

# Class core values

1. Be **respectful** to yourself and others
2. Be **confident** and believe in yourself
3. Always do your **best**
4. Be **cooperative**
5. Be **creative**
6. Have **fun**
7. Be **patient** with yourself while you learn
8. Don't be shy to **ask "stupid" questions**



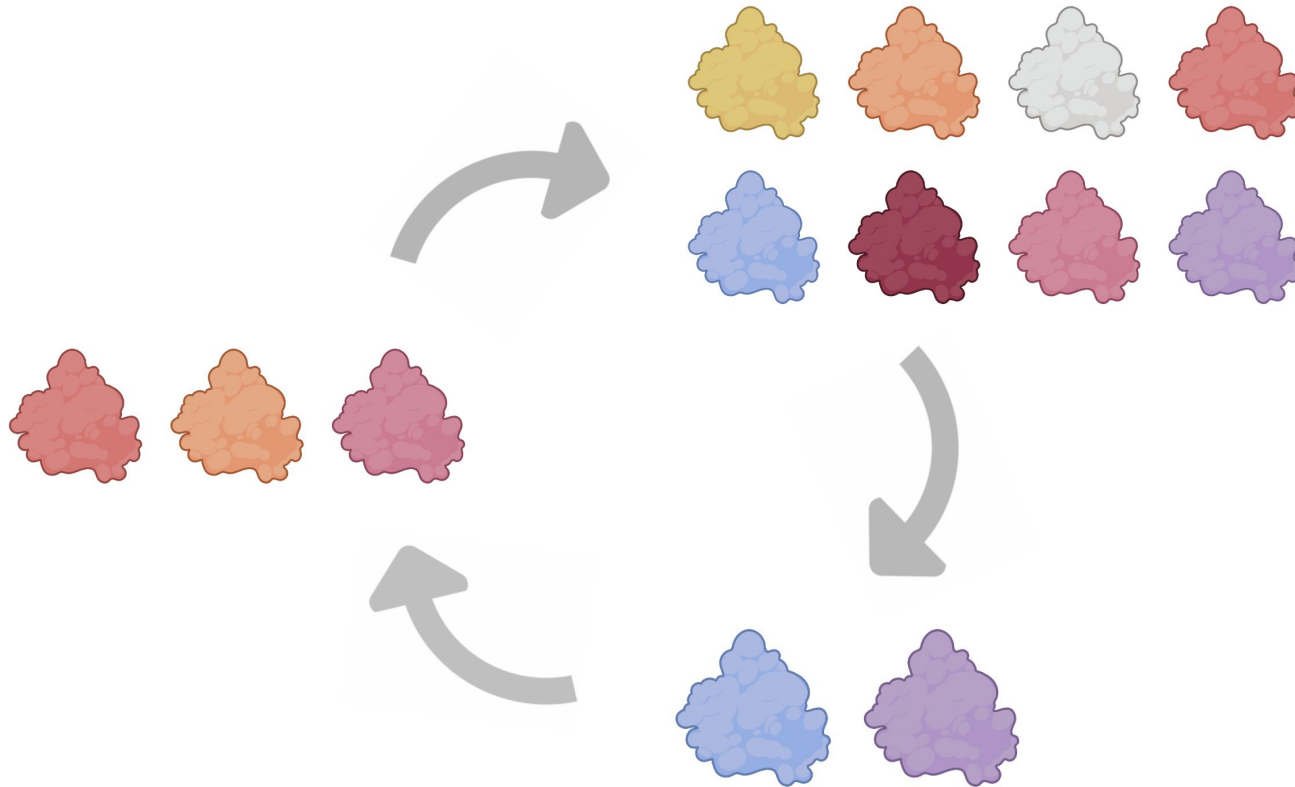
Week 4, Lecture 1

You get what you  
select for ...

# Learning Objectives

1. Describe the concepts of selection and screening
2. Identify the main methods for evolving binders
3. Critically evaluate the use of screening/selection methods for a given application
4. Identify methods for linking phenotype and genotype

# The overall process of directed evolution

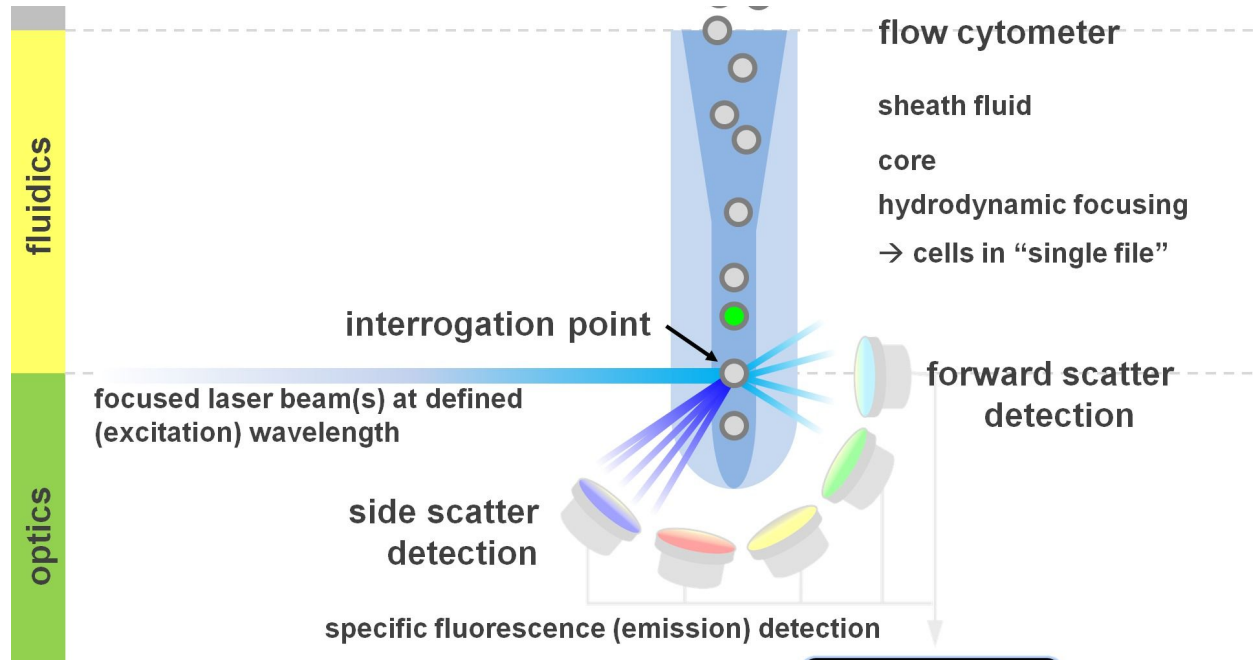


# A key part of directed evolution is choosing the best variants

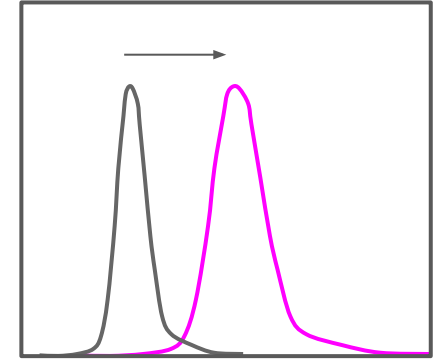
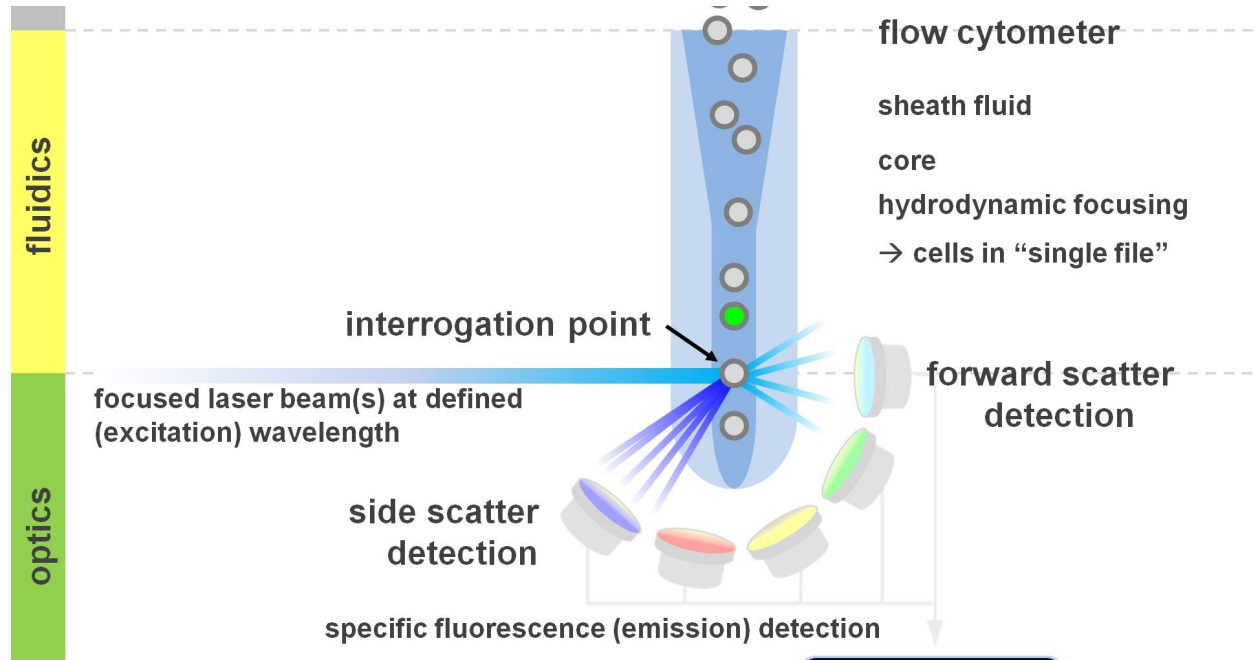


