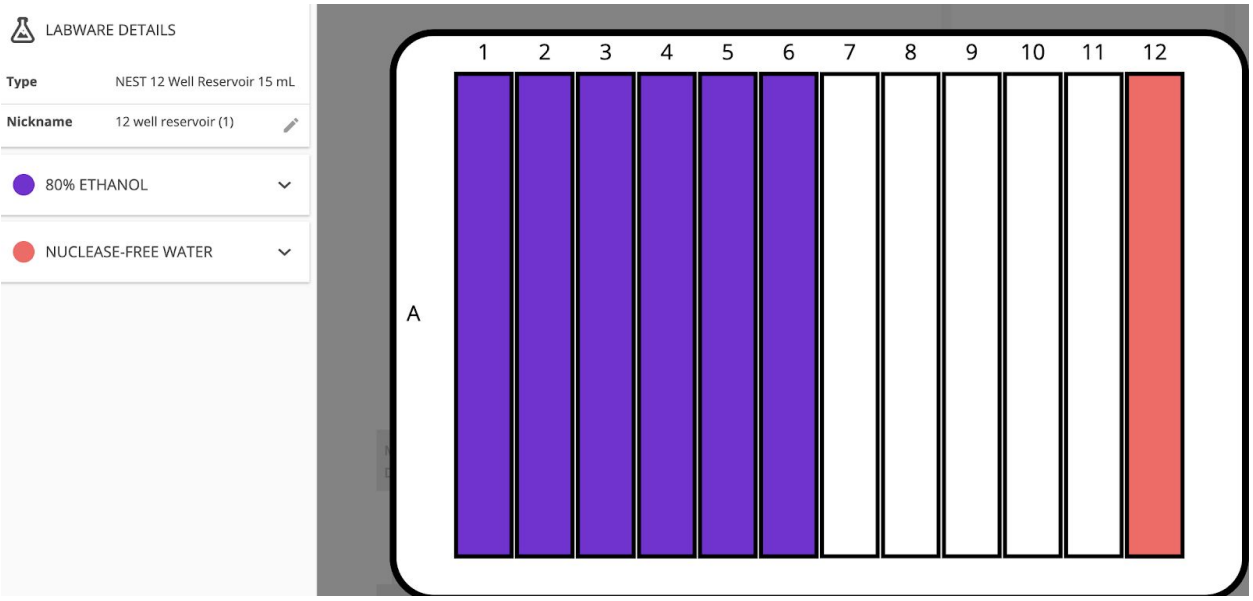
Volume requirements:
Note*: the below volumes account for a 10% overage - the dead volume can be adjusted depending on the calibration of the pipette to the labware, but it is recommended to have an overage of at least 10%*

Reagents	**Volume per sample (µl)**	**Volume for 8 samples (µL)**	**Volume for 48 samples (mL)**	**Volume for 96 samples (mL)**
VDR Lysis buffer + Carrier RNA VDR	240uL + 1uL	2,880 + 12uL	12.4mL + 52uL	26.6mL + 105uL
Isopropanol and CleanNA Particles VDR	280uL + 10uL	3,360uL + 120uL	24.5mL + 520uL	28mL + 1mL
VDR Wash buffer	350	4,200	18.2	35
80% Ethanol	700	8,400	36.4	70
Nuclease-free Water	50	600	2.6	5

Slot 2 - 12 well reservoir



Slot 3 - 12 well reservoir



Before you begin:

1. Pre-cool the Temperature Module in the Opentrons App to 4°C
2. Add the buffers to the appropriate wells in the 12 well reservoirs
3. Place the deep well plate of samples from Station A to on top of the magnetic module in slot 4.
4. Add a 96 well aluminum block and the 96 well PCR plate or PCR strip tubes on top of the temperature module

The final plate of eluates/extractions will be found on top of the temperature module in slot 1. Once the run is complete, please proceed to Station C for RT-qPCR set up.