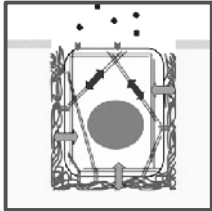




Biochemical and mechanical regulation of extracellular matrix signaling: insights from fibronectin

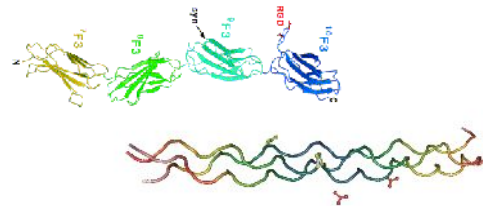
BOSTON
UNIVERSITY

Michael Smith
March 5, 2014

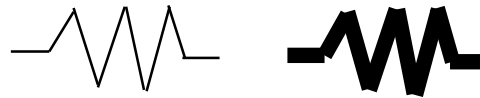


Cell sensory toolbox

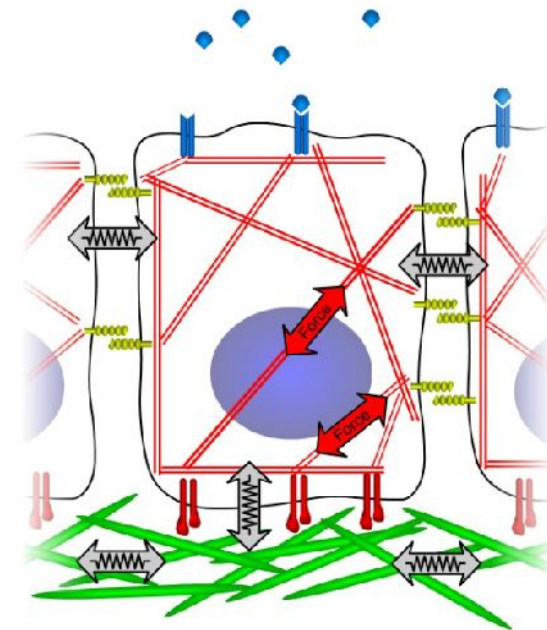
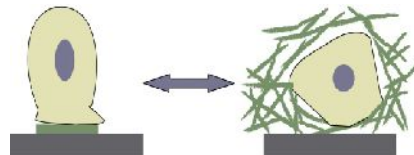
biochemical



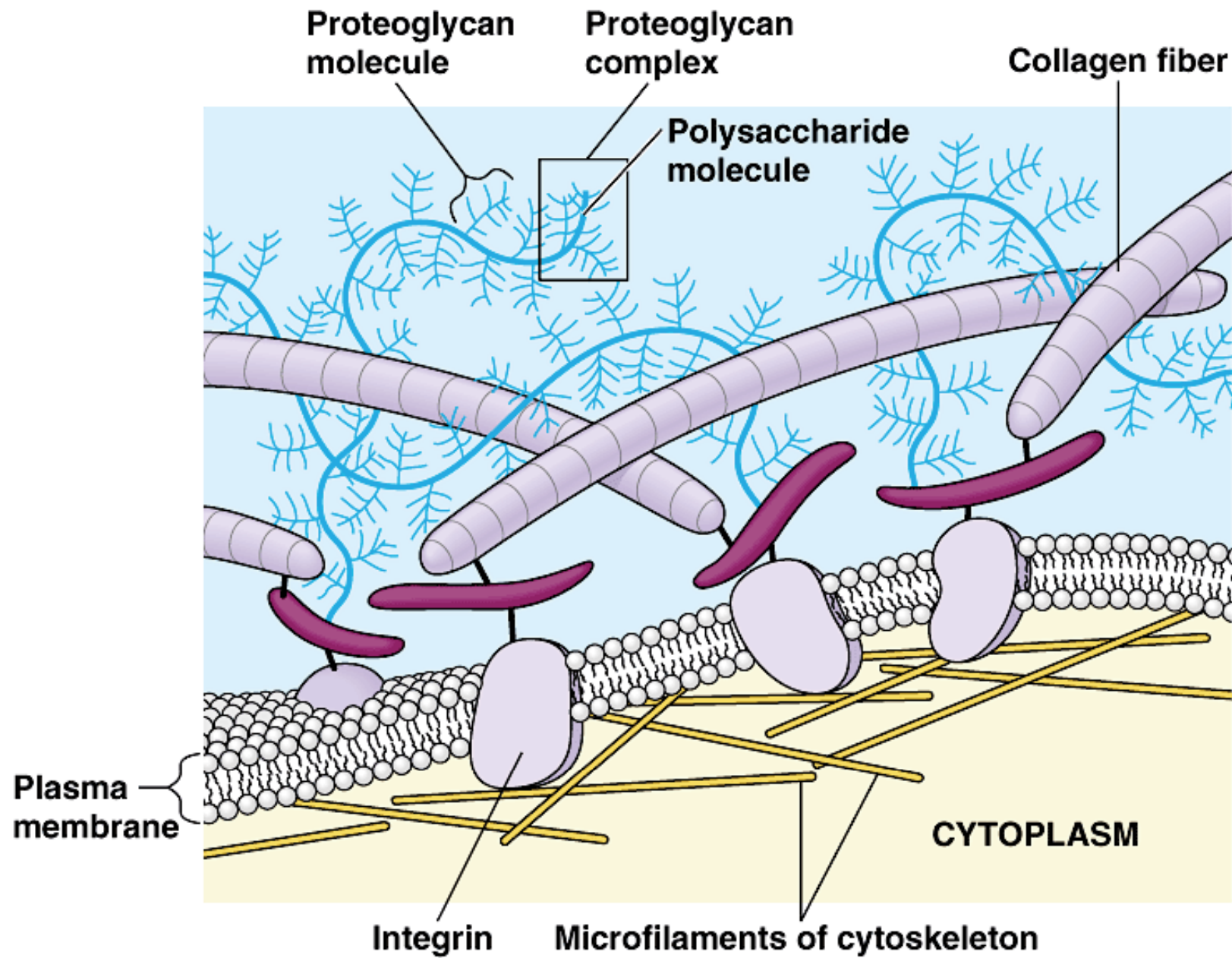
mechanical



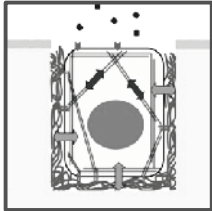
topographical



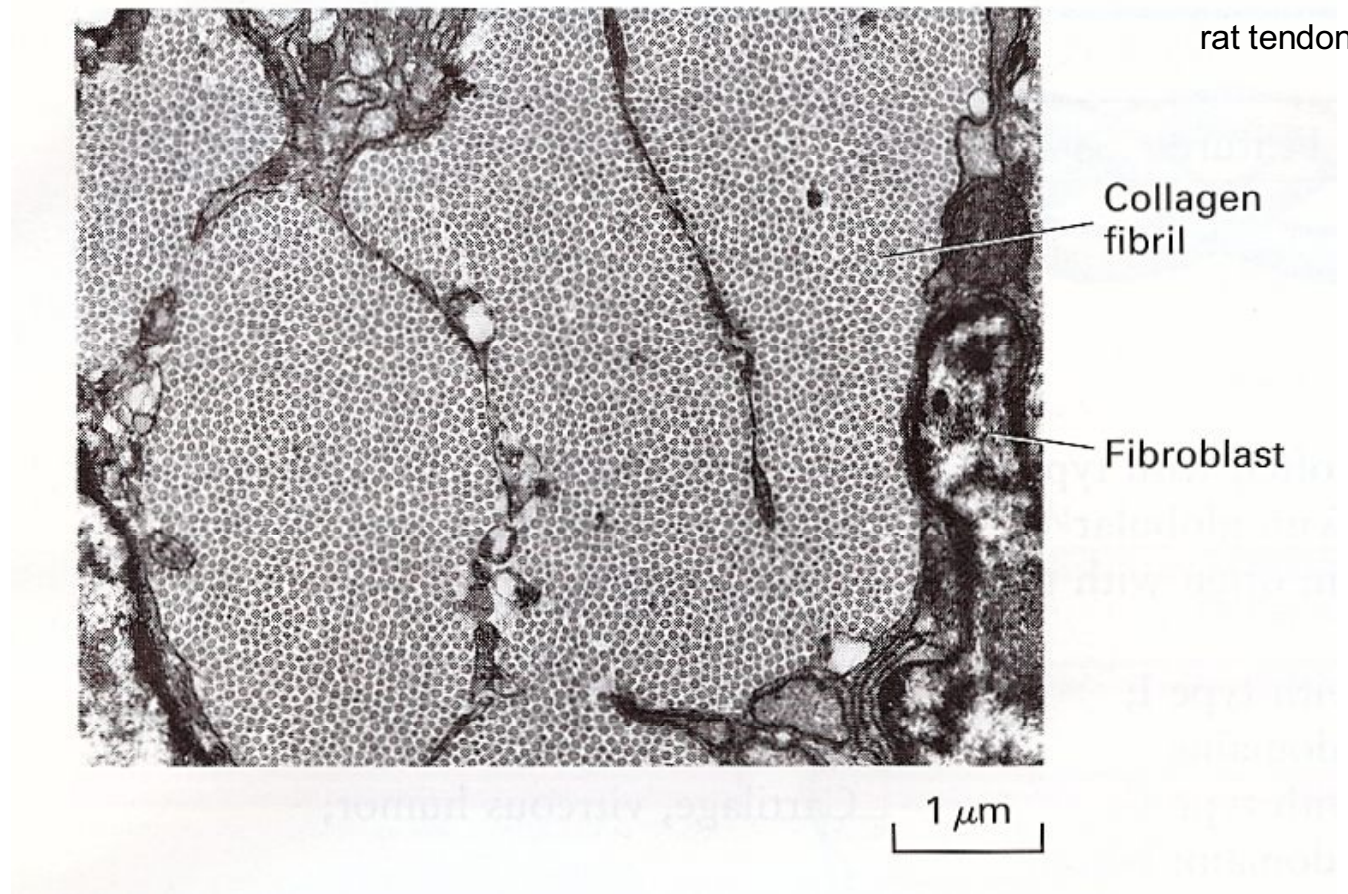
Goal: Determine how extracellular matrix fibers communicate biochemical and mechanical signals to cells



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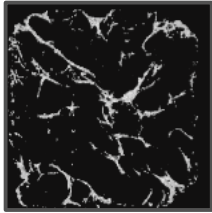


Collagen

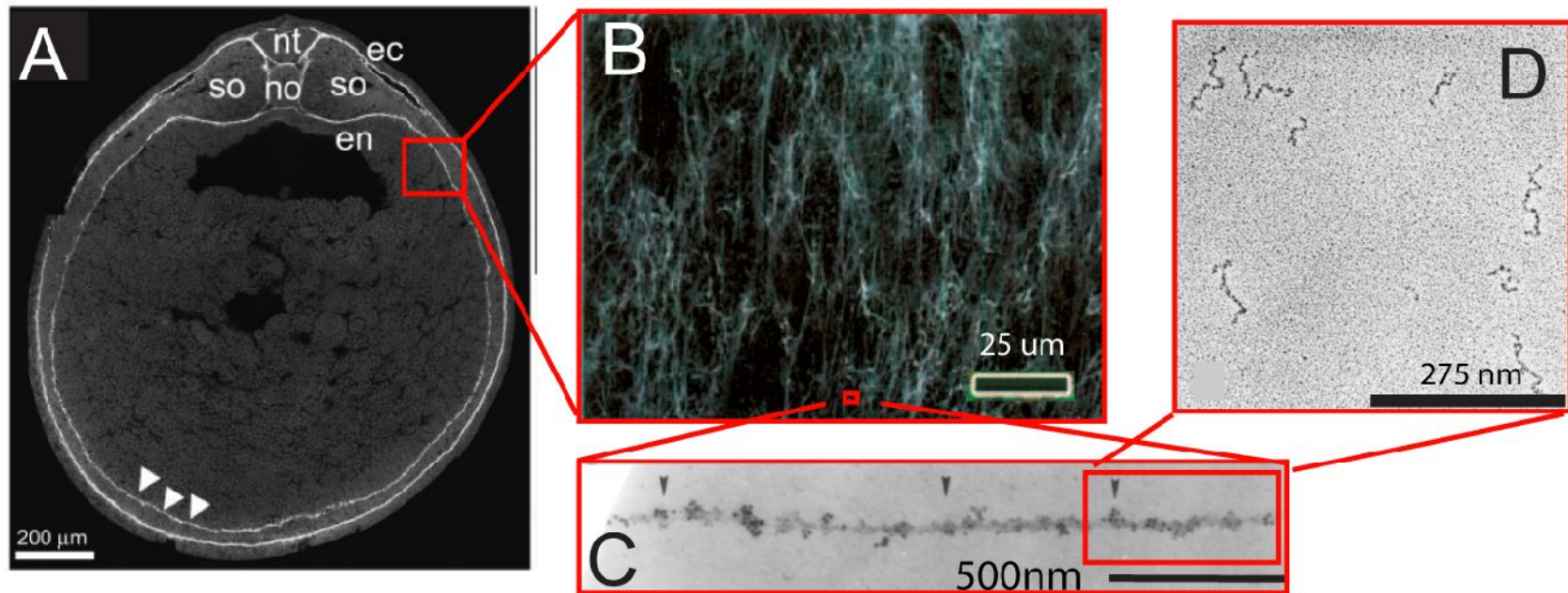


Lodish et al. Molecular Biology of the Cell.

Parry. 1988. BiophysChem. 29:195.



Fibronectin



Davidson, Keller, DeSimone. 2004. *DevDyn*. 231: 888.

Erickson, Carrell. 1983. *JBiolChem*. 258: 14539.

Peters, Portz, Fullenwider, Mosher. 1990. *JCellBiol*. 111: 249.