**Screening** is the process of going through all variants and picking up the best one

# Fluorescence can be used for screening

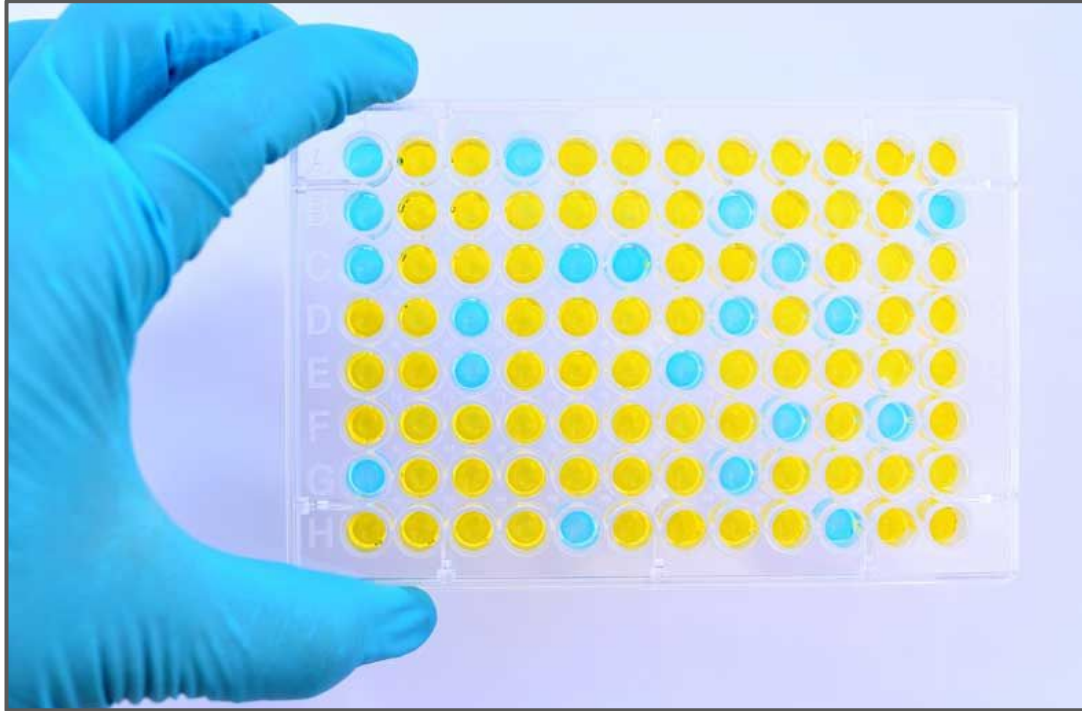


# Fluorescence can be used for screening



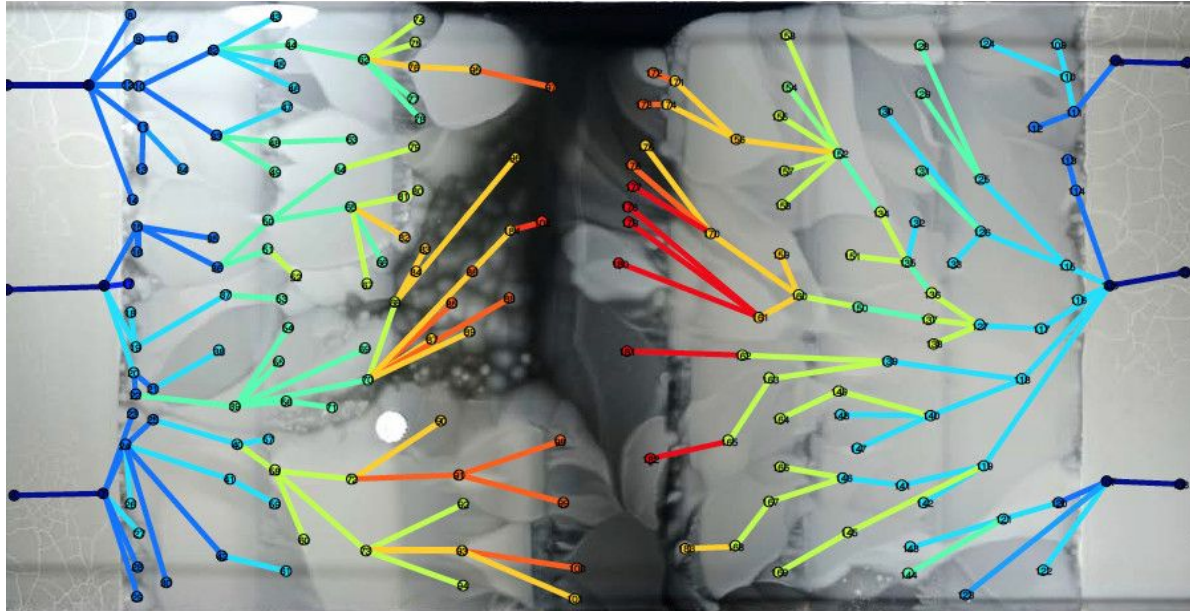


# Many enzymatic assays are screening-based



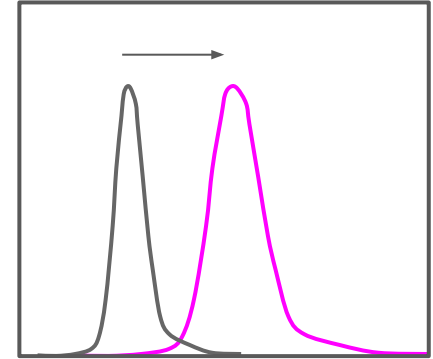
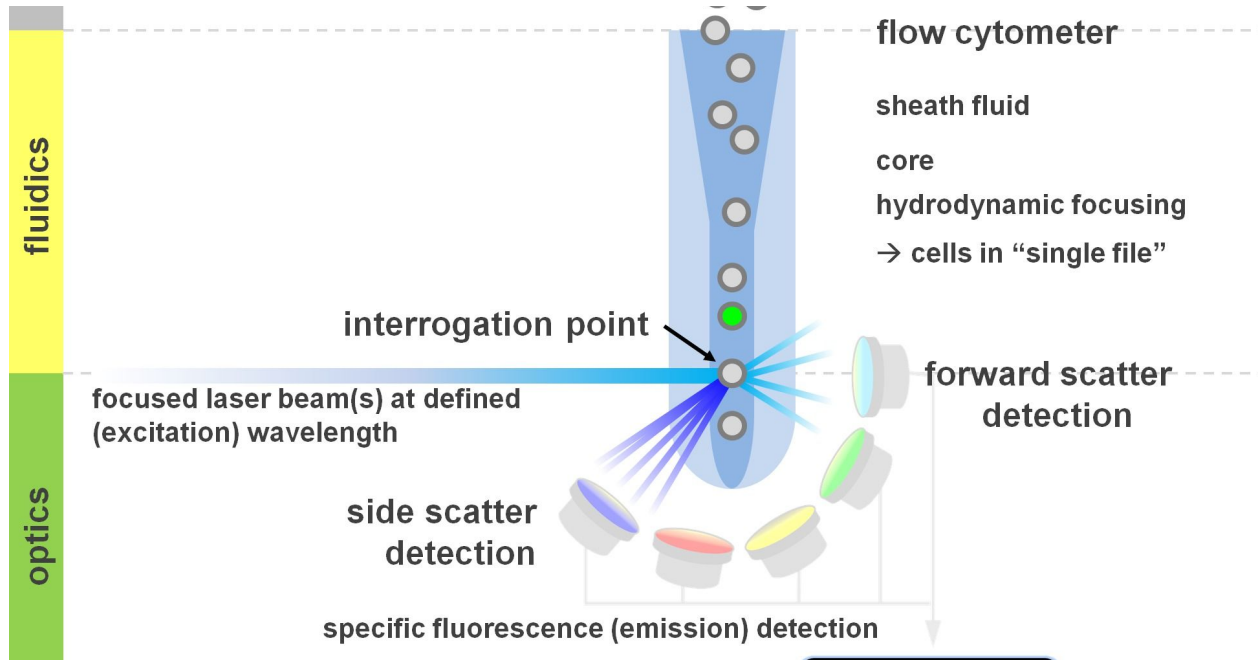
When you perform **selection**, only the fit variants survive

# Antibiotic plates are the most common method for selection

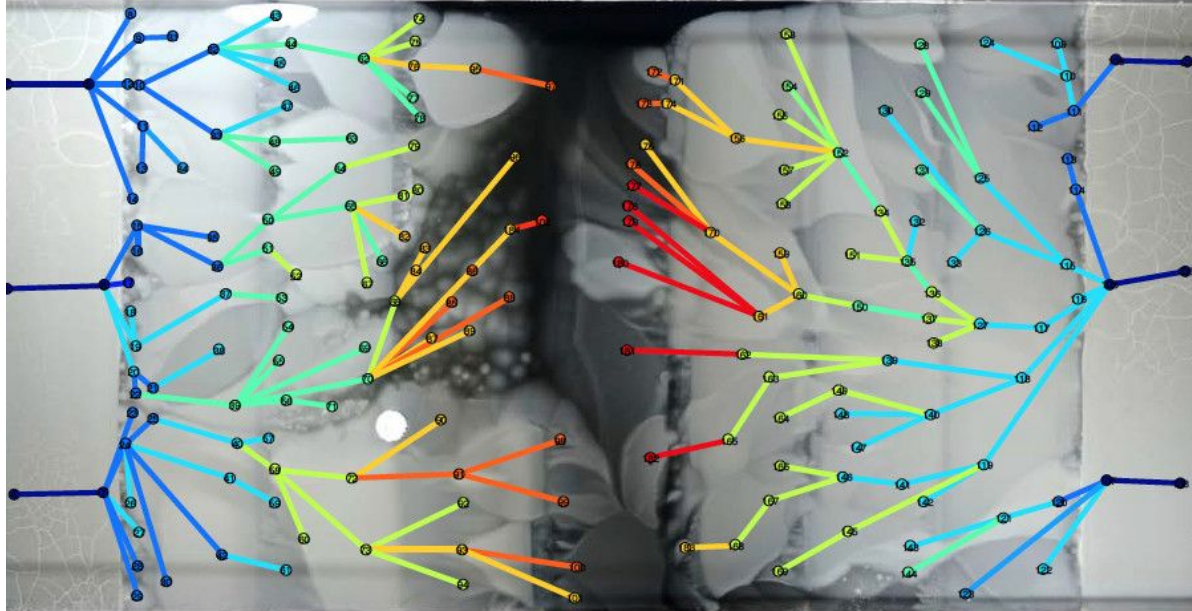


The outcome of evolution studies heavily depend on your selection/screening method

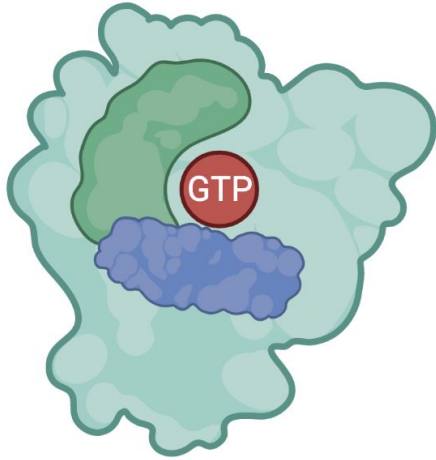
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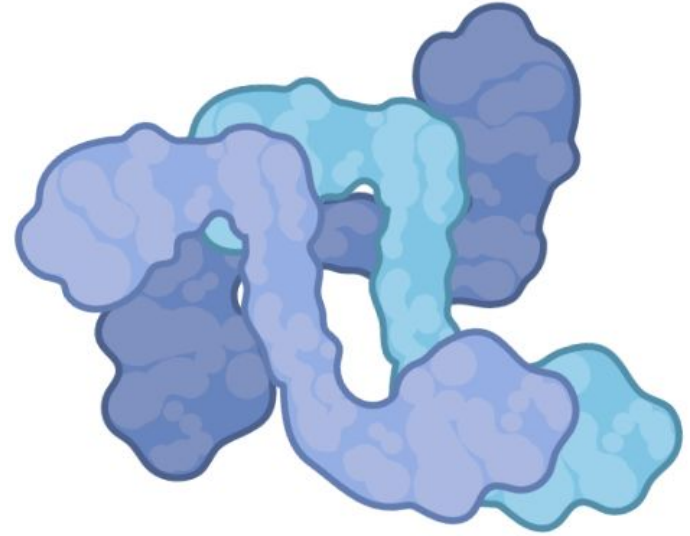
# In-class activity: You get what you select for ...



