Cognitive processes are robust to early environmental conditions in two lizard species

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## Lay summary

Animals need to learn to adapt their behavior, but early developmental conditions can limit this ability. Factors like temperature and maternal stress hormones (glucocorticoids) may interact to influence learning, particularly in ectotherms. We studied how glucocorticoids and temperature during incubation affected learning in two lizard species. Surprisingly, both species maintained similar learning rates regardless of treatment, suggesting they could overcome early challenges. Additionally, color influenced their decision-making, highlighting the importance of color in cognitive tests.

## Abstract

Animals must acquire new information through learning to adjust their behavior adaptively. However, learning ability can be constrained by conditions experienced during early development, when the brain is especially susceptible to environmental conditions. For example, temperature can result in phenotypically plastic adjustments to growth, metabolism, and learning in ectotherms. In vertebrates, thermal conditions can increase the production of glucocorticoid (GCs) - ‘stress’ hormones. Maternal GCs can be transmitted to offspring during development, potentially impacting their learning abilities. GCs and thermal environments are, therefore, predicted to have interactive effects on the development of learning in ectotherms. Here, we investigated the combined effects of prenatal corticosterone (CORT) - the main GC in reptiles - and incubation temperature on associative learning using two species of lizards, *Lampropholis delicata* and *L. guichenoti*. We manipulated CORT levels and temperature in a 2x2 factorial design, and then subjected juveniles to a color-associative learning task. We predicted that elevated CORT and low temperatures would impair associative learning. However, both species showed similar learning rates independently of treatment. Our results suggest that these two species may have evolved mechanisms to maintain learning performance despite prenatal challenges. We also found that color affected decision-making in both species. Overall, we observed a non-learned preference towards blue, underscoring the need to carefully select the color used in cognitive tests involving visual stimuli.

## Introduction

Cognition is a set of processes by which animals gather, preserve, and use information from their environment through perception, learning, memory, and decision-making ([Shettleworth 2010](#ref-shettleworth)). These processes are fundamental to foraging, mate selection, antipredatory strategies, and social behaviors, all of which are crucial for survival and reproduction ([Dukas 2004](#ref-dukas_evolutionary_2004)). Learning - acquiring and consolidating new information ([Dukas 2004](#ref-dukas_evolutionary_2004)) - is essential for coping with environmental changes by enabling individuals to create new associations between events ([Dukas 2004](#ref-dukas_evolutionary_2004); [Leal and Powell 2012](#ref-leal_behavioural_2012); [Buchanan et al. 2013](#ref-buchanan_condition_2013)). However, the capacity to acquire information varies among individuals, shaped by factors such as age, sex, and the developmental environment ([Szuran et al. 1994](#ref-szuran_water_1994); [Lemaire et al. 2000](#ref-lemaire_prenatal_2000); [Zhu et al. 2004](#ref-zhu_prenatal_2004); [Amiel and Shine 2012](#ref-amiel_hotter_2012); [Amiel et al. 2014](#ref-amiel_egg_2014); [Carazo et al. 2014](#ref-carazo_sex_2014); [Noble et al. 2014](#ref-noble_age-dependent_2014)). Interindividual differences in learning can lead to unequal responses to environmental conditions, potentially affecting population dynamics ([Ward-Fear et al. 2016](#ref-ward2016ecological); [Welklin et al. 2024](#ref-welklin2024spatial)). For example, fast learners may better exploit novel resources or avoid new dangerous stimuli, while those with lower learning capabilities might struggle to adapt to environmental changes ([Ward-Fear et al. 2016](#ref-ward2016ecological)). Learning rate could affect their survival and reproductive output, ultimately influencing population growth rates and stability ([Ward-Fear et al. 2016](#ref-ward2016ecological); [Welklin et al. 2024](#ref-welklin2024spatial)). Therefore, understanding the factors that shape learning is crucial to predict how populations will respond to novel conditions.

Variation in learning is usually considered a product of an individual’s capacity to form associations between stimuli through memory formation ([Dukas 2004](#ref-dukas_evolutionary_2004)). However, differences in learning among individuals can also result from innate preferences or perceptual biases that influence how information is acquired and affect the decision-making process ([Toure and Reader 2022](#ref-toure2022colour)). The brain’s integration of information and the establishment of new connections is a complex process involving interactions among various nuclei to generate responses ([Dukas 2004](#ref-dukas_evolutionary_2004)). Consequently, any impact on specific brain regions or their interactions can directly influence learning through memory formation or alterations in decision-making.

Because the brain is highly susceptible to environmental inputs during early stages of life, developmental conditions are especially relevant in shaping cognitive abilities ([Zhu et al. 2004](#ref-zhu_prenatal_2004)). Impacts on brain development can have long-lasting effects on cognition, potentially influencing an individual’s capacity to learn and adapt to new environments ([Lemaire et al. 2000](#ref-lemaire_prenatal_2000); [Zhu et al. 2004](#ref-zhu_prenatal_2004); [Amiel and Shine 2012](#ref-amiel_hotter_2012); [Abayarathna and Webb 2020](#ref-abayarathna_effects_2020)). For example, the state of the mother can influence offspring phenotype beyond genetic transmission through nest-site selection, provisioning, or the transmission of hormones or nutrients (i.e., ‘maternal effects’ [Moore et al. 2019](#ref-moore_mothers_2019)); and the developing brain is also susceptible to these effects.

Glucocorticoids (GCs) are a class of steroid hormones that are particularly relevant in phenotypic plasticity. In vertebrates, GCs regulate metabolism and maintain homeostasis in response to disturbances (i.e., ‘the stress response’ [Sapolsky et al. 2000](#ref-sapolsky_how_2000); [Picard et al. 2014](#ref-picard_mitochondrial_2014)). Under stressful situations, animals react by initiating adaptive physiological and behavioral adjustments mediated by GCs. These GCs can be transmitted directly from parents to their offspring with various effects on offspring phenotype (reviewed by [Crino et al. 2023](#ref-Crino_2023)). Elevation in GCs during early stages of development typically results in altered neurogenesis, brain structure, and metabolic activity that, in most cases, are related to decreased learning abilities ([Lemaire et al. 2000](#ref-lemaire_prenatal_2000); [Zhu et al. 2004](#ref-zhu_prenatal_2004); [Du et al. 2009](#ref-du_dynamic_2009); [Eaton et al. 2015](#ref-eaton2015mild); [Farrell et al. 2015](#ref-farrell_developmental_2015-learn); [Farrell et al. 2016](#ref-farrell_developmental_2016)). For instance, prenatal stress in rats (*Rattus norvegicus*) suppresses neurogenesis in the dentate gyrus, which is associated with impairments in hippocampal-related spatial tasks ([Lemaire et al. 2000](#ref-lemaire_prenatal_2000)). Factors such as sex or the nature of the learning task can also affect the direction and magnitude of the effects of prenatal exposure to elevated GCs ([Szuran et al. 1994](#ref-szuran_water_1994); [Crino et al. 2014](#ref-crino_corticosterone_2014-learn); [Farrell et al. 2015](#ref-farrell_developmental_2015-learn); [Bebus et al. 2016](#ref-bebus_associative_2016); [Farrell et al. 2016](#ref-farrell_developmental_2016)). Because the effects of prenatal GCs on learning can be context-dependent, it is crucial to conduct studies across diverse taxa and experimental conditions to understand these effects fully.

In addition to the environments experienced by parents, offspring also experience potentially stressful environmental conditions that can interact with or amplify parental effects. In ectotherms, the early thermal environment is a mechanism of phenotypic plasticity, influencing a broad spectrum of traits, including growth, metabolism, or cognition ([Amiel and Shine 2012](#ref-amiel_hotter_2012); [Amiel et al. 2014](#ref-amiel_egg_2014); [Dayananda and Webb 2017](#ref-dayananda_incubation_2017); [Noble et al. 2018](#ref-noble_developmental_2018); [Abayarathna and Webb 2020](#ref-abayarathna_effects_2020)). For instance, Port Jackson sharks (*Heterodontus portusjacksoni*) incubated at warmer temperatures took fewer days to master a numerical learning task than those incubated at cooler ([Vila Pouca et al. 2019](#ref-vila_pouca_quantity_2019)). In skinks, high incubation temperatures are generally associated with faster learning rates ([Amiel and Shine 2012](#ref-amiel_hotter_2012); [Amiel et al. 2014](#ref-amiel_egg_2014); [Clark et al. 2014](#ref-clark_colour_2014)); but velvet geckos (*Amalosia lesueurii*) incubated at temperatures beyond their natural thermal range are worse learners than those incubated at colder temperatures ([Dayananda and Webb 2017](#ref-dayananda_incubation_2017); [Abayarathna and Webb 2020](#ref-abayarathna_effects_2020)). Changes in neural structure and function likely mediate the effects of incubation temperature in reptiles, as high temperatures increase neural density and metabolic activity in the brain ([Coomber et al. 1997](#ref-coomber_independent_1997); [Sakata et al. 2000](#ref-sakata_neural_2000); [Amiel et al. 2017](#ref-amiel_effects_2017); [Beltrán et al. 2021](#ref-beltran_are_2021)).

