

Protocol for the Examination of Specimens From Patients With Gastrointestinal Stromal Tumor (GIST)

Version: GIST 4.0.1.0 Protocol Posting Date: June 2017

Includes pTNM requirements from the 8th Edition, AJCC Staging Manual

For accreditation purposes, this protocol should be used for the following procedures and tumor types:

Procedure	Description
Resection	
Tumor Type	Description
Gastrointestinal stromal tumor	

This protocol is NOT required for accreditation purposes for the following:

Procedure				
Biopsy				
Local excision				
Metastasectomy				
Primary resection specimen with no residual cancer (eg, following neoadjuvant therapy)				
Cytologic specimens				

Authors

Javier A. Laurini, MD*; Charles D. Blanke, MD; Kumarasen Cooper, MBChB, DPhil, FRCPath; George D. Demetri, MD; Ronald P. Dematteo, MD; Christopher D.M. Fletcher, MD, FRCPath; John R. Goldblum, MD; Thomas Krausz, MD, FRCPath; Jerzy Lasota, MD, PhD; Alexander Lazar, MD, PhD; Robert G. Maki, MD, PhD; Markku Miettinen, MD, PhD; Amy Noffsinger, MD; Jordan E. Olson, MD; Brian P. Rubin, MD, PhD; Mary K. Washington, MD, PhD

With guidance from the CAP Cancer and CAP Pathology Electronic Reporting Committees.

^{*} Denotes primary author. All other contributing authors are listed alphabetically.

Accreditation Requirements

This protocol can be utilized for a variety of procedures and tumor types for clinical care purposes. For accreditation purposes, only the definitive primary cancer resection specimen is required to have the core and conditional data elements reported in a synoptic format.

- <u>Core data elements</u> are required in reports to adequately describe appropriate malignancies. For accreditation purposes, essential data elements must be reported in all instances, even if the response is "not applicable" or "cannot be determined."
- <u>Conditional data elements</u> are only required to be reported if applicable as delineated in the protocol. For
 instance, the total number of lymph nodes examined must be reported, but only if nodes are present in the
 specimen.
- Optional data elements are identified with "+" and although not required for CAP accreditation purposes, may be considered for reporting as determined by local practice standards.

The use of this protocol is not required for recurrent tumors or for metastatic tumors that are resected at a different time than the primary tumor. Use of this protocol is also not required for pathology reviews performed at a second institution (ie, secondary consultation, second opinion, or review of outside case at second institution).

Synoptic Reporting

All core and conditionally required data elements outlined on the surgical case summary from this cancer protocol must be displayed in synoptic report format. Synoptic format is defined as:

- Data element: followed by its answer (response), outline format without the paired "Data element: Response" format is NOT considered synoptic.
- The data element should be represented in the report as it is listed in the case summary. The response for any data element may be modified from those listed in the case summary, including "Cannot be determined" if appropriate.
- Each diagnostic parameter pair (Data element: Response) is listed on a separate line or in a tabular format to achieve visual separation. The following exceptions are allowed to be listed on one line:
 - o Anatomic site or specimen, laterality, and procedure
 - Pathologic Stage Classification (pTNM) elements
 - Negative margins, as long as all negative margins are specifically enumerated where applicable
- The synoptic portion of the report can appear in the diagnosis section of the pathology report, at the end of the report or in a separate section, but all Data element: Responses must be listed together in one location Organizations and pathologists may choose to list the required elements in any order, use additional methods in order to enhance or achieve visual separation, or add optional items within the synoptic report. The report may have required elements in a summary format elsewhere in the report IN ADDITION TO but not as replacement for the synoptic report ie, all required elements must be in the synoptic portion of the report in the format defined above.

CAP Laboratory Accreditation Program Protocol Required Use Date: March 2018

CAP GIST Protocol Summary of Changes

Version 4.0.1.0

The following data elements were modified:

Regional Lymph Nodes pN: Conditionally required if nodes are present pN0 changed definition to AJCC approved "No regional lymph node metastasis"

___ Cannot be determined

Surgical Pathology Cancer Case Summary

Protocol posting date: June 2017

GASTROINTESTINAL STROMAL TUMOR (GIST): Biopsy

Note: This case summary is recommended for reporting biopsy specimens, but is not required for accreditation purposes.