Defects in mesoderm, neural tube and vascular development in mouse embryos lacking fibronectin

Elizabeth L. George*, Elisabeth N. Georges-Labouesse†, Ramila S. Patel-King, Helen Rayburn and Richard O. Hynes‡

Cell and fibronectin dynamics during branching morphogenesis

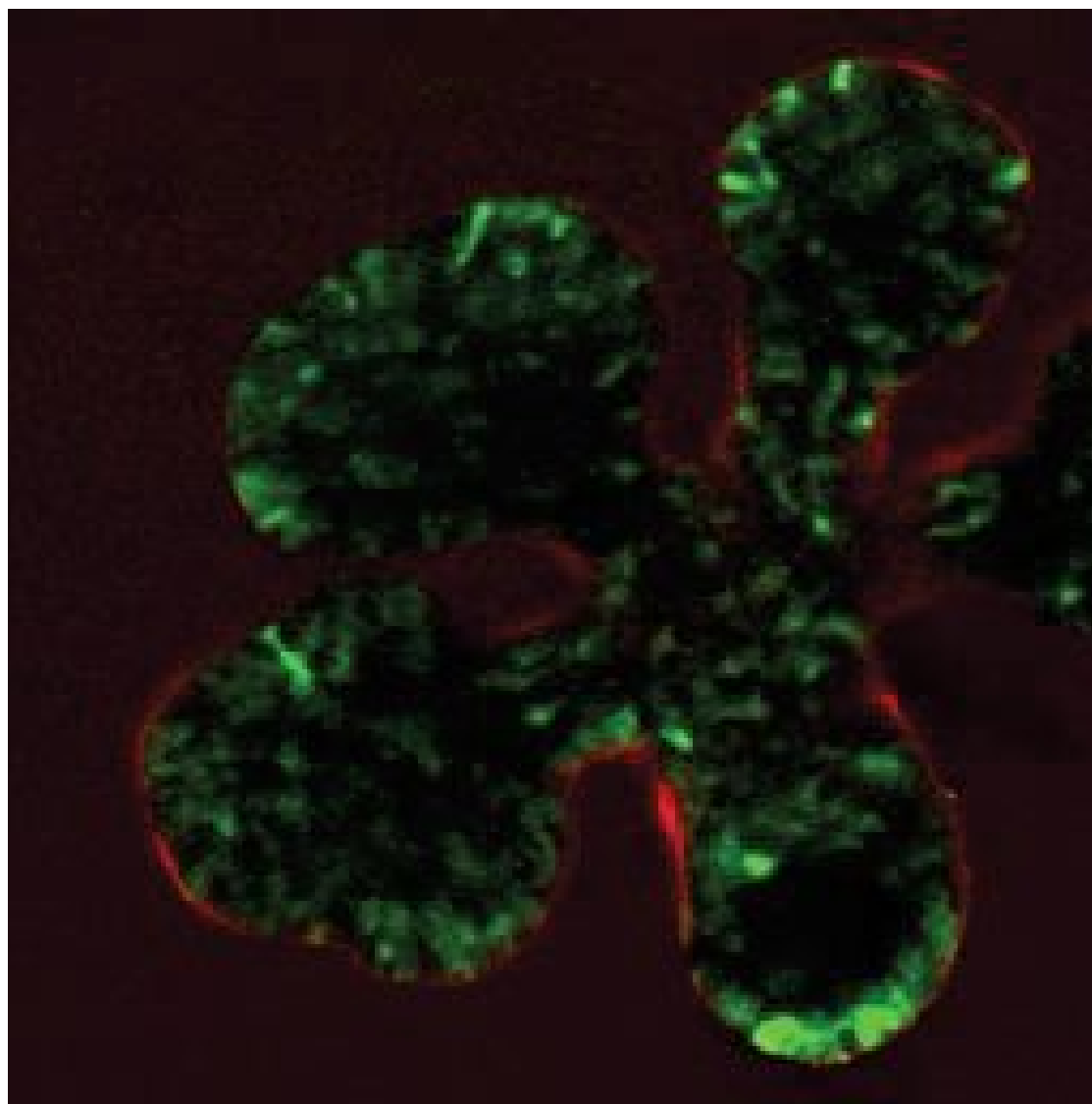
Melinda Larsen, Cindy Wei and Kenneth M. Yamada*

Craniofacial Developmental Biology and Regeneration Branch, National Institute of Dental and Craniofacial Research, National Institutes of Health, 30 Convent Drive, MSC 4370, Bethesda, MD 20892-4370, USA

*Author for correspondence (e-mail: kenneth.yamada@nih.gov)

Accepted 5 June 2006

Journal of Cell Science 119, 3376-3384 Published by The Company of Biologists 2006
doi:10.1242/jcs.03079

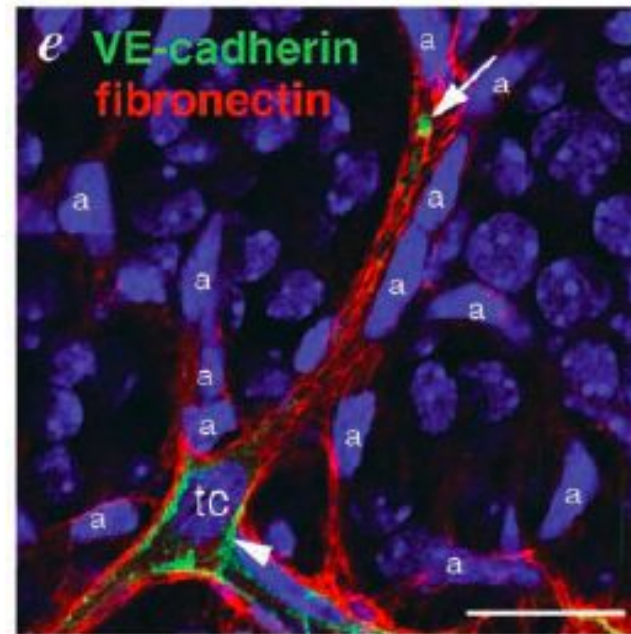


VEGF guides angiogenic sprouting utilizing endothelial tip cell filopodia

Holger Gerhardt,¹ Matthew Golding,² Marcus Fruttiger,³ Christiana Ruhrberg,² Andrea Lundkvist,¹ Alexandra Abramsson,¹ Michael Jeltsch,⁴ Christopher Mitchell,⁵ Kari Alitalo,⁴ David Shima,² and Christer Betsholtz¹

We show here that VEGF-A controls angiogenic sprouting in the early postnatal retina by guiding filopodial extension from specialized endothelial cells situated at the tips of the vascular sprouts. The tip cells

respond to VEGF-A only by guided migration; the proliferative response to VEGF-A occurs in the sprout stalks. These two cellular responses are both mediated by agonistic activity of VEGF-A on VEGF receptor 2.



Proposed strategy to create bioartificial heart

1. Decellularize cadaveric heart

→ create whole-heart scaffold

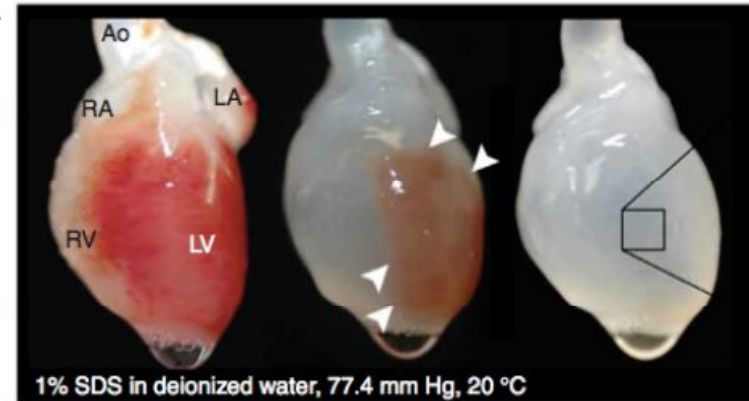
2. Repopulate decellularized heart

→ cardiac and endothelial cells

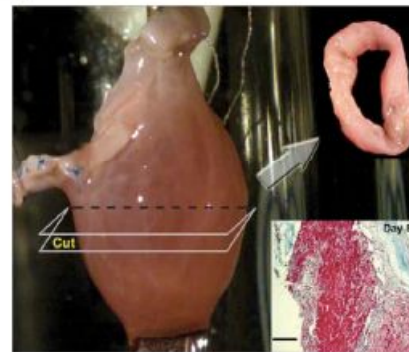
3. Functionalize construct

→ form contractile myocardium

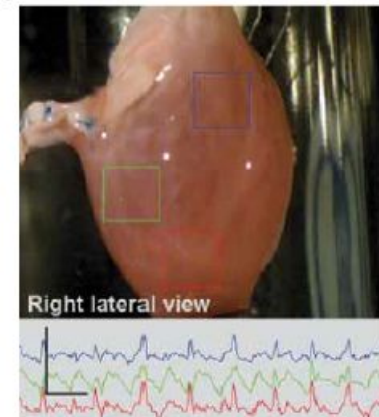
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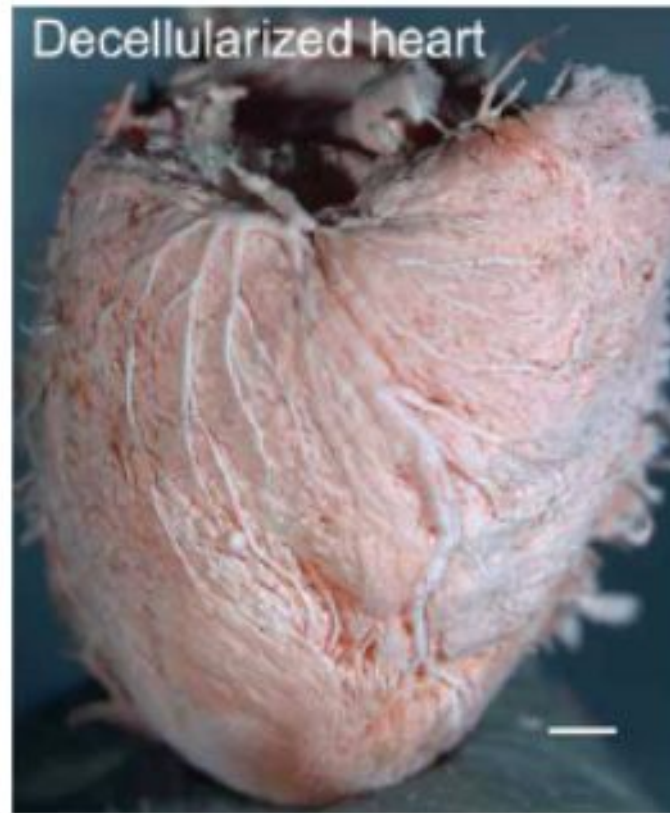


2.



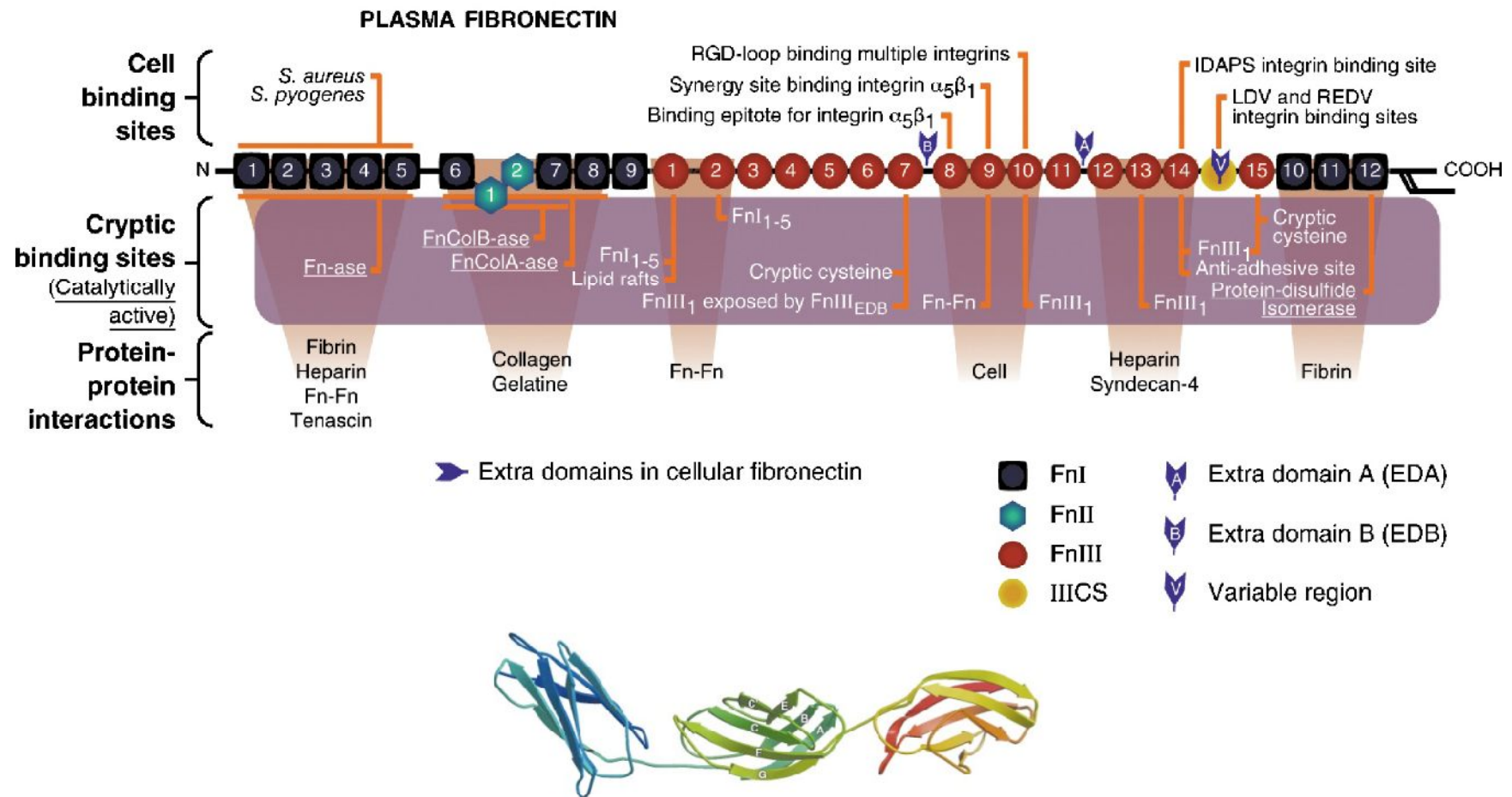
3.





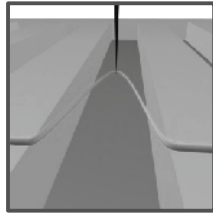
Perfusion-decellularized matrix: using nature's platform to engineer a bioartificial heart

Harald C Ott¹, Thomas S Matthiesen², Saik-Kia Goh², Lauren D Black³, Stefan M Kren², Theoden I Netoff³ & Doris A Taylor^{2,4}

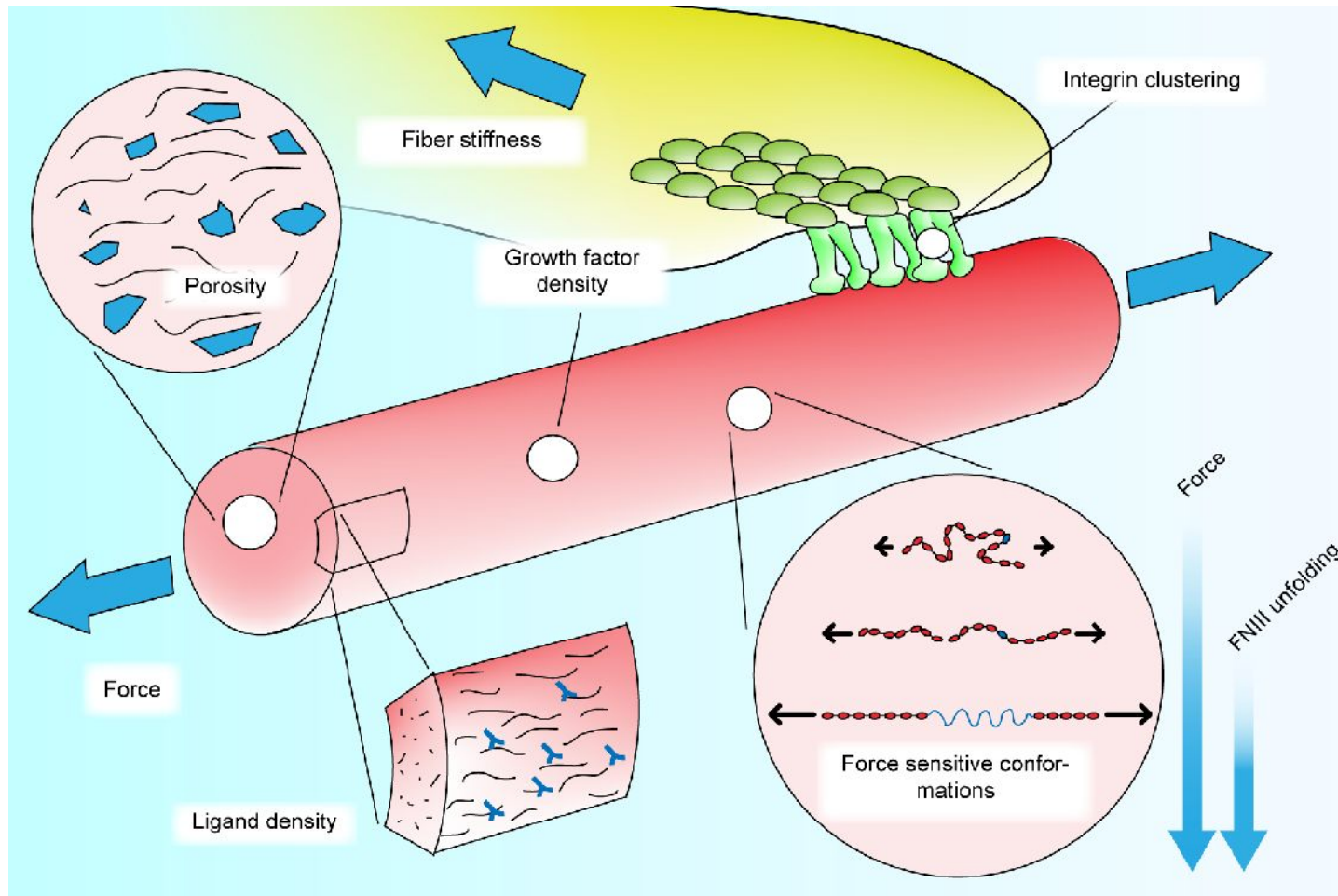


Sharma, Askari, Humphries, Jones, Stuart. 1999. *EMBOJ*. 18: 1468.