GCs can play a pivotal role in determining vertebrate responses to elevated temperatures ([Crino et al. 2023](#ref-Crino_2023)), potentially fostering natural interactions between temperature and GCs. Additionally, GCs and temperatures can act upon similar physiological mechanisms ([Coomber et al. 1997](#ref-coomber_independent_1997); [Lemaire et al. 2000](#ref-lemaire_prenatal_2000); [Sakata et al. 2000](#ref-sakata_neural_2000); [Zhu et al. 2004](#ref-zhu_prenatal_2004); [Du et al. 2009](#ref-du_dynamic_2009); [Amiel et al. 2017](#ref-amiel_effects_2017); [Beltrán et al. 2021](#ref-beltran_are_2021)). Therefore, the effects of GCs and temperature may be interdependent. However, the interactive effects of GCs and temperature on learning abilities remain unexplored, yet understanding of how temperature and GC’s interact is particularly relevant in global climate change ([Vinogradov et al. 2024](#ref-vinogradov2024inbreeding)).

Here, we investigated the combined effects of prenatal corticosterone (CORT) - the main GC in reptiles - and incubation temperature on cognition in two species of skinks, the delicate skink (*Lampropholis delicata*) and the common garden skink (*L. guichenoti*). We manipulated CORT levels in the eggs and then incubated them at two different temperatures in a 2X2 factorial design. Post-incubation, juveniles were subjected to a color-associative task to assess their learning abilities. We hypothesized that prenatal CORT levels and thermal environment would impact the learning of an association task. We predicted that individuals exposed to high levels of CORT or low temperatures would learn slower than control individuals or those incubated at higher temperatures ([Zhu et al. 2004](#ref-zhu_prenatal_2004); [Amiel and Shine 2012](#ref-amiel_hotter_2012); [Eaton et al. 2015](#ref-eaton2015mild); [Amiel et al. 2017](#ref-amiel_effects_2017)). Additionally, we predicted that incubation at high temperatures would mitigate the impact of CORT on learning performance, while cold incubation temperatures were expected to enhance the effects of CORT. The interactive effects of CORT and temperature may occur for two reasons that are not mutually exclusive: first, an increase in temperature leads to an overall rise in neural density ([Amiel et al. 2017](#ref-amiel_effects_2017)), thereby counteracting the impact of CORT (see [Lemaire et al. 2000](#ref-lemaire_prenatal_2000); [Zhu et al. 2004](#ref-zhu_prenatal_2004); [Eaton et al. 2015](#ref-eaton2015mild)); and second, the elevated metabolic rate associated with higher temperatures could accelerate CORT metabolism, resulting in embryos being exposed to CORT for a shorter time. In contrast, glucocorticoids in endotherms are associated with increased energy demands (e.g., Rubalcaba and Jimeno, 2022), which could lead to higher CORT production in lizards incubated at warmer temperatures. However, previous research on *L. delicata* (Crino et al., 2024) has found that cold-incubated lizards had higher baseline CORT levels, suggesting that cooler incubation temperatures may increase the potential effects of CORT exposure.

## Methods

#### Subjects

*Lampropholis delicata* and *L. guichenoti* are small [∼35-55 mm snout-vent length (SVL)], oviparous, and generalist skinks that usually share the same habitat in suburban areas throughout south-eastern Australia ([Chapple et al. 2011](#ref-chapple_know_2011)). Both species have similar breeding periods, but with some differences in reproductive output: while *L. delicata* lays 1-6 eggs in only one clutch per season, *L. guichenoti* clutches are smaller (1-5 eggs per clutch), and they usually lay two clutches per season ([Joss and Minard 1985](#ref-joss1985reproductive); [Chapple et al. 2011](#ref-chapple_know_2011); [Chapple et al. 2014](#ref-chapple_biology_2014)). Previous studies exploring behavioral differences between the two species have found *L. delicata* to be more exploratory than *L. guichenoti* ([Chapple et al. 2011](#ref-chapple_know_2011)). However, no differences in learning were observed between the skinks in an associative learning task ([Bezzina et al. 2014](#ref-bezzina2014does)).

#### Husbandry

*Breeding colony* – We tested juveniles from a 2019-established lab breeding colony. A total of 270 and 180 adults of *L. delicata* and *L. guichenoti*, respectively, were housed in plastic containers (41.5 L x 30.5 W x 21 H cm) with six lizards (2 males and 4 females) per enclosure. Enclosures were lined with non-stick matting, shelter, and several small water dishes. Water is given daily, and they were fed approximately 40 mid-size crickets (*Acheta domestica*) per enclosure three days a week. Crickets were dusted with calcium weekly and multivitamins and calcium biweekly. We used a heat chord and a heat lamp following a 12 h light:12 h dark cycle to ensure a temperature gradient. Room temperatures were set to 22-24 ºC, and the warm side of enclosures was usually at 32-34 ºC.

*Eggs collection and incubation* – Between mid-October 2022 to the end of February 2023, we provided females with a place to lay eggs by means of small boxes (12.5 L x 8.3 W x 5 H cm) containing moist vermiculite. These boxes were placed in one side of the communal enclosures (see above). We checked for eggs in the boxes three days a week (Monday, Wednesday, and Friday). After collection, we measured the length and width of eggs with a digital caliper to the nearest 0.1 mm, and mass using a digital scale (OHAUS, Model spx123; ± 0.001 g error). Then, we assigned individual IDs to the clutch and each egg. Eggs were treated with a CORT or control treatment (see CORT and Temperature Manipulation below) and were placed in individual cups (80 mL) with moist vermiculite (12 parts water to 4 parts vermiculite). The cups were covered with cling wrap to retain moisture and left in LABWIT 2X5D-R1160 incubators at two different temperatures (see CORT and Temperature manipulation below) until hatching. Eggs were then checked three times a week for hatchlings.

*Hatchlings* – Immediately after hatching, we measured lizards’ snout-to-vent length (SVL) and tail length (TL) with a ruler to the nearest mm, and mass using a digital scale (OHAUS, Model spx123; ± 0.001 g error). Hatchlings were then housed individually in small enclosures (18.7L x 13.2W x 6.3H cm) provided with non-stick matting, shelter, and a water dish. All care otherwise follows similar protocols to adults (see above).

Two weeks prior to the training phase (see details below), 1.5 to 5 months old lizards (see details in Statistical Analyses and Supplementary Material) were moved to the experimental arena for acclimatization. The arenas were individual medium-size (41 L x 29.7 W x 22 H cm) plastic containers with a shelter (9 L x 6 W x 1.5 H cm) on one of the sides and a water dish on the other. These arenas were placed in two rooms in 7 different racks that were monitored by 7 different CCTV systems (device model DVR-HP210475), allowing us to record their behavior during the experiment (see details below). Although the conditions in the experimental rooms were identical to the colony room, the number of lizards per species and treatment in each rack was counterbalanced to control for any potential effect of the room or the position of the lizard in the rack. During acclimatization and throughout the experiment, lizards were fed with only one cricket per day dusted with calcium and multivitamins, and water was supplied *ad libitum*. We provided a temperature gradient by means of a heat cord and heat lamps in a 12 h light: 12 h dark cycle.

#### CORT and Temperature Manipulation

To empirically test the effect of early environment, we manipulated CORT concentration in eggs and incubated them under one of two temperature regimes (‘Cold’ - 23 ± 3 ºC or ‘Hot’ - 28 ± 3 ºC) in a 2x2 factorial design ([Fig. 1](#fig-Methods) A). We topically dosed eggs with a CORT solution (10 pg/mL) or a control treatment (100% ethanol). Corticosterone treatments were made by dissolving crystalline corticosterone (Sigma, Cat. No. C2505) in 100% ethanol. To dose eggs, we applied 5µl of solutions to eggshells using a micropipette. We selected these doses based on previous studies publishing yolk CORT concentrations in other oviparous reptiles ([Lovern and Adams 2008](#ref-lovern2008effects); [Hanover et al. 2019](#ref-hanover2019corticosterone)), while also validating that it fell within the range of CORT concentrations in eggs within our population ([Crino et al. 2024](#ref-crino2024eggs)). CORT treatment increased mean yolk CORT levels ~3.7x higher than control eggs ([Crino et al. 2024](#ref-crino2024eggs)). After the treatment, the eggs were incubated in one of the two previously mentioned temperature regimes (‘Cold’ or ‘Hot’) until hatching. These temperatures represent the upper and lower limit of the natural incubation temperatures (Qualls and Shine 2000; Cheetam et al. 2011). The number of eggs per clutch assigned to each hormone and temperature treatment was counterbalanced in both species.

