

Template for Reporting Results of Biomarker Testing of Specimens From Patients With Carcinoma of the Colon and Rectum

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For the Members of the Cancer Biomarker Reporting Workgroup, College of American Pathologists

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CAP Colon and Rectum Biomarker Template Revision History

Version: ColonBiomarkers 1.2.0.1

Summary of Changes

Microsatellite Instability

Changed MSI response from Indeterminate to Cannot be determined

Biomarker Reporting Template

Template web posting date: December 2014

Completion of the template is the responsibility of the laboratory performing the biomarker testing and/or providing the interpretation. When both testing and interpretation are performed elsewhere (eg, a reference laboratory), synoptic reporting of the results by the laboratory submitting the tissue for testing is also encouraged to ensure that all information is included in the patient's medical record and thus readily available to the treating clinical team.

COLON AND RECTUM

Select a single response unless otherwise indicated.

Note: Use of this template is optional.

+ RESULTS

+ Immunohistochemistry (IHC) Testing for Mismatch Repair (MMR) Proteins (select all that apply) (Note A)

- + ☐ MLH1
 - + ☐ Intact nuclear expression
 - + ☐ Loss of nuclear expression
 - + ☐ Cannot be determined (explain): _____
- + ☐ MSH2
 - + ☐ Intact nuclear expression
 - + ☐ Loss of nuclear expression
 - + ☐ Cannot be determined (explain): _____
- + ☐ MSH6
 - + ☐ Intact nuclear expression
 - + ☐ Loss of nuclear expression
 - + ☐ Cannot be determined (explain): _____
- + ☐ PMS2
 - + ☐ Intact nuclear expression
 - + ☐ Loss of nuclear expression
 - + ☐ Cannot be determined (explain): _____
- + ☐ Background nonneoplastic tissue/internal control with intact nuclear expression

+ IHC Interpretation

- + ☐ No loss of nuclear expression of MMR proteins: low probability of microsatellite instability-high (MSI-H)[#]
- + ☐ Loss of nuclear expression of MLH1 and PMS2: testing for methylation of the *MLH1* promoter and/or mutation of *BRAF* is indicated (the presence of a *BRAF* V600E mutation and/or *MLH1* methylation suggests that the tumor is sporadic and germline evaluation is probably not indicated; absence of both *MLH1* methylation and of *BRAF* V600E mutation suggests the possibility of Lynch syndrome, and sequencing and/or large deletion/duplication testing of germline *MLH1* may be indicated)[#]
- + ☐ Loss of nuclear expression of MSH2 and MSH6: high probability of Lynch syndrome (sequencing and/or large deletion/duplication testing of germline *MSH2* may be indicated, and, if negative, sequencing and/or large deletion/duplication testing of germline *MSH6* may be indicated)[#]
- + ☐ Loss of nuclear expression of MSH6 only: high probability of Lynch syndrome (sequencing and/or large deletion/duplication testing of germline *MSH6* may be indicated)[#]
- + ☐ Loss of nuclear expression of PMS2 only: high probability of Lynch syndrome (sequencing and/or large deletion/duplication testing of germline *PMS2* may be indicated)[#]

+ Data elements preceded by this symbol are not required.

There are exceptions to the above IHC interpretations. These results should not be considered in isolation, and clinical correlation with genetic counseling is recommended to assess the need for germline testing.