Select a single response unless otherwise indicated.
Procedure Core needle biopsy Endoscopic biopsy Other (specify): Not specified
Tumor Site (Note A) Specify: Not specified
Histologic Type Gastrointestinal stromal tumor, spindle cell type Gastrointestinal stromal tumor, epithelioid type Gastrointestinal stromal tumor, mixed Gastrointestinal stromal tumor, other (specify):
Mitotic Rate Specify: /5 mm² Cannot be determined (explain):
Note: The required total count of mitoses is per 5 mm ² on the glass slide section. With the use of older model microscopes, 50 HPF is equivalent to 5 mm ² . Most modern microscopes with wider 40X lenses/fields require approximately 20 to 25 HPF to encompass 5 mm ² . If necessary please measure field of view to accurately determine actual number of fields required to be counted on individual microscopes to encompass 5 mm ² .
+ Necrosis + Not identified + Present
Histologic Grade (Note B) G1: Low grade; mitotic rate ≤5/5 mm² G2: High grade; mitotic rate >5/5 mm² GX: Grade cannot be assessed
Risk Assessment (Note C) None Very low risk Low risk Moderate risk High risk Overtly metastatic

Data elements preceded by this symbol are not required for accreditation purposes. These optional elements may be clinically important but are not yet validated or regularly used in patient management.

Biopsy

+ Additional Pathologic Findings + Specify:
Ancillary Studies (Note E)
Note: For molecular genetic and further immunohistochemical study reporting, the CAP GIST Biomarker Template should be used. Pending biomarker studies should be listed in the Comments section of this report.
Immunohistochemical Studies (select all that apply) KIT (CD117) Positive Negative DOG1 (ANO1) Positive Negative Negative Other (specify): Pending
Not performed
+ Molecular Genetic Studies (eg, KIT, PDGFRA, BRAF, SDHA/B/C/D, or NF1 mutational analysis) + Submitted for analysis; results pending + Performed, see separate report: + Performed + Specify method(s) and results: + Not performed
+ Prebiopsy Treatment + No known prebiopsy therapy + Systemic therapy performed (specify type): + Therapy performed, type not specified + Not specified
Treatment Effect (Note F) No known presurgical therapy Not identified Present + Specify percentage of viable tumor:% Cannot be determined
+ Comment(s)

⁺ Data elements preceded by this symbol are not required for accreditation purposes. These optional elements may be clinically important but are not yet validated or regularly used in patient management.

Surgical Pathology Cancer Case Summary

Protocol posting date: June 2017

GASTROINTESTINAL STROMAL TUMOR (GIST): Resection

Note: This case summary is recommended for reporting local excision specimens, but is not required for accreditation purposes.

Select a single response unless otherwise indicated.

Procedure Local excision Resection Specify type (eg, partial gastrectomy): Metastasectomy Other (specify): Not specified
Tumor Site Specify (if known): Not specified
Tumor Size Greatest dimension (centimeters): cm + Additional dimensions (centimeters): x cm Cannot be determined (explain):
Tumor Focality Unifocal Multifocal Specify number of tumors: Specify size of tumors:
Histologic Type Gastrointestinal stromal tumor, spindle cell type Gastrointestinal stromal tumor, epithelioid type Gastrointestinal stromal tumor, mixed Gastrointestinal stromal tumor, other (specify):
Mitotic Rate Specify: /5 mm² Cannot be determined (explain):
Note: The required total count of mitoses is per 5 mm ² on the glass slide section. With the use of older model microscopes, 50 HPF is equivalent to 5 mm ² . Most modern microscopes with wider 40X lenses/fields require approximately 20 to 25 HPF to encompass 5 mm ² . If necessary, please measure field of view to accurately determine actual number of fields required to be counted on individual microscopes to encompass 5 mm ² .
+ Necrosis + Not identified + Present

Data elements preceded by this symbol are not required for accreditation purposes. These optional elements may be clinically important but are not yet validated or regularly used in patient management.

