An introduction to Bayesian modelling with R, JAGS and STAN

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Bayesian modelling is growing fast

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IDEAS AND PERSPECTIVES

Why environmental scientists are becoming Bayesians

- Powerful
- Flexible
- Knowledge synthesis
- Uncertainty

Even if you don't become Bayesian today, this workshop will help you to understand regression and 'mixed-effects' models better.

Practical workshop: we'll see main concepts, but do read the literature!

General

- Data analysis using regression and multilevel/hierarchical models
- Statistical Rethinking: A Bayesian Course with Examples in R and Stan
- Bayesian data analysis
- The BUGS book
- Doing Bayesian data analysis: a tutorial with R, JAGS, and STAN
- Bayesian linear mixed models using Stan: a tutorial
- Bayesian basics
- Philosophy and the practice of Bayesian statistics

Practical workshop: we'll see main concepts, but do read the literature!

Ecology-oriented

- The ecological detective: confronting models with data
- Bayesian methods for ecology
- Models for ecological data
- Introduction to WinBUGS for ecologists
- Applied hierarchical modeling in ecology
- Bayesian Models: A Statistical Primer for Ecologists
- and more...

Bayesian modelling software

- WinBUGS/OpenBUGS
- JAGS
- STAN
- Filzbach
- Nimble
- Many R packages: MCMCpack, MCMCglmm, LaplacesDemon, r-inla, etc (see Bayesian task view)

We'll focus on JAGS and STAN

- Fast, powerful, and most popular
- Similar to BUGS
- Easy to start
- Open-ended modelling: deal with complex models too
- But look for specific implementations of your analysis (e.g. hSDM, rstanarm)
- Once concepts understood, switching software not difficult

JAGS has to be installed independently

```
http://mcmc-jags.sourceforge.net/
Use latest version (4.2.0)
```

Calling JAGS from R

- rjags
- R2jags
- runjags
- jagsUI
- dclone
- rube

Calling STAN from R

- rstan
- rstanarm
- rethinking
- brms

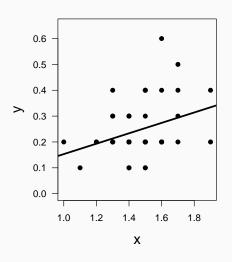
We will also need these R packages:

```
library(arm)
library(R2jags)
library(ggmcmc)
library(shinystan)
library(rube)
library(lme4)
library(rstan)
```

and their dependencies

The very basics: linear regression

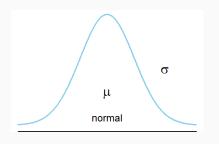
The very basics: linear regression



$$y_i = a + bx_i + \epsilon_i$$

How many parameters?

The very basics: linear regression



$$y_i = a + bx_i + \epsilon_i$$

 $\epsilon_i \sim N(0, \sigma^2)$

Or also

$$y_i \sim N(\mu_i, \sigma^2)$$

 $\mu_i = a + bx_i$

Our dataset: tree heights and DBH

http://tinyurl.com/treesdata

- One species
- 10 plots
- 1000 trees
- Number of trees per plot ranging from 4 to 392

```
trees <- read.csv("trees.csv")
summary(trees[, 1:3])</pre>
```

```
plot dbh height

Min.: 1.0 Min.: 5.06 Min.: 13.40

1st Qu.: 1.0 1st Qu.:17.69 1st Qu.:29.68

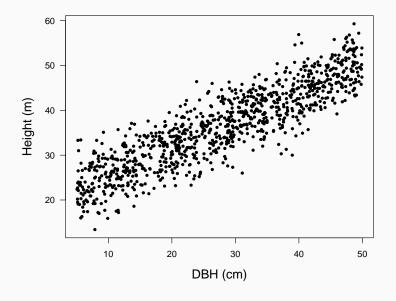
Median: 2.0 Median: 28.62 Median: 36.55

Mean: 2.7 Mean: 27.88 Mean: 36.51

3rd Qu.: 4.0 3rd Qu.:38.97 3rd Qu.:43.33

Max.: 10.0 Max.: 49.92 Max.: 59.30
```

What's the relationship between DBH and height?



First step: linear regression (lm)

Interpretation?

Always centre continuous variables

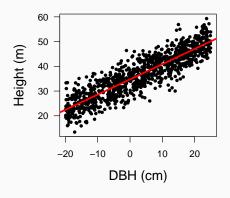
summary(trees\$dbh)

```
Min. 1st Qu. Median Mean 3rd Qu. Max. 5.06 17.69 28.62 27.88 38.96 49.92
```

```
trees$dbh.c <- trees$dbh - 25
```

So, all parameters will be referred to a 25 cm DBH tree.