+ Microsatellite Instability (MSI) (Note A)

- + ☐ MSI – stable (MSS)
- + ☐ MSI – low (MSI-L)
 - + ☐ 1% - 29% of the National Cancer Institute (NCI) or mononucleotide markers exhibit instability
 - + ☐ 1 of the NCI or mononucleotide markers exhibit instability
 - + ☐ Other (specify): _____
- + ☐ MSI – high (MSI-H)
 - + ☐ ≥30% of the NCI or mononucleotide markers exhibit instability
 - + ☐ 2 or more of the NCI or mononucleotide markers exhibit instability
 - + ☐ Other (specify): _____
- + ☐ MSI – cannot be determined

+ Loci Testing

- + ☐ Mononucleotide panel
 - + BAT-25
 - + ☐ Stable
 - + ☐ Unstable
 - + ☐ Cannot be determined (explain): _____
 - + ☐ Not performed
 - + BAT-26
 - + ☐ Stable
 - + ☐ Unstable
 - + ☐ Cannot be determined (explain): _____
 - + ☐ Not performed
 - + NR-21
 - + ☐ Stable
 - + ☐ Unstable
 - + ☐ Cannot be determined (explain): _____
 - + ☐ Not performed
 - + NR-24
 - + ☐ Stable
 - + ☐ Unstable
 - + ☐ Cannot be determined (explain): _____
 - + ☐ Not performed
 - + Mono-27
 - + ☐ Stable
 - + ☐ Unstable
 - + ☐ Cannot be determined (explain): _____
 - + ☐ Not performed
- + ☐ NCI panel
 - + BAT-25
 - + ☐ Stable
 - + ☐ Unstable
 - + ☐ Cannot be determined (explain): _____
 - + ☐ Not performed
 - + BAT -26
 - + ☐ Stable
 - + ☐ Unstable
 - + ☐ Cannot be determined (explain): _____
 - + ☐ Not performed
 - + D2S123
 - + ☐ Stable
 - + ☐ Unstable

- + ☐ Cannot be determined (explain): _____
- + ☐ Not performed
- + D5S346
- + ☐ Stable
- + ☐ Unstable
- + ☐ Cannot be determined (explain): _____
- + ☐ Not performed
- + D17S250
- + ☐ Stable
- + ☐ Unstable
- + ☐ Cannot be determined (explain): _____
- + ☐ Not performed
- + ☐ Other (specify): _____
- + ☐ Stable
- + ☐ Unstable
- + ☐ Cannot be determined (explain): _____

+ *MLH1* Promoter Methylation Analysis (Note B)

- + ☐ *MLH1* promoter hypermethylation present
- + ☐ *MLH1* promoter hypermethylation absent
- + ☐ Cannot be determined (explain): _____

+ *KRAS* Mutational Analysis (Note C)

- + ☐ No mutation detected
- + ☐ Mutation identified (select all that apply)
 - + Codon 12
 - + ☐ Gly12Asp (GGT>GAT)
 - + ☐ Gly12Val (GGT>GTT)
 - + ☐ Gly12Cys (GGT>TGT)
 - + ☐ Gly12Ser (GGT>AGT)
 - + ☐ Gly12Ala (GGT>GCT)
 - + ☐ Gly12 Arg (GGT>CGT)
 - + ☐ Codon 12 mutation, not otherwise specified
 - + ☐ Other codon 12 mutation (specify): _____
 - + Codon 13
 - + ☐ Gly13Asp (GGC>GAC)
 - + ☐ Gly13Arg (GGC>CGC)
 - + ☐ Gly13Cys (GGC>TGC)
 - + ☐ Gly13Ala (GGC>GCC)
 - + ☐ Gly13Val (GGC>GTC)
 - + ☐ Codon 13 mutation, not otherwise specified
 - + ☐ Other codon 13 mutation (specify): _____
 - + Codon 61
 - + ☐ Gln61Leu (CAA>CTA)
 - + ☐ Gln61His (CAA>CAC)
 - + ☐ Codon 61 mutation, not otherwise specified
 - + ☐ Other codon 61 mutation (specify): _____
 - + Codon 146
 - + ☐ Ala146Thr (G436A) (GCA>ACA)
 - + ☐ Codon 146 mutation, not otherwise specified
 - + ☐ Other codon 146 mutation (specify): _____
 - + ☐ Other codon (specify): _____
- + ☐ Cannot be determined (explain): _____

+ *NRAS* Mutational Analysis (Note C)