Histologic Grade (Note B) G1: Low grade; mitotic rate ≤5/5 mm² G2: High grade; mitotic rate >5/5 mm² GX: Grade cannot be assessed
Risk Assessment (Note C) None Very low risk Low risk Moderate risk High risk Overtly malignant/metastatic Cannot be determined
Margins Cannot be assessed Uninvolved by GIST Distance of tumor from closest margin (millimeters or centimeters): mm or cm Specify margin (if known): Involved by GIST Specify margin(s) (if known):
Regional Lymph Nodes (Note D)
No lymph nodes submitted or found
Lymph Node Examination (required only if lymph nodes are present in specimen)
Number of Lymph Nodes Involved: Number cannot be determined (explain):
Number of Lymph Nodes Examined: Number cannot be determined (explain):
Pathologic Stage Classification (pTNM, AJCC 8 th Edition) (Note G) Note: Reporting of pT, pN, and (when applicable) pM categories is based on information available to the pathologist at the time the report is issued. Only the applicable T, N, or M category is required for reporting; their definitions need not be included in the report. The categories (with modifiers when applicable) can be listed on 1 line or more than 1 line. TNM Descriptors (required only if applicable) (select all that apply) m (multiple) r (recurrent) y (posttreatment)
Primary Tumor (pT) pTX: Primary tumor cannot be assessed pT0: No evidence of primary tumor pT1: Tumor 2 cm or less pT2: Tumor more than 2 cm but not more than 5 cm pT3: Tumor more than 5 cm but not more than 10 cm pT4: Tumor more than 10 cm in greatest dimension
Regional Lymph Nodes (pN) (Note D) (required only if lymph nodes submitted in this case)# pN0: No regional lymph node metastasis pN1: Regional lymph node metastasis # When no lymph nodes are present (as is often the case with resection for GIST), the pathologic 'N' category is not assigned

+ Data elements preceded by this symbol are not required for accreditation purposes. These optional elements may be clinically important but are not yet validated or regularly used in patient management.

(pNX is not used for GIST) and should not be reported.

Distant Metastasis (pM) (Note D) (required only if confirmed pathologically in this case)
pM1: Distant metastasis
Specify site(s), if known:
+ Additional Pathologic Findings
+ Specify:
Ancillary Studies (Note E)
Note: For molecular genetic and further immunohistochemical study reporting, the CAP GIST Biomarker Template should be used. Pending biomarker studies should be listed in the Comments section of this report.
Immunohistochemical Studies
KIT (CD117)
Positive
Negative
DOG1 (ANO1)
Positive
Negative Other (specify):
Pending
Not performed
Not perioritied
+ Molecular Genetic Studies (eg, KIT, PDGFRA, BRAF, SDHA/B/C/D, or NF1 mutational analysis)
+ Submitted for analysis; results pending
+ Performed, see separate report:
+ Performed
+ Specify method(s) and results:
+ Not performed
+ Preresection Treatment (select all that apply)
+ No known preresection therapy
+ Previous biopsy or surgery (specify):
+ Systemic therapy performed (specify type):
+ Therapy performed, type not specified + Not specified
+ Not specified
Treatment Effect (Note F)
No known presurgical therapy
Not identified
Present
+ Specify percentage of viable tumor:%
Cannot be determined
+ Comment(s)

⁺ Data elements preceded by this symbol are not required for accreditation purposes. These optional elements may be clinically important but are not yet validated or regularly used in patient management.

Explanatory Notes

A. Location

Gastrointestinal stromal tumors may occur anywhere along the entire length of the tubal gut, as well as in extravisceral locations, which include the omentum, mesentery, pelvis, and retroperitoneum.¹⁻³ Typically, they arise from the wall of the gut and extend inward toward the mucosa, outward toward the serosa, or in both directions. Lesions that involve the wall of the gastrointestinal (GI) tract frequently cause ulceration of the overlying mucosa. Infrequently, lesions invade through the muscularis mucosae to involve the mucosae. Mucosal invasion is an adverse prognostic factor in numerous studies. Because the anatomic location along the GI tract affects prognosis, with location in the stomach having a more favorable prognosis, it is very important to specify anatomic location as precisely as possible.⁴

B. Histologic Grade

Histologic grading, an important component of soft tissue sarcoma staging, is not well suited to GISTs, because most of these tumors have low or relatively low mitotic rates below the thresholds used for grading of soft tissue tumors, and because GISTs often manifest aggressive features with mitotic rates below the thresholds used for soft tissue tumor grading (the lowest tier of mitotic rates for soft tissue sarcomas being 10 mitoses per 10 HPF). In GIST staging, the grade is determined entirely by mitotic activity.