Linear regression with centred DBH



Let's make it Bayesian

Things we'll need

- A function describing the model (including **priors**)
- Data
- Choose parameters to save
- Define initial values for MCMC chains
- Decide number of iterations (and burnin)

Remember our model structure

$$y_i \sim N(\mu_i, \sigma^2)$$

 $\mu_i = \alpha + \beta x_i$

In this case:

Height_i
$$\sim N(\mu_i, \sigma^2)$$

 $\mu_i = \alpha + \beta DBH_i$

 α : expected height when DBH = 0

 β : how much height increases with every unit increase of DBH

JAGS uses precision instead of variance

$$\tau = \frac{1}{\sigma^2}$$

So, residual variance $\sigma^2=100$ expressed as $\tau=0.01$.

Specify the model as an R function

```
Height<sub>i</sub> \sim N(\mu_i, \sigma^2)

\mu_i = \alpha + \beta DBH_i
```

```
bayes.lm <- function(){

# LIKELIHOOD
for (i in 1:length(height)){
  height[i] ~ dnorm(mu[i], tau) # tau=precision (inverse var)
  mu[i] <- alpha + beta*dbhc[i] # expected height ~ dbhc
}</pre>
```

We need priors for every parameter!

```
bayes.lm <- function(){

# LIKELIHOOD
for (i in 1:length(height)){
   height[i] ~ dnorm(mu[i], tau)  # tau = precision (inverse var)
   mu[i] <- alpha + beta*dbhc[i]  # expected height ~ dbhc
}

# PRIORS (vague or weakly informative)
alpha ~ dunif(1, 100)  # prior avg height of 25-cm-DBH tree
beta ~ dunif(0, 10)  # how much do we expect height to scale with DBH?
tau <- 1/(sigma*sigma)  # tau = 1/sigma*2
sigma ~ dunif(0, 50)  # residual standard deviation
}</pre>
```

A note on priors

Avoid 'non-informative' priors (see this and this)
Use weakly informative (e.g. bounded Uniform, Normal with reasonable parameters, Cauchy...)
or strongly informative priors based on previous knowledge and common sense.
Some tips for setting priors:

- https:
 //github.com/stan-dev/stan/wiki/Prior-Choice-Recommendations
- http://www.mrc-bsu.cam.ac.uk/wp-content/uploads/bugsbook_ chapter5.pdf

Usually good idea to try different priors and evaluate posterior sensitivity Or run model without likelihood (priors only).

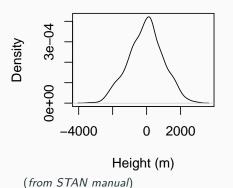
Example: estimating people height across countries

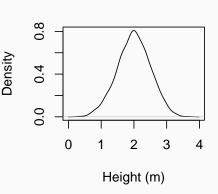
Unreasonable prior

main="", xlab="Height (m)")

Reasonable prior

```
plot(density(rnorm(1000, 0, 1000)), plot(density(rnorm(1000, 2, 0.5)),
                                          main="", xlab="Height (m)")
```





We already have our model definition

```
bayes.lm <- function(){</pre>
  # LIKELIHOOD
  for (i in 1:length(height)){
   height[i] ~ dnorm(mu[i], tau)
                                    # tau = precision (inverse of vari
   mu[i] <- alpha + beta*dbhc[i]</pre>
                                    # centred diameter
  # PRIORS (vaque or weakly informative)
  alpha ~ dunif(1, 100) # prior for average height of a 25-cm-DBH t
  beta ~ dunif(0, 10) # how much do we expect height to scale wit
 tau <- 1/(sigma*sigma) # tau = 1/sigma^2
  sigma ~ dunif(0, 50) # residual standard deviation
```

Next step: create list with data

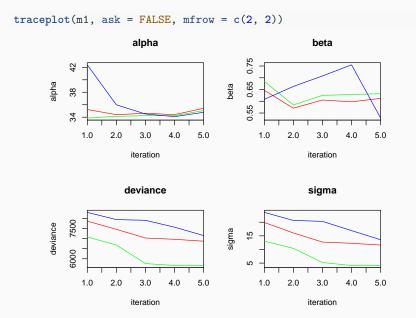
```
Data = known values
```

Next step: choose parameters to save

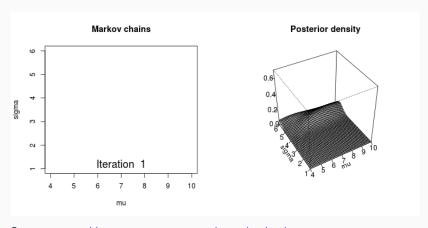
```
params <- c("alpha", "beta", "sigma")</pre>
```

Traceplots: viewing MCMC in action

Obviously we haven't achieved convergence yet



Viewing MCMC in action (II)

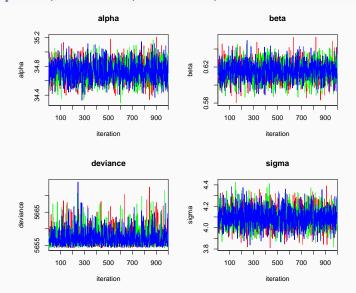


Source: http://mbjoseph.github.io/2013/09/08/metropolis.html

Let's run JAGS for longer

Use traceplots to asess convergence

traceplot(m1, ask = FALSE, mfrow = c(2, 2))