Scenario 1

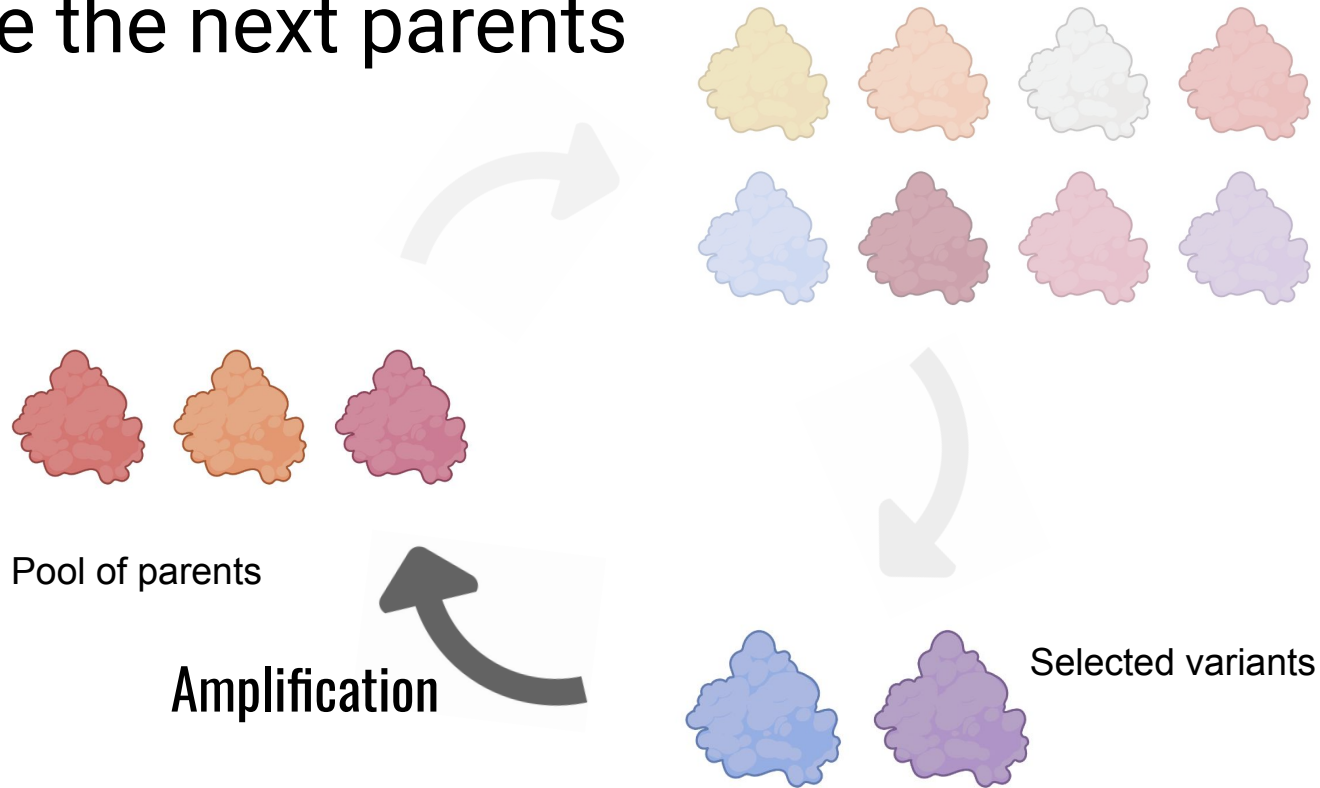


Scenario 2



Scenario 3

# The selected variants need to be amplified to create the next parents





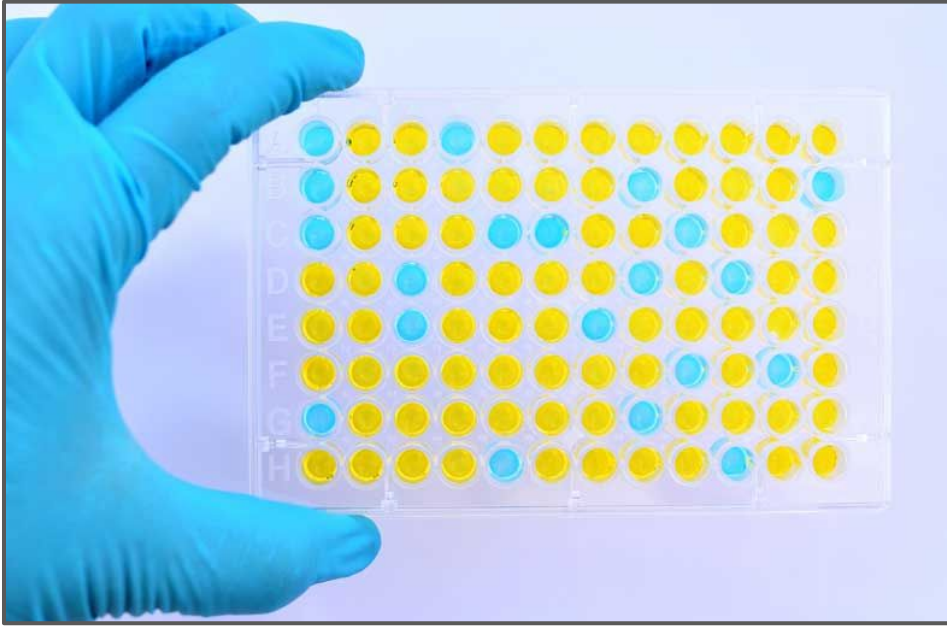
**Selection** happens at the protein level

**Selection** happens at the protein level, but  
**amplification** is at the DNA level

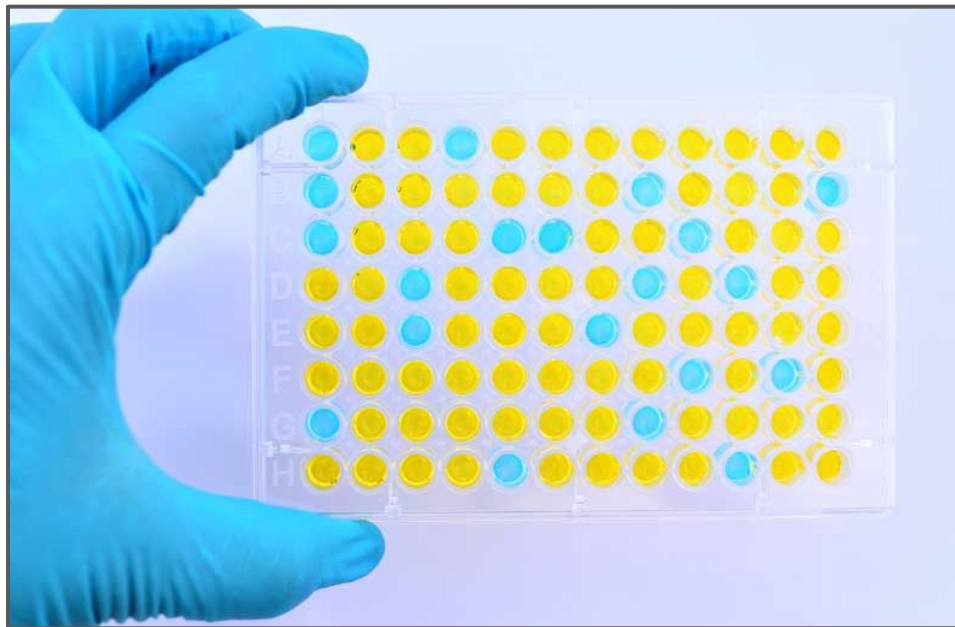
**Selection** happens at the protein level, but  
**amplification** is at the DNA level

We need a way to link the phenotype (protein) to  
genotype (DNA)

# In low-throughput assays ...



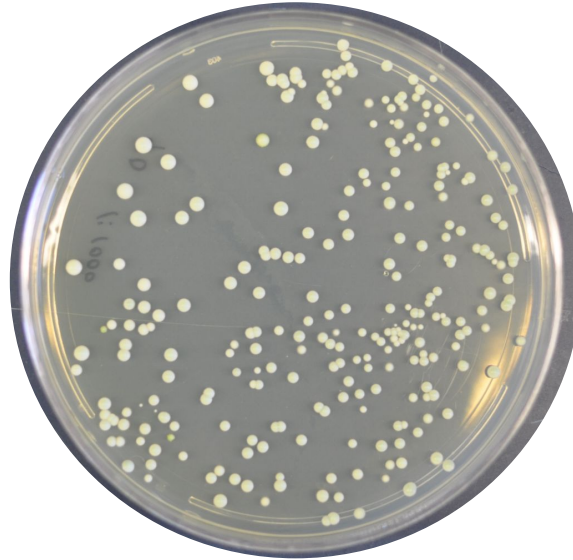
# In low-throughput assays, you can take note of the phenotype-genotype link



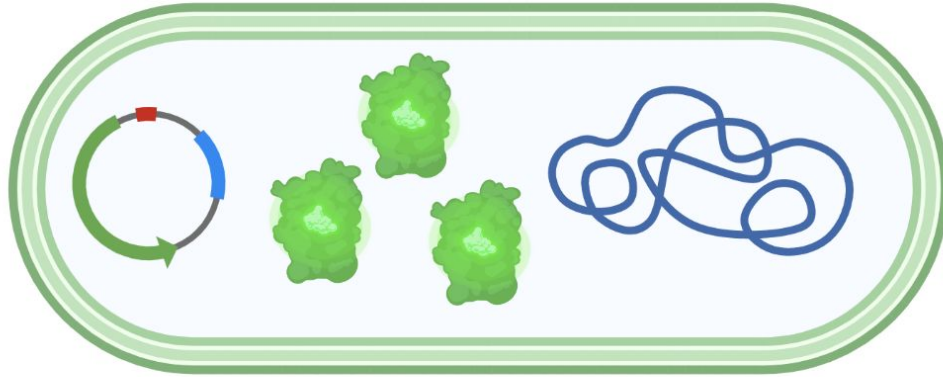
Row ID	Activity	Sequence
A1	90%	<b>LKNMGFTHILKDFSA</b>
A2	25%	<b>LKQMGFSHILKDWSA</b>
A3	40%	<b>IRNMGYTHIVKDFSA</b>
...	...	...
H12	35%	<b>LRNCGWTHIIKDETV</b>