#### Learning

To estimate learning skills, we tested each lizard’s ability to locate a food reward in a color-associative learning task ([Fig. 1](#fig-Methods) C, D). First, we performed a training phase where lizards had to learn to eat from white 3D-printed PLA ramps (9 L x 4 W x 5 H cm) identical in size and shape to the ones used during the associative task ([Fig. 1](#fig-Methods) B). We divided the training phase into three stages: the first stage where lizards had to eat a small, frozen cricket (*A. domestica*) from an opaque petri dish (3D x 1.6H cm) placed in the middle of their enclosure ([Fig. 1](#fig-Methods) C, Stage 1); the second stage where the petri dish with the cricket was placed on top of the white 3D printed ramps ([Fig. 1](#fig-Methods) C, Stage 2); and the third where the cricket was left inside a well (3D x 1.75H cm) on the top of the ramp ([Fig. 1](#fig-Methods) C, Stage 3). Every trial began when we left the feeding block (petri dish, ramp, or both) inside the enclosure and finished one hour later when we took it away. At the end of each trial, we recorded whether the cricket had been consumed. A trial was considered successful if the lizard could locate and consume the reward, while completion of each stage required the lizards to eat the crickets in at least 5 out of 6 trials to ensure lizards were feeding consistently. This phase lasted 38 days until all the lizards learned to eat from the ramp; only in one case did we decide not to use the lizard because its behavior inconsistent throughout the course of the training phase.

During the associative learning phase, we trained lizards to associate color with a food reward ([Fig. 1](#fig-Methods) D). This test was like the third stage of the training phase, but here, lizards were presented with three 3D-printed ramps that differed in color. Ramps were green, red, and blue, as previous studies demonstrate that squamates can discriminate between these colors ([Baden and Osorio 2019](#ref-Baden_Osorio_2019_Vert_vision)). The food reward (small, frozen, *A. domestica* crickets) was placed inside the wells of the three ramps, covering two of the crickets with 3D-printed lids (3D x 0.5H cm) so prey was only accessible in “the correct” ramp. This way, we controlled for prey chemical cues, as the lids had a series of small holes on the top to allow the release of those chemicals. To control for potential color preference that could bias our results, we split the subjects into two groups counterbalanced by treatment and species: in one group the correct choice (i.e., the ramp with the non-covered frozen cricket) was blue, while the other group was assigned the red ramp as correct. In all trials, the position of the ramps was changed randomly to ensure subjects were using color rather than spatial cues for the association. Lizards were tested in this task once a day for 35 days.

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The experiment occurred between the 6th of March to the 17th of May 2023, and tests were performed between 1100 to 1200 when the lizards were active. Trials in the learning phase were recorded with CCTV systems, always using the same camera per individual. All the videos were analysed by the same observer (PR) who was blind to the treatment of the lizards. We recorded whether the animal chose the correct ramp first or not. We considered that a choice was made if the head of the lizard was inside the well of one of the ramps. We considered a trial to have failed if there was no choice after one hour of recording. These trials were scored as NA. We excluded from our analyses those individuals that performed inconsistently, as defined as not choosing in less than 20 out of 35 trials (~57%). We considered each lizard’s first trial to be the first one where a choice was made.

#### Statistical analyses

We performed analyses for each species separately. Preliminary analyses showed a significant effect of the color. As such, we decided to split the data by color (blue or red). Therefore, we ran a total of four different Bayesian multilevel models using the *brm* function from the brms package ([Bürkner 2017](#ref-burkner2017brms)) in R (version 2.8.2) ([R Core Team 2021](#ref-R)). We ran four parallel MCMC chains of 3000 iterations for each model, with a warmup period of 1000 iterations. We checked that all MCMC chains converged (Rhat < 1.2) and mixed effectively to ensure we had >1000 effective samples from the posterior distribution.

We modelled correct choice [correct (1) or not (0)] as the response variable, and trial, incubation temperature (Cold versus Hot), hormone (CORT versus Control), and the three-way interaction as fixed factors. The error structure was modeled using a Bernoulli distribution with a logit link function [family = Bernoulli(link = ‘logit’)]. We included each lizard’s random intercept and slope (trial) in our models. We also incorporated the clutch identity as a random factor. *L. delicata* lays one clutch per breeding season while *L. guichenoti* lays up to two ([Joss and Minard 1985](#ref-joss1985reproductive); [Chapple et al. 2011](#ref-chapple_know_2011); [Chapple et al. 2014](#ref-chapple_biology_2014)). Since egg collection was done during half of the breeding season, each clutch likely came from a unique mother, so clutch identity captures potential maternal effects. In addition, previous studies indicate that clutches are typically fertilized by a single male, although sperm storage can occur ([Kar et al. 2024](#ref-kar2024heritability)). Considering our partial split-clutch design and the expectation that maternal effects are likely more pronounced than paternal effects in these species, incorporating clutch as a random factor should effectively account for parental condition.

Learning in lizards can be age-dependent ([Noble et al. 2014](#ref-noble_age-dependent_2014)), and given that incubation temperature will affect hatching time ([Zhang et al. 2023](#ref-zhang2023developmental)), we explored the effects of age on learning by including it as a predictor. Lizard ages at the beginning of the experiment ranged from 41 to 148 days old in *L. delicata* and 48 to 132 in *L. guichenoti*. However, when this variable was included in the models, we did not find any significant effect of age (see *Supplementary Material*). As such, we present models without age as a fixed effect.

Learning rates were the estimated slopes of ‘trial’ and its interaction with hormone and temperature treatments. We used the posterior distributions of parameters to test for differences in learning rates between treatments and species. Slopes directly measure the change in the probability of choosing the correct dish across trials. Learning rates greater than zero, indicating that lizards were improving over trails, were considered evidence of learning, while those less or equal to zero were not.

Decision-making, in contrast, is considered the average probability of choosing correctly in a given trial. We estimated the predicted probability of choosing the correct ramp in the first trial using the intercepts from our posterior distributions following the formula:

In our case, we were interested in estimating the probability of correct choice at the early stages of learning [P(1)] because perception can play a role in the learning process ([Toure and Reader 2022](#ref-toure2022colour)). It also allowed us to test if lizards were biased towards the assigned color.

We calculated the pmcmc to test the hypothesis that learning rates and contrasts were different from zero, and that the probability of choosing correctly in the first trial was higher than expected by chance (0.33). We considered statistical significance as pmcmc < 0.05.

## Results

We started with 96 lizards, 48 per species and 12 per treatment per species. However, due to mortality (n = 11), failing to pass the training stage (n = 1), or lack of motivation during the learning tasks (n = 3), we had a final sample size of 81 lizards. Final sample sizes per treatment and species are listed in [Fig. 2](#fig-deli) and [Fig. 3](#fig-guich) (figures for both species with the raw data are included in the *Supplementary Material*). Mean slopes (denoted as β throughout) per treatment for both species are provided in Table 1 in the *Supplementary Material*.

#### Early developmental environments do not influence visual-sensory systems to impact decision-making

*Lampropholis delicata*: individuals given the blue ramp as the correct choice had an 89.5% increase in the probability of choosing the right ramp on trial 1 compared to lizards assigned to the red ramps (mean PBlue = 0.657, 95% CI PBlue = [0.361 , 0.896]; mean PRed = 0.347, 95% CI PRed = [0.069 , 0.835]), suggesting a bias towards blue. A blue bias was also supported by initial choices differing significantly from chance, and this was consistent across all treatments (i.e., P > 0.33; Table 2 in *Supplementary Material*). However, this difference in the probability of correctly choosing between red and blue ramps was not significant, likely due to small sample sizes (Contrast between probabilities: PBlue - PRed = 0.310, pmcmc = 0.23, see [Fig. 2](#fig-deli)).

Decision-making by *L. delicata* was not impacted by the CORT (PControl - PCORT = 0.188, pmcmc = 0.20), incubation temperature (PHot - PCold = 0.237, pmcmc = 0.14) or their interaction ([(PControl-Hot - PCORT-Hot) - (PControl-Cold - PCORT-Cold)] = 0.072, pmcmc = 0.20) when red ramps were the correct choice (see [Fig. 2](#fig-deli) A). When blue ramps were the correct choice, there was no significant effect of hormone (PControl - PCORT = 0.118, pmcmc = 0.39), temperature (PHot - PCold = -0.073, pmcmc = 0.65) or their interaction ([(PControl-Hot - PCORT-Hot) - (PControl-Cold - PCORT-Cold)] = 0.005, pmcmc = 0.75) either (see [Fig. 2](#fig-deli) D).

*Lampropholis guichenoti*: when the correct ramps were blue, *L. guichenoti* had a 115.7% increase in the probability of choosing the right ramp on trial 1 compared to red ramps (mean PBlue = 0.553, 95% CI PBlue = [0.201 , 0.927]; mean PRed = 0.256, 95% CI PRed = [0.028 , 0.806]). However, the difference was not significant (Contrast between probabilities: PBlue - PRed = 0.297, pmcmc = 0.16, see [Fig. 3](#fig-guich)).

Decision-making by *L. guichenoti* when the red ramps are the correct choice, was not affected by the hormone treatment (PControl - PCORT = 0.188, pmcmc = 0.20), temperature treatment (PHot - PCold = 0.230, pmcmc = 0.10) or their interaction ([(PControl-Hot - PCORT-Hot) - (PControl-Cold - PCORT-Cold)] = 0.125, pmcmc = 0.25; see [Fig. 3](#fig-guich) A). When blue ramps were the correct choice, neither CORT (βControl - βCORT = 0.231, pmcmc = 0.21) incubation temperature (βHot - βCold = 0.118, pmcmc = 0.43; ), or their interaction ([(PControl-Hot - PCORT-Hot) - (PControl-Cold - PCORT-Cold)] = -0.037) have significant effects ([Fig. 3](#fig-guich) D).