GX: Grade cannot be assessed

G1: Low grade; mitotic rate ≤5/5 mm² G2: High grade; mitotic rate >5/5 mm²

The mitotic count should be initiated on an area that on screening magnification shows the highest level of mitotic activity and be performed as consecutive high-power fields (HPF). Stringent criteria should be applied when counting mitotic figures; pyknotic, dyskaryotic or apoptotic nuclei should not be regarded as mitosis.

Note: The required total count of mitoses is per 5 mm² on the glass slide section. With the use of older model microscopes, 50 HPF is equivalent to 5 mm². Most modern microscopes with wider 40X lenses/fields require approximately 20 to 25 HPF to encompass 5 mm². If necessary, please measure field of view to accurately determine actual number of fields required to be counted on individual microscopes to encompass 5 mm².

C. Risk Assessment

Because GISTs can recur many years after initial excision, we now regard most GISTs as having at least some potential for distant metastasis. This concept was originally the result of a National Cancer Institute-sponsored consensus conference that was held in 2002.¹ More specific data generated by large follow-up studies refined the biologic potential assessment.⁴⁴8 Criteria obtained from those data were adopted in a National Cancer Care Network (NCCN) Task Force report on GIST.⁴ We have adopted the criteria for risk stratification, as indicated in Table 1.⁴⁴8 The scheme includes anatomic site as a factor, because small bowel GISTs carry a higher risk of progression than gastric GISTs of similar size and mitotic activity. For anatomic sites not listed in this table, such as esophagus, mesentery, and peritoneum, or in the case of "insufficient data," it is best to use risk criteria for jejunum/ileum.

Tumor Parameters		Risk of Progressive Disease# (%)			
Mitotic Rate	Size	Gastric	Duodenum	Jejunum/Ileum	Rectum
≤5 per 5 mm²	≤2 cm	None (0%)	None (0%)	None (0%)	None (0%)
	>2 - ≤5 cm	Very low (1.9%)	Low (8.3%)	Low (4.3%)	Low (8.5%)
	>5 - ≤10 cm	Low (3.6%)	(Insufficient data)	Moderate (24%)	(Insufficient data)
	>10 cm	Moderate (10%)	High (34%)	High (52%)	High (57%)
>5 per 5 mm²	≤2 cm	None##	(Insufficient data)	High##	High (54%)
	>2 - ≤5 cm	Moderate (16%)	High (50%)	High (73%)	High (52%)
	>5 - ≤10 cm	High (55%)	(Insufficient data)	High (85%)	(Insufficient data)
	>10 cm	High (86%)	High (86%)	High (90%)	High (71%)

Adapted with permission from Miettinen and Lasota. 7 Copyright 2006 by Elsevier.

Data based on long-term follow-up of 1055 gastric, 629 small intestinal, 144 duodenal, and 111 rectal GISTs from the pre-imatinib era.^{4-6,8}

Note: See Note B, "Histologic Grade," regarding the number of high power fields to evaluate.

D. Metastasis

Gastrointestinal stromal tumors generally metastasize to a very limited subset of anatomic sites. They rarely metastasize to lymph nodes, which is important to note because lymphadenectomy is unnecessary except in rare circumstances when an enlarged or otherwise suspicious lymph node is encountered. Gastrointestinal stromal tumors metastasize predominantly to the liver or to the peritoneal surfaces, where there can be disseminated intra-abdominal disease presenting as innumerable metastatic nodules. Very rarely, GISTs metastasize to the lungs. This situation is associated with rectal location or very advanced disease. Metastasis to bone has also been documented, but it is very rare.

E. Ancillary Studies

Immunohistochemistry

Because of the advent of small-molecule kinase inhibitor therapy in the treatment of GIST (see the following), it has become imperative to distinguish GIST from its histologic mimics, mainly leiomyoma, leiomyosarcoma, schwannoma, and desmoid fibromatosis. ^{10,11} Immunohistochemistry is instrumental in the workup of GIST. Approximately 95% of GISTs are immunoreactive for KIT (CD117). ¹² Most KIT-negative GISTs are gastric or extra-visceral GISTs that are positive for the *platelet-derived growth factor receptor A (PDGFRA)* mutation. ¹³ KIT immunoreactivity is usually strong and diffuse but can be more focal in unusual cases (Figure 1, A and B). It is not unusual for GISTs to exhibit dot-like perinuclear staining (Figure 1, C), while less commonly, some cases exhibit membranous staining (Figure 1, D). These patterns do not clearly correlate with mutation type or response to therapy. Approximately 70% of GISTs are positive for CD34, 30% to 40% are positive for smooth muscle actin, 5% are positive for S100 (usually focal), 5% are positive for desmin (usually focal), and 1% to 2% are positive for keratin (weak/focal). ¹

[#] Defined as metastasis or tumor-related death.

