Figures

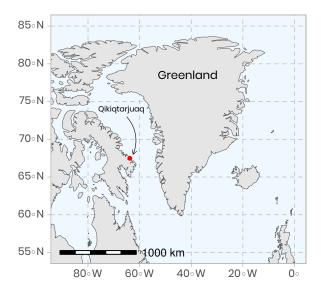


Figure 1: Location of the ice camp located near the Qikiqtarjuaq Island in the Baffin Bay. Projection used: EPSG-4326.

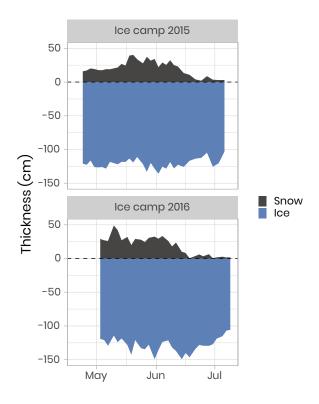


Figure 2: Temporal evolution of the snow and sea-ice thickness for both ice camp missions. The dashed horizontal line represents the snow/ice interface.

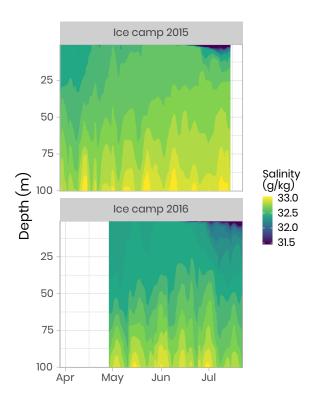


Figure 3: Temporal evolution of the salinity in the first 100 meters of the water column for both campaigns. Note that for visualization, salinity below 31.5 g kg $^{-1}$ have been binned to 31.5 g kg $^{-1}$.

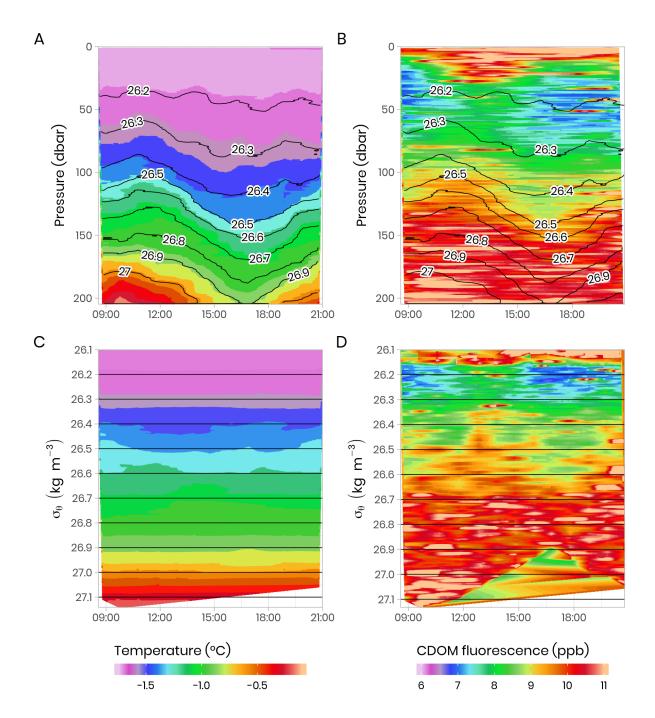


Figure 4: Temporal evolution of physical (temperature) and bio-optical (CDOM fluorescence) variables with superimposed lines of potential density anomaly (σ_{θ} , kg m⁻³) during a 13-h tidal cycle. Surface tidal height versus time at Qikiqtarjuaq is shown in blue. (A-B) Plotted versus pressure coordinates (equivalent to depth in meters). (C-D) The same data plotted versus potential density anomaly σ_{θ} coordinates (kg m⁻³). The tidal survey was performed on 2015-06-09.

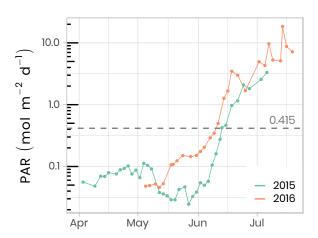


Figure 5: Temporal evolution of daily photosynthetically available radiation (PAR) at the seaice/water interface (1.3 m depth) for both ice camp missions. The horizontal dashed line shows the 0.415 mol photons m⁻² d⁻¹ threshold often used in the literature as the minimum light requirement for primary production.