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#### Early developmental environments do not impact the rates of learning across species

*Lampropholis delicata*: Learning rates for those *L. delicata* assigned to red ramps did not show any significant effects of CORT (βControl - βCORT = -0.014, pmcmc = 0.76), incubation temperature (βHot - βCold = -0.004, pmcmc = 0.94), or their interaction ([(βControl-Hot - βCORT-Hot) - (βControl-Cold - βCORT-Cold)] = 0.061, pmcmc = 0.17) ([Fig. 2](#fig-deli) B, C). Similarly, those assigned to blue were also not affected by CORT (βControl - βCORT = 0.000, pmcmc = 0.98), temperature (βHot - βCold = 0.040, pmcmc = 0.32), or their interaction ([(βControl-Hot - βCORT-Hot) - (βControl-Cold - βCORT-Cold)] = 0.041, pmcmc = 0.41) ([Fig. 2](#fig-deli) E, F).

*Lampropholis guichenoti*: Learning rates in *L. guichenoti* were not influenced by hormone treatment (βControl - βCORT = -0.009, pmcmc = 0.84), temperature treatment (βHot - βCold = -0.032, pmcmc = 0.49), or their interaction ([(βControl-Hot - βCORT-Hot) - (βControl-Cold - βCORT-Cold)] = 0.026, pmcmc = 0.66) when red ramps were the correct choice ([Fig. 3](#fig-guich) B, C). Similarly, CORT (βControl - βCORT = -0.046, pmcmc = 0.31), incubation temperature (βHot - βCold = 0.040, pmcmc = 0.38), or their interaction ([(βControl-Hot - βCORT-Hot) - (βControl-Cold - βCORT-Cold)] = 0.016, pmcmc = 0.79) were not significant for *L. guichenoti* assigned to blue ramps (see [Fig. 3](#fig-guich) E, F).

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#### Contrasting impacts of early developmental environments on decision-making and learning in Lampropholis delicata and Lampropholis guichenoti

Overall, decision-making did not differ between species when the red ramps (mean P*L. delicata* = 0.347, 95% CI P*L. delicata* = [0.069 , 0.835]; mean P*L. guichenoti* = 0.256, 95% CI P*L. guichenoti* = [0.028 , 0.806]; Contrast between probabilities: P*L. delicata* - P*L. guichenoti* = 0.091, pmcmc = 0.52) or the blue ramps were the correct choice (mean P*L. delicata* = 0.657, 95% CI P *L. delicata* = [0.361 , 0.896]; mean P*L. guichenoti* = 0.553, 95% CI P*L. guichenoti* = [0.201 , 0.927]; P*L. delicata* - P*L. guichenoti* = 0.105, pmcmc = 0.61).

We also found no significant differences in learning rates between species when the red ramp (mean β*L. delicata* = 0.077, 95% CI β*L. delicata* = [0.026 , 0.135]; mean β*L. guichenoti* = 0.092, 95% CI β*L. guichenoti* = [0.023 , 0.171]; Contrast between slopes: β*L. delicata* - β*L. guichenoti* = -0.016, pmcmc = 0.72) or blue ramp was the correct choice (mean β*L. delicata* = 0.023, 95% CI β*L. delicata* = [-0.041 , 0.090]; mean β*L. guichenoti* = 0.063, 95% CI β*L. guichenoti* = [-0.030 , 0.149]; Contrast between slopes: β*L. delicata* - β*L. guichenoti* = -0.040, pmcmc = 0.40).

## Discussion

We predicted that elevated CORT levels in eggs or incubating them at colder temperatures would decrease learning rates, and that warmer incubation temperatures would suppress the negative effects of CORT on learning. In contrast to our predictions, both species learned to associate a color with a food reward at the same rate regardless of the experimental treatment. These findings suggest that both species may have developed mechanisms to buffer against early environmental stressors and maintain learning performance on colour-associative tasks despite prenatal challenges. While we did not find any effect of treatment on decision-making, there was a clear bias towards the color blue irrespective of the treatment, though this pattern differed between the species. We discuss the implications of these findings below.

#### Learning is not impacted by prenatal corticosterone exposure

Prenatal CORT was predicted to have a negative effect on learning ([Lemaire et al. 2000](#ref-lemaire_prenatal_2000); [Zhu et al. 2004](#ref-zhu_prenatal_2004); [Eaton et al. 2015](#ref-eaton2015mild); [Farrell et al. 2015](#ref-farrell_developmental_2015-learn); [Bebus et al. 2016](#ref-bebus_associative_2016)). However, we found no significant difference in learning rates between hormone treatments in either species. Our results are consistent with other experiments that showed no influence of prenatal GCs on learning abilities ([Szuran et al. 1994](#ref-szuran_water_1994); [Bebus et al. 2016](#ref-bebus_associative_2016); [Farrell et al. 2016](#ref-farrell_developmental_2016)) and suggest that *L. delicata* and *L. guichenoti* may have developed strategies to buffer the impacts of prenatal stress.

Alternatively, our observations could be obscured by other variables that are known to influence the extent and direction of the effects of GCs ([Szuran et al. 1994](#ref-szuran_water_1994); [Crino et al. 2014](#ref-crino_corticosterone_2014-learn); [Farrell et al. 2015](#ref-farrell_developmental_2015-learn); [Bebus et al. 2016](#ref-bebus_associative_2016); [Farrell et al. 2016](#ref-farrell_developmental_2016)). GCs are known to have hormetic effects ([Du et al. 2009](#ref-du_dynamic_2009)). Generally, low concentrations or short exposure to GCs have been shown to improve the rate at which animals learn while high concentrations or exposure to GCs during long periods have the opposite effects ([Du et al. 2009](#ref-du_dynamic_2009); [McEwen 2012](#ref-mcewen_brain_2012)). Thus, high elevations in CORT could affect learning through permanent changes in brain function (programmatic effects), while lower doses can result in lifelong elevation of baseline CORT, affecting learning through activational effects. Our findings indicate that associative learning in *L. delicata* and *L. guichenoti* is robust to the programmatic effects of elevated CORT exposure. Similarly, exposing embryos at different stages of development could impact the effects of CORT. However, we cannot exclude the possibility that concentrations different from the one used here or exposure at different developmental times does affect learning.

Conversely, the absence of significant effects of prenatal CORT on learning could be due to the cognitive task employed. Some studies show that the impact of GCs can vary between brain regions ([Lemaire et al. 2000](#ref-lemaire_prenatal_2000)). GC actions involve genomic and nongenomic mechanisms that implicate different types of receptors that can be distributed differentially in the various regions of the brain ([McEwen 2012](#ref-mcewen_brain_2012)), and may be related to the learning performance in distinct tasks. For instance, in European starlings (*Sturnus vulgaris*), males stressed during early stages of development performed worse in a visual associative learning task, but show no differences with control birds with acoustic stimuli ([Farrell et al. 2015](#ref-farrell_developmental_2015-learn)). Similarly, scrub-jays (*Aphelocoma coerulescens*) with lower CORT levels as nestlings performed better on an associative learning test as adults but not on a reversal-learning task ([Bebus et al. 2016](#ref-bebus_associative_2016)). Our results suggest that prenatal CORT does not affect the brain regions involved in coding associative learning in *L. delicata* and *L. guichenoti*, but we cannot rule out the possibility that other cognitive domains, such as spatial learning, might be affected. Future studies should examine the differential effects of early-life stress on different cognitive domains.

#### Incubation temperature does not affect learning

We predicted hot-incubated lizards would perform better in the associative learning task, since most studies in other species demonstrate enhanced learning abilities when eggs are incubated at higher temperatures ([Amiel and Shine 2012](#ref-amiel_hotter_2012); [Amiel et al. 2014](#ref-amiel_egg_2014); [Clark et al. 2014](#ref-clark_colour_2014)). These studies employed incubation temperatures within natural nesting thermal limits. In contrast, in those studies where cold-incubated lizards outperformed hot incubated ones, the incubation temperatures employed in the hot condition were far above the natural thermal range of the species ([Dayananda and Webb 2017](#ref-dayananda_incubation_2017); [Abayarathna and Webb 2020](#ref-abayarathna_effects_2020)). As such, it is unclear how such these extreme conditions relate to wild environments. In *L. delicata* and *L. guichenoti*, associative learning appears robust to incubation temperature because of the lack of detectable effect temperature had on learning. Our results, therefore, do not align with previous studies. However, the effect of prenatal temperature on cognition, and brain physiology and structure has been investigated only in a small number of species (see [Coomber et al. 1997](#ref-coomber_independent_1997); [Sakata et al. 2000](#ref-sakata_neural_2000); [Amiel and Shine 2012](#ref-amiel_hotter_2012); [Amiel et al. 2014](#ref-amiel_egg_2014); [Clark et al. 2014](#ref-clark_colour_2014); [Amiel et al. 2017](#ref-amiel_effects_2017); [Dayananda and Webb 2017](#ref-dayananda_incubation_2017); [Abayarathna and Webb 2020](#ref-abayarathna_effects_2020)), limiting our understanding on how early thermal environment can affect cognitive abilities. Our results suggest that the impact of incubation temperature on learning may not be as widespread across reptiles as we think.