**Containment:** Keeping the activity inside the cell where the DNA is!

# Antibiotic resistance is an example of contained activities

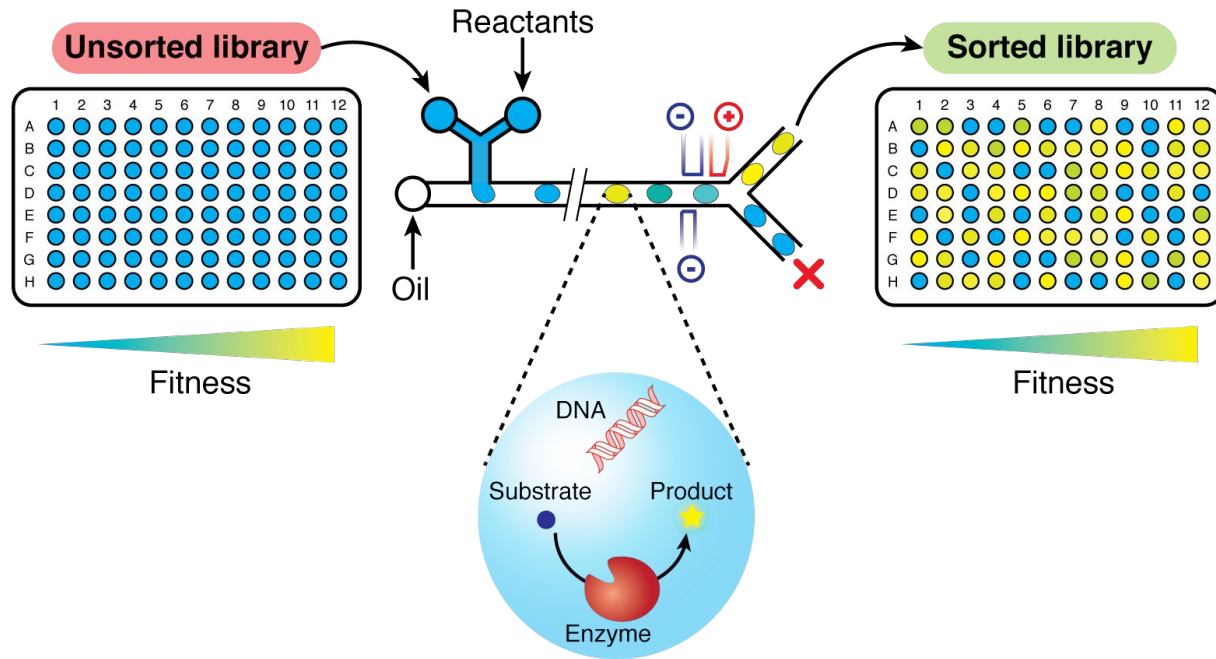


Fluorescent protein signals are contained within the cytosol



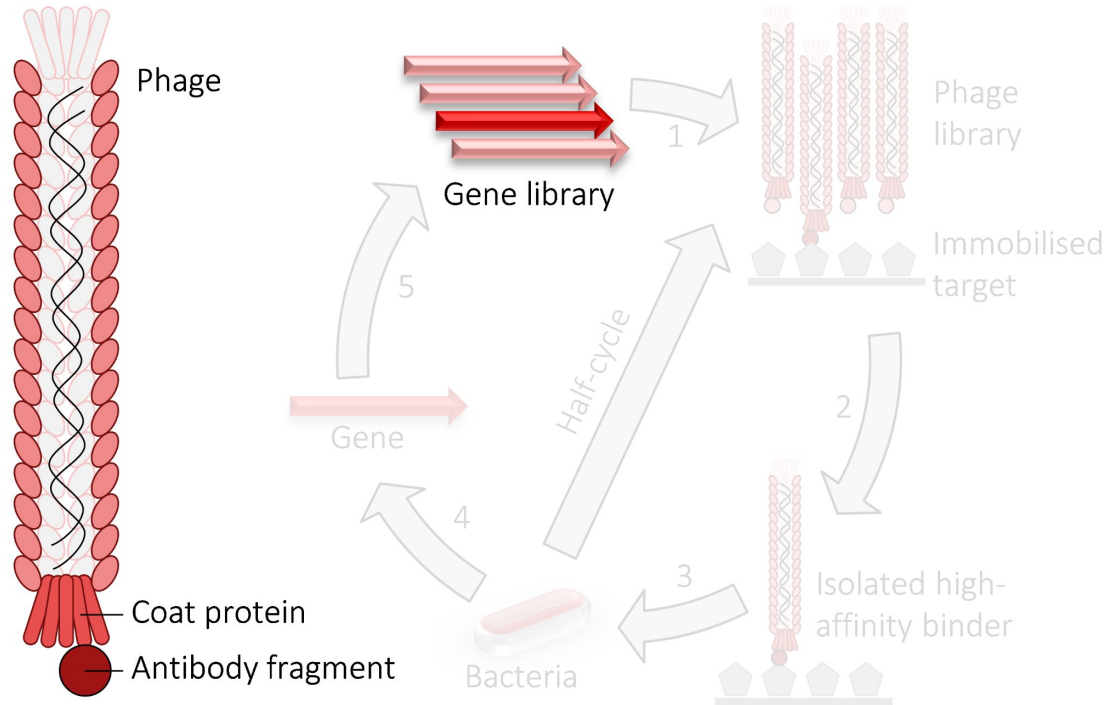


# Oil-water emulsions can be used to contain the phenotype and genotype within the same droplet

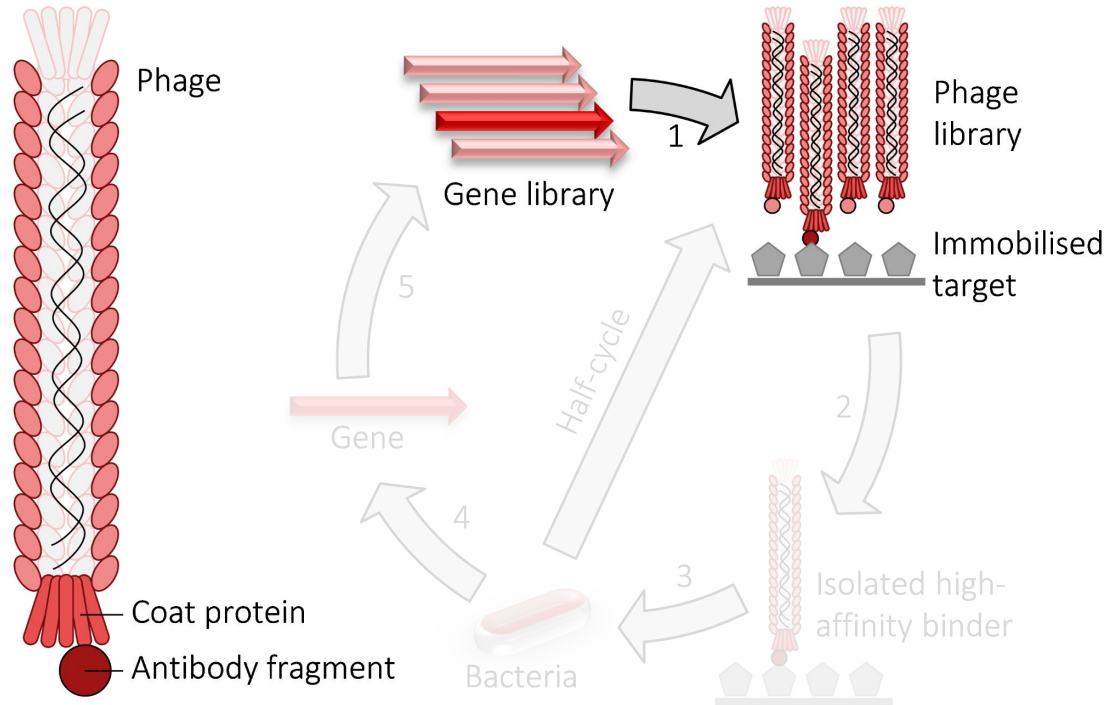


**Display:** Covalently linking the protein to the DNA source

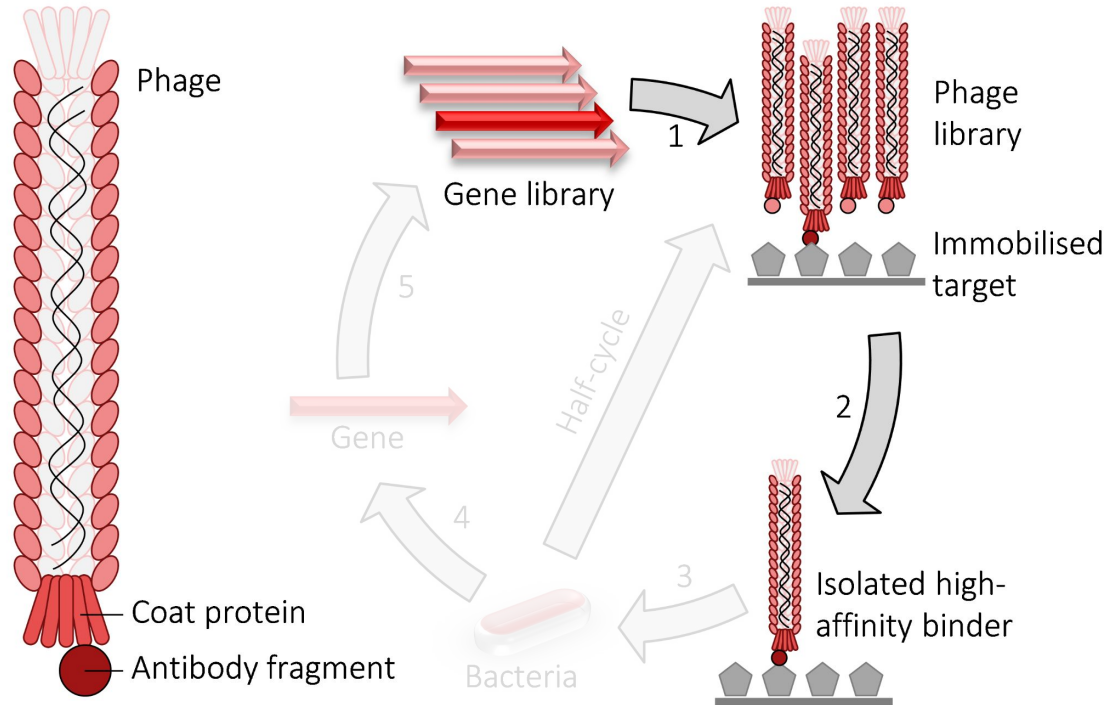
# *Phage display* is one the most commonly used display methods



