## **BiOCamLib**

BiOCamLib is the OCaml foundation upon which a number of the bioinformatics tools I developed are built.

It mostly consists of a library — you'll need to clone this repository if you want to manually compile other programs I've developed, notably SiNPle or KPop. You might also use the library for your own programs, if you are familiar with OCaml and patient enough to read the code.

As a bonus, BiOCamLib comes bundled with a few programs:

· Octopus , which is a high-throughput program to compute the transitive closure of strings. For instance, the input

```
A duh
b C
c f e
duh zz x
```

will result in the output

```
Α
         duh
                   ZZ
h
         С
```

(tab-separated) once processed by Octopus . This is useful to cluster things.

- Parallel, which allows you to split and process an input file chunk-wise using the reader/workers/writer model implemented in BiOCamLib.Tools.Parallel. You can see it as a demonstration of the capabilities of the library, but I also often use it as a useful tool to solve real-life problems.
- FASTools, which is a Swiss-knife tool for the manipulation of FASTA/FASTQ files. It supports all formats (FASTA, single- and paired-end FASTQ, interleaved FASTQ) and a simpler tabular format whereby FASTA/FASTQ records are represented as tabseparated lines. It facilitates format interconversions and other manipulations.

## Installing Octopus, Parallel, and FASTools



Note that the only operating systems we officially support are Linux and MacOS. 🛕



OCaml is highly portable and you might be able to manually compile/install everything successfully on other platforms (for instance, Windows) but you will have to do it yourself.

There are several possible ways of installing the software on your machine: through conda; by downloading pre-compiled binaries (Linux and MacOS x86\_64 only); or manually.

#### Conda channel



### **Pre-compiled binaries**

You can download pre-compiled binaries for Linux and MacOS x86\_64 from our releases.

### Manual install

Alternatively, you can install Octopus , Parallel , and FASTools manually by cloning and compiling the BiOCamLib sources. You'll need an up-to-date distribution of the OCaml compiler and the Dune package manager for that. Both can be installed through OPAM, the official OCaml distribution system. Once you have a working OPAM distribution you'll also have a working OCaml compiler, and Dune can be installed with the command

```
$ opam install dune
```

if it is not already present. Make sure that you install OCaml version 4.12 or later.

Then go to the directory into which you have downloaded the latest BiOCamLib sources, and type

\$ ./BUILD

That should generate the executables Octopus, Parallel, and FASTools. Copy them to some favourite location in your PATH, for instance ~/.local/bin.

## Command line options for Octopus

This is the full list of command line options available for the program Octopus . You can visualise the list by typing

```
$ Octopus -h
```

in your terminal. You will see a header containing information about the version:

```
This is the Octopus program (version 0.3)
(c) 2016-2023 Paolo Ribeca, paolo.ribeca@gmail.com>
```

followed by detailed information. The general form(s) the command can be used is:

```
Octopus [OPTIONS]
```

#### Miscellaneous

Option	Argument(s)	Effect	Note(s)
-V version		print version and exit	
-h help		print syntax and exit	

# Command line options for Parallel

This is the full list of command line options available for the program Parallel . You can visualise the list by typing

```
$ Parallel -h
```

in your terminal. You will see a header containing information about the version:

```
This is the Parallel program (version 0.4)
(c) 2019-2022 Paolo Ribeca, paolo Ribeca
```

followed by detailed information. The general form(s) the command can be used is:

```
Parallel [OPTIONS] -- [COMMAND TO PARALLELIZE AND ITS OPTIONS]
```

### Command to parallelize

Option	Argument(s)	Effect	Note(s)
		consider all the subsequent parameters as the command to be executed in parallel. At least one command must be specified	(mandatory)

### Input/Output

Option	Argument(s)	Effect	Note(s)
-lines-per-block	<positive_integer></positive_integer>	number of lines to be processed per block	default= <mark>10000</mark>
-i input	<input_file></input_file>	name of input file	default= <mark>stdin</mark>
-o output	<output_file></output_file>	name of output file	default= <mark>stdout</mark>

### Miscellaneous

Option	Argument(s)	Effect	Note(s)
-t  threads	<positive_integer></positive_integer>	number of concurrent computing threads to be spawned (default automatically detected from your configuration)	default= <mark>nproc</mark>
-v  verbose		set verbose execution	default= <mark>false</mark>
-d debug		output debugging information	default= <u>false</u>
-h help		print syntax and exit	

## **Command line options for FASTools**

This is the full list of command line options available for the program FAST001s . You can visualise the list by typing

```
$ FASTools -h
```

in your terminal. You will see a header containing information about the version:

```
This is the FASTools program (version 0.4)
(c) 2022 Paolo Ribeca, <paolo.ribeca@gmail.com>
```

followed by detailed information. The general form(s) the command can be used is:

```
FASTools [OPTIONS]
```

**Working mode.** Executed delayed in order of specification, default=compact.

Option	Argument(s)	Effect	Note(s)
compact -ccompact		put each FASTA/FASTQ record on one tab-separated line	
expand -e expand		split each tab-separated line into one or more FASTA/FASTQ records	
match -mmatch	<regexp></regexp>	select matching sequence names in FASTA/FASTQ records or tab-separated lines. For paired-end files, the pair matches when at least one name matches	
revcom -rrevcom		reverse-complement sequences in FASTA/FASTQ records or tab-separated lines	
dropq -ddropq		drop qualities in FASTA/FASTQ records or tab-separated lines	

**Input/Output.** Executed delayed in order of specification, default=<u>-</u>F.

Option	Argument(s)	Effect	Note(s)
-f fasta	<fasta_file_name></fasta_file_name>	process FASTA input file containing sequences	
-F		process FASTA sequences from standard input	
-s single- end	<fastq_file_name></fastq_file_name>	process FASTQ input file containing single-end sequencing reads	
-8		process single-end FASTQ sequencing reads from standard input	
-p paired- end	<fastq_file_name1> <fastq_file_name2></fastq_file_name2></fastq_file_name1>	process FASTQ input files containing paired- end sequencing reads	
- P		process interleaved FASTQ sequencing reads from standard input	
-t tabular	<tabular_file_name></tabular_file_name>	process input file containing FAST[A Q] records as tab-separated lines	
-Т		process FAST[A Q] records in tabular form from standard input	
-linter	'none' 'DNA' 'dna' 'protein'	sets linter for sequence. All non-base (for DNA) or non-AA (for protein) characters are converted to unknowns	default= <mark>none</mark>
linter- keep-dashes	<bool></bool>	sets whether the linter should keep dashes appearing in sequences or convert them to unknowns	default= <mark>false</mark>
-o output	<output_file_name></output_file_name>	set the name of the output file. Files are kept open, and it is possible to switch between them by repeatedly using this option. Use '/dev/stdout' for standard output	default= <mark>/dev/stdout</mark>
-0 paired- end-output	<output_file_name_1> <output_file_name_2></output_file_name_2></output_file_name_1>	set the names of paired-end FASTQ output files. Files are kept open, and it is possible to switch between them by repeatedly using this option. Use '/dev/stdout' for standard output	default= <mark>/dev/stdout</mark> <u>/dev/stdout</u>
flush flush- output		flush output after each record (global option)	default= <mark>do not flush</mark>

### Miscellaneous

Option	Argument(s)	Effect	Note(s)
-v verbose		set verbose execution (global option)	default= <mark>false</mark>
-h help		print syntax and exit	