Station C: Allplex 2019-nCov Detection Assay - Seegene

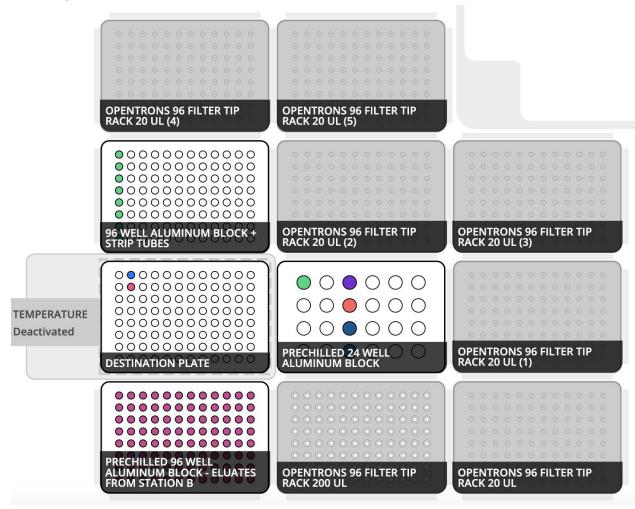
Code parameters:

- Change the sample number on line 14 (default is 94, max is 94)
- Change whether or not if the mastermix is created on the robot or manually on line 15 (default is True)
- Tip rack tracking can be changed from False to True on line 16 (default is False)

Pipettes:

- P20 multichannel on the right mount
- P300 single channel on the left mount

Deck Layout:



Labware and module requirements:

- 1 x Temperature Module
- 6 x 20µl tipracks

- 1 x 200µl tiprack
- 3 x 96 well aluminum blocks (1 x **prechilled*** in slot 1, 1 x on top of the Temperature Module in slot 4, 1 x in slot 7)
- 1 x 24 well **prechilled*** aluminum block **[holds 1.5 2mL tubes with mastermix mastermix components]**
- 5 x 1.5-2mL tubes (if you select **False** for mastermix creation on deck, then there are 1 x 1.5-2mL tubes)
- 2 x strip tubes on top of the 96 well aluminum block in slot 7
- 1 x 96 well plate or strip tubes [Input holds eluates/extractions from Station B)
- 1 x RT-PCR Plate (can be 96 well plate or PCR tubes) [Output]

Volume components

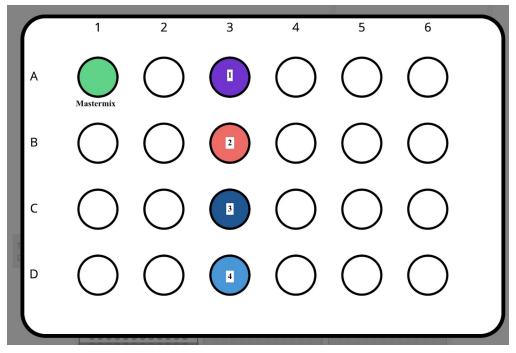
Note: the belows volumes account for a dead volume - the dead volume can be adjusted depending on the calibration of the pipette to the labware, but it's best to have a dead volume of at least 10%

Mastermix Component	What the component is	Volume for 1 sample (µI)	Volume for 8 samples (µI)	Volume for 48 samples (μΙ)	Volume for 96 samples (μΙ)	
1	nCov MOM probe	5	60	260	500	
2	H20	5	60	260	500	
3	5x Real time one-step buffer	5	60	260	500	
4	Real time one-step enzyme	2	24	105	200	

^{*}Prechilled means the aluminum block has been chilled in the -20C before beginning the protocol

Slot 5 - 24 well aluminum block tube layout

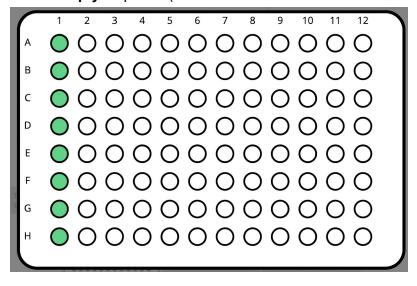
Note: mastermix tube in A1 is loaded empty unless mastermix is created manually