#### Visual-sensory bias is not dependent on early environment

We found strong evidence for a visual/sensory bias towards choosing the blue feeding ramp during the initial stages of decision-making. While this effect appeared stronger in *L. delicata* compared to *L. guichenoti*, the bias did not result from experiencing different early environmental conditions. What created an initial bias towards blue? One possibility might be that the bias towards blue ramps was a byproduct of the training phase where the blue ramp was more similar to the white training ramp than the green or red ramps. However, this hypothesis is unlikely given that the light spectrum and the perceived chromatic differences between the ramps used in the associative task and the ones used during training phase meant that the white training phase ramp was more similar to the green color rather than blue (see *Supplementary Material*). It seems more likely that lizards innately prefer the blue ramps. Some animals bias their attention towards colors they are familiar with ([Putman et al. 2017](#ref-putman2017fear) - the ‘species confidence hypothesis’). For instance, dark and light blue T-shirts were associated with lower flight initiation distances and higher capture rates in Western fence lizards where blue is used in intraspecific communication ([Putman et al. 2017](#ref-putman2017fear)). Nevertheless, to the best of our knowledge, there has been no reported color bias in *L. delicata* or *L. guichenoti* before; and blue coloration is not considered to be involved in intraspecific communication in these species ([Torr and Shine 1996](#ref-torr1996patterns); [Chapple et al. 2014](#ref-chapple_biology_2014)). Regardless, our results demonstrate that, at least in *L. delicata*, there is a bias toward blue colors, highlighting the need to consider the colors used in associative learning paradigms carefully.

#### Learning rates between species did not vary

We did not see any difference in learning rates when comparing both species. Since both species occupy similar habitats and have similar ecology (Chapple et al. 2011; 2013; 2014), it may not be surprising that cognitive abilities are similar between both species. Nonetheless, previous studies have shown that *L. guichenoti* is less prone to explore novel environments than *L. delicata*, which may be related to the success of *L. delicata* as an invasive species compared to *L. guichenoti* (Chapple et al., 2011). Our results show that the ability of *L. delicata* to colonize new areas seems not to be related to learning abilities. Our findings support previous studies that found similar learning abilities in *L. delicata* and *L. guichenoti* ([Bezzina et al. 2014](#ref-bezzina2014does)). In the experiment conducted by Bezzina et al. ([2014](#ref-bezzina2014does)), both species failed to complete the associative learning task under the authors’ criterion, while in our experiment, both species completed the task, exhibiting similar learning rates. Complexity, experimental design, or the criterion employed to define learning could be the major cause of the discrepancies between ours and Bezzina et al. ([2014](#ref-bezzina2014does)). Developing common strategies and approaches to assess animal learning could help understand how learning abilities are shaped in different taxa or environments.

#### Conclusion

Our results revealed that associative learning abilities and decision-making in *L. delicata* and *L. guichenoti* are resilient to prenatal CORT and temperature. This outcome contrasts with our initial predictions, indicating that the learning skills of these lizards may be more robust than anticipated under varying early life conditions. We also found significant effects of the color employed in the task on learning rates in most *L. delicata* and some *L. guichenoti*. These results seem to be a consequence of an innate color bias and highlight the importance of carefully selecting the color employed when testing cognition using visual stimuli.

Future research should continue exploring the potential effects of prenatal corticosterone (CORT) and temperature on other cognitive functions. Furthermore, it is crucial to explore how these treatments influence brain function at a neurological level. Investigating these aspects will help us understand these species’ cognitive and physiological mechanisms underpinning adaptability and offer insights into how early developmental factors shape long-term cognitive outcomes.

## Acknowledgements

We thank the help and assistance of our lab technicians Benjamin Durant and Michelle Stephens for taking care of the lizards. We are also grateful to ACTHA for the grant for the 3D printed ramps, and we also thank ANU MakerSpace, where we designed and built the prototypes of the 3D printed ramps. Finally, we wish to acknowledge the anonymous reviewers for their valuable feedback on the manuscript.

## Funding

This work was supported by a National Australian University fellowship (PR), the Australian Research Council (grant no. DP210101152) to DN and CRF, and the ACT Herpetological Association grant to PR.

## Authors’ contributions

PR: conceptualization, methodology, data collection, data curation, formal analysis, writing—original draft, writing—review and editing; DCL: data collection, writing—review and editing; OC: conceptualization, methodology, writing—review and editing; KW: conceptualization, writing—review and editing; CRF: conceptualization, methodology, funding acquisition, writing—review and editing; BM: data collection, writing—review and editing; AP: data collection, writing—review and editing; DN: conceptualization, methodology, funding acquisition, project administration, resources, supervision, writing—review and editing. All authors gave final approval for publication and agreed to be held accountable for the work performed therein.

## Data accessibility

All data, data description, and R code are available in public repository <https://github.com/Pablo-Recio/CORT_Temp_learning>.

## 

## Satements and declarations

#### Competing Interests

We declare we have no competing interests

#### Conflict of Interest

We declare we have no conflict of interest

#### Ethical Approval

All experimental procedures and laboratory housing complied with Australian law and were approved by the Australian National University Animal Experimentation Ethics Committee (A2022\_33).

## References

Abayarathna T, Webb JK. 2020. Effects of incubation temperatures on learning abilities of hatchling velvet geckos. Animal Cognition. 23(4):613–620. doi:[10.1007/s10071-020-01365-4](https://doi.org/10.1007/s10071-020-01365-4). [accessed 2023 Jun 21]. <http://link.springer.com/10.1007/s10071-020-01365-4>.

Amiel JJ, Bao S, Shine R. 2017. The effects of incubation temperature on the development of the cortical forebrain in a lizard. Animal Cognition. 20(1):117–125. doi:[10.1007/s10071-016-0993-2](https://doi.org/10.1007/s10071-016-0993-2). [accessed 2023 Jun 21]. <http://link.springer.com/10.1007/s10071-016-0993-2>.

Amiel JJ, Lindström T, Shine R. 2014. Egg incubation effects generate positive correlations between size, speed and learning ability in young lizards. Animal Cognition. 17(2):337–347. doi:[10.1007/s10071-013-0665-4](https://doi.org/10.1007/s10071-013-0665-4). [accessed 2023 Jun 21]. <http://link.springer.com/10.1007/s10071-013-0665-4>.

Amiel JJ, Shine R. 2012. Hotter nests produce smarter young lizards. Biology Letters. 8(3):372–374. doi:[10.1098/rsbl.2011.1161](https://doi.org/10.1098/rsbl.2011.1161). [accessed 2023 Jun 21]. <https://royalsocietypublishing.org/doi/10.1098/rsbl.2011.1161>.

Baden T, Osorio D. 2019. The retinal basis of vertebrate color vision. Annual Review of Vision Science.:177–200.

Bebus SE, Small TW, Jones BC, Elderbrock EK, Schoech SJ. 2016. Associative learning is inversely related to reversal learning and varies with nestling corticosterone exposure. Animal Behaviour. 111:251–260. doi:[10.1016/j.anbehav.2015.10.027](https://doi.org/10.1016/j.anbehav.2015.10.027). [accessed 2023 Jun 21]. <https://linkinghub.elsevier.com/retrieve/pii/S0003347215003991>.

Beltrán I, Herculano-Houzel S, Sinervo B, Whiting MJ. 2021. Are ectotherm brains vulnerable to global warming? Trends in Ecology & Evolution. 36(8):691–699. doi:[10.1016/j.tree.2021.04.009](https://doi.org/10.1016/j.tree.2021.04.009). [accessed 2023 Jun 21]. <https://linkinghub.elsevier.com/retrieve/pii/S0169534721001233>.

Bezzina CN, Amiel JJ, Shine R. 2014. Does invasion success reflect superior cognitive ability? A case study of two congeneric lizard species (lampropholis, scincidae). PLoS One. 9(1):e86271.

Buchanan KL, Grindstaff JL, Pravosudov VV. 2013. Condition dependence, developmental plasticity, and cognition: Implications for ecology and evolution. Trends in Ecology & Evolution. 28(5):290–296. doi:[10.1016/j.tree.2013.02.004](https://doi.org/10.1016/j.tree.2013.02.004). [accessed 2023 Jun 21]. <https://linkinghub.elsevier.com/retrieve/pii/S0169534713000566>.

Bürkner P-C. 2017. Brms: An r package for bayesian multilevel models using stan. Journal of statistical software. 80:1–28.

Carazo P, Noble DWA, Chandrasoma D, Whiting MJ. 2014. Sex and boldness explain individual differences in spatial learning in a lizard. Proceedings of the Royal Society B: Biological Sciences. 281(1782):20133275. doi:[10.1098/rspb.2013.3275](https://doi.org/10.1098/rspb.2013.3275). [accessed 2023 May 25]. <https://royalsocietypublishing.org/doi/10.1098/rspb.2013.3275>.