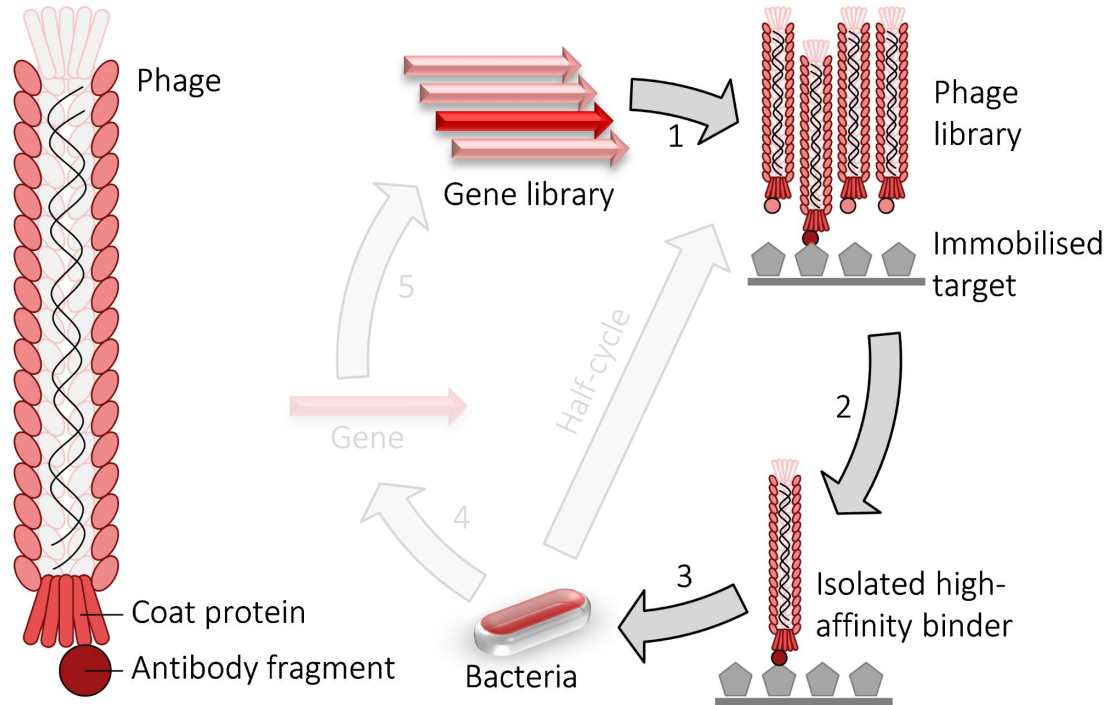
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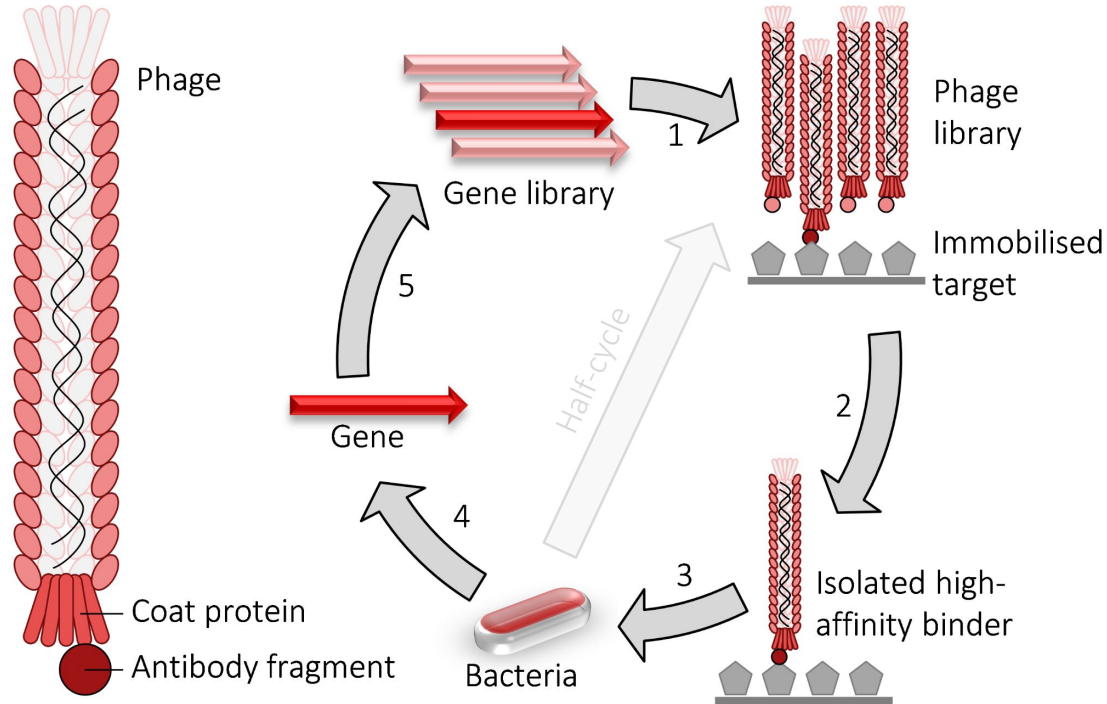
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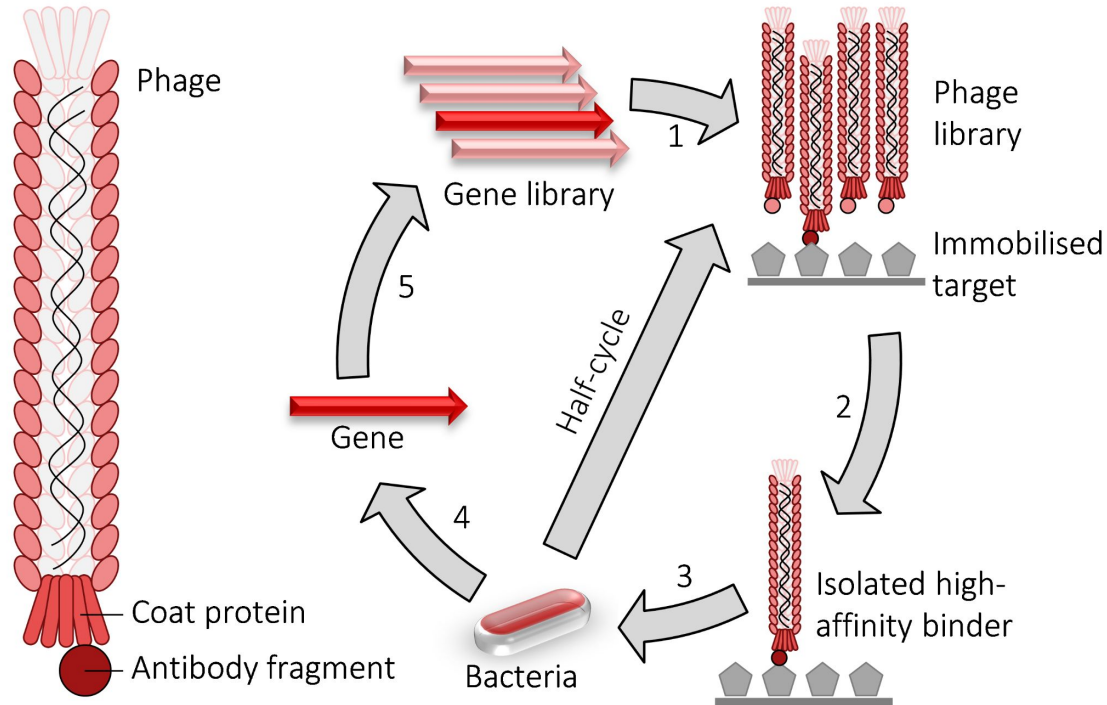
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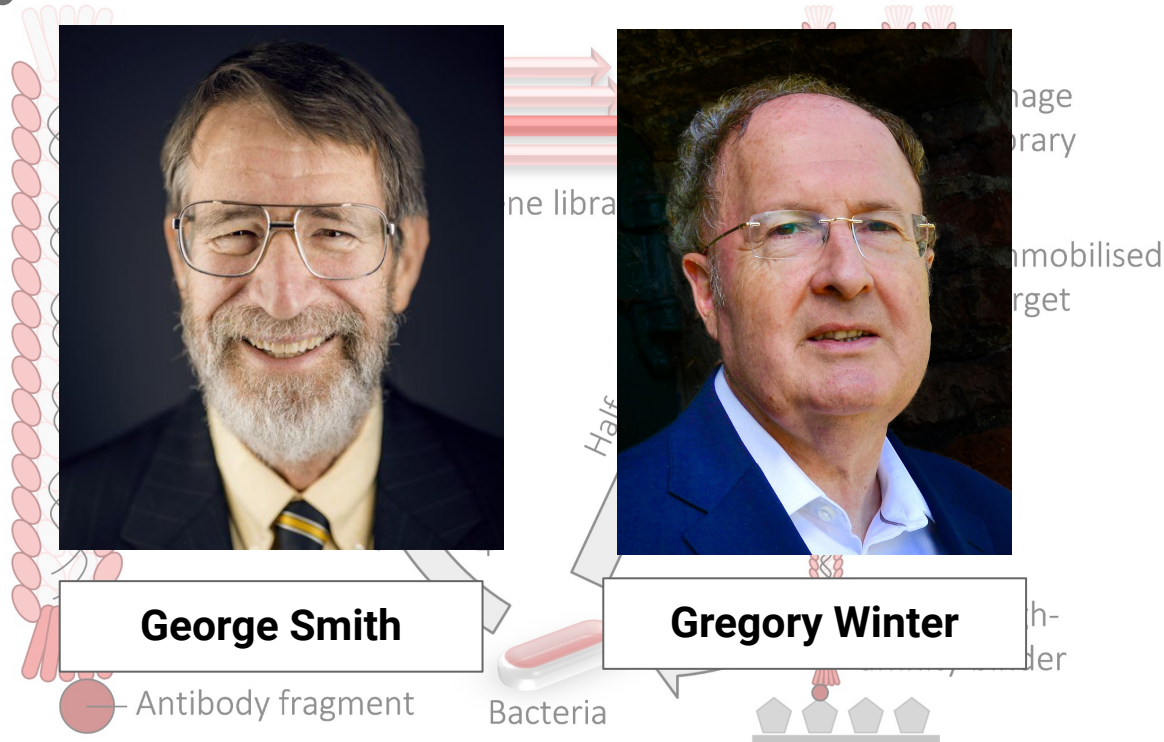
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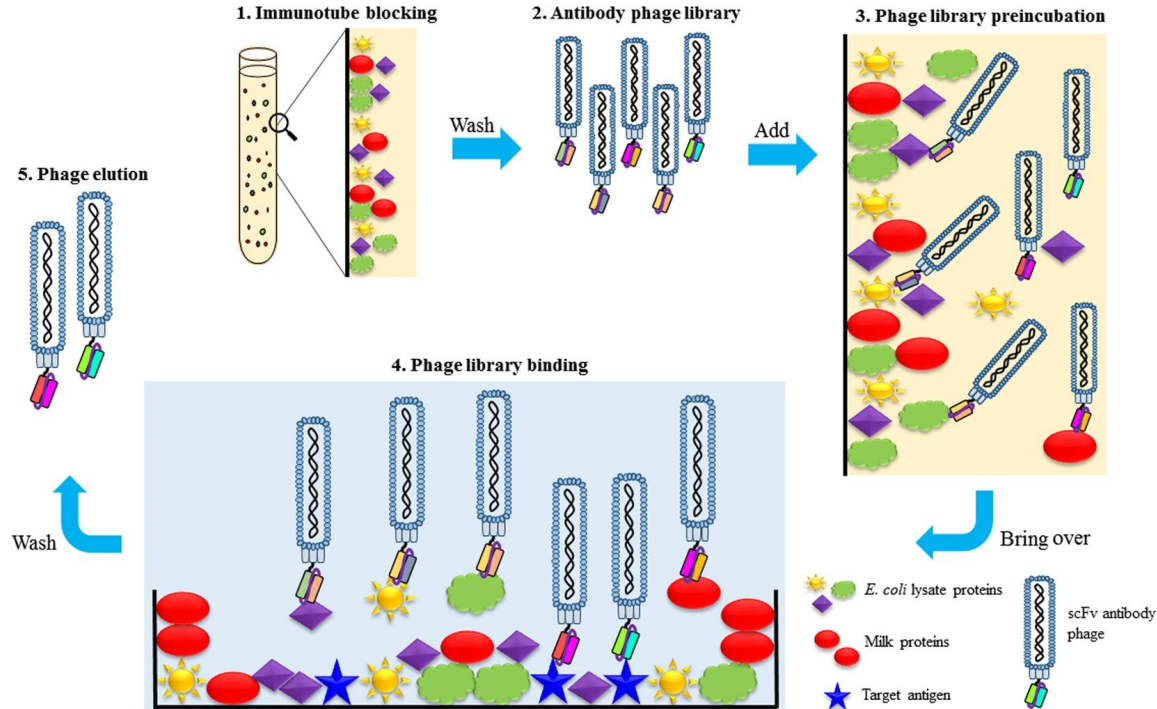
**George Smith**