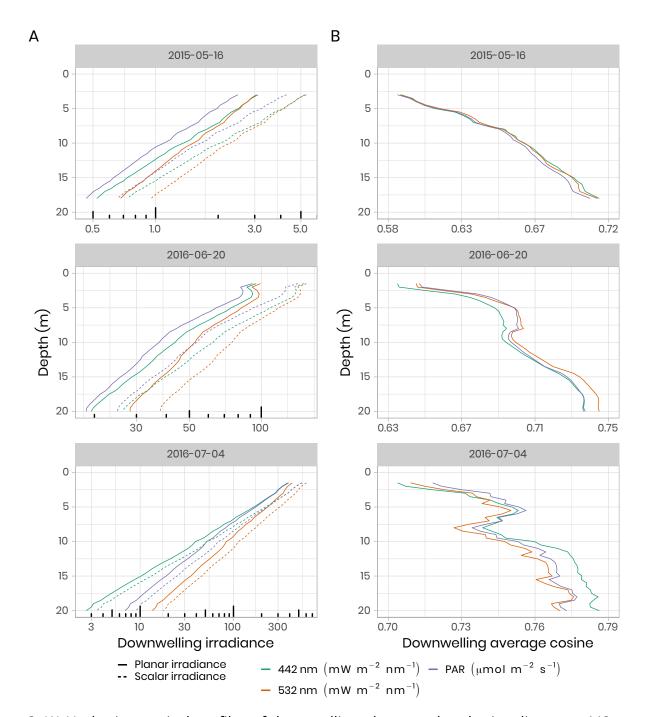


Figure 6: (A) Under-ice vertical profiles of downwelling planar and scalar irradiance at 442 nm, 532 nm and for PAR. (B) Calculated downwelling average cosine (unitless) was measured beneath snow-covered sea ice on 16 May 2015, beneath bare ice on 20 June 2016 and beneath a melt pond on 4 July 2016. Note the log scale for the irradiance measurements (A).

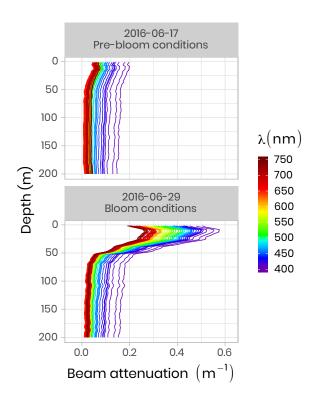


Figure 7: Beam attenuation coefficients (*c*, m⁻¹) measured in 2016 using an ACS before and during the phytoplankton bloom. Note that the colors of the lines correspond to wavelength frequencies.

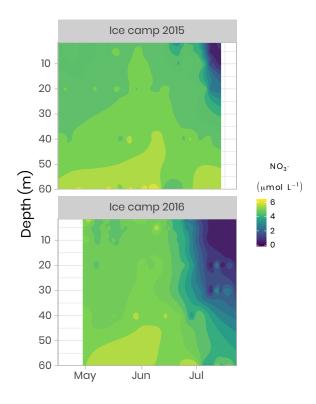


Figure 8: Temporal evolution of the nitrates in the first 60 m of the water column for both ice camp missions.

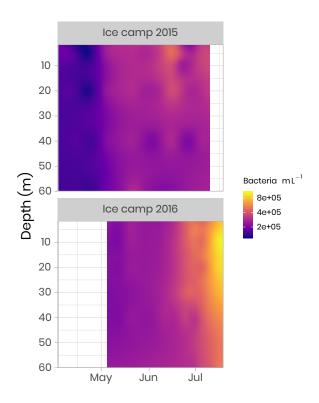


Figure 9: Concentration of bacteria in the water column at the ice camp in 2015 and 2016.