Results: parameter estimates

Inference for Bugs model at "C:/Users/FRS/AppData/Local/Temp/RtmpgxpkpY
3 chains, each with 10000 iterations (first 5000 discarded), n.thin =
n.sims = 3000 iterations saved

	mu.vect	sd.vect	2.5%	97.5%	Rhat	n.eff
alpha	34.735	0.135	34.473	34.998	1.001	2100
beta	0.616	0.010	0.596	0.635	1.001	3000
sigma	4.098	0.091	3.926	4.283	1.001	2000
deviance	5657.283	2.514	5654.454	5663.867	1.002	1400

For each parameter, n.eff is a crude measure of effective sample size, and Rhat is the potential scale reduction factor (at convergence, Rhat=

```
DIC info (using the rule, pD = var(deviance)/2)
pD = 3.2 and DIC = 5660.4
```

DIC is an estimate of expected predictive error (lower deviance is bett

Compare with simple.lm

Results pretty similar, because of vague priors

A plot of the whole model

Bugs model at "C:/Users/FRS/AppData/Local/Temp/RtmpgxpkpY/model1cf45e321f74.txt", fit using jags, 3 chains, each with 10000 iterations (first 5000 discarded)



medians and 80% intervals



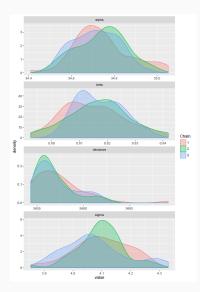


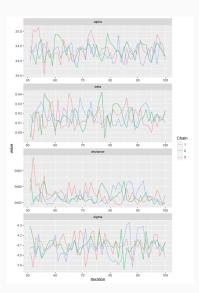
Model checking

Using ggmcmc to produce diagnostic plots

```
suppressPackageStartupMessages(library(ggmcmc))
m1.mcmc <- as.mcmc(m1) # Get list of MCMC values
m1.tidy = ggs(m1.mcmc) # Produce tidy data frame
ggmcmc(m1.tidy)
Plotting histograms
Plotting density plots
Plotting traceplots
Plotting running means
Plotting comparison of partial and full chain
Plotting autocorrelation plots
Plotting crosscorrelation plot
Plotting Potential Scale Reduction Factors
Plotting Geweke Diagnostic
Plotting caterpillar plot
```

ggmcmc output sample





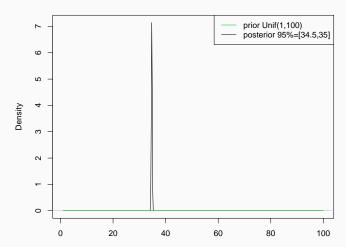
shinystan: assess model interactively

```
library(shinystan)
launch_shinystan(as.shinystan(m1.mcmc))
```

Comparing prior and posterior

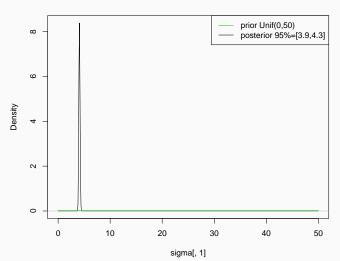
Comparing prior and posterior

Prior / Posterior Plot

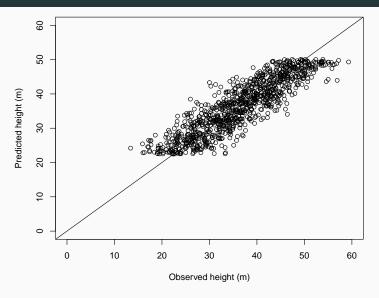


Comparing prior and posterior (sigma)

Prior / Posterior Plot

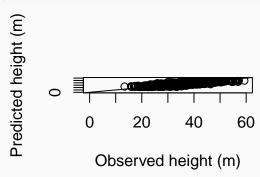


Observed vs Predicted values



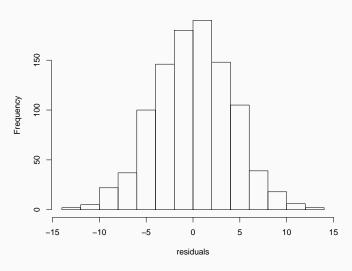
How would you do this?