- + ☐ No mutation detected
- + ☐ Mutation identified (select all that apply)
 - + Codon 12
 - + ☐ Gly12Asp (GGT>GAT)
 - + ☐ Gly12Val (GGT>GTT)
 - + ☐ Gly12Cys (GGT>TGT)
 - + ☐ Gly12Ser (GGT>AGT)
 - + ☐ Gly12Ala (GGT>GCT)
 - + ☐ Gly12Arg (GGT>CGT)
 - + ☐ Codon 12 mutation, not otherwise specified
 - + ☐ Other codon 12 mutation (specify): _____
 - + Codon 13
 - + ☐ Specific codon 13 mutation (specify): _____
 - + ☐ Codon 13 mutation, not otherwise specified
 - + Codon 61
 - + ☐ Gln61Lys (CAA>AAA)
 - + ☐ Gln61Arg (CAA>CGA)
 - + ☐ Codon 61 mutation, not otherwise specified
 - + ☐ Other codon 61 mutation (specify): _____
 - + ☐ Other codon (specify): _____
- + ☐ Cannot be determined (explain): _____

+ **BRAF Expression (by immunohistochemistry) (Note B)**

- + ☐ Positive cytoplasmic expression
- + ☐ Negative for cytoplasmic expression
- + ☐ Cannot be determined (explain): _____

+ **BRAF Mutational Analysis (Note B)**

- + ☐ No mutations detected
- + ☐ BRAF V600E (c.1799T>A) mutation
- + ☐ Other BRAF mutation (specify): _____
- + ☐ Cannot be determined (explain): _____

+ **PIK3CA Mutational Analysis (Note D)**

- + ☐ No mutations detected
- + ☐ Exon 9 mutation present (specify): _____
- + ☐ Exon 20 mutation present (specify): _____
- + ☐ Cannot be determined (explain): _____

+ **PTEN Expression (by immunohistochemistry) (Note E)**

- + ☐ Positive cytoplasmic and/or nuclear expression
- + ☐ Negative for cytoplasmic and nuclear expression
- + ☐ Cannot be determined (explain): _____

+ **PTEN Mutational Analysis**

- + ☐ No mutation detected
- + ☐ Exon 1-9 mutation present (specify): _____
- + ☐ Cannot be determined (explain): _____

+ **Multiparameter Gene Expression/Protein Expression Assay**

- + Specify type: _____
- + Results:
 - + ☐ Low risk
 - + ☐ Moderate risk
 - + ☐ High risk

+ Recurrence score: _____

+ METHODS

+ Dissection Method(s) (select all that apply) (Note F)

- + ☐ Laser capture microdissection
 - + Specify test name#: _____
- + ☐ Manual under microscopic observation
 - + Specify test name#: _____
- + ☐ Manual without microscopic observation
 - + Specify test name#: _____
- + ☐ Cored from block
 - + Specify test name#: _____
- + ☐ Whole tissue section (no tumor enrichment procedure employed)
 - + Specify test name#: _____

If more than 1 dissection method used, please specify which test was associated with each selected dissection method.

+ Microsatellite Instability (MSI)

+ Number of MSI markers tested (specify): _____

+ Cellularity

+ Percent tumor cells present in specimen: _____%

+ Whole Genome or Exome Sequencing

- + ☐ Whole genome sequencing (specify): _____
- + ☐ Whole exome sequencing (specify): _____

+ MLH1 Promoter Methylation

+ Testing Method

- + ☐ Methylation-specific real-time polymerase chain reaction (PCR)
- + ☐ Other (specify): _____

+ KRAS Mutational Analysis

+ Codons Assessed (select all that apply)

- + ☐ 12
- + ☐ 13
- + ☐ 61
- + ☐ 146

+ Testing Method(s) (select all that apply)

- + ☐ Direct Sanger sequencing
- + ☐ Pyrosequencing
- + ☐ High-resolution melting analysis
- + ☐ PCR, allele-specific hybridization
- + ☐ Real-time PCR
- + ☐ Other (specify): _____

Please specify in Comments section if different testing methods are used for different codons.