**Gregory Winter**



# Phage display is one the most commonly used display methods

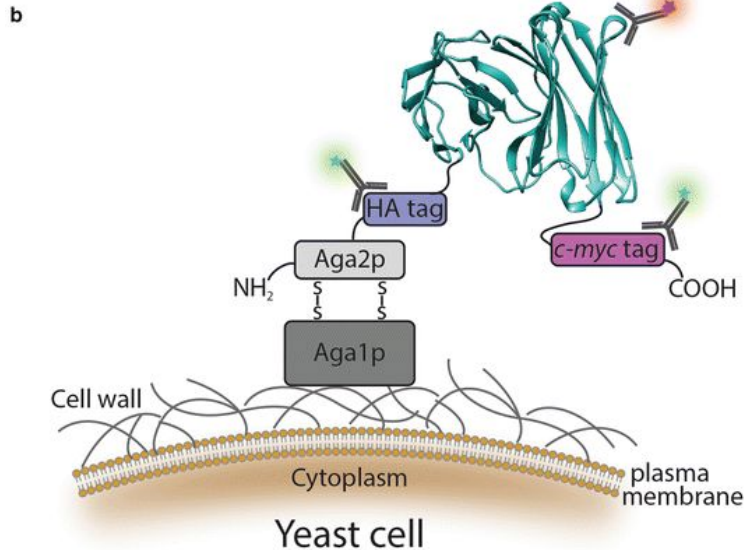


# *Yeast surface display* is the method of choice for most protein engineering approaches

a



NH<sub>2</sub> — Aga2p — HA tag — Protein of interest — c-myc tag — COOH



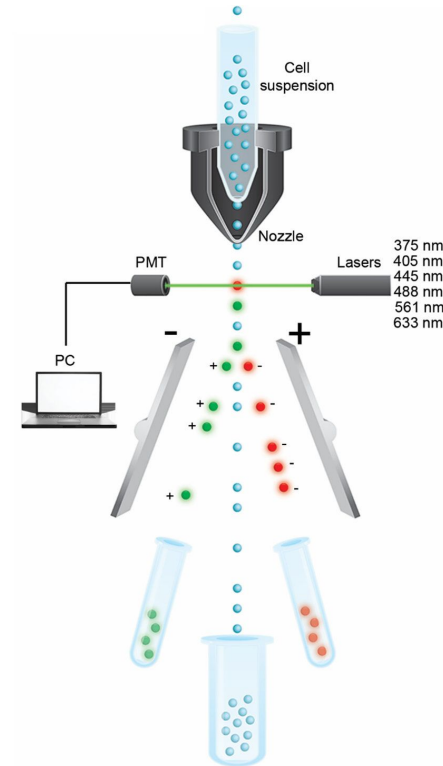
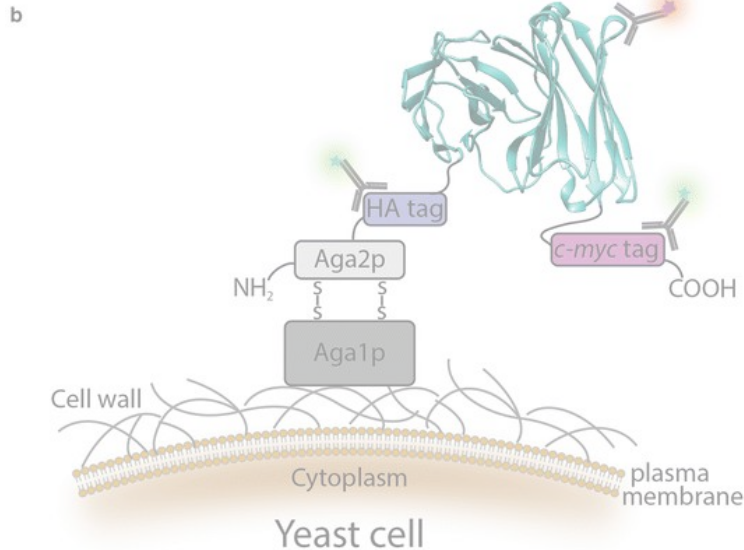
# Yeast surface display can be linked with FACS to obtain best binders

a

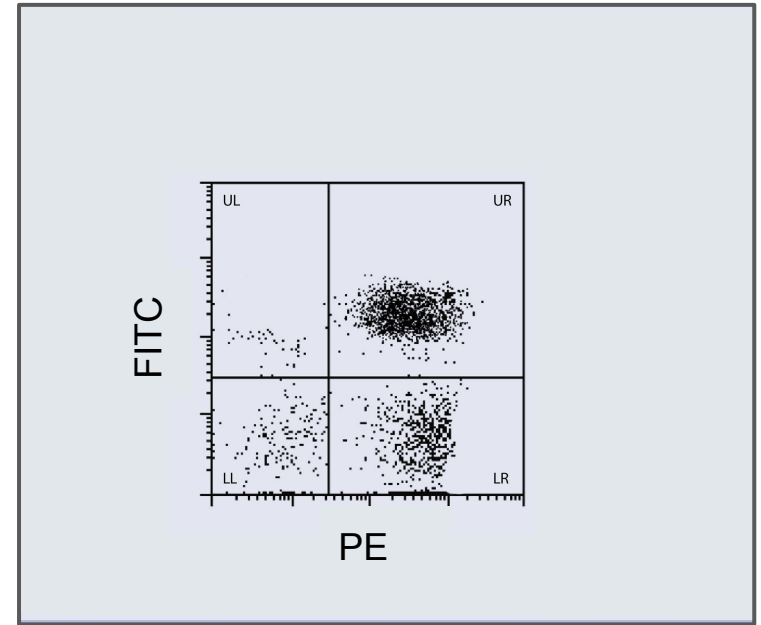
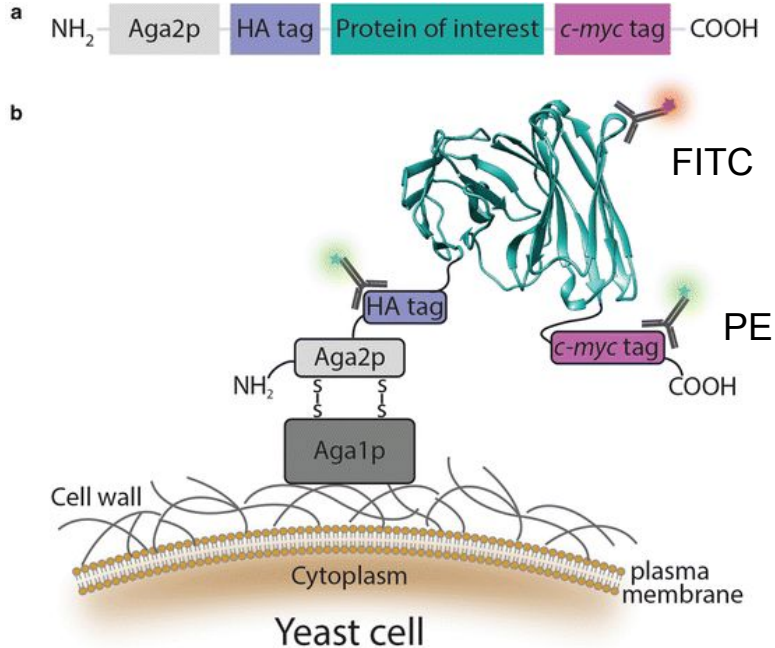


NH<sub>2</sub>—Aga2p—HA tag—Protein of interest—c-myc tag—COOH

This diagram shows a linear protein construct. It starts with an NH<sub>2</sub> terminus, followed by a grey box labeled 'Aga2p', a purple box labeled 'HA tag', a teal box labeled 'Protein of interest', a pink box labeled 'c-myc tag', and ends with a COOH terminus.

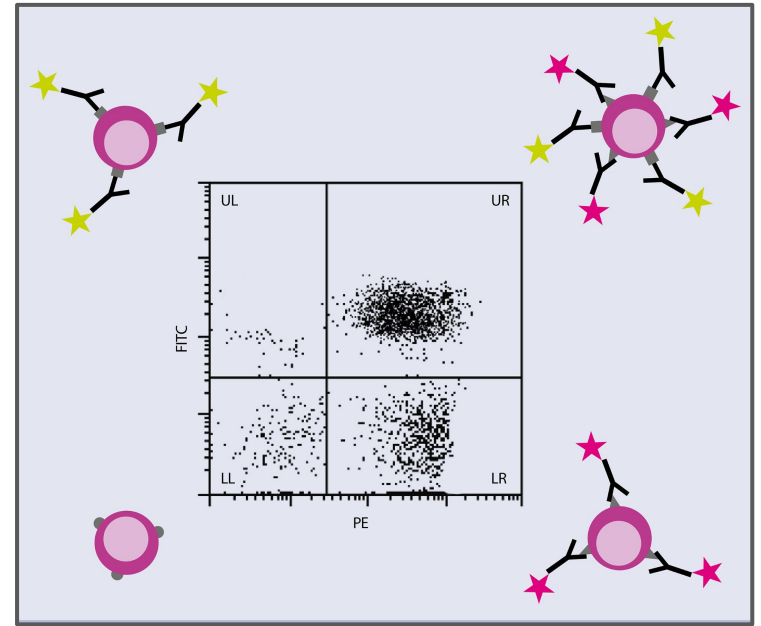
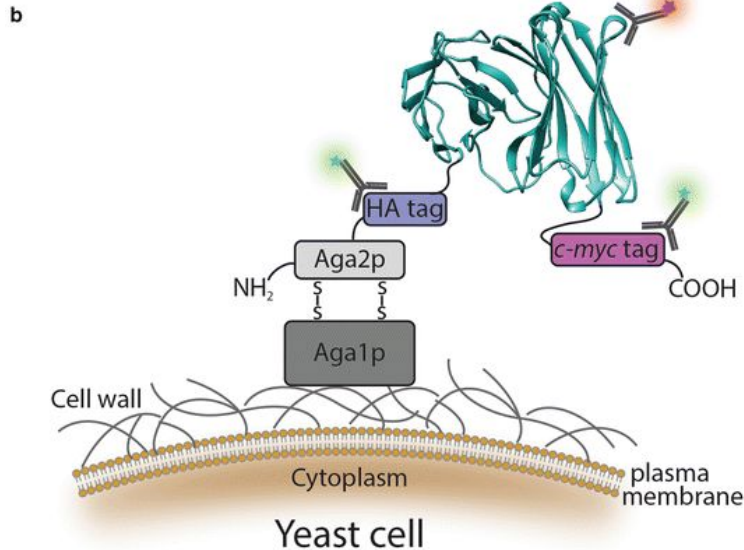