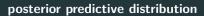
Observed vs Predicted values



Residuals

Histogram of residuals





Now using Normal vague priors

```
bayes.lm.N <- function(){</pre>
  # LIKELIHOOD
  for (i in 1:length(height)){
   height[i] ~ dnorm(mu[i], tau)
                                    # tau = precision (inverse of vari
   mu[i] <- alpha + beta*dbhc[i]</pre>
                                    # centred diameter
  # PRITORS
  alpha ~ dnorm(0, 0.01) # prior for intercept
  beta ~ dnorm(0, 0.01)
                             # prior for beta (slope)
  tau <- 1/(sigma*sigma) # tau = 1/sigma^2
  sigma ~ dunif(0, 50)
                             # residual standard deviation
```

Calling JAGS

Results with Normal priors on intercept and slope

Inference for Bugs model at "C:/Users/FRS/AppData/Local/Temp/RtmpgxpkpY 3 chains, each with 4000 iterations (first 2000 discarded), n.thin = 2 n.sims = 3000 iterations saved

	mu.vect	sd.vect	2.5%	97.5%	Rhat	n.eff
alpha	34.724	0.136	34.463	34.991	1.001	3000
beta	0.616	0.010	0.595	0.635	1.001	3000
sigma	4.095	0.091	3.922	4.274	1.003	720
deviance	5657.288	2.471	5654.452	5663.709	1.001	3000

For each parameter, n.eff is a crude measure of effective sample size, and Rhat is the potential scale reduction factor (at convergence, Rhat=

```
DIC info (using the rule, pD = var(deviance)/2)
pD = 3.1 and DIC = 5660.3
```

DIC is an estimate of expected predictive error (lower deviance is bett

Very similar

Let's try strongly informative prior on alpha

```
bayes.lm.N2 <- function(){</pre>
  # LIKELIHOOD
  for (i in 1:length(height)){
   height[i] ~ dnorm(mu[i], tau)
                                    # tau = precision (inverse of vari
   mu[i] <- alpha + beta*dbhc[i]</pre>
                                    # centred diameter
  # PRITORS
  alpha ~ dnorm(10, 10) # prior for intercept
  beta ~ dnorm(0, 0.01)
                            # prior for beta (slope)
  tau <- 1/(sigma*sigma)
                            # tau = 1/sigma^2
  sigma ~ dunif(0, 50)
                            # residual standard deviation
```

Calling JAGS

Posteriors: between prior and likelihood

For each parameter, n.eff is a crude measure of effective sample size, and Rhat is the potential scale reduction factor (at convergence, Rhat=

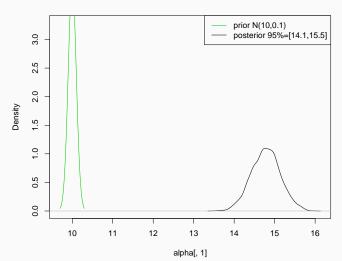
DIC info (using the rule, pD = var(deviance)/2) pD = 593.0 and DIC = 9412.5

sigma 19.919 0.560 18.860 21.035 1.001 3000 deviance 8819.536 34.430 8751.074 8886.390 1.001 3000

DIC is an estimate of expected predictive error (lower deviance is bett

Prior vs Posterior

Prior / Posterior Plot



Bayesian inference

Bayes theorem: prior, likelihood, posterior

$$P(Hypothesis|Data) = \frac{P(Data|Hypothesis)P(Hypothesis)}{P(Data)}$$

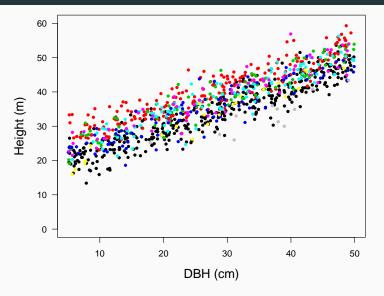
$$P(\textit{Hypothesis}|\textit{Data}) \propto P(\textit{Data}|\textit{Hypothesis}) \times P(\textit{Hypothesis})$$

Posterior \propto Likelihood \times Prior

Hence, prior influence decreases with size of data set.

Varying-intercept models

Accounting for plot effects

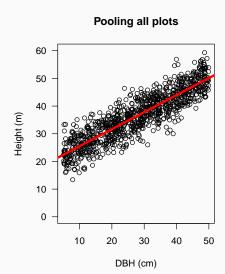


Do it yourself using Im

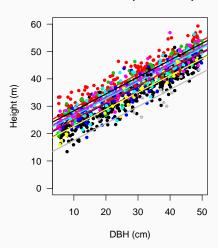
```
lm.plot <- lm(height ~ factor(plot) + dbh.c, data = trees)</pre>
lm(formula = height ~ factor(plot) + dbh.c, data = trees)
             coef.est coef.se
(Intercept) 32.13
                      0.16
factor(plot)2 6.50
                      0.26
factor(plot)3 4.36
                      0.35
factor(plot)4 1.93
                      0.36
factor(plot)5 3.64
                      0.34
factor(plot)6 4.20
                      0.42
factor(plot)7 -0.18
                      0.67
factor(plot)8 -5.31
                      0.89
factor(plot)9 5.44
                      1.09
factor(plot)10 2.26
                      1.37
dbh.c
             0.62
                      0.01
n = 1000, k = 11
residual sd = 3.04, R-Squared = 0.88
```

Interpretation?