+ NRAS Mutational Analysis

+ Codons Assessed (select all that apply)

- + ☐ 12
- + ☐ 13

+ Data elements preceded by this symbol are not required.

+ ____ 61

+ Testing Method(s) (select all that apply)

+ ____ Direct Sanger sequencing

+ ____ Pyrosequencing

+ ____ High-resolution melting analysis

+ ____ PCR, allele-specific hybridization

+ ____ Real-time PCR

+ ____ Other (specify): _____

Please specify in Comments section if different testing methods are used for different codons.

+ **BRAF** Mutational Analysis

+ Mutations Assessed (select all that apply)

+ ____ V600E

+ ____ Other *BRAF* V600 mutation (specify): _____

+ ____ Other (specify): _____

+ Testing Method(s) (select all that apply)

+ ____ Direct Sanger sequencing

+ ____ PCR, allele-specific hybridization

+ ____ Pyrosequencing

+ ____ Real-time PCR

+ ____ Immunohistochemistry for V600E gene product

+ ____ Other (specify): _____

+ **PIK3CA** Mutational Analysis

+ Testing Method(s)

+ ____ Direct Sanger sequencing

+ ____ Other (specify): _____

+ **PTEN** Expression and Mutational Analysis

+ Testing Method(s) (select all that apply)

+ ____ Immunohistochemistry (specify antibody): _____

+ ____ In situ hybridization (specify probe): _____

+ ____ Direct Sanger sequencing

+ ____ Duplication/deletion testing (MLPA)

+ ____ Other (specify): _____

+ **COMMENT(S)**

Note: Fixative type, time to fixation (cold ischemia time), and time of fixation should be reported if applicable in this template or in the original pathology report.

Gene names should follow recommendations of The Human Genome Organisation (HUGO) Nomenclature Committee (www.genenames.org; accessed February 10, 2015).

All reported gene sequence variations should be identified following the recommendations of the Human Genome Variation Society (www.hgvs.org/mutnomen/; accessed February 10, 2015).

Explanatory Notes

A. Mismatch Repair Testing: Microsatellite instability and Immunohistochemistry

Detection of defective mismatch repair in colorectal carcinomas is important for detection of Lynch syndrome (hereditary nonpolyposis colorectal cancer syndrome [HNPCC]), which accounts for approximately 2% to 3% of all colorectal carcinomas and has clinical implications for treatment of the affected patient and family members. Microsatellite instability (MSI) testing can be used to cost-effectively screen colorectal cancer patients for possible Lynch syndrome. Patients with a microsatellite instability-high (MSI-H) phenotype that indicates mismatch repair deficiency in their cancer may have a germline mutation in one of several DNA mismatch repair (MMR) genes (eg, *MLH1*, *MSH2*, *MSH6*, or *PMS2*) or an altered *EPCAM* (*TACSTD1*) gene. After appropriate genetic counseling, patients may want to consider testing to identify the causative heritable abnormality. An MSI-H phenotype is more frequently observed in sporadic colorectal cancer (about 15% of cases) due to somatic abnormalities, usually hypermethylation of the *MLH1* gene promoter. The specificity of MSI testing can be increased by using it primarily on at-risk populations, such as colorectal cancer patients younger than 50 years, or patients with a strong family history of Lynch-associated tumors (eg, colorectal, endometrial, gastric, or upper urinary tract urothelial carcinoma),¹ but with sacrifice of sensitivity, since a sizeable minority of cases lacks these clinical characteristics.