^{##} Denotes small number of cases.

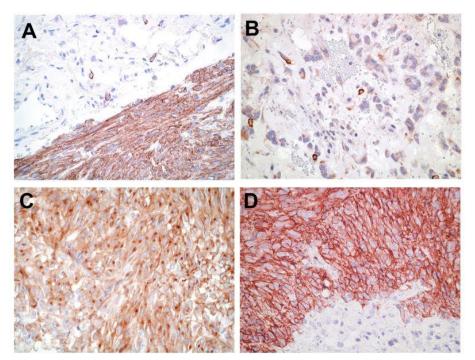
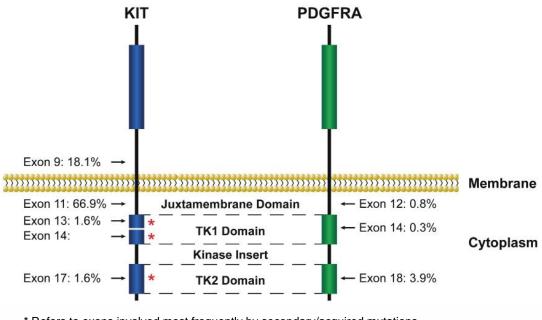


Figure 1. Patterns of KIT staining in gastrointestinal stromal tumor (GIST). A. Diffuse and strong immunoreactivity in a typical GIST. B. Focal and weak pattern in an epithelioid gastric GIST with a *PDGFRA* mutation. C. Dot-like perinuclear staining. D. Membranous pattern. (Original magnification X400.)

Molecular Analysis

Approximately 85% of GISTs possess activating mutations in the *KIT* gene, whereas another 10% have activating mutations in the *PDGFRA* gene. 14-17 These mutations result in virtually full-length KIT proteins that exhibit ligand-independent activation. *KIT* and *PDGFRA* each contain 21 exons. However, mutations cluster within "hotspots": exons 9, 11, 13, and 17 in *KIT*, and exons 12, 14, and 18 in *PDGFRA* (Figure 2). About 5% to 10% of GISTs appear to be negative for both *KIT* and *PDGFRA* mutations. The most recent NCCN Task Force on GIST strongly encourages that *KIT* and *PDGFRA* mutational analysis be performed if imatinib therapy is begun for unresectable or metastatic disease and that mutational analysis be considered for patients with primary disease, particularly those with high-risk tumors. *KIT* and *PDGFRA* mutation status can be determined easily from paraffin-embedded tissue. Secondary or acquired mutations can be associated with development of tumor resistance in the setting of long-term imatinib mesylate treatment. These are usually point mutations that occur most commonly in *KIT* exons 13, 14, and 17.18 The clinical utility of these mutations is an evolving concept, but it is important not to confuse them with the primary or initial mutation in GIST.



^{*} Refers to exons involved most frequently by secondary/acquired mutations.

Figure 2. Locations and frequency of activating *KIT* and *PDGFRA* mutations in GIST. Adapted with permission from Heinrich et al.¹⁴ Copyright 2003 by the American Society of Clinical Oncology. All rights reserved.

KIT and *PDGFRA* are excellent targets for small-molecule tyrosine kinase inhibitors, and two compounds of this class, imatinib mesylate (Gleevec, Novartis Pharmaceuticals, Basel, Switzerland) and sunitinib malate (Sutent, Pfizer Pharmaceuticals, New York, New York), have shown efficacy in clinical trials and have been approved by the US Food and Drug Administration for the treatment of GIST.^{9,19,20} Because different treatments may have more efficacy in genetic subsets of GIST, the molecular era of GIST analysis has arrived, and oncologists may want to know the mutation status of each GIST, because this may impact which drug each patient should receive.^{14,21} Secondary resistance mutations may also affect drug selection as their significance is further defined.

