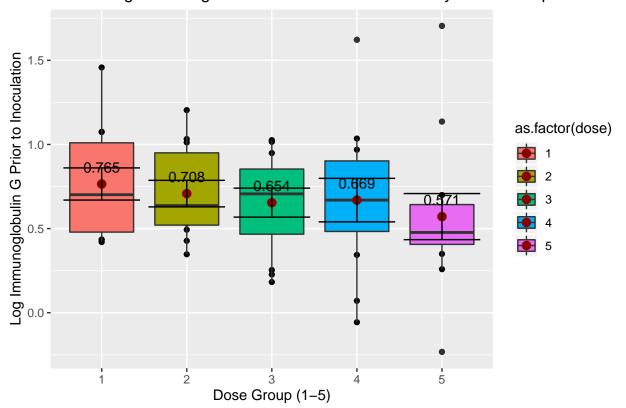
# Vaccine Project

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3/2/2021

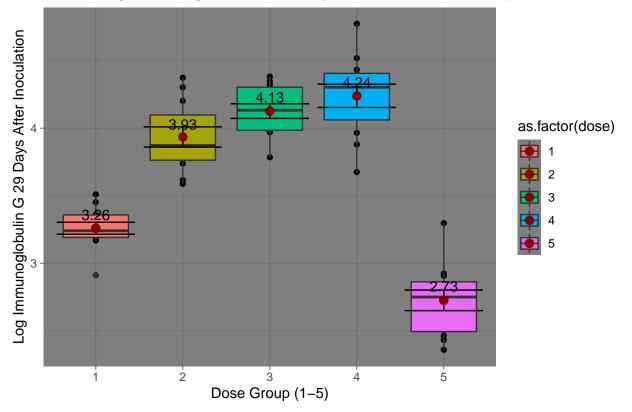
## No summary function supplied, defaulting to `mean\_se()

## Plot of Log Immunoglobulin G Prior to Inoculation By Dose Group



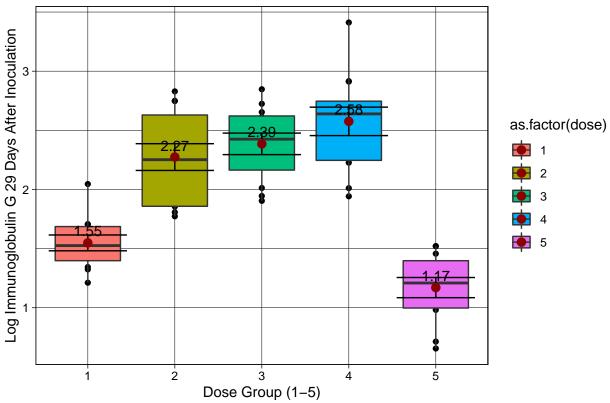
## No summary function supplied, defaulting to `mean\_se()