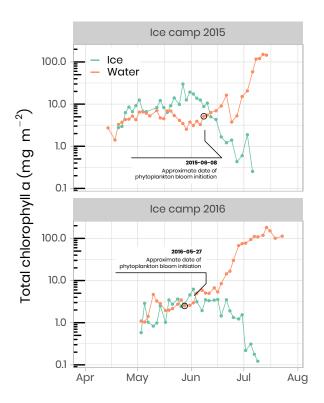


Figure 10: Temporal evolution of chlorophyll a in ice and water (depth-integrated) for both ice camp missions. Note that the water chlorophyll a have been integrated over the first 100 m of the water column whereas the ice chlorophyll a was measured on the bottom 0-10 cm of the ice cores. The details of the calculations to determine the approximate dates of phytoplankton bloom initiation can be found in Oziel et al. (2019).

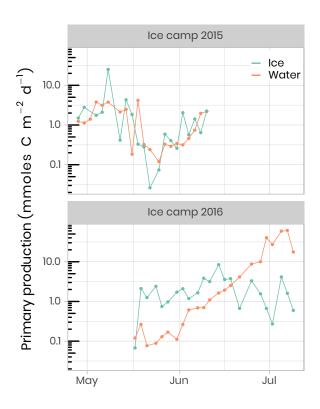


Figure 11: Temporal evolution of primary production a in ice and water (depth-integrated) for both ice camp missions.

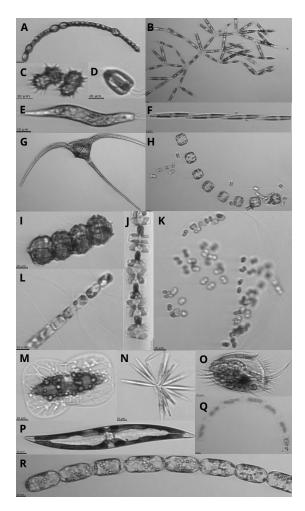


Figure 12: Images of protists sampled with the IFCB. Scale bar on images is 10 μm. Note that images are not to scale. (A) *Anabaena* sp. (B) *Nitzschia frigida* (C) *Polarella glacialis* (D) Flagellate (E) Euglena (F) *Pseudo-nitzschia* sp. (G) *Ceratium* sp. (H) *Thalassiosira nordenskioeldii* with *Attheya septentrionalis* (I) *Peridiniella catenata* (J) *Navicula pelagica* (K) *Phaeocystis* sp. colony (L) *Chaetoceros* sp. (M) *Entomoneis* sp. (N) *Synedropsis hyperborea* (O) Ciliate (P) Pennate diatom (Q) *Eucampia* sp. (R) *Melosira* sp.

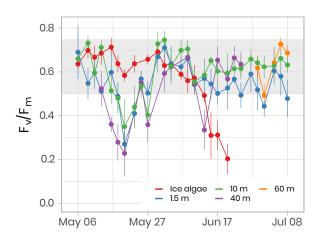


Figure 13: Temporal evolution of F_v/F_m for ice (last cm) and water underneath the ice (depths 1.5 m, 10 m, 40 m) samples for the ice camp 2016 between May 6th and July 8th. F_v/F_m monitoring on ice samples stopped on Day 172-June 20th because the Chl a fluorescence signal was not reliable anymore. F_v/F_m monitoring on 40 m and 60 m depth samples was limited between May 13th and June 24th and between June 29th-July 08th, respectively. The gray shaded area represents the range at which the algae are optimally growing.

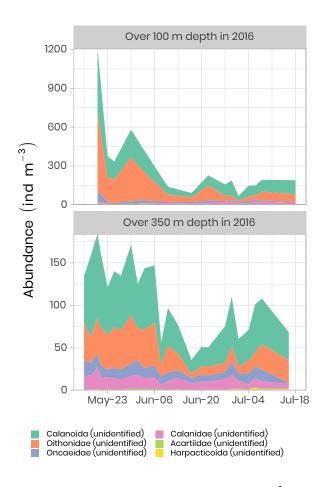


Figure 14: Time series of the abundance of the copepods (ind m⁻¹) measured over the first 100 m and 350 m of the water column in 2016 using the zooscan. For visualization, only the six most abundant groups are presented in decreasing order of importance. Note the different y-axes in both panels.