Chapple DG, Miller KA, Chaplin K, Barnett L, Thompson MB, Bray RD. 2014. Biology of the invasive delicate skink (Lampropholis delicata) on Lord Howe Island. Australian Journal of Zoology. 62(6):498. doi:[10.1071/ZO14098](https://doi.org/10.1071/ZO14098). [accessed 2023 May 25]. <http://www.publish.csiro.au/?paper=ZO14098>.

Chapple DG, Miller KA, Kraus F, Thompson MB. 2013. Divergent introduction histories among invasive populations of the delicate skink (l ampropholis delicata): Has the importance of genetic admixture in the success of biological invasions been overemphasized? Diversity and Distributions. 19(2):134–146.

Chapple DG, Simmonds SM, Wong BBM. 2011. Know when to run, know when to hide: Can behavioral differences explain the divergent invasion success of two sympatric lizards?: Invasion Success of Two Sympatric Lizards. Ecology and Evolution. 1(3):278–289. doi:[10.1002/ece3.22](https://doi.org/10.1002/ece3.22). [accessed 2023 May 25]. <https://onlinelibrary.wiley.com/doi/10.1002/ece3.22>.

Cheetham E, Doody JS, Stewart B, Harlow P. 2011. Embryonic mortality as a cost of communal nesting in the delicate skink. Journal of Zoology. 283(4):234–242.

Clark BF, Amiel JJ, Shine R, Noble DWA, Whiting MJ. 2014. Colour discrimination and associative learning in hatchling lizards incubated at ‘hot’ and ‘cold’ temperatures. Behavioral Ecology and Sociobiology. 68(2):239–247. doi:[10.1007/s00265-013-1639-x](https://doi.org/10.1007/s00265-013-1639-x). [accessed 2023 Jun 21]. <http://link.springer.com/10.1007/s00265-013-1639-x>.

Coomber P, Crews D, Gonzalez-Lima F. 1997. Independent effects of incubation temperature and gonadal sex on the volume and metabolic capacity of brain nuclei in the leopard gecko (Eublepharis macularius), a lizard with temperature-dependent sex determination. The Journal of Comparative Neurology. 380(3):409–421. doi:[10.1002/(SICI)1096-9861(19970414)380:3<409::AID-CNE9>3.0.CO;2-6](https://doi.org/10.1002/(SICI)1096-9861(19970414)380:3%3c409::AID-CNE9%3e3.0.CO;2-6). [accessed 2023 Jun 1]. [https://onlinelibrary.wiley.com/doi/10.1002/(SICI)1096-9861(19970414)380:3<409::AID-CNE9>3.0.CO;2-6](https://onlinelibrary.wiley.com/doi/10.1002/(SICI)1096-9861(19970414)380:3%3c409::AID-CNE9%3e3.0.CO;2-6).

Crino OL, Bonduriansky R, Martin LB, Noble DWA. 2023. A conceptual framework for understanding stressinduced physiological and transgenerational effects on population responses to climate change. Evolution Letters.

Crino OL, Driscoll SC, Ton R, Breuner CW. 2014. Corticosterone exposure during development improves performance on a novel foraging task in zebra finches. Animal Behaviour. 91:27–32. doi:[10.1016/j.anbehav.2014.02.017](https://doi.org/10.1016/j.anbehav.2014.02.017). [accessed 2023 Jun 21]. <https://linkinghub.elsevier.com/retrieve/pii/S000334721400102X>.

Crino O, Wild KH, Friesen CR, Leibold DC, Laven N, Peardon AY, Recio P, Noble DW, et al. 2024. From eggs to adulthood: Sustained effects of early developmental temperature and corticosterone exposure on physiology and body size in an australian lizard.

Dayananda B, Webb JK. 2017. Incubation under climate warming affects learning ability and survival in hatchling lizards. Biology Letters. 13(3):20170002. doi:[10.1098/rsbl.2017.0002](https://doi.org/10.1098/rsbl.2017.0002). [accessed 2023 Jun 21]. <https://royalsocietypublishing.org/doi/10.1098/rsbl.2017.0002>.

Du J, Wang Y, Hunter R, Wei Y, Blumenthal R, Falke C, Khairova R, Zhou R, Yuan P, Machado-Vieira R, et al. 2009. Dynamic regulation of mitochondrial function by glucocorticoids. Proceedings of the National Academy of Sciences. 106(9):3543–3548. doi:[10.1073/pnas.0812671106](https://doi.org/10.1073/pnas.0812671106). [accessed 2023 Jun 21]. <https://pnas.org/doi/full/10.1073/pnas.0812671106>.

Dukas R. 2004. Evolutionary Biology of Animal Cognition. Annual Review of Ecology, Evolution, and Systematics. 35(1):347–374. doi:[10.1146/annurev.ecolsys.35.112202.130152](https://doi.org/10.1146/annurev.ecolsys.35.112202.130152). [accessed 2023 Jun 21]. <https://www.annualreviews.org/doi/10.1146/annurev.ecolsys.35.112202.130152>.

Eaton L, Edmonds E, Henry T, Snellgrove D, Sloman K. 2015. Mild maternal stress disrupts associative learning and increases aggression in offspring. Hormones and behavior. 71:10–15.

Farrell TM, Morgan A, MacDougall-Shackleton SA. 2016. Developmental stress impairs performance on an association task in male and female songbirds, but impairs auditory learning in females only. Animal Cognition. 19(1):1–14. doi:[10.1007/s10071-015-0908-7](https://doi.org/10.1007/s10071-015-0908-7). [accessed 2023 Jun 21]. <http://link.springer.com/10.1007/s10071-015-0908-7>.

Farrell TM, Neuert MAC, Cui A, MacDougall-Shackleton SA. 2015. Developmental stress impairs a female songbird’s behavioural and neural response to a sexually selected signal. Animal Behaviour. 102:157–167. doi:[10.1016/j.anbehav.2015.01.018](https://doi.org/10.1016/j.anbehav.2015.01.018). [accessed 2023 Jun 21]. <https://linkinghub.elsevier.com/retrieve/pii/S0003347215000287>.

Hanover AM, Husak JF, Lovern M. 2019. Corticosterone in lizard egg yolk is reduced by maternal diet restriction but unaltered by maternal exercise. Physiological and Biochemical Zoology. 92(6):573–578.

Joss J, Minard J. 1985. On the reproductive cycles of lampropholis guichenoti and l. Delicata (squamata: Scincidae) in the sydney region. Australian Journal of Zoology. 33(5):699–704.

Kar F, Nakagawa S, Noble DW. 2024. Heritability and developmental plasticity of growth in an oviparous lizard. Heredity. 132(2):67–76.

Leal M, Powell BJ. 2012. Behavioural flexibility and problem-solving in a tropical lizard. Biology Letters. 8(1):28–30. doi:[10.1098/rsbl.2011.0480](https://doi.org/10.1098/rsbl.2011.0480). [accessed 2023 Jun 21]. <https://royalsocietypublishing.org/doi/10.1098/rsbl.2011.0480>.

Lemaire V, Koehl M, Le Moal M, Abrous DN. 2000. Prenatal stress produces learning deficits associated with an inhibition of neurogenesis in the hippocampus. Proceedings of the National Academy of Sciences. 97(20):11032–11037. doi:[10.1073/pnas.97.20.11032](https://doi.org/10.1073/pnas.97.20.11032). [accessed 2023 Jun 21]. <https://pnas.org/doi/full/10.1073/pnas.97.20.11032>.

Lovern MB, Adams AL. 2008. The effects of diet on plasma and yolk steroids in lizards (anolis carolinensis). Integrative and comparative biology. 48(3):428–436.

McEwen BS. 2012. Brain on stress: How the social environment gets under the skin. Proceedings of the National Academy of Sciences. 109(supplement\_2):17180–17185. doi:[10.1073/pnas.1121254109](https://doi.org/10.1073/pnas.1121254109). [accessed 2023 Jun 21]. <https://pnas.org/doi/full/10.1073/pnas.1121254109>.

Moore MP, Whiteman HH, Martin RA. 2019. A mother’s legacy: The strength of maternal effects in animal populations. Nakagawa S, editor. Ecology Letters. 22(10):1620–1628. doi:[10.1111/ele.13351](https://doi.org/10.1111/ele.13351). [accessed 2023 Jun 21]. <https://onlinelibrary.wiley.com/doi/10.1111/ele.13351>.

Noble DWA, Byrne RW, Whiting MJ. 2014. Age-dependent social learning in a lizard. Biology Letters. 10(7):20140430. doi:[10.1098/rsbl.2014.0430](https://doi.org/10.1098/rsbl.2014.0430). [accessed 2023 Jun 21]. <https://royalsocietypublishing.org/doi/10.1098/rsbl.2014.0430>.

Noble DWA, Stenhouse V, Schwanz LE. 2018. Developmental temperatures and phenotypic plasticity in reptiles: A systematic review and meta-analysis: Incubation temperature and plasticity. Biological Reviews. 93(1):72–97. doi:[10.1111/brv.12333](https://doi.org/10.1111/brv.12333). [accessed 2023 Jun 21]. <https://onlinelibrary.wiley.com/doi/10.1111/brv.12333>.