Single vs varying intercept



Different intercept for each plot



Let's make it Bayesian

Things we'll need

- A function describing the model (including **priors**)
- Data
- Choose parameters to save
- Define initial values for MCMC chains
- Decide number of iterations (and burnin)

Bayesian varying-intercept model with no pooling

```
varint.nopool <- function(){</pre>
  # I.TKFI.THOOD
  for (i in 1:length(height)){
   height[i] ~ dnorm(mu[i], tau) # tau = precision (inverse of vari
   mu[i] <- alpha[plot[i]] + beta*dbhc[i] # centred diameter</pre>
  # PRIORS
  #alpha ~ dnorm(0, .001) # previous model
  for (j in 1:10){
    alpha[j] ~ dnorm(0, .001) # Plot effects drawn from Normal distrib
                               # with large **fixed** variance
  beta ~ dnorm(0, .001)
 tau \leftarrow 1/(sigma*sigma) # tau = 1/sigma^2
  sigma ~ dunif(0, 50)
```

This fits same model as lm.plot

Running JAGS in parallel (multiple cores)

Results of Bayesian varying intercept model without pooling

Inference for Bugs model at "varint.nopool", fit using jags,
3 chains, each with 4000 iterations (first 2000 discarded), n.thin = 2
n.sims = 3000 iterations saved

2.5% 97.5% Rhat n.eff mu.vect sd.vect alpha[1] 32.126 31.821 32.427 1.001 3000 0.154 alpha[2] 38.631 0.202 38.228 39.033 1.002 3000 alpha[3] 36.487 0.328 1700 35.830 37.109 1.002 alpha[4] 34.050 0.322 33.397 34.663 1.001 2200 alpha[5] 35.754 0.309 35.149 36.346 1.002 1400 alpha[6] 36.339 0.389 35.550 37.094 1.002 1100 alpha[7] 31.934 0.632 30.746 33.168 1.001 3000 alpha[8] 26.810 0.870 25.174 28.493 1.001 3000 alpha[9] 37.519 1.089 35.460 39.709 1.001 3000 alpha[10] 34.341 1.367 31.708 36.983 1.001 3000 beta 0.617 0.008 0.603 0.631 1.003 950 3.047 0.067 2.919 3.183 1.003 680 sigma deviance 5064.489 4.777 5056.981 5075.523 1.002 1000

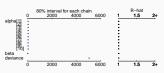
For each parameter, n.eff is a crude measure of effective sample size, 67 ,

Same results as lm.plot

```
lm(formula = height ~ factor(plot) + dbh.c, data = trees)
             coef.est coef.se
(Intercept)
             32.13
                      0.16
factor(plot)2 6.50
                      0.26
factor(plot)3 4.36
                      0.35
factor(plot)4 1.93
                      0.36
factor(plot)5 3.64
                      0.34
factor(plot)6 4.20
                      0.42
factor(plot)7 -0.18
                      0.67
factor(plot)8 -5.31
                      0.89
factor(plot)9 5.44
                      1.09
factor(plot)10 2.26
                      1.37
dbh.c
              0.62
                      0.01
n = 1000, k = 11
residual sd = 3.04, R-Squared = 0.88
```

Plot whole model

Bugs model at "varint.nopool", fit using jags, 3 chains, each with 4000 iterations (first 2000 discarded)











The varying-intercept model seems better

Based on Deviance Information Criterion (DIC)

DIC(m1) = 5660

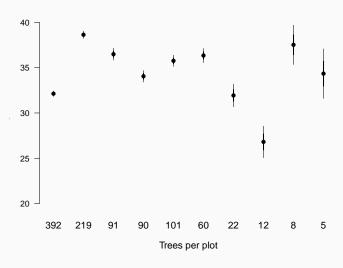
DIC(m2) = 5076

But note DIC (as AIC and BIC) have been criticised

WAIC and cross-validation are preferred alternatives.

Estimation of plot effects improves with sample size

Average height of a 25-cm DBH tree in each plot



Varying-intercepts with pooling (shrinkage)

Multilevel model with varying intercepts

$$y_i = a_j + bx_i + \varepsilon_i$$

 $a_j \sim N(0, \tau^2)$
 $\varepsilon_i \sim N(0, \sigma^2)$

In our example:

Height_i = plot_j + bDBH_i +
$$\varepsilon_i$$

plot_j ~ $N(0, \tau^2)$
 $\varepsilon_i \sim N(0, \sigma^2)$

Fitting mixed models with Imer

```
mixed <- lmer(height ~ dbh.c + (1|plot), data = trees)
lmer(formula = height ~ dbh.c + (1 | plot), data = trees)
           coef.est coef.se
(Intercept) 34.43 1.08
dbh.c 0.62 0.01
Error terms:
 Groups Name Std.Dev.
 plot (Intercept) 3.35
 Residual 3.04
---
number of obs: 1000, groups: plot, 10
AIC = 5116.3, DIC = 5096.3
deviance = 5102.3
```