F. Treatment Effect

Gastrointestinal stromal tumors respond well to the newer targeted systemic therapies, imatinib mesylate and sunitib malate. The types of treatment effects that have been seen are hypocellularity, myxoid stroma, fibrosis, and necrosis. Nests of viable tumor cells are virtually always seen. Because all of these histologic features can be seen in untreated GISTs, it is not possible to know whether they are due to treatment or not. As a practical compromise, we think it is best to report the percentage of viable tumor after treatment.

G. TNM and Stage Groupings

The American Joint Committee on Cancer (AJCC) and International Union Against Cancer (UICC) GIST staging system is recommended.²² The staging system should **not** be applied to pediatric GIST, familial GIST (germline mutant *KIT* or *PDGFRA*) or syndromic GIST (GISTs arising in the setting of neurofibromatosis type 1, Carney triad, or Carney dyad also known as Carney-Stratakis syndrome).

TNM Descriptors

For identification of special cases of TNM or pTNM classifications, the "m" suffix and "y" and "r" prefixes are used. Although they do not affect the stage grouping, they indicate cases needing separate analysis.

The "m" suffix indicates the presence of multiple primary tumors in a single site and is recorded in parentheses: pT(m)NM.

<u>The "y" prefix</u> indicates those cases in which classification is performed during or after initial multimodality therapy (ie, neoadjuvant chemotherapy, radiation therapy, or both chemotherapy and radiation therapy). The cTNM or pTNM category is identified by a "y" prefix. The ycTNM or ypTNM categorizes the extent of tumor actually present at the time of that examination. The "y" categorization is not an estimate of tumor before multimodality therapy (ie, before initiation of neoadjuvant therapy).

The "r" prefix indicates a recurrent tumor when staged after a documented disease-free interval and is identified by the "r" prefix: rTNM.

T Category Considerations

In the case of ruptured tumors, estimates of tumor size can be obtained from radiologic data, if available.

N Category Considerations

Regional nodal metastasis is extremely rare in GIST, and there is no routine indication for lymph node biopsy or lymph node dissection. In the absence of information on regional lymph node status, N0/pN0 is appropriate; NX should not be used.

M Category Considerations

Most GISTs metastasize to intra-abdominal soft tissues, liver, or both. Intra-abdominal metastasis refers to tumor involvement in the abdominal cavity away from the primary mass. Such metastasis is usually to the serosal surfaces of the abdomen, pelvis, and retroperitoneum. Multiple primary tumors can be seen in the setting of neurofibromatosis type 1 or familial GIST syndrome and should not be considered intra-abdominal metastasis. Rare cases of multiple independent GISTs at different GI locations have been reported. In the absence of a primary gastrointestinal GIST, solitary omental, mesenteric, pelvic, or retroperitoneal GISTs should be considered primary tumors because extra-gastrointestinal GISTs have been described. Liver metastasis implies the presence of metastatic tumor inside the liver parenchyma as 1 or more nodules. Adherence to liver capsule, even if extensive, as sometimes seen in gastric GISTs, should not be considered liver metastasis.

Stage Groupings:

Although T, N and M definitions are identical for all GISTs, separate stage grouping schemes are provided for gastric and small intestinal tumors. Primary omental GISTs should follow the gastric GIST staging group scheme. GISTs arising in other locations (ie, mesentery, esophagus, colon, and rectum) are to follow the small intestinal group staging scheme.