Picard M, Juster R-P, McEwen BS. 2014. Mitochondrial allostatic load puts the ’gluc’ back in glucocorticoids. Nature Reviews Endocrinology. 10(5):303–310. doi:[10.1038/nrendo.2014.22](https://doi.org/10.1038/nrendo.2014.22). [accessed 2023 Jun 21]. <https://www.nature.com/articles/nrendo.2014.22>.

Putman BJ, Drury JP, Blumstein DT, Pauly GB. 2017. Fear no colors? Observer clothing color influences lizard escape behavior. PLoS One. 12(8):e0182146.

Qualls FJ, Shine R. 2000. Post-hatching environment contributes greatly to phenotypic variation between two populations of the australian garden skink, lampropholis guichenoti. Biological Journal of the Linnean Society. 71(2):315–341.

R Core Team. 2021. R: A language and environment for statistical computing. Vienna, Austria: R Foundation for Statistical Computing.

Rubalcaba JG, Jimeno B. 2022. Body temperature and activity patterns modulate glucocorticoid levels across lizard species: A macrophysiological approach. Frontiers in Ecology and Evolution. 10:1032083.

Sakata JT, Coomber P, Gonzalez-Lima F, Crews D. 2000. Functional connectivity among limbic brain areas: Differential effects of incubation temperature and gonadal sex in the leopard gecko, eublepharis macularius. Brain, Behavior and Evolution.:139–151.

Sapolsky RM, Romero LM, Munck AU. 2000. How Do Glucocorticoids Influence Stress Responses? Integrating Permissive, Suppressive, Stimulatory, and Preparative Actions. 21(1).

Shettleworth SJ. 2010. Cognition, evolution, and behaviour. 2nd ed. Shettleworth, editor. Oxford University Press.

Szuran T, Zimmermann E, Welzl H. 1994. Water maze performance and hippocampal weight of prenatally stressed rats. Behavioural Brain Research. 65(2):153–155. doi:[10.1016/0166-4328(94)90100-7](https://doi.org/10.1016/0166-4328(94)90100-7). [accessed 2023 Jun 21]. <https://linkinghub.elsevier.com/retrieve/pii/0166432894901007>.

Torr GA, Shine R. 1996. Patterns of dominance in the small scincid lizard lampropholis guichenoti. Journal of Herpetology.:230–237.

Toure MW, Reader SM. 2022. Colour biases in learned foraging preferences in trinidadian guppies. Ethology. 128(1):49–60.

Vila Pouca C, Gervais C, Reed J, Michard J, Brown C. 2019. Quantity discrimination in Port Jackson sharks incubated under elevated temperatures. Behavioral Ecology and Sociobiology. 73(7):93. doi:[10.1007/s00265-019-2706-8](https://doi.org/10.1007/s00265-019-2706-8). [accessed 2023 Jun 21]. <http://link.springer.com/10.1007/s00265-019-2706-8>.

Vinogradov I, Zang C, Mahmud-Al-Hasan M, Head M, Jennions M. 2024. Inbreeding and high developmental temperatures affect cognition and boldness in guppies (poecilia reticulata). Proceedings of the Royal Society B. 291(2031):20240785.

Ward-Fear G, Pearson D, Brown G, Rangers B, Shine R. 2016. Ecological immunization: In situ training of free-ranging predatory lizards reduces their vulnerability to invasive toxic prey. Biology letters. 12(1):20150863.

Welklin JF, Sonnenberg BR, Branch CL, Heinen VK, Pitera AM, Benedict LM, Whitenack LE, Bridge ES, Pravosudov VV. 2024. Spatial cognitive ability is associated with longevity in food-caching chickadees. Science. 385(6713):1111–1115.

Zhang RY, Wild KH, Pottier P, Carrasco MI, Nakagawa S, Noble DW. 2023. Developmental environments do not affect thermal physiological traits in reptiles: An experimental test and meta-analysis. Biology Letters. 19(5):20230019.

Zhu Z, Li X, Chen W, Zhao Y, Li H, Qing C, Jia N, Bai Z, Liu J. 2004. Prenatal stress causes gender-dependent neuronal loss and oxidative stress in rat hippocampus. Journal of Neuroscience Research. 78(6):837–844. doi:[10.1002/jnr.20338](https://doi.org/10.1002/jnr.20338). [accessed 2023 May 25]. <https://onlinelibrary.wiley.com/doi/10.1002/jnr.20338>.

Fig 1— Experimental design of the study. Panel A shows the early environment manipulation. In panel B, the measurements of the 3D-printed ramps that were employed in the habituation and learning tasks. Panel C shows the habituation process in three different stages. In panel D, the associative task is done with the three different 3D-printed ramps. White lids in D show the ramps where the food reward was not attainable.

Fig 2— Results for *Lampropholis delicata* for both color groups red (A, B, C) and blue (D, E, F). Panels A and D show the predicted probability of choosing the correct ramp in the first trial (Decision first trial). In panels B and E, the distribution of the estimates of slopes per each treatment. In all A, B, D, and E the x-axis represents the slope estimate, and in the y-axis are the density of the estimates. The different colors indicate the different treatments. Points and bars represent the mean and standard deviation of the mean of the estimates, respectively. Dashed lines indicate value 0.33 (the probability of choosing correctly by chance) in panels A, D, and 0 in panels B, E. Asterisks indicate significant results (pmcmc < 0.05). Panels C and F show the predicted probability of choosing the correct ramp first over trials. The lines represent the mean predicted probability of choosing the correct ramp for each trial, and the shaded areas indicate the standard deviation of the mean; both were obtained using the slope and intercept estimates from the posterior distributions. The different colors indicate the different treatments.

Fig 3— Results for *Lampropholis guichenoti* for both color groups red (A, B, C) and blue (D, E, F). Panels A and D show the predicted probability of choosing the correct ramp in the first trial (Decision first trial). In panels B and E, the distribution of the estimates of slopes per each treatment. In all A, B, D, and E the x-axis represents the slope estimate, and in the y-axis are the density of the estimates. The different colors indicate the different treatments. Points and bars represent the mean and standard deviation of the mean of the estimates, respectively. Dashed lines indicate value 0.33 (the probability of choosing correctly by chance) in panels A, D, and 0 in panels B, E. Asterisks indicate significant results (pmcmc < 0.05). Panels C and F show the predicted probability of choosing the correct ramp first over trials. The lines represent the mean predicted probability of choosing the correct ramp for each trial, and the shaded areas indicate the standard deviation of the mean; both were obtained using the slope and intercept estimates from the posterior distributions. The different colors indicate the different treatments.

# Suplementary Material

#### Estimates of learning rates for all the different treatments per each task, species, and group.

Table 1. Estimates of learning rates for all the different treatments per each task, species, and group. Mean shows the arithmetic means of the estimates obtained from the posteriors of the model, and 95% CI indicates the 95% confidence interval of the mean. All pmcmc tested the hypothesis that the mean equals zero. In bold, those values that are significant (pmcmc <0.05).

|  | | Red | | | Blue | | |
| --- | --- | --- | --- | --- | --- | --- | --- |
| Species | Treatment | Mean | 95% CI | pmcmc | Mean | 95% CI | pmcmc |
| *L. delicata* | CORT-Cold | **0.101** | **[0.055 , 0.152]** | **< 0.001** | 0.013 | [-0.035 , 0.064] | 0.60 |
|  | Control-Cold | **0.056** | **[0.018 , 0.096]** | **< 0.05** | -0.007 | [-0.056 , 0.042] | 0.77 |
|  | CORT-Hot | **0.066** | **[0.023 , 0.110]** | **< 0.05** | 0.032 | [-0.013 , 0.083] | 0.17 |
|  | Control-Hot | **0.082** | **[0.037 , 0.131]** | **< 0.001** | **0.053** | **[0.002 , 0.107]** | **< 0.05** |
| *L. guichenoti* | CORT-Cold | **0.119** | **[0.054 , 0.190]** | **< 0.05** | **0.070** | **[0.012 , 0.129]** | **< 0.05** |
|  | Control-Cold | **0.097** | **[0.032 , 0.171]** | **< 0.05** | 0.015 | [-0.058 , 0.091] | 0.65 |
|  | CORT-Hot | **0.074** | **[0.014 , 0.139]** | **< 0.05** | **0.102** | **[0.036 , 0.177]** | **< 0.05** |
|  | Control-Hot | **0.078** | **[0.016 , 0.145]** | **< 0.05** | **0.064** | **[0.004 , 0.125]** | **< 0.05** |

#### Color preference

To test if lizards were biased towards the assigned color as our preliminary analyses suggested, we employed the values from our posterior distributions. We estimated the predicted probability of choosing the correct ramp first in the first trial (see Statistical analyses) and tested the hypothesis that this estimated probability was higher than expected by chance (i.e. 0.33) using pmcmc. If the estimated probability is above 0.33 we consider it as an indication that there was a preference towards that color that could be affecting learning rates.