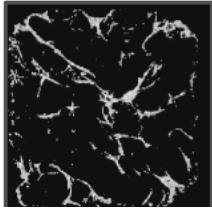
Hytonen, Smith, Vogel. 2010. *Cellular Mechanotransduction: Diverse Perspectives from Molecules to Tissues*, Chapter 13. Cambridge University Press (Editors: Mofrad, Kamm).



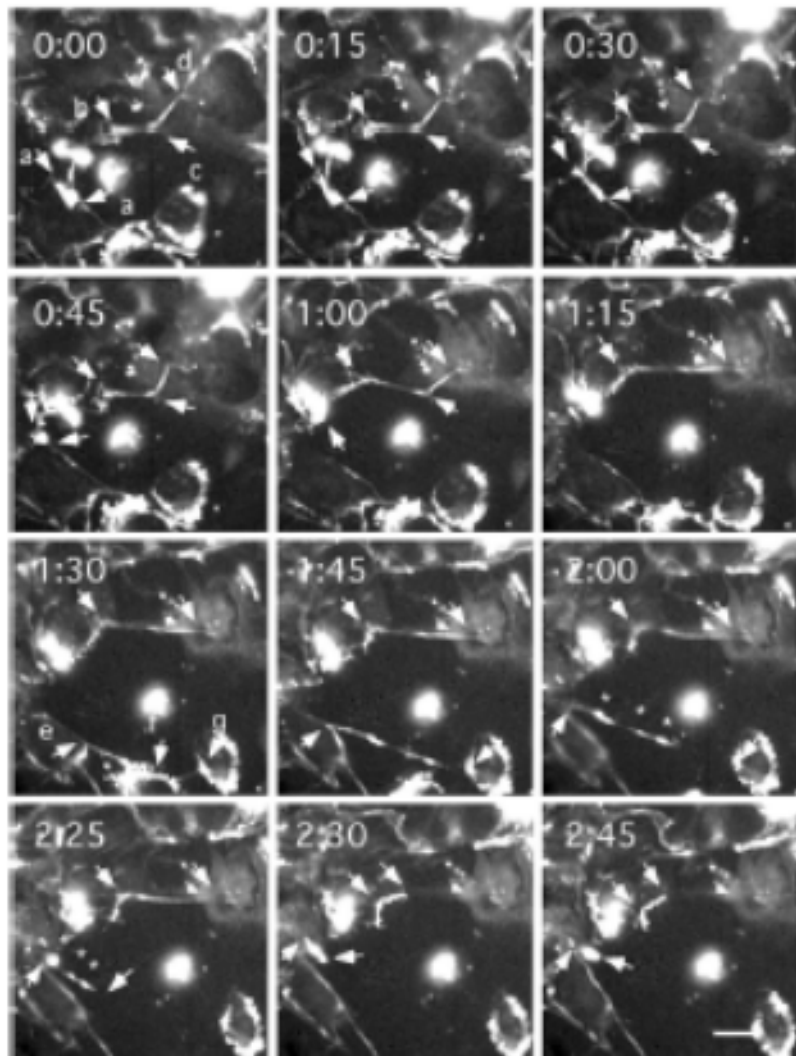
Challenge 1



Bradshaw, Smith. 2013. *ActaBiomater.* In Press.



Challenge 2



Ohashi, Kiehart, Erickson. 1999. *ProcNatlAcadSci.* 96: 2153.

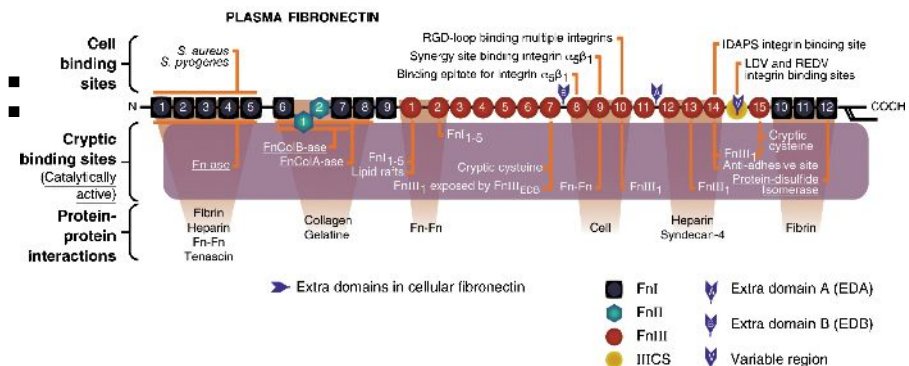
Temperature:
time

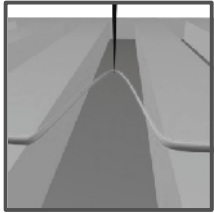
Force: time

Chemical:
denaturant, allosteric
partners, surface
chemistry

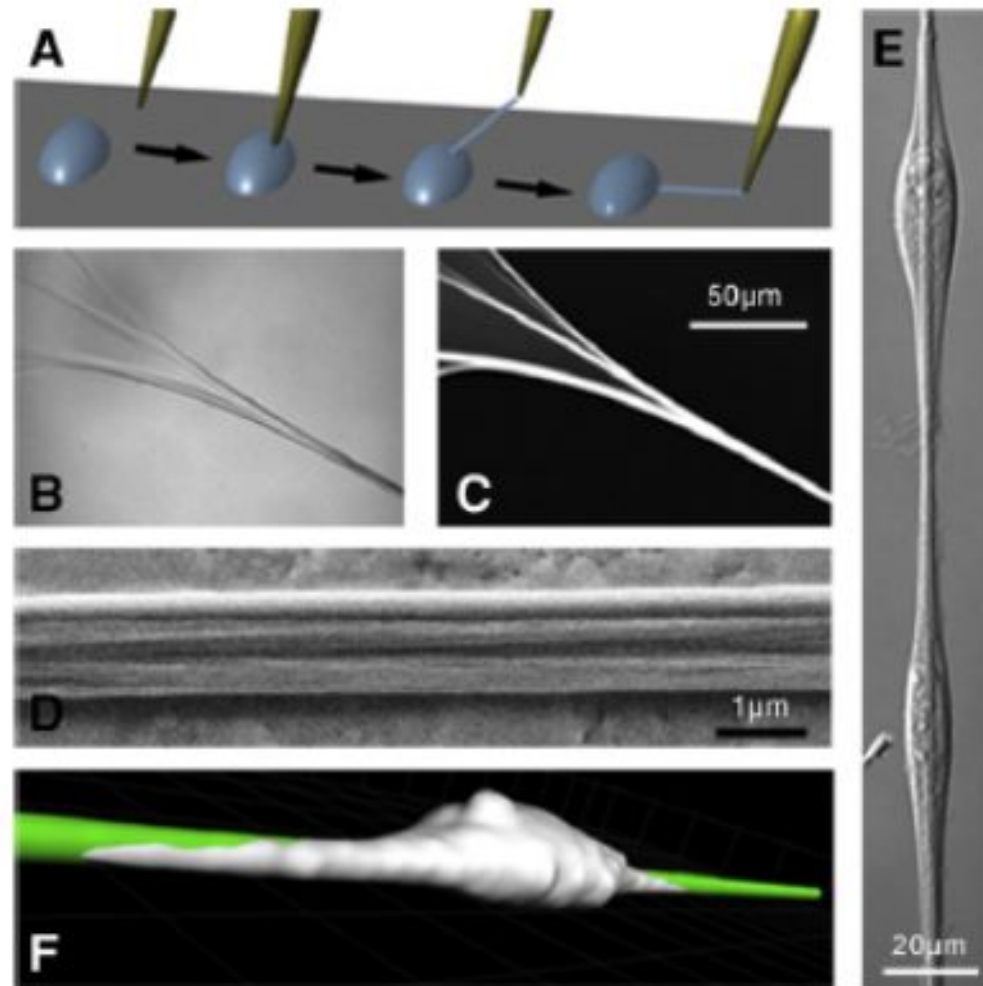
Conformation

Function:





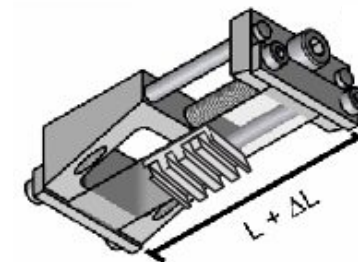
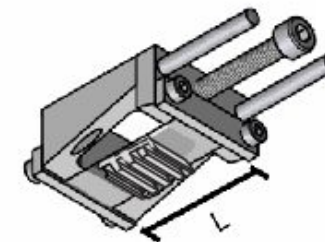
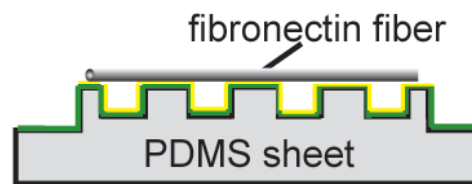
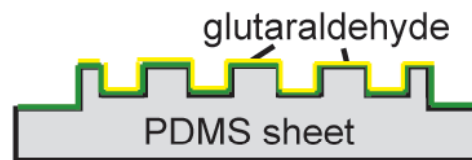
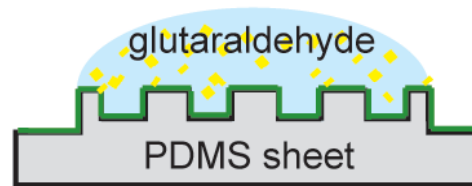
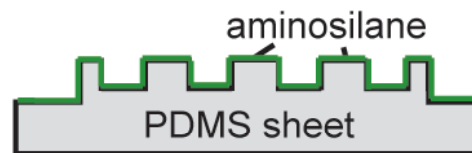
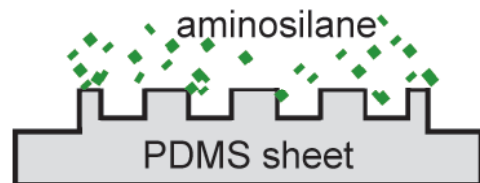
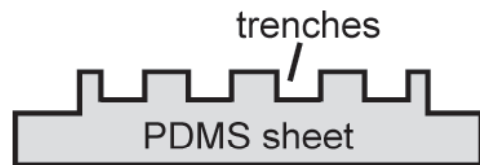
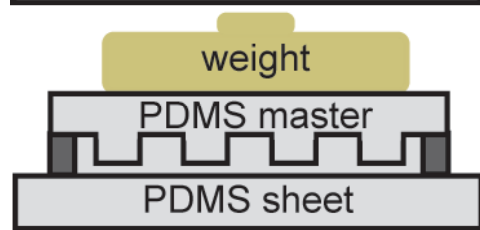
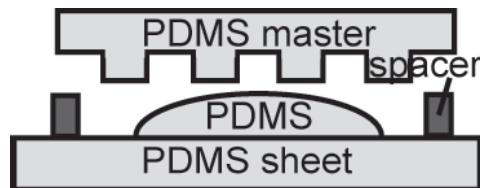
Fabricating fibronectin fibers

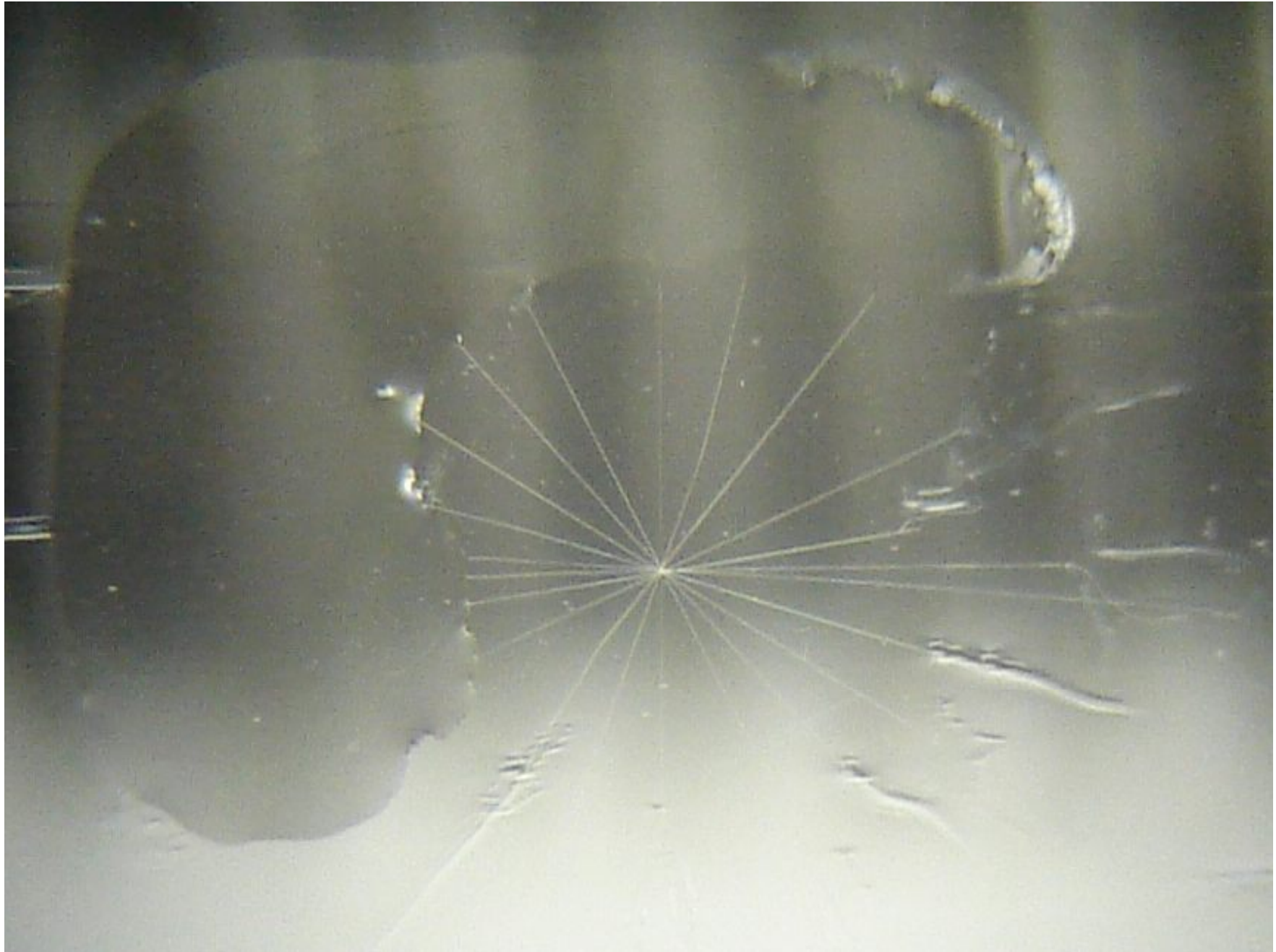


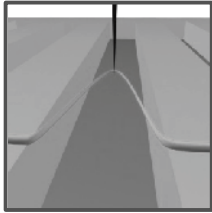
Ejim, Blunn, Brown. 1993. Biomaterials. 14: 743.