References

- 1. Fletcher CD, Berman JJ, Corless C, et al. Diagnosis of gastrointestinal stromal tumors: a consensus approach. *Hum Pathol.* 2002;33(5):459-465.
- 2. Miettinen M, Lasota J. Gastrointestinal stromal tumors: definition, clinical, histological, immunohistochemical, and molecular genetic features and differential diagnosis. *Virchows Arch.* 2001;438(1):1-12.
- 3. Reith JD, Goldblum JR, Lyles RH, Weiss SW. Extragastrointestinal (soft tissue) stromal tumors: an analysis of 48 cases with emphasis on histologic predictors of outcome. *Mod Pathol.* 2000;13(5):577-585.
- 4. Miettinen M, Sobin LH, Lasota J. Gastrointestinal stromal tumors of the stomach: a clinicopathologic, immunohistochemical, and molecular genetic study of 1765 cases with long-term follow-up. *Am J Surg Pathol.* 2005;29(1):52-68.
- 5. Miettinen M, Furlong M, Sarlomo-Rikala M, Burke A, Sobin LH, Lasota J. Gastrointestinal stromal tumors, intramural leiomyomas, and leiomyosarcomas in the rectum and anus: a clinicopathologic, immunohistochemical, and molecular genetic study of 144 cases. *Am J Surg Pathol.* 2001;25(9):1121-1133.
- 6. Miettinen M, Kopczynski J, Makhlouf HR, et al. Gastrointestinal stromal tumors, intramural leiomyomas, and leiomyosarcomas in the duodenum: a clinicopathologic, immunohistochemical, and molecular genetic study of 167 cases. *Am J Surg Pathol.* 2003;27(5):625-641.
- 7. Miettinen M, Lasota J. Gastrointestinal stromal tumors: pathology and prognosis at different sites. *Semin Diagn Pathol.* 2006;23(2):70-83.
- 8. Miettinen M, Makhlouf H, Sobin LH, Lasota J. Gastrointestinal stromal tumors of the jejunum and ileum: a clinicopathologic, immunohistochemical, and molecular genetic study of 906 cases before imatinib with long-term follow-up. *Am J Surg Pathol.* 2006;30(4):477-489.
- 9. Demetri GD, Benjamin RS, Blanke CD, et al; NCCN Task Force. NCCN Task Force report: management of patients with gastrointestinal stromal tumor (GIST)--update of the NCCN clinical practice guidelines. *J Natl Compr Canc Netw.* 2007;5(Suppl 2):S1-S29.
- 10. Hornick JL, Fletcher CD. Immunohistochemical staining for KIT (CD117) in soft tissue sarcomas is very limited in distribution. *Am J Clin Pathol*. 2002;117(2):188-193.
- 11. Miettinen M, Sobin LH, Sarlomo-Rikala M. Immunohistochemical spectrum of GISTs at different sites and their differential diagnosis with a reference to CD117 (KIT). *Mod Pathol*. 2000;13(10):1134-1142.
- 12. Sarlomo-Rikala M, Kovatich AJ, Barusevicius A, Miettinen M. CD117: a sensitive marker for gastrointestinal stromal tumors that is more specific than CD34. *Mod Pathol.* 1998;11(8):728-734.
- 13. Medeiros F, Corless CL, Duensing A, et al. KIT-negative gastrointestinal stromal tumors: proof of concept and therapeutic implications. *Am J Surg Pathol.* 2004;28(7):889-894.
- 14. Heinrich MC, Corless CL, Demetri GD, et al. Kinase mutations and imatinib response in patients with metastatic gastrointestinal stromal tumor. *J Clin Oncol*. 2003;21(23):4342-4349.
- 15. Heinrich MC, Corless CL, Duensing A, et al. PDGFRA activating mutations in gastrointestinal stromal tumors. *Science*. 2003;299(5607):708-710.
- 16. Hirota S, Isozaki K, Moriyama Y, et al. Gain-of-function mutations of c-kit in human gastrointestinal stromal tumors. *Science*. 1998;279(5350):577-580.
- 17. Rubin BP, Singer S, Tsao C, et al. KIT activation is a ubiquitous feature of gastrointestinal stromal tumors. *Cancer Res.* 2001;61(22):8118-8121.
- 18. Heinrich MC, Corless CL, Blanke CD, et al. Molecular correlates of imatinib resistance in gastrointestinal stromal tumors. *J Clin Oncol.* 2006;24(29):4764-4774.
- 19. Demetri GD. Targeting the molecular pathophysiology of gastrointestinal stromal tumors with imatinib: mechanisms, successes, and challenges to rational drug development. *Hematol Oncol Clin North Am.* 2002;16(5):1115-1124.
- 20. Demetri GD, van Oosterom AT, Garrett CR, et al. Efficacy and safety of sunitinib in patients with advanced gastrointestinal stromal tumour after failure of imatinib: a randomised controlled trial. *Lancet*. 2006;368(9544):1329-1338.
- 21. Corless CL, Schroeder A, Griffith D, et al. PDGFRA mutations in gastrointestinal stromal tumors: frequency, spectrum and in vitro sensitivity to imatinib. *J Clin Oncol.* 2005;23(23):5357-5364.
- 22. Amin MB, Edge SB, Greene FL, et al, eds. *AJCC Cancer Staging Manual*. 8th ed. New York, NY: Springer; 2017.