MSI testing of tumor DNA is generally performed with at least 5 microsatellite markers, generally mononucleotide or dinucleotide repeat markers. In 1998, a National Institutes of Health consensus panel proposed that laboratories use a 5-marker panel consisting of 3 dinucleotide and 2 mononucleotide repeats for MSI testing.² Recent data suggests that dinucleotide repeats may have lower sensitivity and specificity for identifying tumors with an MSI-H phenotype. As a consequence, there has been a move towards including more mononucleotides and fewer dinucleotides in MSI testing panels. Many laboratories now use a commercially available kit for MSI testing that utilizes 5 mononucleotide markers.

MSI testing is frequently done in conjunction with immunohistochemical (IHC) testing for DNA MMR protein expression (ie, *MLH1*, *MSH2*, *MSH6*, and *PMS2* expression). If DNA MMR IHC has not been performed, this testing should be recommended for any case that shows an MSI-H phenotype, because this information will help identify the gene that is most likely to have a germline mutation (eg, a patient whose tumor shows loss of *MSH2* and *MSH6* expression, but retention of *MLH1* and *PMS2* expression, is likely to have an *MSH2* germline mutation). If the results of DNA MMR IHC and MSI testing are discordant (eg, MSI-H phenotype with normal IHC or abnormal IHC with MSS phenotype), then the laboratory should make sure that the same sample was used for MSI and IHC testing and that there was no sample mix-up. However, MSI-H may not occur in colorectal cancers of patients with germline *MSH6* mutation. Intact expression of all 4 proteins indicates that MMR enzymes tested are intact but does not entirely exclude Lynch syndrome, as approximately 5% of families may have a missense mutation (especially in *MLH1*) that can lead to a nonfunctional protein with retained antigenicity. Defects in lesser-known MMR enzymes may also lead to a similar result, but this situation is rare.

Any positive reaction in the nuclei of tumor cells is considered as intact expression (normal), and it is common for intact staining to be somewhat patchy. An interpretation of expression loss in tumor cells should be made only if a positive reaction is seen in internal control cells, such as the nuclei of stromal, inflammatory, or nonneoplastic epithelial cells. Loss of expression of *MLH1* may be due to Lynch syndrome or methylation of the *MLH1* promoter region (as occurs in sporadic MSI colorectal carcinoma). Genetic testing is ultimately required for this distinction, although a specific *BRAF* gene mutation (V600E) is present in many sporadic cases, but not familial cancers. Loss of *MSH2* expression strongly suggests Lynch syndrome. *PMS2* loss is often associated with loss of *MLH1* and is only independently meaningful if *MLH1* is intact. *MSH6* is similarly related to *MSH2*. One should also keep in mind that nucleolar staining or complete loss of *MSH6* staining has been described in colorectal cancer cases with prior radiation or chemotherapy,^{3,4} and a significant reduction of *MSH6* staining has been described in a small percentage of colorectal carcinomas with somatic mutations of the coding region microsatellites of the *MSH6* gene in *MLH1*/*PMS2*-deficient carcinomas.⁵

B. *MLH1* Promoter Hypermethylation Analysis and *BRAF* Mutational Analysis

Defective mismatch repair in sporadic colorectal cancer is most often due to inactivation of the *MLH1* gene promoter by hypermethylation (epigenetic silencing). The V600E mutation of the *BRAF* gene may be present in

up to 70% of tumors with hypermethylation of the *MLH1* promoter. In colorectal cancer, this mutation has been associated with a limited clinical response to epidermal growth factor receptor (*EGFR*) targeted therapies (cetuximab or panitumumab). Analysis for somatic mutations in the V600E hot spot in *BRAF* may also be indicated for tumors that show MSI-H, as this mutation has been found in sporadic MSI-H tumors, but not in Lynch-associated cancers with *MLH1* or *MSH2* mutations.⁶ *BRAF* V600E mutations have been described in probands with monoallelic *PMS2* mutations.⁷ Direct testing of *MLH1* promoter hypermethylation and/or the use of *BRAF* V600E mutational analysis prior to germline genetic testing in patients with MSI-H tumors and loss of *MLH1* by IHC may be a cost-effective means of identifying patients with sporadic tumors for whom further testing is not indicated.⁸