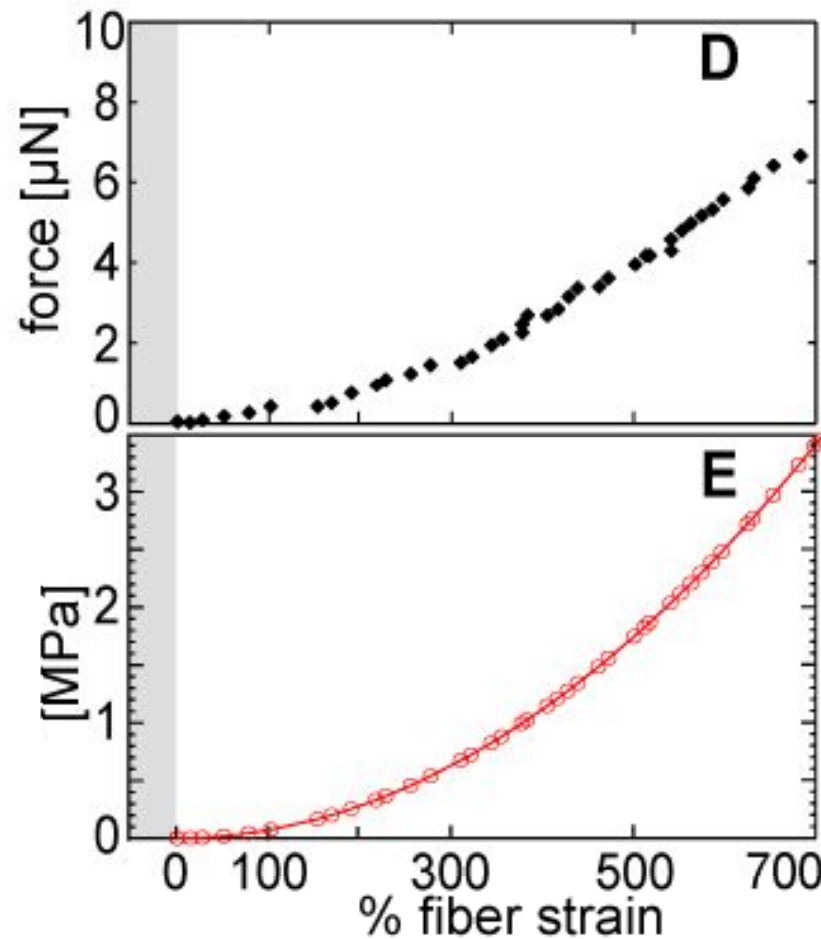
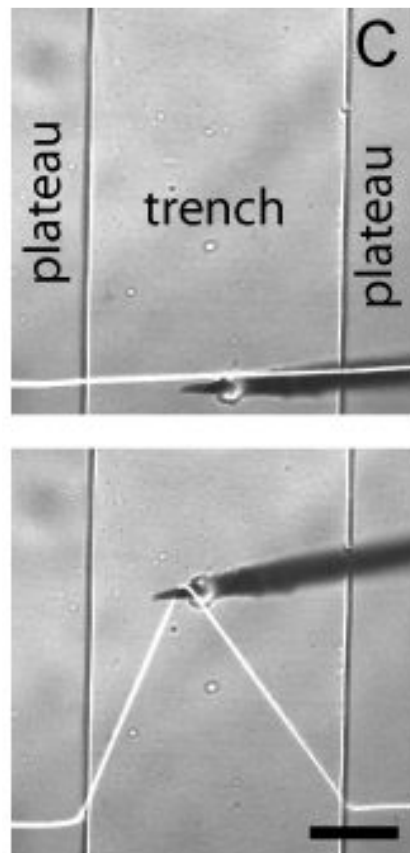
Fibronectin extensibility



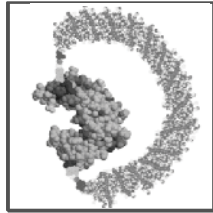




Fn mechanical properties



Klotzsch, Smith, Kubow, Muntwyler, Little, Beyeler, Gourdon, Nelson, Vogel. PNAS. 106: 18267.

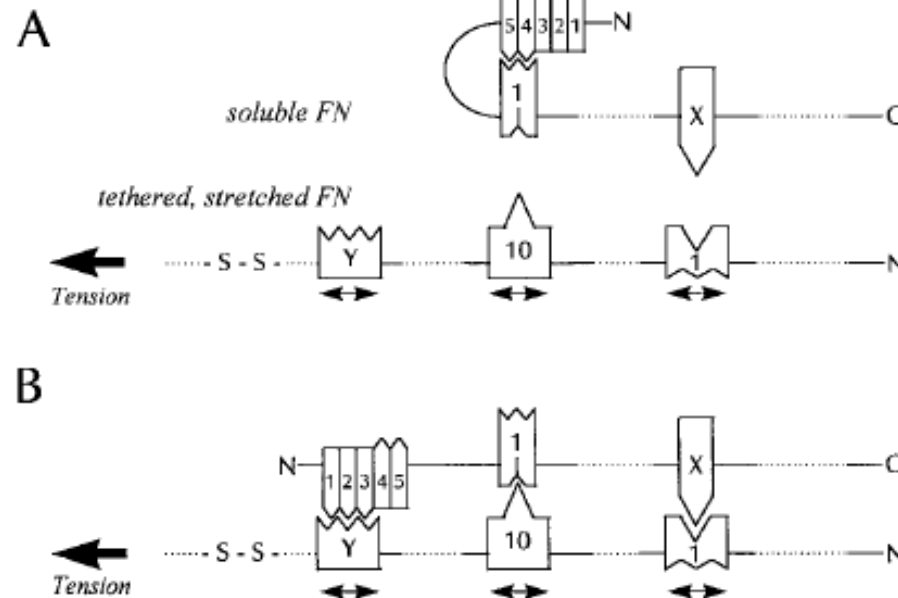


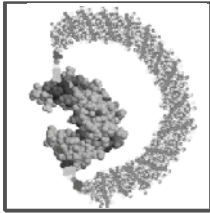
Strain-dependent biochemistry?

Rho-mediated Contractility Exposes a Cryptic Site in Fibronectin and Induces Fibronectin Matrix Assembly

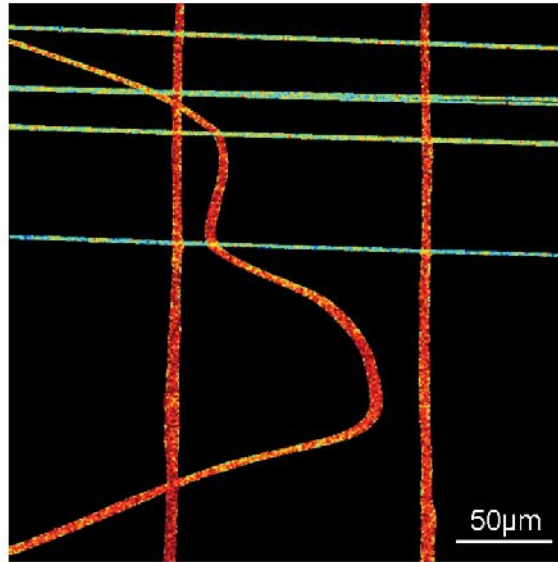
Cuiling Zhong, Magdalena Chrzanowska-Wodnicka, James Brown, Amy Shaub, Alexey M. Belkin, and Keith Burridge

Department of Cell Biology and Anatomy, and Lineberger Comprehensive Cancer Center, University of North Carolina, Chapel Hill, North Carolina

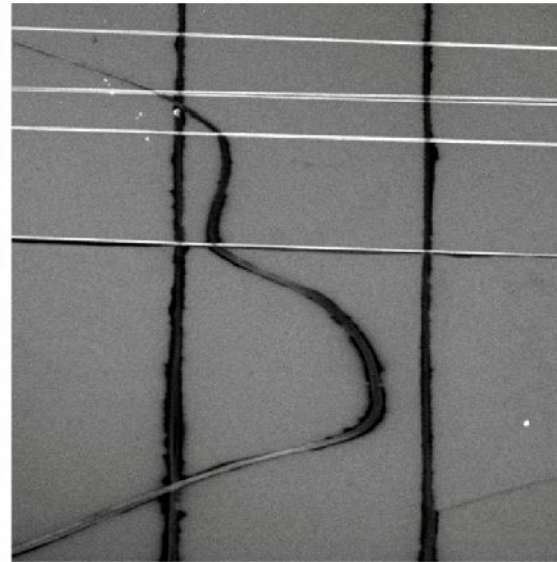




Strain-dependent biochemistry?

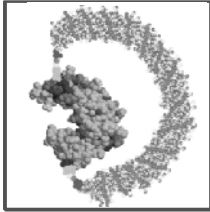


I_A/I_D

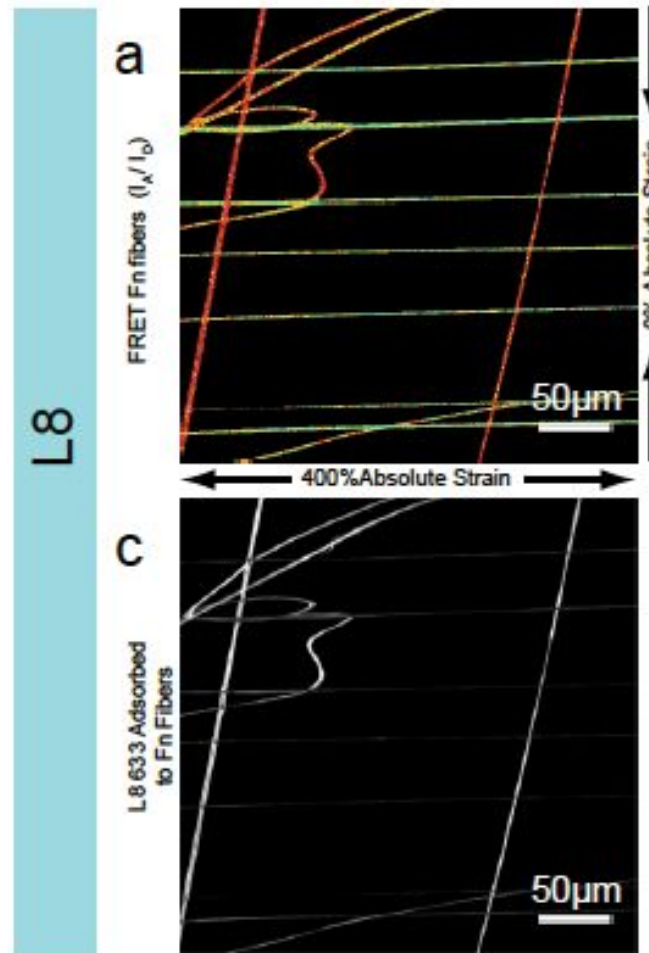


Alexa633 fibronectin
adsorbed to the sample

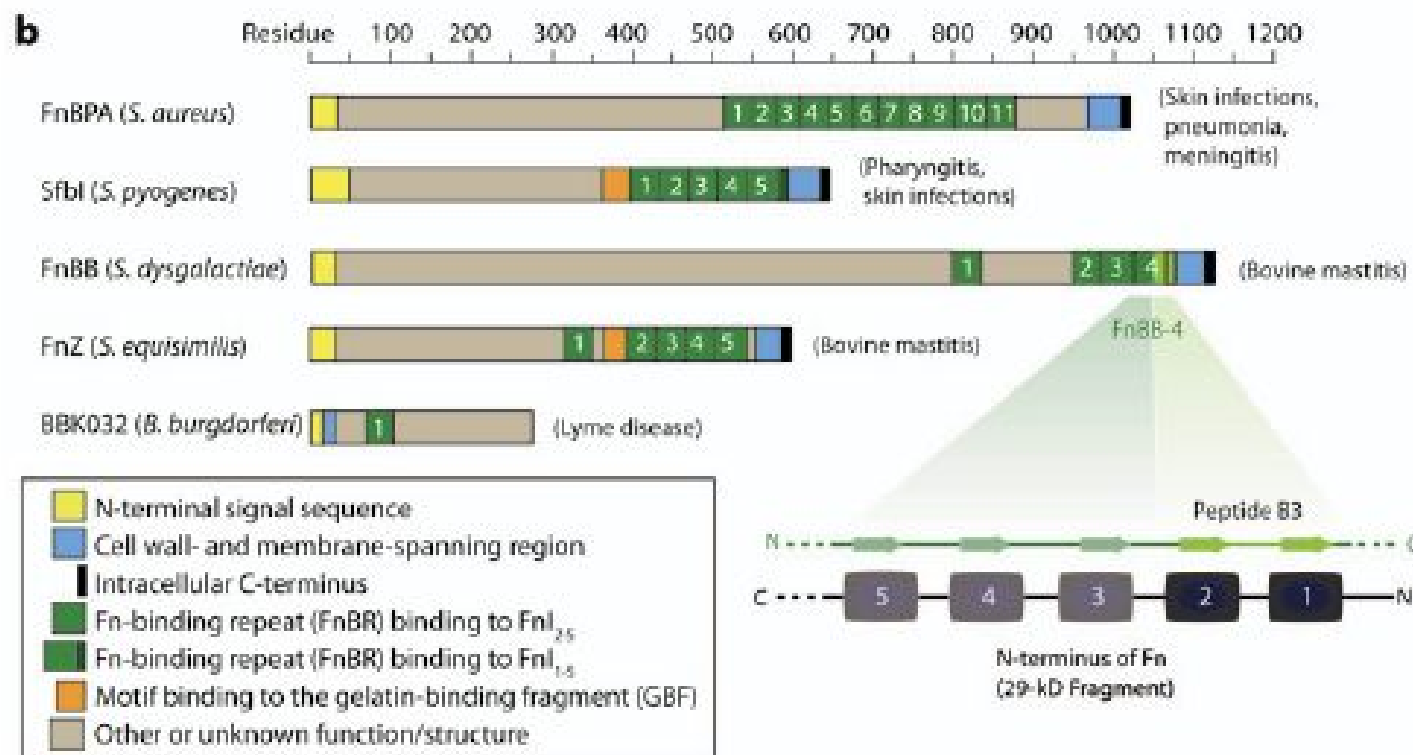
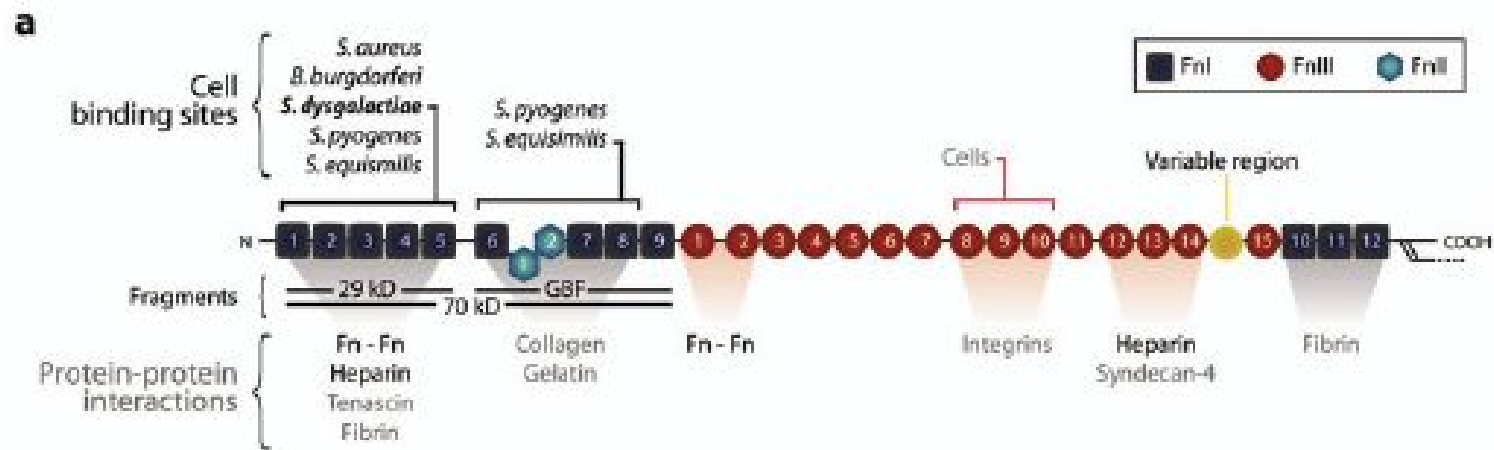
Little, Smith, Ebner, Vogel. 2008. MatrixBiol. 27: 451.