# *Yeast surface display* can be linked with FACS to obtain best binders



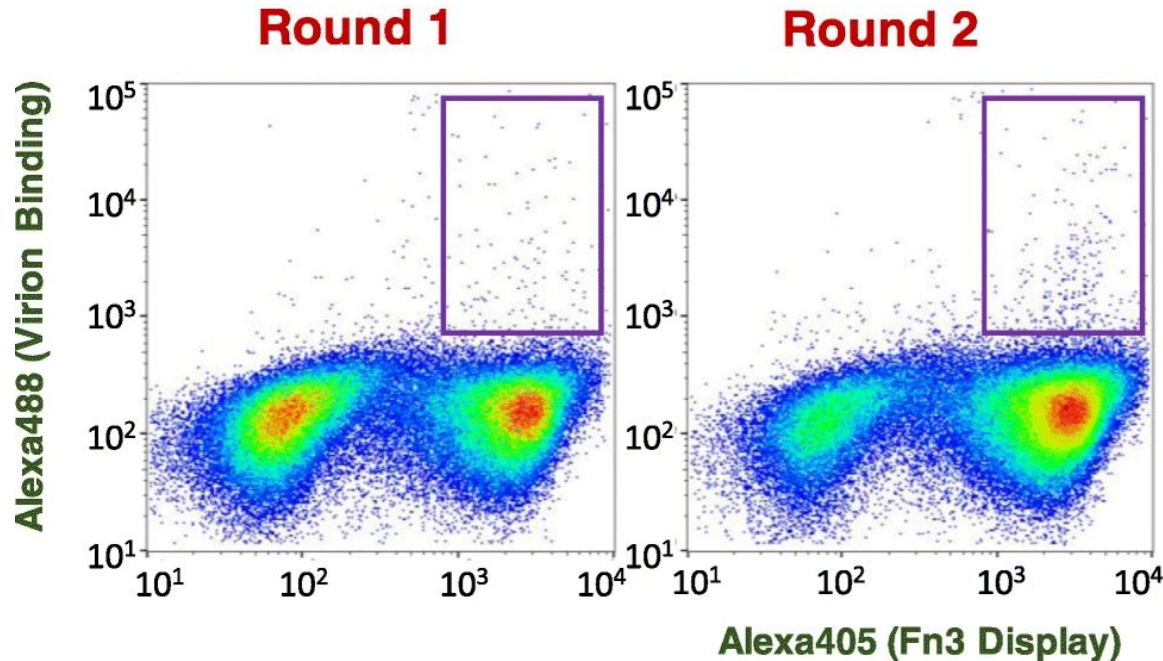
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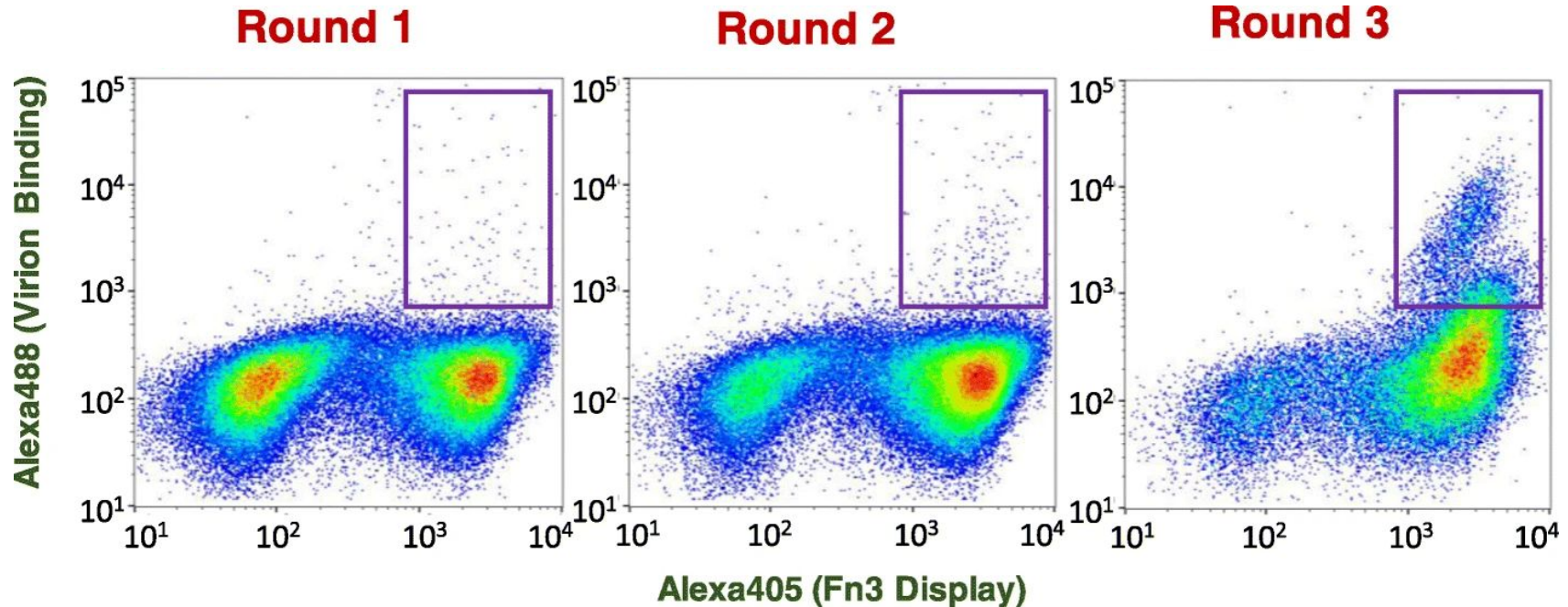




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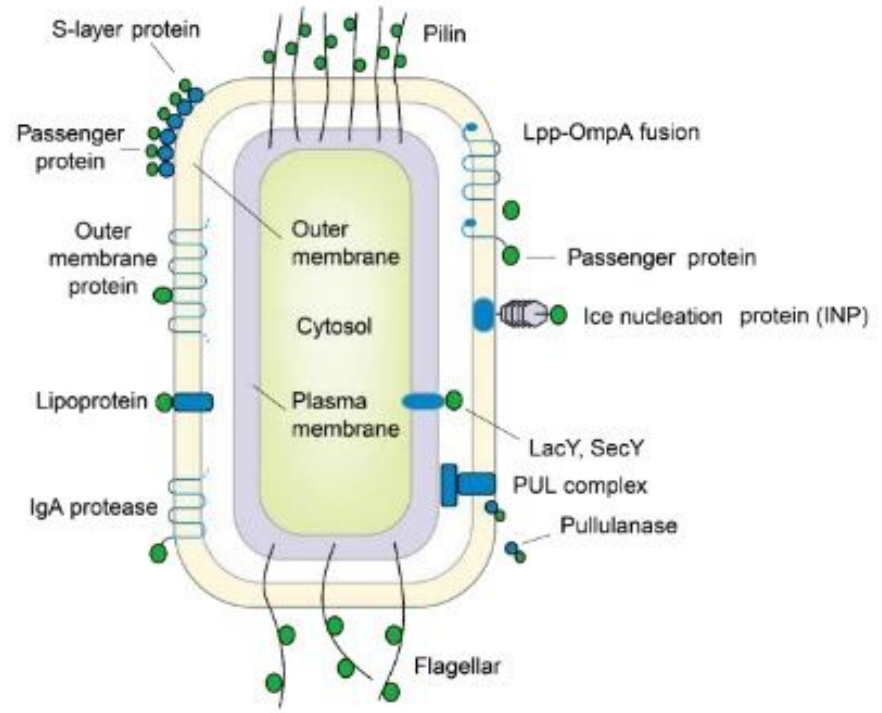


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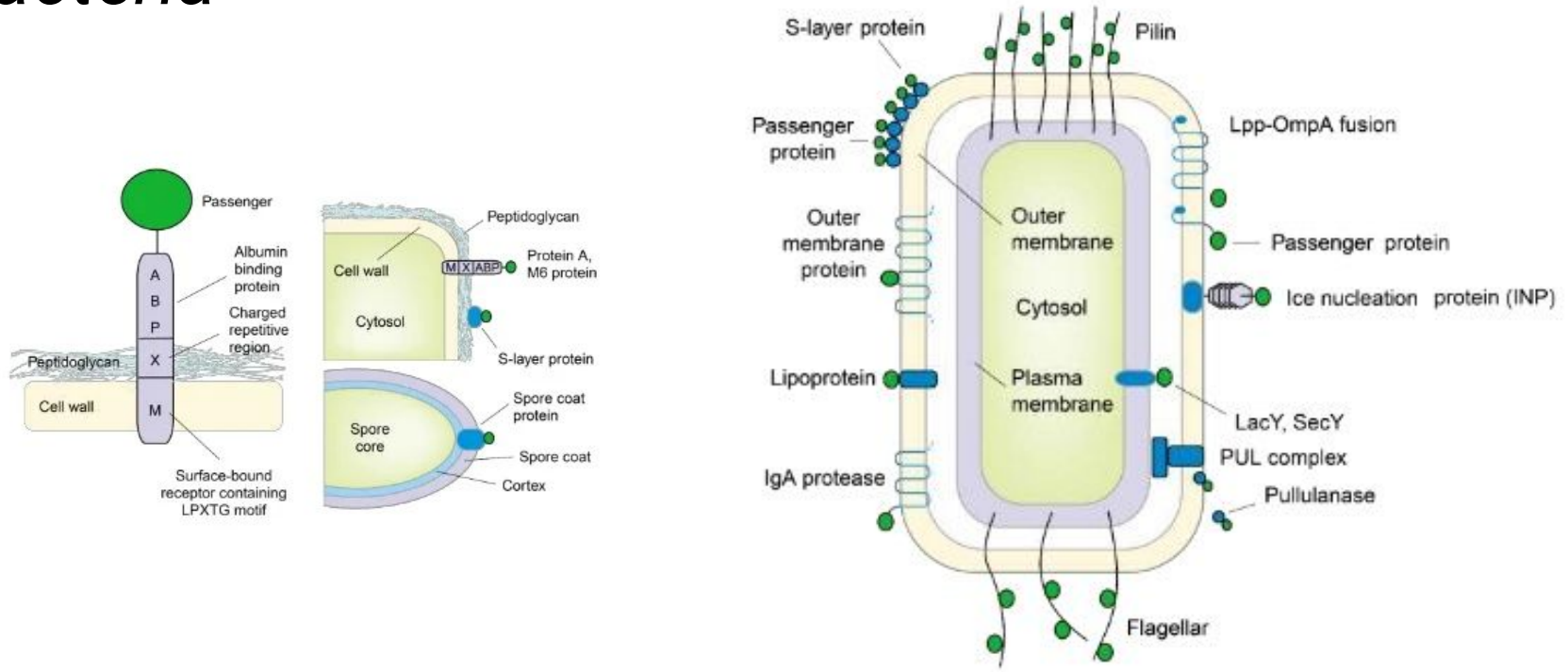




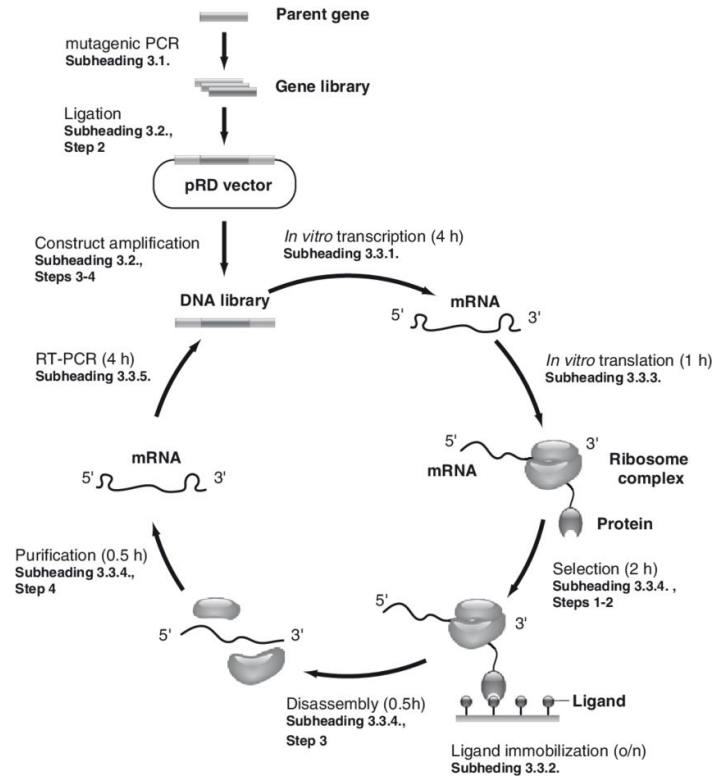
# Proteins can be displayed on the surface of *bacteria*