Imer coefficients

coef(mixed) \$plot (Intercept) dbh.c 32.13118 0.6169271 2 38.61479 0.6169271 3 36.46547 0.6169271 4 34.06404 0.6169271 5 35.75313 0.6169271 6 36.30517 0.6169271 32.04003 0.6169271 8 27.30620 0.6169271 9 37.27097 0.6169271 10 34.39546 0.6169271

attr(,"class")
[1] "coef.mer"

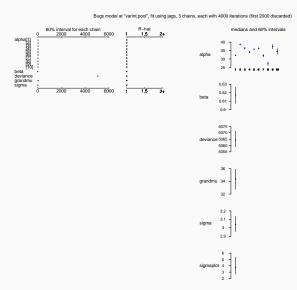
Bayesian varying-intercept model with pooling across plots

```
varint.pool <- function(){</pre>
    # I.TKEI.THOOD
 for (i in 1:length(height)){
   height[i] ~ dnorm(mu[i], tau) # tau = precision (inverse of vari
   mu[i] <- alpha[plot[i]] + beta*dbhc[i] # centred diameter</pre>
    # PRIORS
 for (j in 1:10){
    alpha[j] ~ dnorm(grandmu, tauplot) # Now we are estimating the plo
  grandmu ~ dnorm(0, .001) # Overall mean height across all plots
  tauplot <- 1/(sigmaplot*sigmaplot)</pre>
  sigmaplot ~ dunif(0, 20) # between-plot variance
  beta ~ dnorm(0, .001)
 tau <- 1/(sigma*sigma)
  sigma ~ dunif(0, 50) # residual variance
```

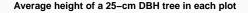
Results of multilevel model with pooling

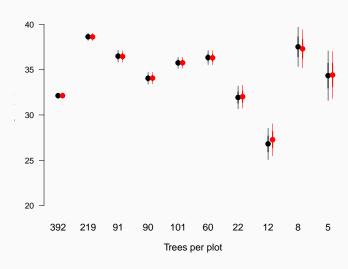
```
Inference for Bugs model at "varint.pool", fit using jags,
3 chains, each with 4000 iterations (first 2000 discarded), n.thin = 2
n.sims = 3000 iterations saved
                           2.5% 97.5% Rhat n.eff
         mu.vect sd.vect
alpha[1]
                  0.156
                          31.827
                                  32.440 1.001
                                              3000
         32.133
alpha[2] 38.616 0.205
                          38.211
                                  39.010 1.001
                                              2800
alpha[3] 36.455
                                              1400
                  0.317
                          35.839
                                  37.090 1.002
alpha[4] 34.067
                  0.318
                          33.436
                                  34.689 1.002
                                              1700
alpha[5] 35.760
                  0.303
                          35.174
                                  36.352 1.001
                                              3000
alpha[6] 36.308
                  0.389
                          35.544
                                  37.093 1.002
                                              1800
alpha[7]
          32.022
                  0.636
                          30.818
                                  33.296 1.001
                                              3000
alpha[8] 27.273
                  0.884
                          25.561
                                  28.951 1.001
                                              3000
alpha[9] 37.299
                  1.056
                          35.230
                                  39.347 1.001
                                              2700
alpha[10] 34.414
                  1.290
                          31.822
                                  36.969 1.001
                                              3000
beta
           0.617
                  0.007
                           0.602
                                   0.631 1.002
                                              1900
          34.372
                  1.280
                          31.696
                                  36.863 1.001
                                              2500
grandmu
           3.044
                  0.070
                           2.913
                                   3.184 1.001
                                              3000
sigma
sigmaplot
           3.927
                  1.162
                           2.353
                                   6.881 1.001
                                              3000
deviance
         5064.690
                  4.881 5056.896 5075.604 1.002
                                              1300
```

A plot of the whole model



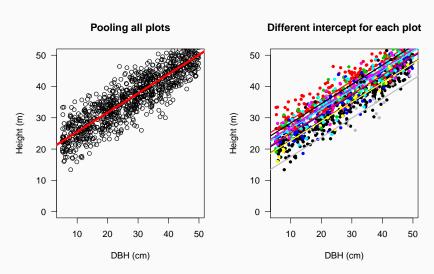
Comparing plot coefficients





A gradient from complete to no pooling

The multilevel model (with pooling) is somewhere between the complete-pooling (single intercept) and the no-pooling (one intercept for each plot, without shrinkage) models.