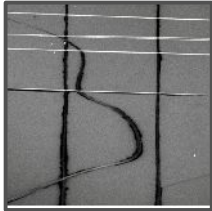


Strain-dependent biochemistry?

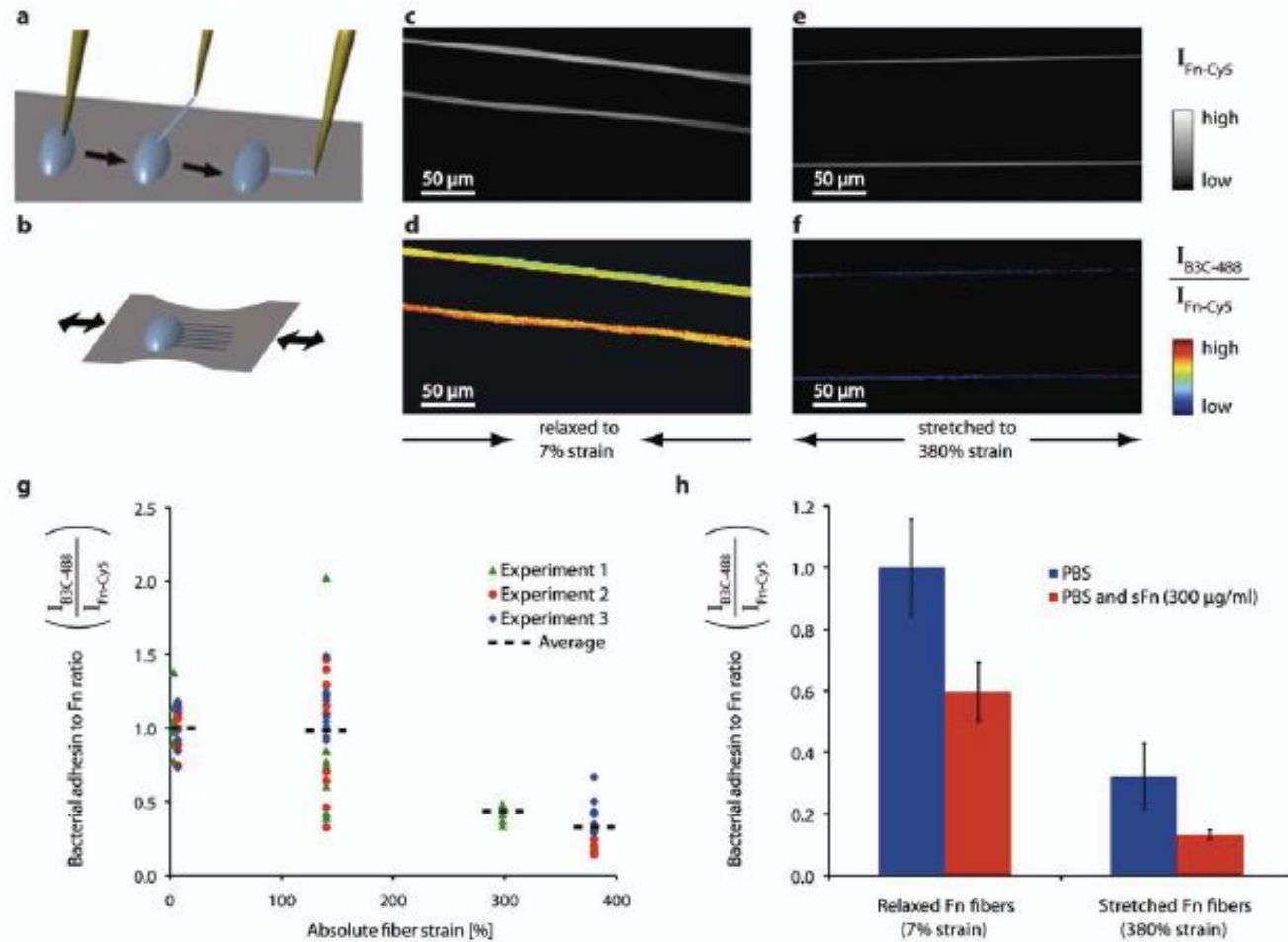


Little, Schwaertlander, Smith, Gourdon, Vogel. 2009. *NanoLett.* 9(12): 4158-4167.





Relevance to disease



M. Chabria, S. Hertig, M.L. Smith, V. Vogel. 2010. *NatCommun.*

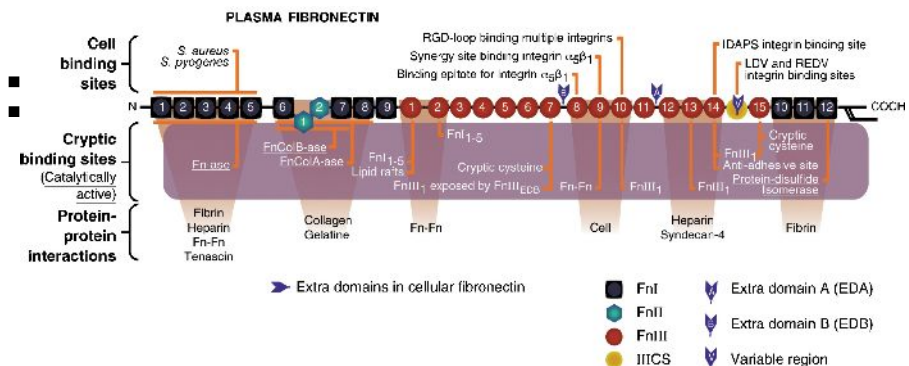
Temperature:
time

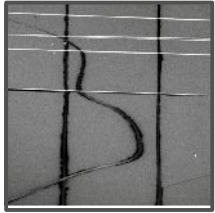
Force: time

Chemical:
denaturant, allosteric
partners, surface
chemistry

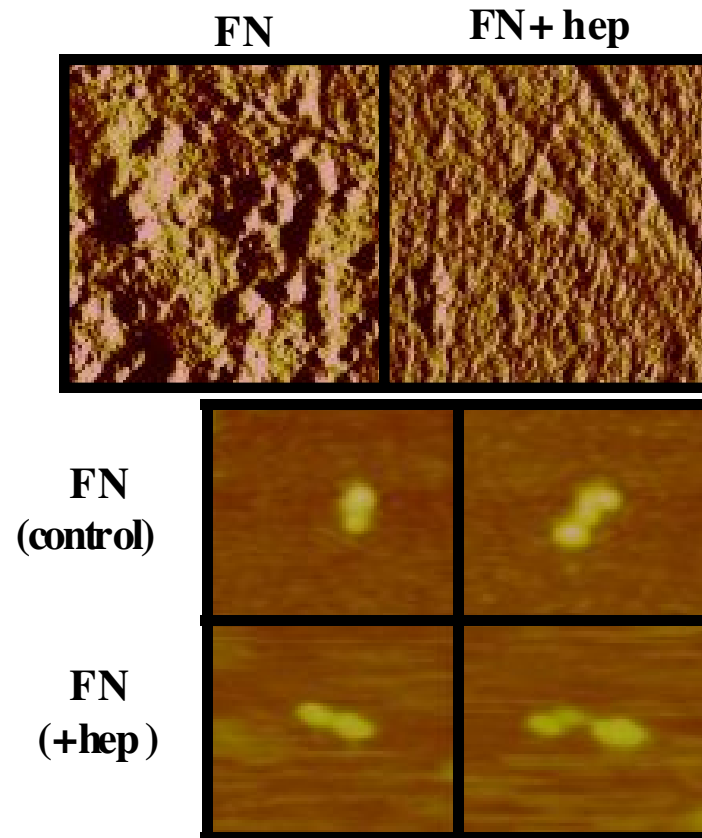
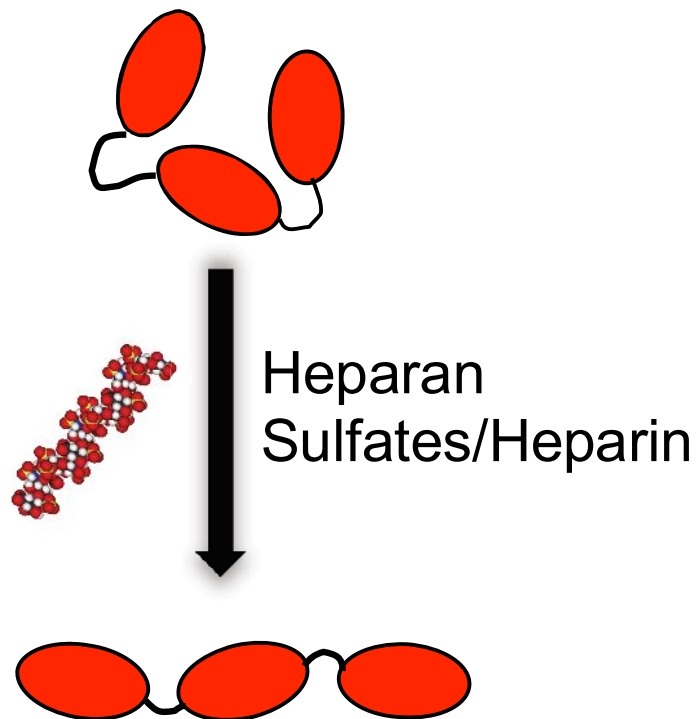
Conformation

Function:

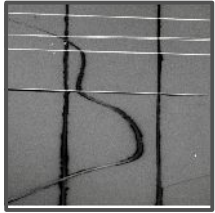




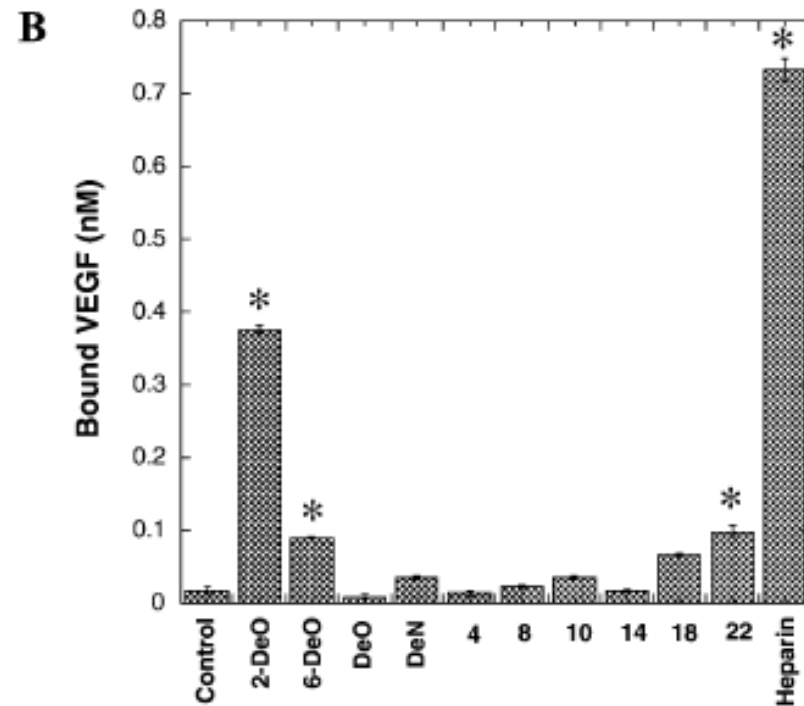
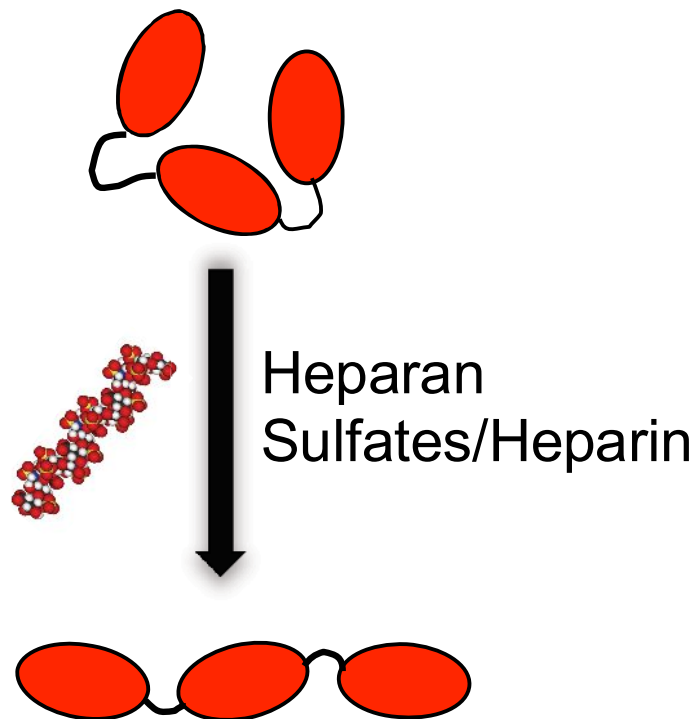
Allosteric binding?