Slot 7 96 well aluminum block setup

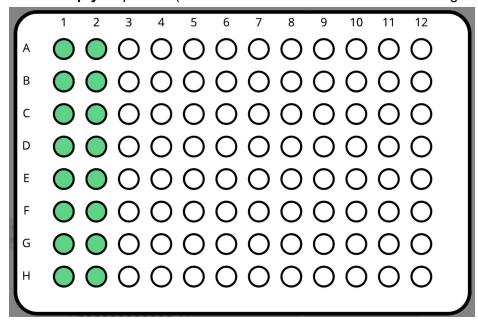
For less than 72 sample throughput:

Load 1 **empty** strip tube (these will be used for mastermix during the run)



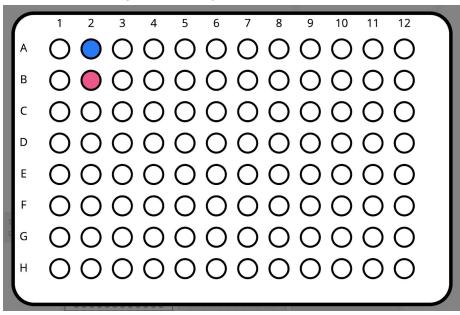
For 72 or greater sample throughput:

Load 2 **empty** strip tubes (these will be used for the mastermix during the run)

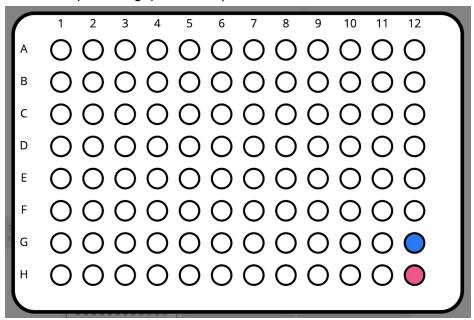


Slot 4 Destination Plate setup

For 8 sample throughput: Add 8 µl of each control



For 94 sample throughput: Add 8 µl of each control



Before you begin:

- 1. Pre-cool the Temperature Module in the Opentrons App to 4°C
- 2. Eluates (extractions) from Station B are loaded onto a **Prechilled** 96 well aluminum block on slot 1.
- 3. Add the Mastermix tube (loaded empty if choosing true for mastermix creation) and mastermix component tubes to the **Prechilled** 24 well aluminum block in slot 5.
- 4. Add empty strip tube(s) to the 96 well aluminum block in slot 7
- 5. Add the 8 µl of control to each corresponding tube in the destination plate
- 6. Check again to make sure each component is added and the Temperature Module had pre-cooled.

The final destination RT-PCR plate will be in Slot 4 on top of the Temperature Module. Once the protocol is complete, the plate will be ready to be sealed, spun down, and loaded onto an RT-PCR machine.

8 sample plate layout:

Seegen e setup	1		2	3	4	5	6	7	8	9	10	11	12
A	1	+ control											
В	2	- Control											
С	3												
D	4												
E	5												
F	6												

G	7			
Н	8			

94 sample plate layout:

	1	2	3	4	5	6	7	8	9	10	11	12
Α	1	9	17	25	33	41	49	57	65	73	81	89
В	2	10	18	26	34	42	50	58	66	74	82	90
С	3	11	19	27	35	43	51	59	67	75	83	91
D	4	12	20	28	36	44	52	60	68	76	84	92
Е	5	13	21	29	37	45	53	61	69	77	85	93
F	6	14	22	30	38	46	54	62	70	78	86	94
G	7	15	23	31	39	47	55	63	71	79	87	+ control
Н	8	16	24	32	40	48	56	64	72	80	88	- Control