Growing the hierarchy: adding plot-level predictors

Model with group-level predictors

We had:

$$y_i = a_j + bx_i + \varepsilon_i$$
$$a_j \sim N(0, \tau^2)$$
$$\varepsilon_i \sim N(0, \sigma^2)$$

Now

$$y_i = a_j + bx_i + \varepsilon_i$$
 $a_j \sim N(\mu_j, \tau^2)$
 $\mu_j = \gamma + \delta \cdot predictor_j$
 $\varepsilon_i \sim N(0, \sigma^2)$

Reading plot data

```
plotdata <- read.csv("plotdata.csv")
temp.c <- plotdata$temp - 15</pre>
```

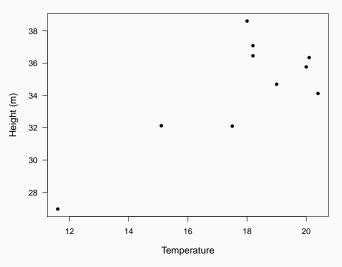
Model with group-level predictors

```
group.preds <- function(){</pre>
  # I.TKFI.THOOD
  for (i in 1:length(height)){
    height[i] ~ dnorm(mu[i], tau)
    mu[i] <- alpha[plot[i]] + beta*dbhc[i]</pre>
  # PRIORS
  for (j in 1:10){
    alpha[j] ~ dnorm(grandmu + beta.temp*tempc[j], tauplot)
  beta.temp ~ dnorm(0, .001) # slope for temperature effects
  grandmu ~ dnorm(0, .001)
  tauplot <- 1/(sigmaplot*sigmaplot)</pre>
  sigmaplot ~ dunif(0, 20)
  beta ~ dnorm(0, .001)
  tau <- 1/(sigma*sigma)
  sigma ~ dunif(0, 50)
```

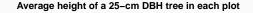
```
data <- list(height = trees$height,
             dbhc = trees$dbh.c,
             plot = trees$plot,
             tempc = temp.c)
params = c("alpha","beta","sigma", "grandmu", "sigmaplot", "beta.temp")
m4 <- jags.parallel(data,
           model.file = group.preds,
           parameters.to.save = params,
           n.chains = 3,
           inits = NULL,
           n.iter = 4000,
           n.burnin = 2000)
```

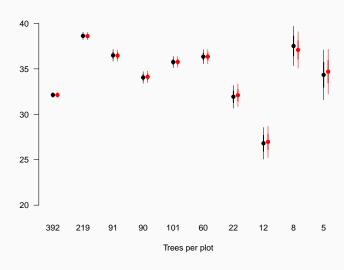
Average heights among plots related to temperature

Average height of a 25-cm DBH tree in the plot

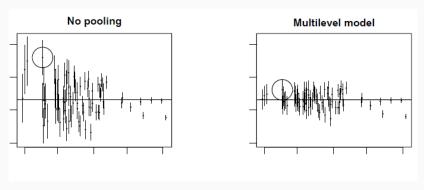


Adding plot-level predictors (pooling) may improve parameter estimation





Adding plot-level predictors (pooling) may improve parameter estimation



From Gelman & Hill p. 253

Slopes can also vary...

- and coefficients be estimated with pooling
- but taking into account correlation between slopes and intercepts
- (see e.g. Gelman & Hill 2007, ch. 13)

Recapitulating...

So what's a multilevel/hierarchical model?

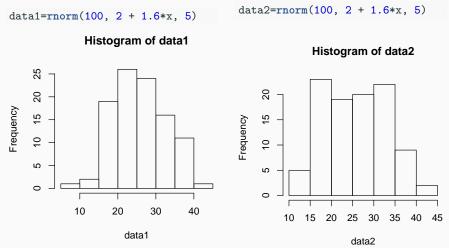
Parameters/coefficients are given a probability model (with their own hyperparameters estimated from data).

Intercepts and/or slopes may vary, and can be modelled (sometimes including their own predictors).

Advantages of hierarchical Bayes

- Perfect for structured data (space-time)
- Predictors enter at the appropriate level
- Accommodate variation in treatment effects
- More efficient inference of regression parameters
- Using all the data to perform inferences for groups with small sample size
- Predictions fully accounting for uncertainty and variability
- Prior information

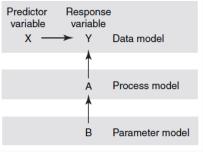
Datasets are stochastic realisations of a process



These two datasets are different, even though they arise from same process.

Data inform about process, but have 'noise' too.

Hierarchical Bayes: data, process, parameters



Clark et al. 2006, Clark 2007

```
f(\text{data,process,parameters})

\propto f(\text{data|process, parameters})

\times f(\text{process|parameters})

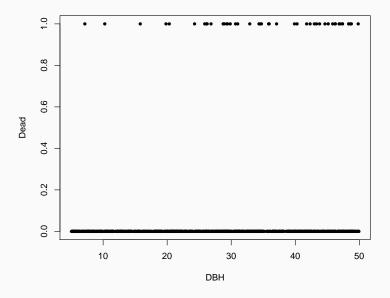
\times f(\text{parameters}).
```

Exercise

Does sex influence height?