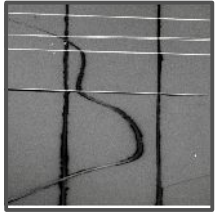
Mitsi, Hong, Costello, Nugent. 2006. *Biochemistry*. 45.



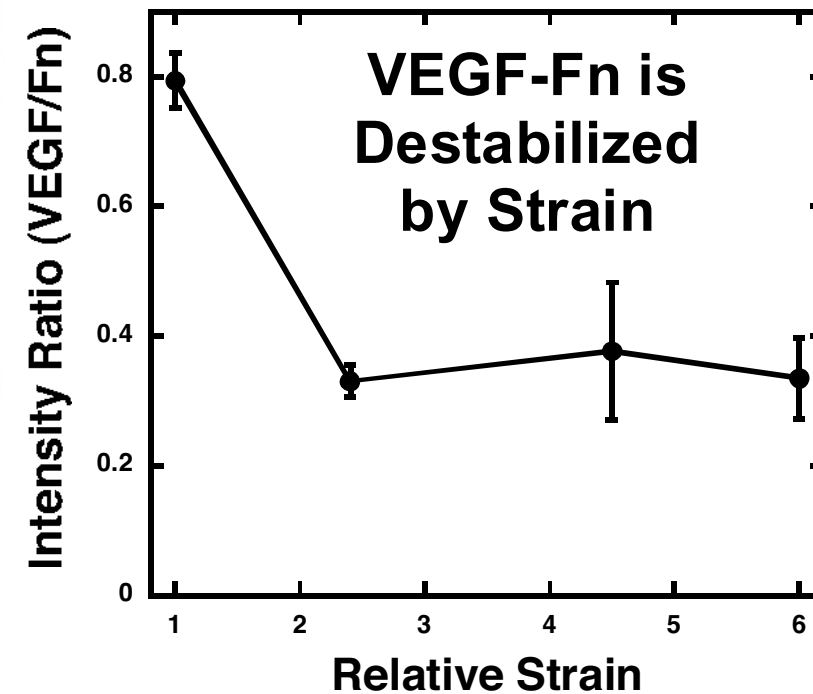
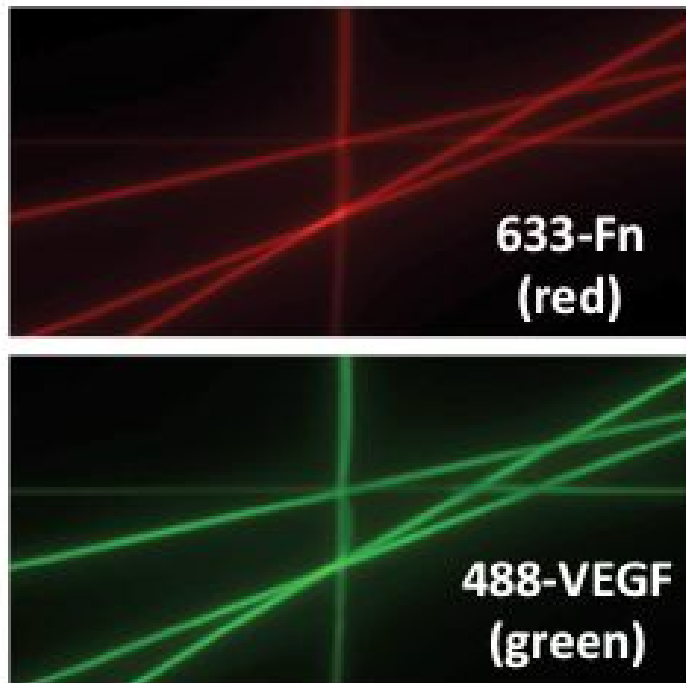
Allosteric binding?



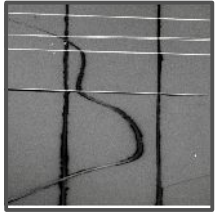
Mitsi, Hong, Costello, Nugent. 2006. *Biochemistry*. 45.



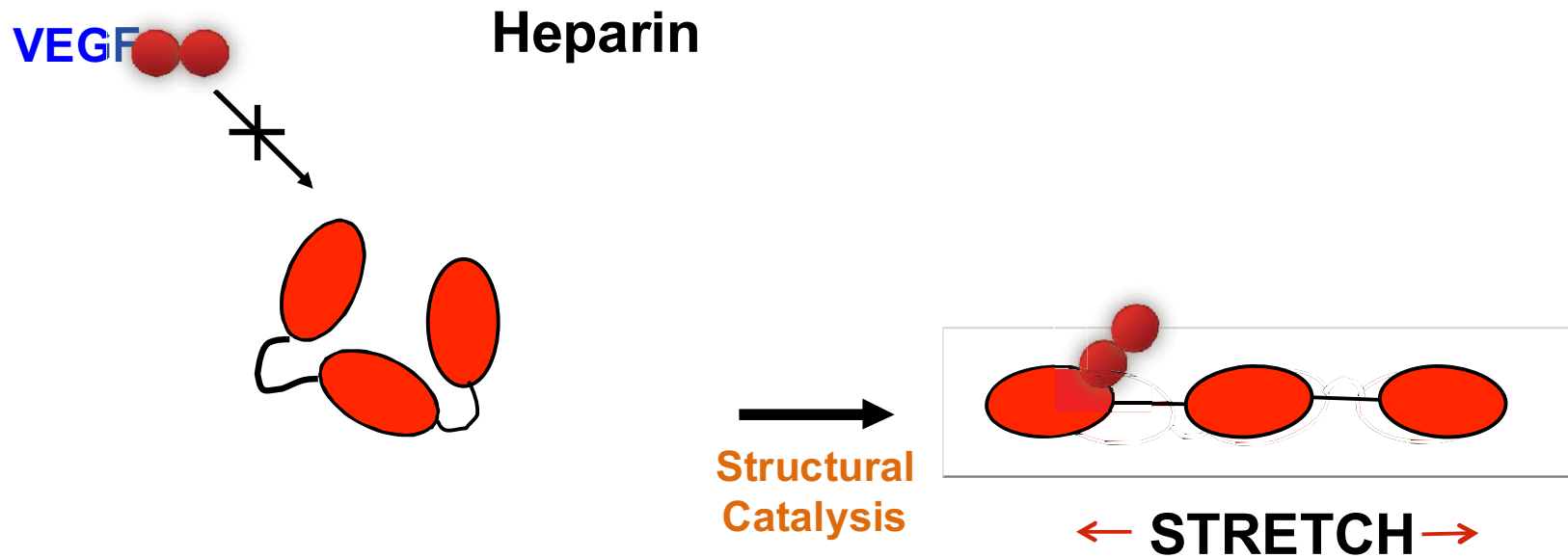
Strain-dependent biochemistry?

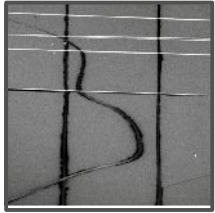


Hubbard, Nugent, Smith. Unpublished.

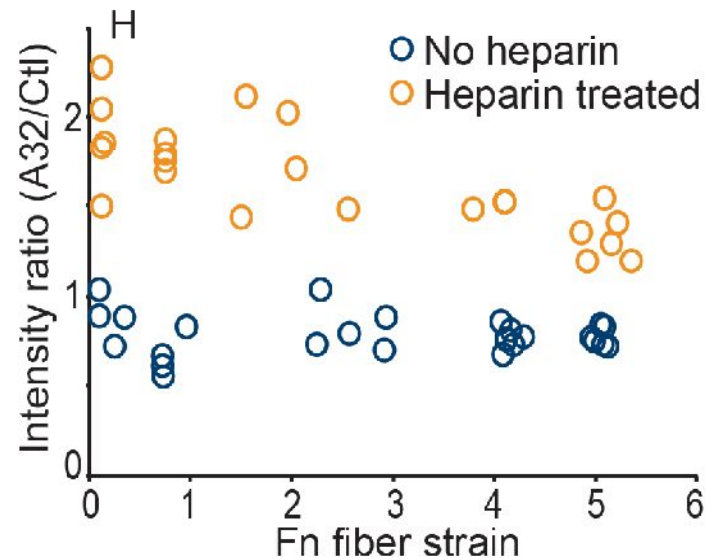
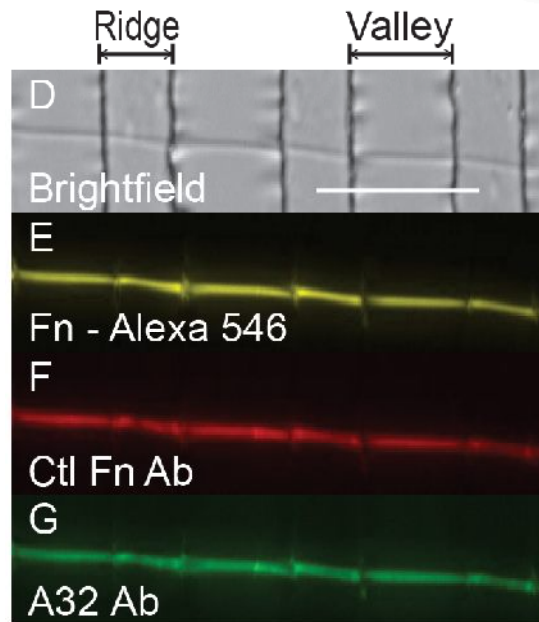
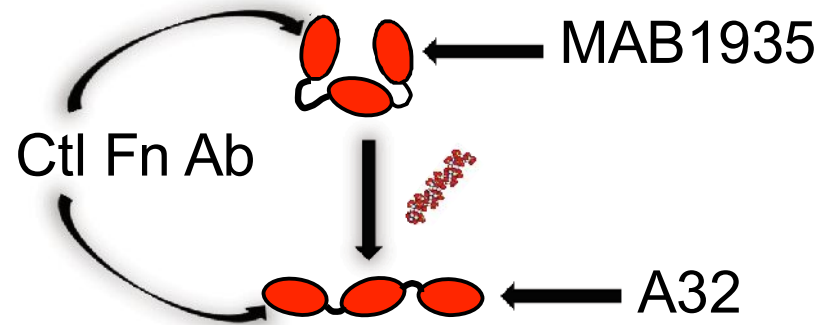


Strain-dependent biochemistry?





Allosteric versus mechanical regulation



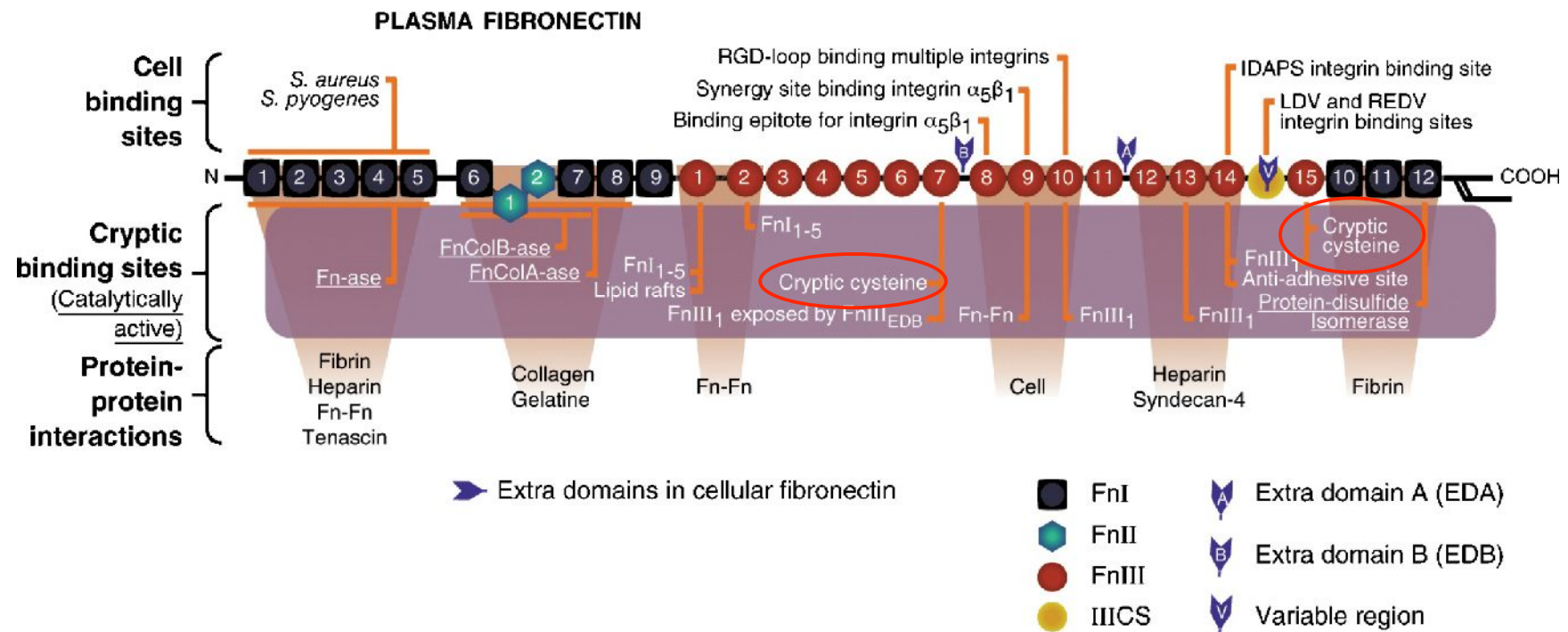
Hubbard, Buczek-Thomas, Nugent, Smith. 2013. *Matrix Biol. In Press.*

Enthalpic and Entropic contributions to extension:





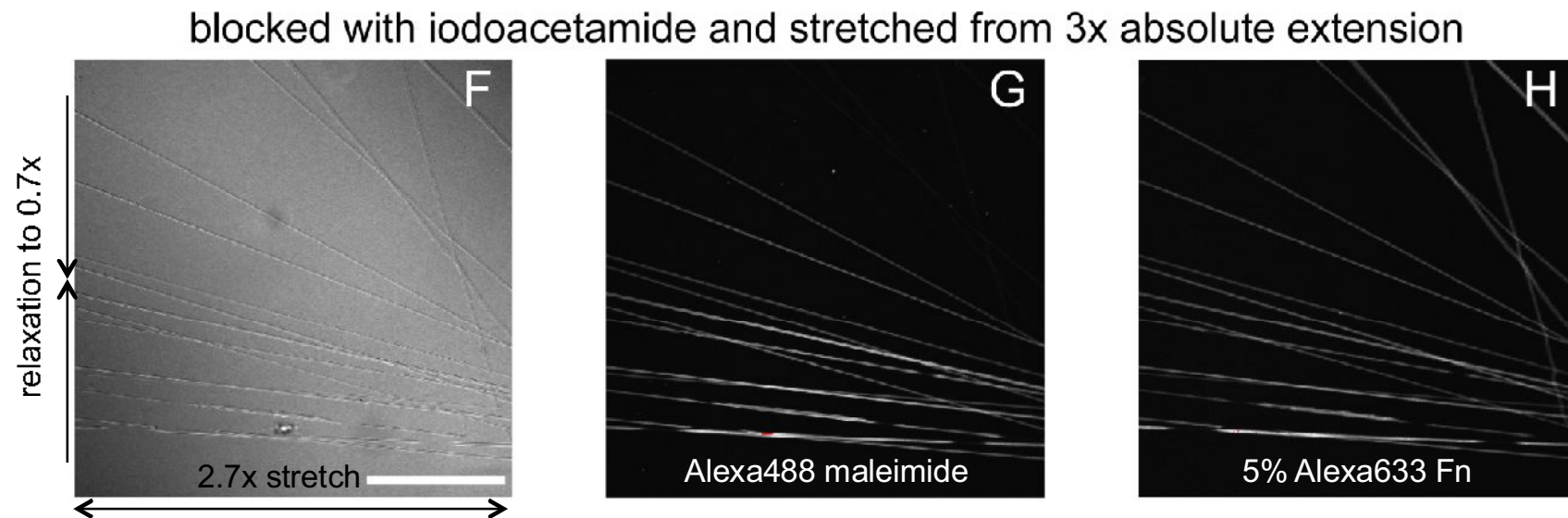
How do fibers extend?