Table 2. Probability of choosing the correct ramp in the first trial when the correct ramp was blue (Prob Blue) or red (Prob Red) for each species and each treatment. pmcmc tested the hypothesis that the probability is >0.33. In bold, those values that are significant (pmcmc <0.05)

| Species | Treatment | Prob Red | pmcmc Red | Prob Blue | pmcmc Blue |
| --- | --- | --- | --- | --- | --- |
| *L. delicata* | CORT-Cold | 0.153 | 0.97 | **0.636** | **< 0.05** |
|  | Control-Cold | 0.304 | 0.63 | **0.752** | **< 0.001** |
|  | CORT-Hot | 0.353 | 0.45 | **0.561** | **< 0.05** |
|  | Control-Hot | 0.577 | 0.13 | **0.681** | **< 0.05** |
| *L. guichenoti* | CORT-Cold | 0.079 | 1.00 | 0.369 | 0.41 |
|  | Control-Cold | 0.204 | 0.89 | **0.618** | **< 0.05** |
|  | CORT-Hot | 0.246 | 0.81 | 0.506 | 0.09 |
|  | Control-Hot | 0.496 | 0.26 | **0.718** | **< 0.05** |

On average, we found that, for both species, the proportion of correct choices in the first trial was significantly above chance when the correct ramp was blue for *L. delicata* (mean Prob choice = 0.657, pmcmc < 0.05) but not for *L. guichenoti* (mean Prob choice = 0.553, pmcmc = 0.14). When the correct choice was red, it was not significant for neither species (*L. delicata*: mean Prob choice = 0.347, pmcmc = 0.55; *L. guichenoti*: mean Prob choice = 0.347, pmcmc = 0.55).

#### Light spectrum

To test if the bias towards blue was something acquired during training, we compared the light spectrum of the ramps used in the associative task and the white ones used during habituation. We took three measurements of ten ramps per color with a spectophotometer, and then analyzed the spectrum and the perceived differences in color using the package pavo. The spetrum of each type of ramp are shown in [Fig. 4](#fig-spectrum); the perceived chromatic constrasts between ramps are shown in [Fig. 5](#fig-perceived1) and [Fig. 6](#fig-perceived2).

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| Fig 4— Light spectrum of the ramps used in the associative task and the white ones used during habituation. The different colors represent the different ramps. |

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| A graph of different colors  Description automatically generated  Fig 5— Perceived chromatic contrasts between ramps. |

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| A diagram of different colors and sizes  Description automatically generated  Fig 6— Perceived achromatic contrasts between feeders. |

#### Checking the models plots

Model formula for task is:  
Choice ~ Trial \* cort \* temp + (1 + Trial|lizard\_id)  
Plots for the different models of the associative task:  
1.- *L. delicata*  
1.a.- Red

Estimate Est.Error Q2.5 Q97.5  
R2 0.1695644 0.02196313 0.1255778 0.2110525

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1.b.- Blue

Estimate Est.Error Q2.5 Q97.5  
R2 0.07757601 0.02088415 0.03926128 0.1200952

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|  |

2.- *L. guichenoti*  
2.a.- Red

Estimate Est.Error Q2.5 Q97.5  
R2 0.1858621 0.02466494 0.1359576 0.2330882

|  |
| --- |
|  |
|  |
|  |

2.b.- Blue

Estimate Est.Error Q2.5 Q97.5  
R2 0.1367205 0.02551682 0.08796974 0.1865906

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#### Models with age included

Table 3. Results for *L. delicata* assigned to red ramps when the model included the age:

| Predictors | Estimate | Est.Error | l-95% CI | u-95% CI | Rhat | Bulk\_ESS | Tail\_ESS |
| --- | --- | --- | --- | --- | --- | --- | --- |
| Intercept | -2.16 | 0.66 | -3.49 | -0.89 | 1.00 | 3402.24 | 3887.20 |
| age.start | 0.00 | 0.02 | -0.03 | 0.04 | 1.00 | 5418.98 | 5776.45 |
| trial\_associative | 0.10 | 0.02 | 0.06 | 0.15 | 1.00 | 3692.82 | 4207.48 |
| cortControl | 1.09 | 0.86 | -0.60 | 2.83 | 1.00 | 3201.98 | 3890.41 |
| tempHot | 1.28 | 0.93 | -0.47 | 3.15 | 1.00 | 2934.34 | 3471.54 |
| trial\_associative:cortControl | -0.05 | 0.03 | -0.11 | 0.01 | 1.00 | 3572.26 | 4634.15 |
| trial\_associative:tempHot | -0.04 | 0.03 | -0.10 | 0.03 | 1.00 | 3574.66 | 4356.48 |
| cortControl:tempHot | -1.26 | 1.21 | -3.73 | 1.08 | 1.00 | 3074.88 | 4021.52 |
| trial\_associative:cortControl:tempHot | 0.06 | 0.04 | -0.03 | 0.15 | 1.00 | 3628.87 | 4183.50 |

Table 4. Results for *L. delicata* assigned to blue ramps when the model included the age:

| Predictors | Estimate | Est.Error | l-95% CI | u-95% CI | Rhat | Bulk\_ESS | Tail\_ESS |
| --- | --- | --- | --- | --- | --- | --- | --- |
| Intercept | 0.50 | 0.58 | -0.64 | 1.64 | 1.00 | 3044.27 | 4311.87 |
| age.start | -0.00 | 0.01 | -0.01 | 0.01 | 1.00 | 5860.40 | 5710.61 |
| trial\_associative | 0.01 | 0.02 | -0.04 | 0.06 | 1.00 | 3068.62 | 4168.88 |
| cortControl | 0.70 | 0.82 | -0.88 | 2.34 | 1.00 | 2870.11 | 3815.82 |
| tempHot | -0.34 | 0.79 | -1.93 | 1.22 | 1.00 | 2598.17 | 3477.53 |
| trial\_associative:cortControl | -0.02 | 0.04 | -0.09 | 0.05 | 1.00 | 2915.98 | 3893.50 |
| trial\_associative:tempHot | 0.02 | 0.03 | -0.05 | 0.09 | 1.00 | 2593.93 | 3764.25 |
| cortControl:tempHot | -0.98 | 1.13 | -3.23 | 1.25 | 1.00 | 2537.70 | 3924.03 |
| trial\_associative:cortControl:tempHot | 0.04 | 0.05 | -0.06 | 0.14 | 1.00 | 2642.40 | 4091.36 |

Table 5. Results for *L. guichenoti* assigned to red ramps when the model included the age:

| Predictors | Estimate | Est.Error | l-95% CI | u-95% CI | Rhat | Bulk\_ESS | Tail\_ESS |
| --- | --- | --- | --- | --- | --- | --- | --- |
| Intercept | -3.06 | 0.84 | -4.87 | -1.55 | 1.00 | 2641.91 | 2963.70 |
| age.start | -0.00 | 0.02 | -0.04 | 0.03 | 1.00 | 5914.10 | 5309.44 |
| trial\_associative | 0.12 | 0.04 | 0.06 | 0.20 | 1.00 | 2411.12 | 2897.00 |
| cortControl | 1.28 | 1.15 | -0.97 | 3.62 | 1.00 | 2479.35 | 3043.41 |
| tempHot | 1.65 | 1.14 | -0.57 | 3.97 | 1.00 | 2336.65 | 2802.60 |
| trial\_associative:cortControl | -0.02 | 0.05 | -0.13 | 0.08 | 1.00 | 2444.48 | 3222.97 |
| trial\_associative:tempHot | -0.05 | 0.05 | -0.15 | 0.04 | 1.00 | 2253.20 | 2466.35 |
| cortControl:tempHot | -1.03 | 1.53 | -4.17 | 1.91 | 1.00 | 2353.37 | 3274.21 |
| trial\_associative:cortControl:tempHot | 0.03 | 0.07 | -0.11 | 0.17 | 1.00 | 2319.80 | 3341.04 |

Table 6. Results for *L. guichenoti* assigned to blue ramps when the model included the age:

| Predictors | Estimate | Est.Error | l-95% CI | u-95% CI | Rhat | Bulk\_ESS | Tail\_ESS |
| --- | --- | --- | --- | --- | --- | --- | --- |
| Intercept | -0.61 | 0.73 | -2.07 | 0.85 | 1.00 | 4476.16 | 4928.86 |
| age.start | 0.01 | 0.01 | -0.02 | 0.03 | 1.00 | 5957.56 | 5181.70 |
| trial\_associative | 0.07 | 0.03 | 0.01 | 0.13 | 1.00 | 4004.25 | 4029.03 |
| cortControl | 1.32 | 1.00 | -0.70 | 3.29 | 1.00 | 3729.95 | 4199.07 |
| tempHot | 0.38 | 0.94 | -1.61 | 2.22 | 1.00 | 4055.41 | 4399.38 |
| trial\_associative:cortControl | -0.05 | 0.05 | -0.14 | 0.04 | 1.00 | 3437.88 | 3686.90 |
| trial\_associative:tempHot | 0.03 | 0.04 | -0.05 | 0.12 | 1.00 | 3667.39 | 4011.10 |
| cortControl:tempHot | -1.23 | 1.34 | -3.86 | 1.52 | 1.00 | 3524.34 | 4053.85 |
| trial\_associative:cortControl:tempHot | 0.02 | 0.06 | -0.12 | 0.14 | 1.00 | 3340.33 | 4029.47 |

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| Fig 7— Distribution of the age of the lizards by treatment and species |