Bayesian logistic regression

Relationship between tree size and mortality



```
logreg <- function(){</pre>
  # LIKELIHOOD
  for (i in 1:length(dead)){
    dead[i] ~ dbern(pdeath[i])
    logit(pdeath[i]) <- mu + beta*dbhc[i]</pre>
  # PRIORS
  mu ~ dnorm(0, .001)
  beta ~ dnorm(0, .001)
```

Inference for Bugs model at "C:/Users/FRS/AppData/Local/Temp/RtmpgxpkpY 3 chains, each with 4000 iterations (first 2000 discarded), n.thin = 2 n.sims = 3000 iterations saved

 mu.vect
 sd.vect
 2.5%
 97.5%
 Rhat n.eff

 beta
 0.056
 0.015
 0.026
 0.085
 1.051
 46

 mu
 -3.487
 0.247
 -3.953
 -3.052
 1.049
 46

 deviance
 346.378
 10.771
 343.749
 352.567
 1.001
 3000

For each parameter, n.eff is a crude measure of effective sample size, and Rhat is the potential scale reduction factor (at convergence, Rhat=

DIC info (using the rule, pD = var(deviance)/2) pD = 58.0 and DIC = 404.4

DIC is an estimate of expected predictive error (lower deviance is bett

Compare with glm

STAN

STAN models are defined similarly to JAGS (but see STAN manual for differences)

```
bayes.lm <- function(){</pre>
  # LIKELIHOOD
  for (i in 1:length(height)){
   height[i] ~ dnorm(mu[i], tau)
                                    # tau = precision (inverse of vari
   mu[i] <- alpha + beta*dbhc[i]</pre>
                                    # centred diameter
  # PRIORS (vaque or weakly informative)
  alpha ~ dunif(1, 100) # prior for average height of a 25-cm-DBH t
  beta ~ dunif(0, 10) # how much do we expect height to scale wit
  tau <- 1/(sigma*sigma) # tau = 1/sigma^2
  sigma ~ dunif(0, 50) # residual standard deviation
```

```
bayes.lm.stan <- "
 data {
    int<lower=0> N;
   vector[N] dbhc;
   vector[N] height;
 parameters {
   real<lower=0, upper=100> alpha;
    real<lower=0, upper=10> beta;
    real<lower=0, upper=50> sigma;
 model {
   height ~ normal(alpha + beta * dbhc, sigma);
```

Fitting model in STAN

Similar results as JAGS

lp__ 1731

1

Inference for Stan model: 57b6d06b34f74072ecc985613f1ce936.
3 chains, each with iter=2000; warmup=1000; thin=1;
post-warmup draws per chain=1000, total post-warmup draws=3000.

	mear	n se_mear	ı sd	2.5%	25%	50%	75%	97.5
alpha	34.74	0.00	0.13	34.49	34.65	34.73	34.83	34.9
beta	0.62	0.00	0.01	0.60	0.61	0.62	0.62	0.6
sigma	4.10	0.00	0.09	3.94	4.04	4.09	4.16	4.2
lp	-1905.63	0.03	1.12	-1908.55	-1906.10	-1905.34	-1904.82	-1904.3
	n_eff Rh	nat						
alpha	2500	1						
beta	2497	1						
sigma	2553	1						

Samples were drawn using NUTS(diag_e) at Wed Nov 09 17:43:53 2016. For each parameter, n_eff is a crude measure of effective sample size, and Rhat is the potential scale reduction factor on split chains (at convergence, Rhat=1).

Lots of example models in STAN

https://github.com/stan-dev/example-models/wiki

To fit 'standard' Bayesian GLMs and GLMMs, use rstanarm

RStanArm

Bayesian applied regression modeling via Stan

RStanArm allows users to specify models via the customary R commands, where

- models are specified with formula syntax,
- · data is provided as a data frame, and
- additional arguments are available to specify priors.

Bayesian modelling in one line of code!

and results make sense:)

```
stan_glm(formula = height ~ dbh.c, family = gaussian, data = trees,
   prior = normal(0, 10), prior_intercept = normal(30, 30))
Estimates:
           Median MAD_SD
(Intercept) 34.7 0.1
dbh.c 0.6 0.0
sigma 4.1 0.1
Sample avg. posterior predictive
distribution of y (X = xbar):
        Median MAD SD
mean_PPD 36.5 0.2
Observations: 1000 Number of unconstrained parameters: 3
```



Less-tech version

More-tech version

Everything

R Users Will Now Inevitably Become Bayesians

26

There are several reasons why everyone isn't using Bayesian methods for regression modeling. One reason is that Bayesian modeling requires more thought: you need pesky things like priors, and you can't assume that if a procedure runs without throwing an error that the answers are valid. A second reason is that MCMC sampling — the bedrock of practical Bayesian modeling — can be slow compared to closed-form or MLE procedures. A third reason is that existing Bayesian solutions have either been highly-specialized (and thus inflexible), or have required knowing how to use a generalized tool like BUGS, JAGS, or Stan. This third reason has recently been shattered in the R world by not one but two packages: brms and rstanarm. Interestingly, both of these packages are elegant front ends to Stan, via rstan and shinystan.

END



Slides and source code available at https://github.com/Pakillo/Bayes-R-JAGS-intro