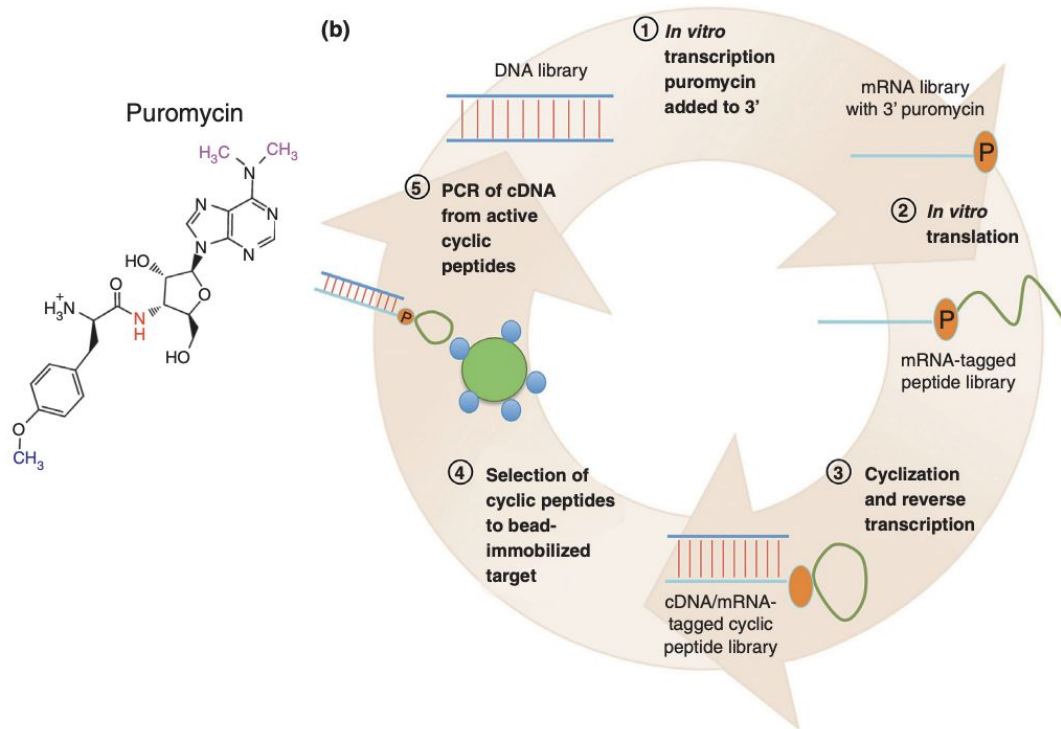
# Proteins can be displayed on the surface of *bacteria*



# Ribosome display can also be used for displaying proteins



# *mRNA display* is attracting attentions as a method for selecting peptide binders



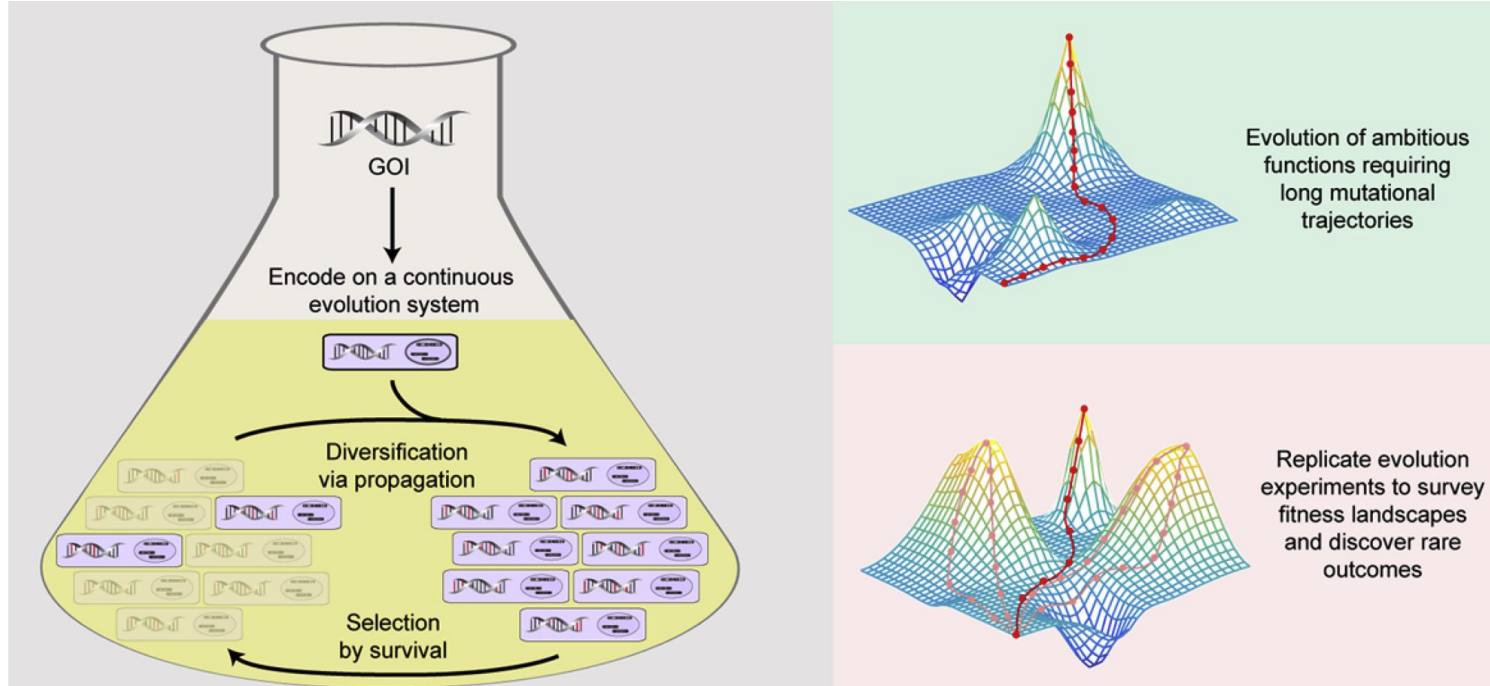
# Each method has its limitations/applications

	Phage display	Eukaryotic display	Prokaryotic display	Ribosome display	mRNA/cDNA display
Host organism	Filamentous phages, M13, T4, T7, lambda, phagemid	<i>S. cerevisiae</i> , <i>P. pastoris</i>	<i>E. coli</i> , <i>B. subtilis</i> , <i>L. bacillus</i> , <i>S. camosus</i>	<i>In vitro</i>	<i>In vitro</i>
Library size					
Highest affinity					
$K_d$ (M) <sup>a</sup>					
Typical enrichment factor per round					
Nucleic acid selected					
Transformation required					
Library form					
Proteins to be displayed					
Covalent link					
Surface anchorage					
Post translational machinery					

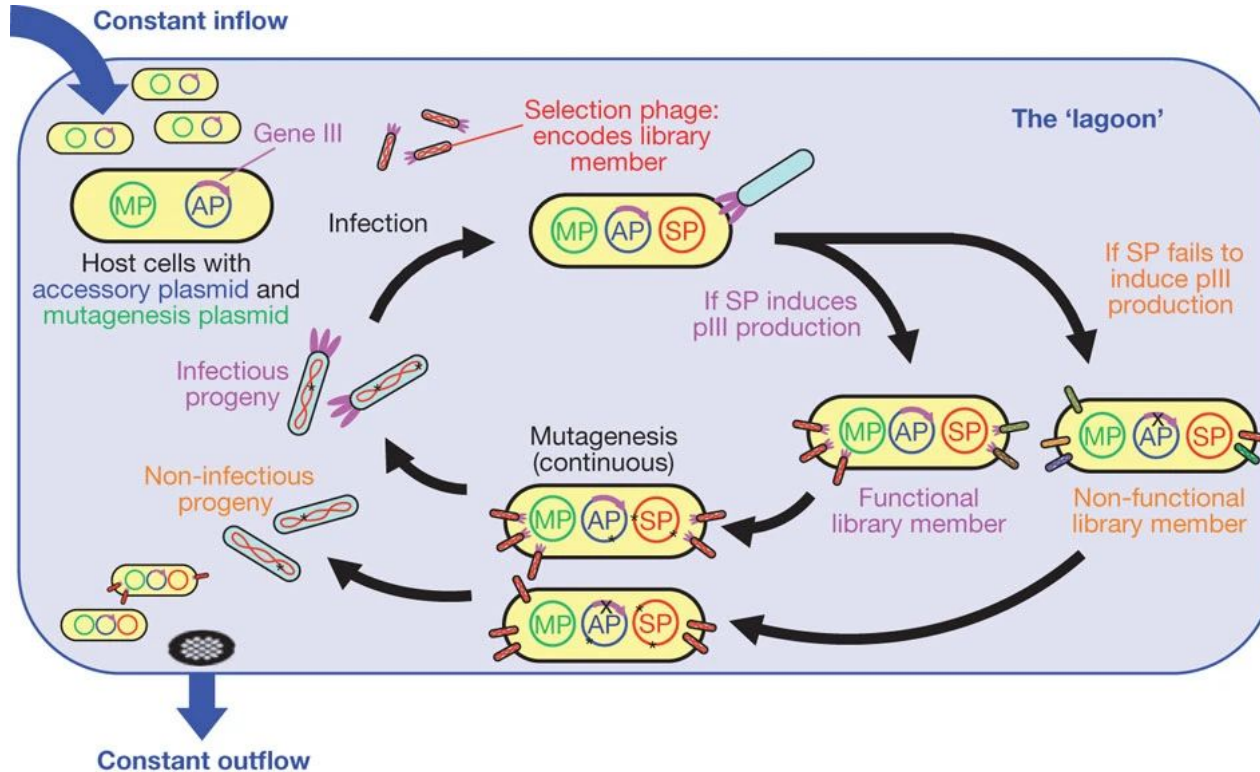
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Library size	$10^{9a}$	$10^7$	$10^{8-10}$	$10^{13-14}$	$10^{13-14}$
Highest affinity	$10^{-11}$	$10^{-14}$	$10^{-13}$	$10^{-12}$	$10^{-10}$
$K_d$ (M) <sup>a</sup>					
Typical enrichment factor per round	$10^{2-4}$	$10^{2-3}$	$10^{2-3}$	$10^{-3}$	$10^{1-3}$
Nucleic acid selected	DNA	DNA	DNA	mRNA	mRNA/cDNA
Transformation required	Yes	Yes	Yes	No	No
Library form	Plasmid	Plasmid	Plasmid	PCR fragment or mRNA	mRNA/cDNA, plasmid
Proteins to be displayed	Soluble, nontoxic, compatible with crossing membranes	Soluble and membrane, nontoxic, compatible with crossing membranes	Soluble and membrane, nontoxic, compatible with crossing membranes	Most proteins including cytotoxic, chemically modified and membrane proteins	Soluble, including cytotoxic, chemically modified
Covalent link	No	No	No	No	Yes (synthetic)
Surface anchorage	Capsid proteins	Agglutination proteins, flocculation proteins	Lpp-OmpA, autotransporter proteins, ice nucleation proteins	Ribosome	<i>In vitro</i>
Post translational machinery	Simple	Sophisticated	Moderate	Moderate	Simple

# Continuous evolution methods combine diversification, selection and amplification



# Phage-Assisted Continuous Evolution (PACE)





# For the next lecture:

1. Pre-class assessment for the next lecture  
Needs to be done before the start of class, will be available after this class
2. Post-class assignment  
Write up questions for our panelists
3. Second journal: Will be discussed next week

# Next lecture:

## *The challenging case of enzymes*