C. RAS Mutational Analysis

The presence of a *KRAS* mutation has been shown to be associated with lack of clinical response to therapies targeted at *EGFR*, such as cetuximab⁹ and panitumumab.¹⁰ While clinical guidelines for *KRAS* mutational analysis are evolving, current provisional recommendations from the American Society of Clinical Oncology are that all patients with stage IV colorectal carcinoma who are candidates for anti-*EGFR* antibody therapy should have their tumor tested for *KRAS* mutations.¹¹ Anti-*EGFR* antibody therapy is not recommended for patients whose tumors show mutations in *KRAS* codon 12, 13, or 61, but data on codon 146 are currently insufficient. A recent study has shown that *NRAS* mutation, like *KRAS* mutation, has influence on response to anti-*EGFR* therapy.¹² Although more studies are needed, these findings may lead to broad *KRAS* and *NRAS* panels to include codons 12, 13, 61, and 146 of both genes.

D. PIK3CA Mutational Analysis

PIK3CA mutations activate the *PI3K-PTEN-AKT* pathway that is downstream from both the *EGFR* and the *RAS-RAF-MAPK* pathways. *PIK3CA* mutation and subsequent activation of the *AKT* pathway has been shown to play an important role in colorectal carcinogenesis and have been associated with *KRAS* mutation¹³ and microsatellite instability.¹⁴ *PIK3CA* mutation has further been associated with poor survival in resectable stage I to III colon cancer, with the adverse effect of *PIK3CA* mutation potentially limited to patients with *KRAS* wild-type tumors.¹⁵ *PIK3CA* mutations have been associated with resistance to anti-*EGFR* therapy in several studies,^{16,17} but not in others.¹⁸ The reasons for the discrepancy are not clear. Mutations of exons 1, 9, and 20 of the *PIK3CA* gene represent >95% of known mutations.

A European consortium recently suggested that only *PIK3CA* exon 20 mutations are associated with a lack of cetuximab activity in *KRAS* wild-type tumors and with a shorter median progression-free survival and overall survival.¹⁷ By contrast, exon 9 *PIK3CA* mutations are associated with *KRAS* mutations and do not have an independent effect on cetuximab efficacy.¹⁷ More studies are needed to establish the prognostic and predictive roles of *PIK3CA* exon-9 and exon-20 mutations.

E. PTEN Mutational Analysis

The role of *PTEN* loss in colorectal cancer prognosis and therapy is unclear. It has been suggested that loss of *PTEN* expression, as determined by immunohistochemistry, is associated with lack of benefit from cetuximab in metastatic colorectal cancer.¹⁹⁻²² Loss of *PTEN* has been found to co-occur with *KRAS*, *BRAF*, and *PIK3CA* mutations.^{19,22} The recorded frequency of loss of *PTEN* expression varies from 19% to 36%, with some studies reporting an effect on response rate and survival, whereas others found an effect only on progression-free or overall survival. Moreover, data on the loss of *PTEN* expression are not concordant in primary and metastatic tissues.²¹ There is currently no standardized method for *PTEN* expression analysis by immunohistochemistry.

F. Dissection Method

Please denote the manner in which the tissue was dissected and specify the biomarker test only if different dissection methods are used for different tests.

1. Laser capture microdissection (LCM): Use of a laser-equipped microscope to isolate and retrieve specific cells of interest from a histopathologic region of interest.
2. Manual under microscopic observation: hematoxylin and eosin (H&E) slide is examined under a light microscope and marked by a pathologist for subsequent tumor dissection and retrieval.

3. Manual without microscopic observation: H&E slide is examined without a microscope and marked by a pathologist for subsequent tumor dissection and retrieval.
4. Cored from block: Area of interest is cored from a paraffin-embedded tissue block.
5. Whole tissue section: No tumor enrichment procedure employed for tissue retrieval.

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