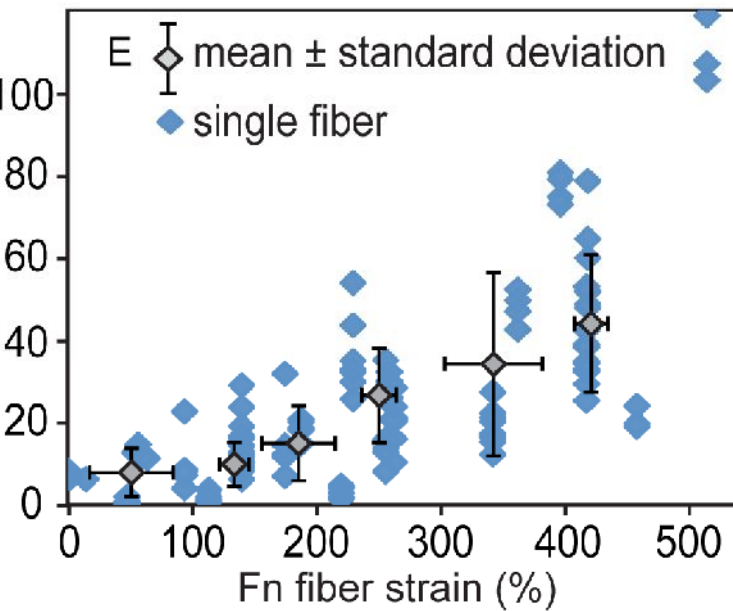
Vogel. 2006. *AnnuRevBiophysBiomolStruct.* 35:459.



How do fibers extend?



Johnson, Tang, Carag, Speicher, Discher. 2007. Science. 317:663.



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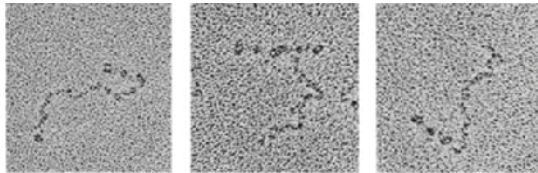


Linking nm to μm length scales

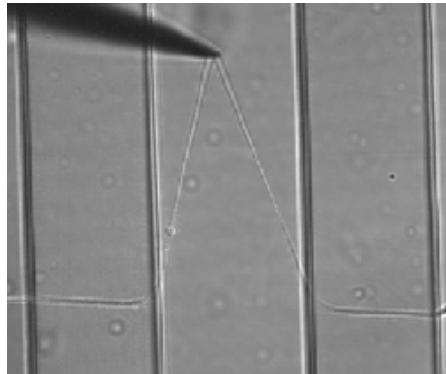
molecular properties

+

intermolecular arrangement



Molecules are probabilistic



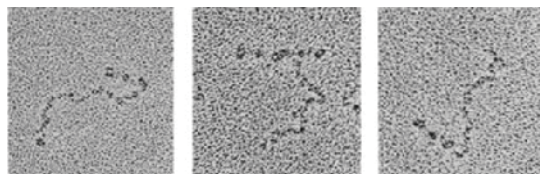
Fibers are deterministic



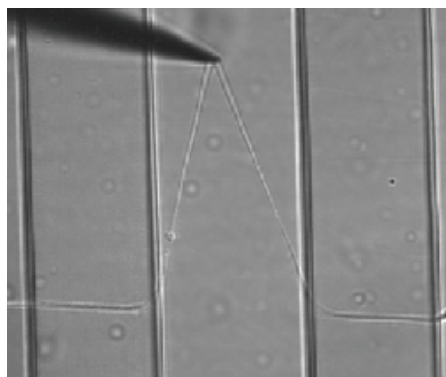
Multi-scale model

Benefits: predictions of intermolecular arrangement, molecular force, molecular conformation, ligand density, and porosity

Challenge:



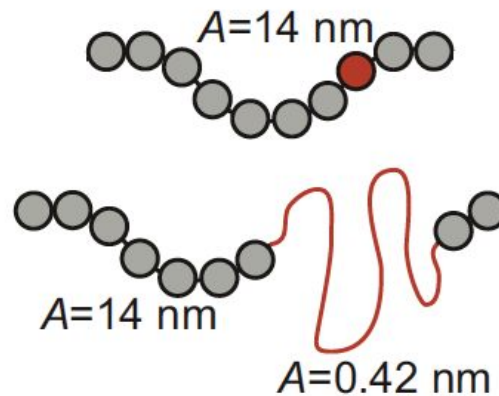
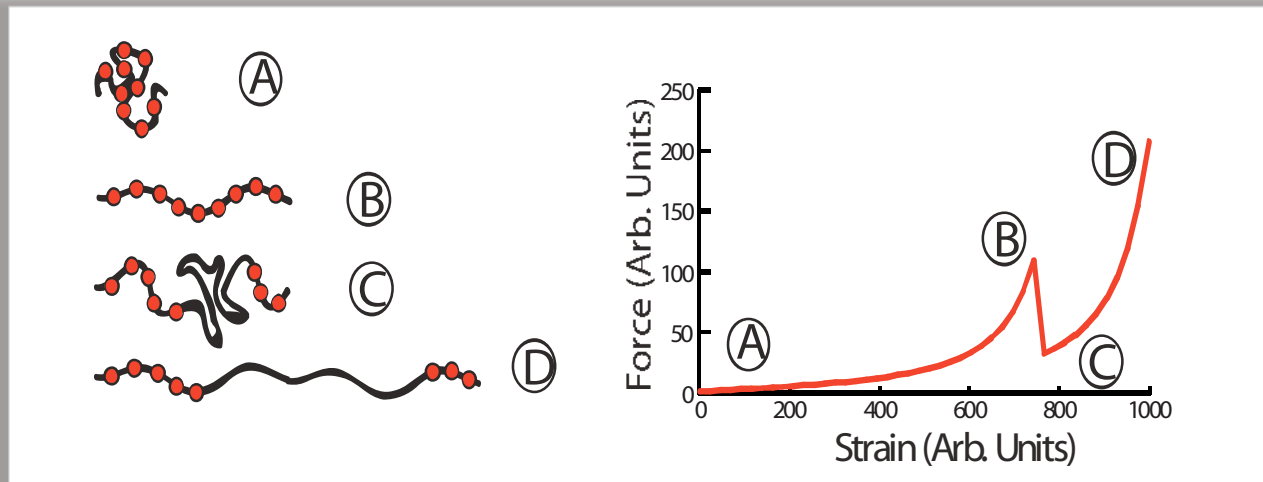
Molecules are probabilistic

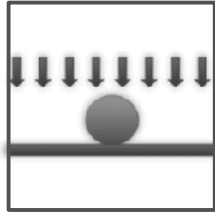


Fibers are deterministic



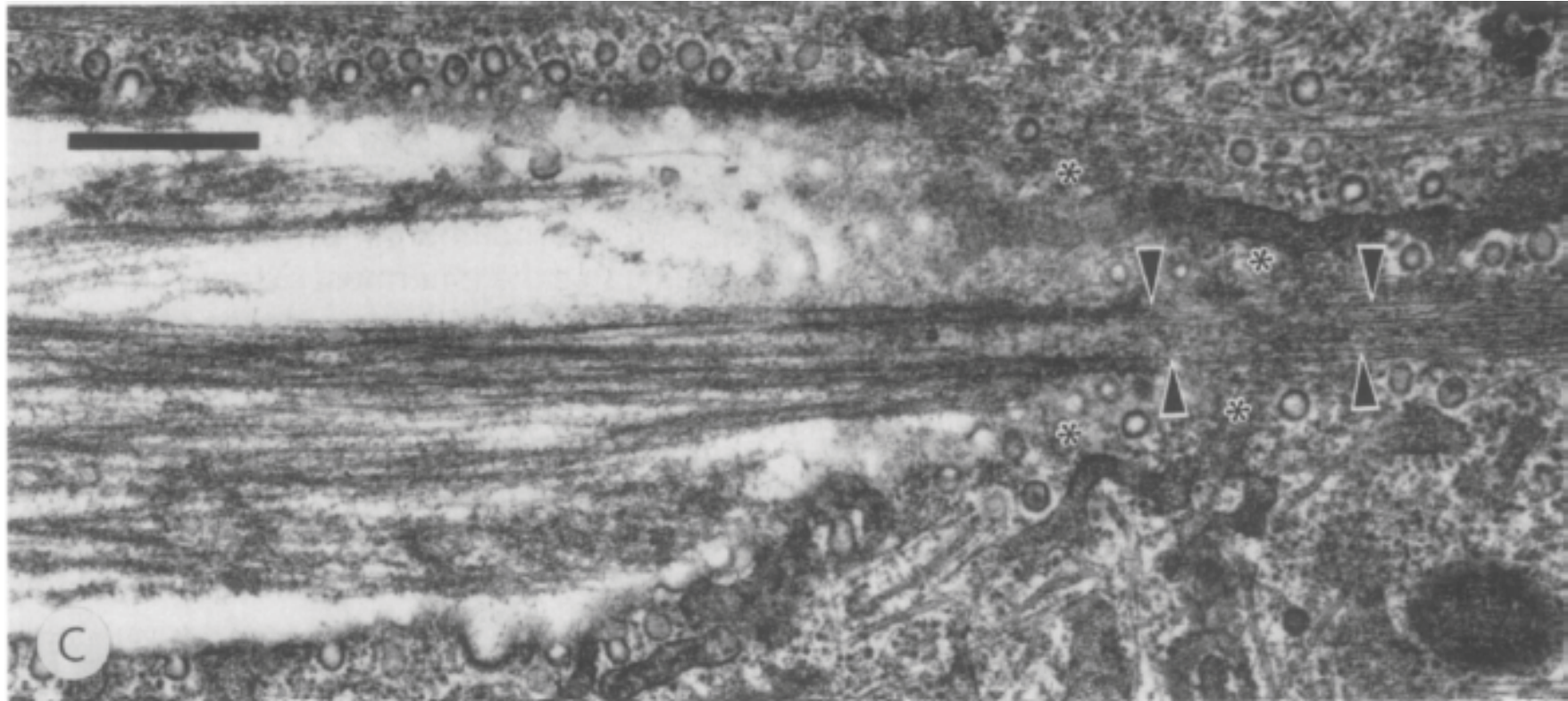
In silico: single molecule



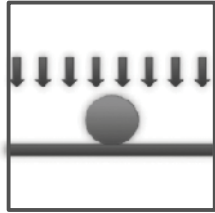


Fibronectin fiber architecture

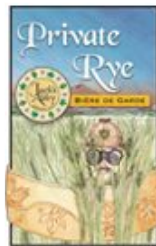
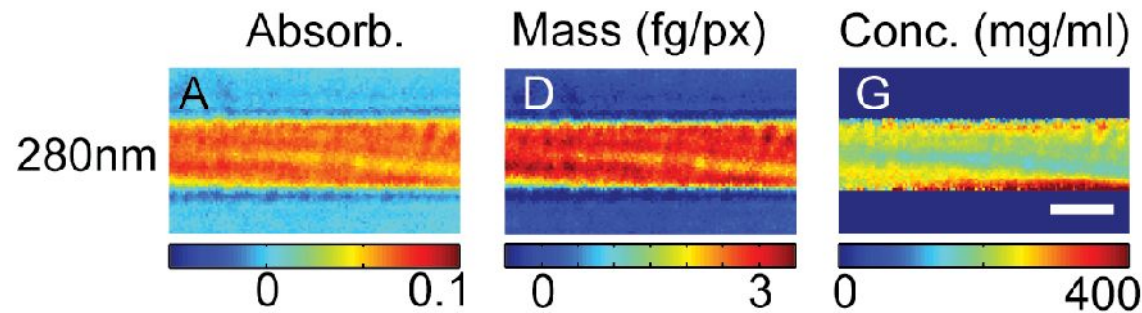
transmission electron microscopy



Singer. 1979. Cell. 16: 675.

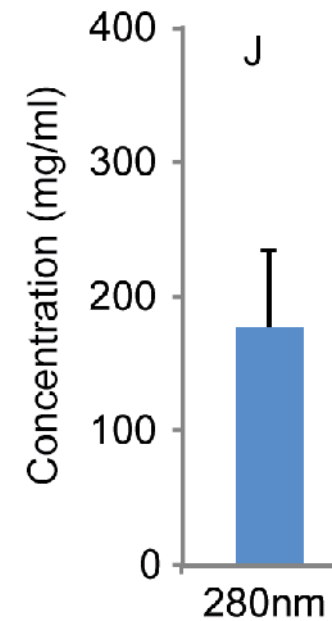


UV absorption microscopy



's Law

$$OD_{wavelength} = e_{wavelength} * c * l$$



Bradshaw, Cheung, Ehrlich, Smith. 2012. PLoSComputBiol. 8(12): e1002845.

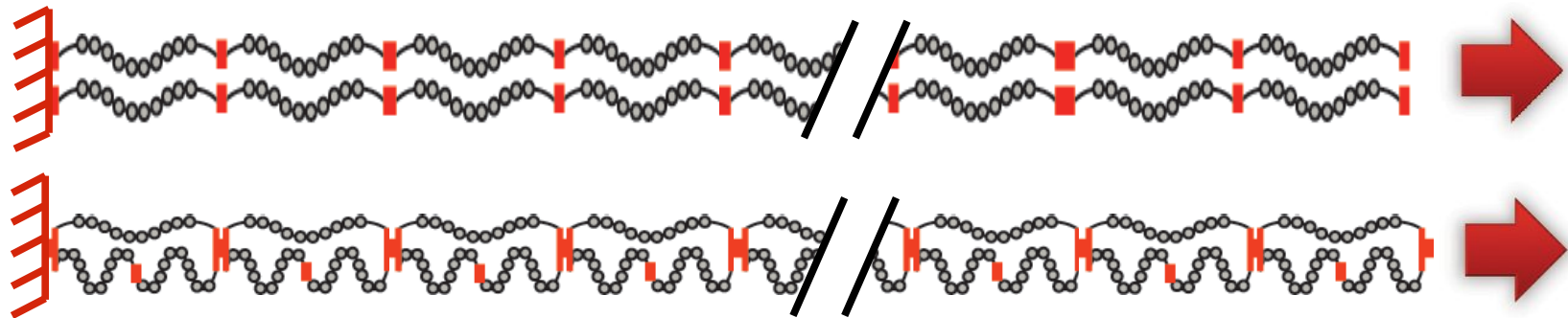


In silico: model fibronectin fiber

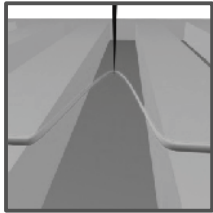
$$f = (k_B T / A) \left(z / L + 1 / (4(1 - z / L)^2) - 1 / 4 \right) \quad (1) \quad \text{Worm like chain equation.}$$

$$U = \sum_i \left[\left(\frac{k_b T}{A_i} \right) \left(\frac{z_i^2}{2L_i} + \frac{L_i}{4} \left(1 - \frac{z_i}{L_i} \right)^{-1} - \frac{z_i}{4} \right) \right] \quad (2) \quad \text{Integrate (1) and sum over all molecules to find potential energy.}$$

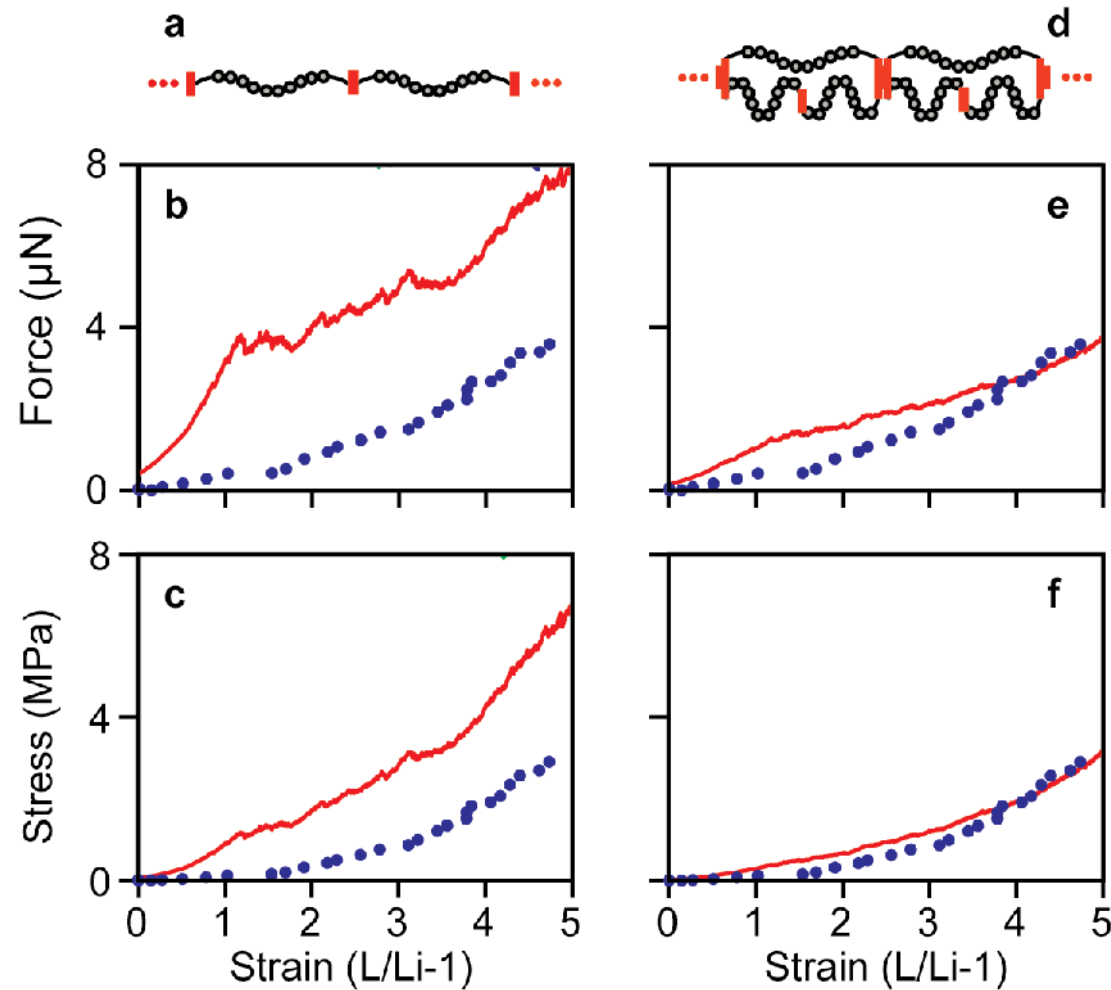
$$P_u = (k_u^0 * \Delta t) (\exp(f * \Delta x_u / k_B T)) \quad (3) \quad \text{The probability of an unfolding event.}$$



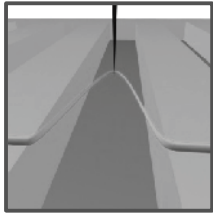
Bradshaw, Smith. 2011. *BiophysJ.* 101: 1740.



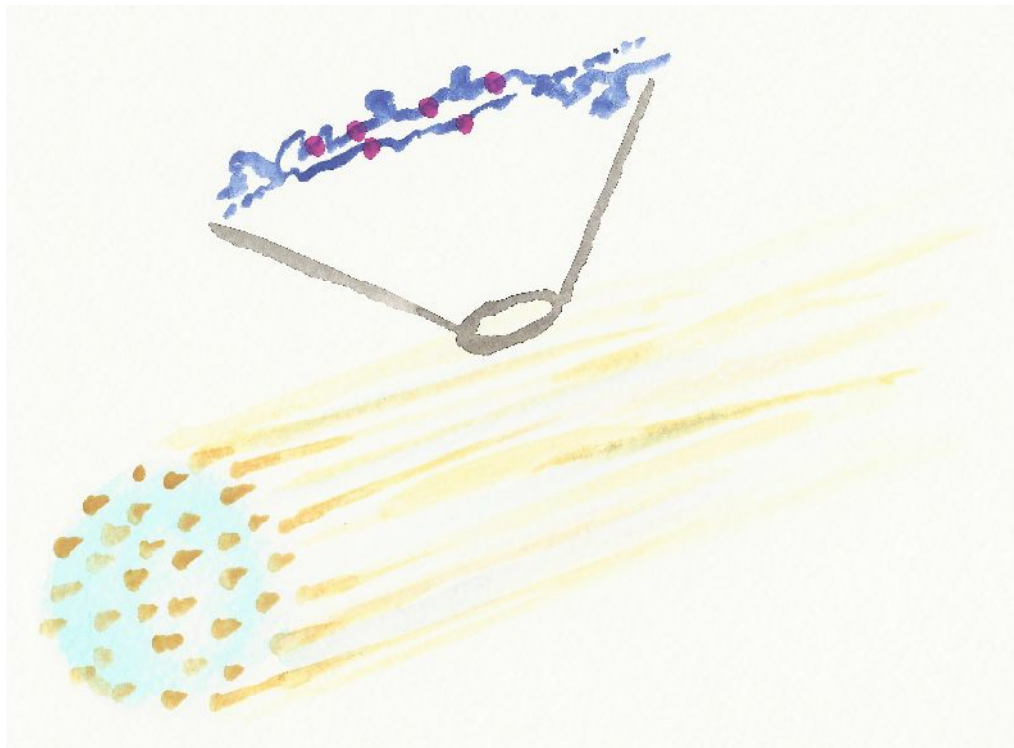
Mechanical comparison



Bradshaw, Cheung, Ehrlich, Smith. 2012. *PLoS Comput Biol.* 8(12): e1002845.



Insight: ligands and pores

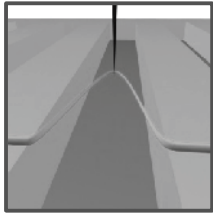


Architecture predictions:

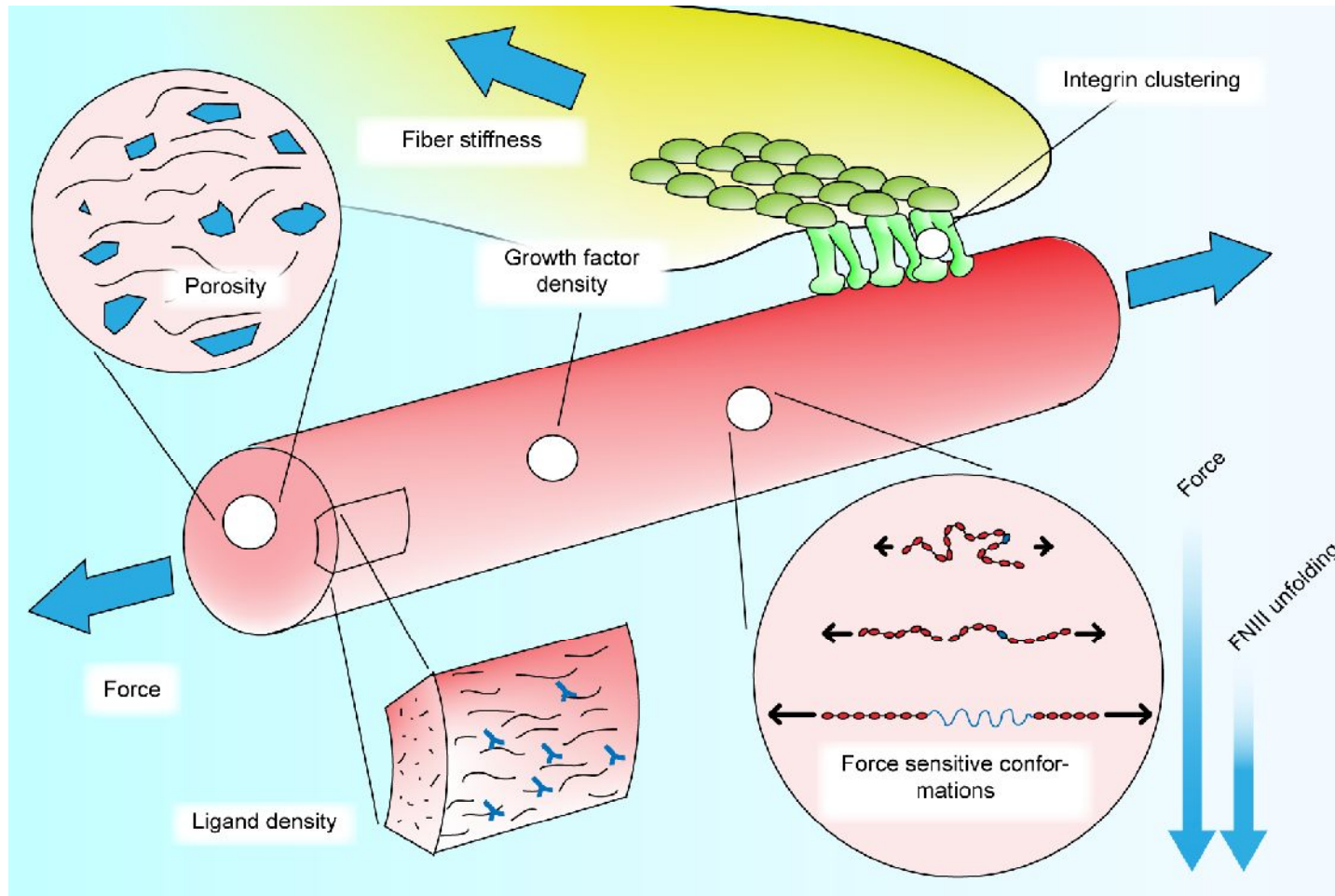
~2500 RGD/ μm^2

~15 nm spacing
between nanofibers

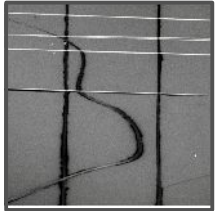
~these properties are
strain dependent!



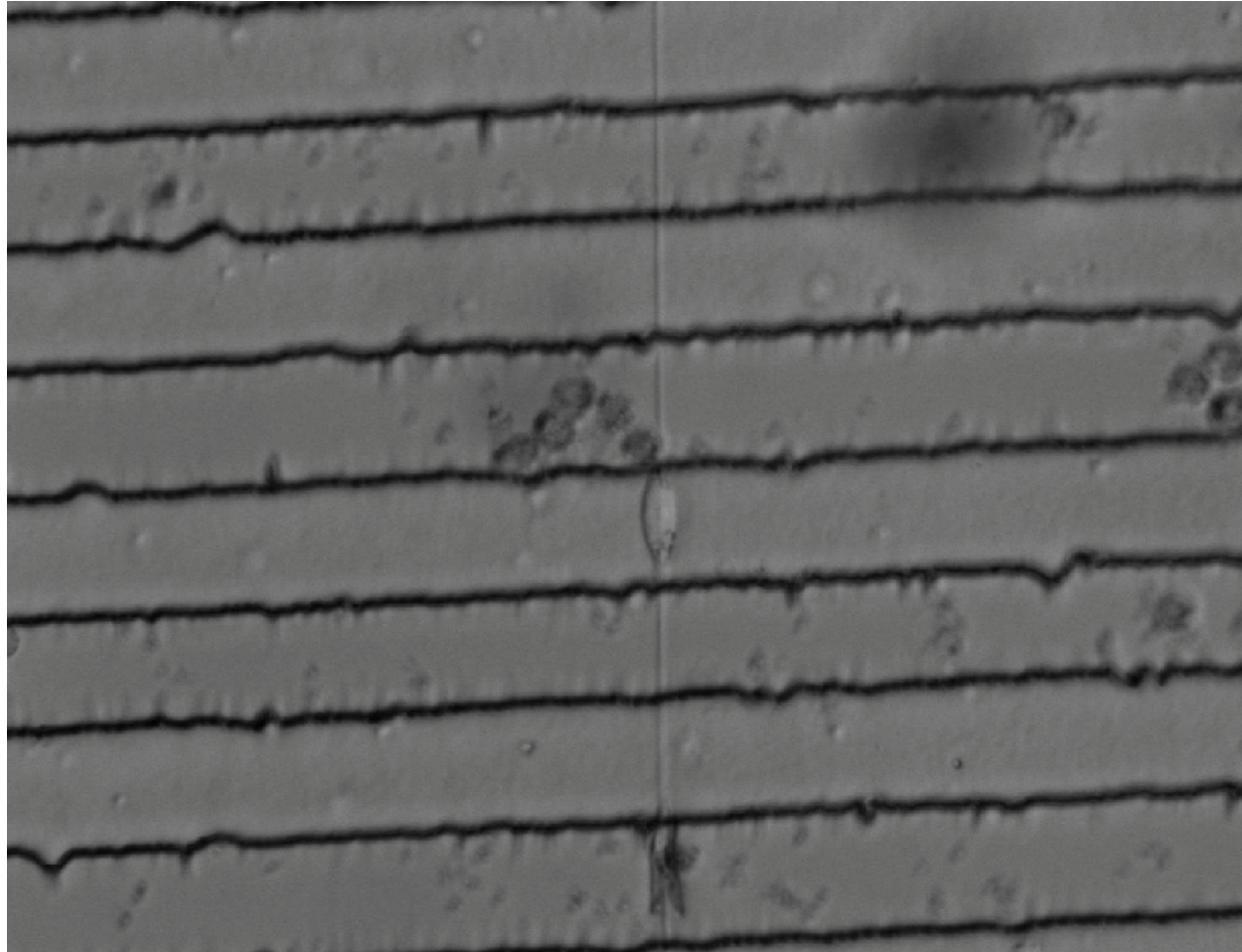
Insight:



Bradshaw, Smith. 2013. *ActaBiomater.* In Review.



Direct regulation of cell behavior





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