### Plot of Log Immunoglobulin G 29 Days After to Inoculation By Dose Group



## No summary function supplied, defaulting to `mean\_se()

## Plot of Virus Neutralizing Titers By Dose Group



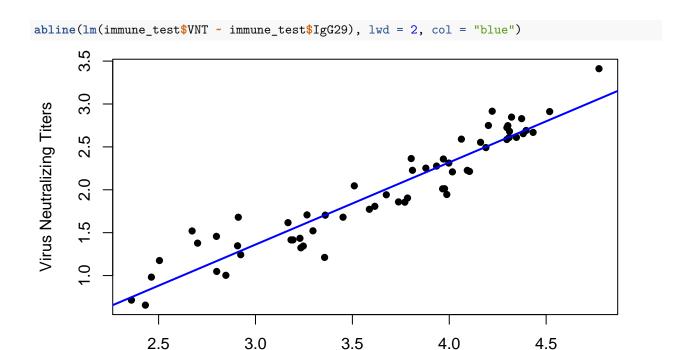
```
var = c("IgGpre", "IgG29", "VNT")
outlist = c()
fn_list <- function(x){
   outlist = c(outlist, mean(x), sd(x), IQR(x), median(x))
}
l = tapply(immune_test[, var[1]], immune_test[, 4], fn_list)
IgGPRE_df = as.data.frame(1)

for (i in 1:NROW(IgGPRE_df)){
   rownames(IgGPRE_df)[i] <- pasteO("Dose Group: ", i)
}
colnames(IgGPRE_df) <- c("MEAN SD IQR MEDIAN")
IgGPRE_df</pre>
```

```
IQR
##
                   MEAN
                               SD
## Dose Group: 1 0.7650815, 0.3308874, 0.5304835, 0.7014505
## Dose Group: 2 0.7079494, 0.2729822, 0.4294729, 0.6368983
## Dose Group: 3 0.6543556, 0.2984409, 0.3871994, 0.7063217
## Dose Group: 4 0.6692356, 0.4479995, 0.4191508, 0.6692866
## Dose Group: 5 0.5712575, 0.4746484, 0.2365727, 0.4771336
s = tapply(immune_test[, var[2]], immune_test[, 4], fn_list)
IgG29_df = as.data.frame(s)
for (i in 1:NROW(IgG29 df)){
 rownames(IgG29_df)[i] <- paste0("Dose Group: ", i)</pre>
                                             IQR MEDIAN")
colnames(IgG29_df) <- c("MEAN</pre>
                                     SD
```

```
IgG29_df
## Dose Group: 1 3.2591991, 0.1539724, 0.1665585, 3.2404413
## Dose Group: 2 3.9347705, 0.2598049, 0.3348957, 3.8722545
## Dose Group: 3 4.1264243, 0.1874612, 0.3191539, 4.1329675
## Dose Group: 4 4.2393304, 0.2988536, 0.3444530, 4.3030041
## Dose Group: 5 2.7254719, 0.2638021, 0.3682269, 2.7497923
t = tapply(immune_test[, var[3]], immune_test[, 4], fn_list)
VNT_df = as.data.frame(t)
for (i in 1:NROW(VNT df)){
  rownames(VNT_df)[i] <- paste0("Dose Group: ", i)</pre>
colnames(VNT_df) <- c("MEAN</pre>
                                     SD
                                                            MEDIAN")
                                                 IQR
VNT_df
##
                    MEAN
                                 SD
                                             IQR
## Dose Group: 1 1.5482880, 0.2316119, 0.2883617, 1.5255049
## Dose Group: 2 2.2738550, 0.3919578, 0.7718572, 2.2517141
## Dose Group: 3 2.3854777, 0.3149157, 0.4578352, 2.4250514
## Dose Group: 4 2.5756446, 0.4165343, 0.5004806, 2.6390034
## Dose Group: 5 1.1703048, 0.2953554, 0.4001061, 1.2091465
plot(immune_test$IgGpre, immune_test$IgG29, pch = 16, xlab = "Log Immunoglobulin G Prior to Inoculation
     ylab = "Log Immunoglobulin G 29 Days After Inoculation")
abline(lm(immune_test$IgG29 ~ immune_test$IgGpre), lwd = 2, col = "firebrick")
Log Immunoglobulin G 29 Days After Inoculation
      2
      4.0
      2
      က
      3.0
      2
                                                           1.0
                     0.0
                                        0.5
                                                                             1.5
```

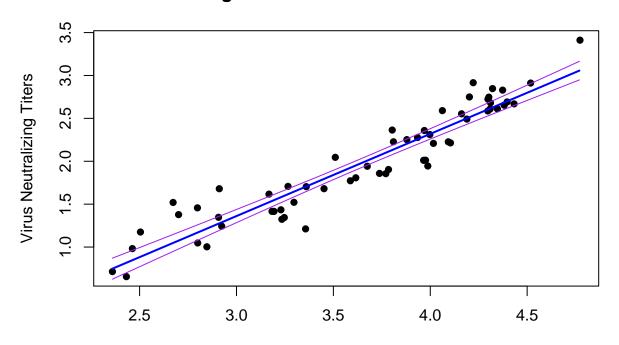
Log Immunoglobulin G Prior to Inoculation



## Log Immunoglobulin G 29 Days After Inoculation

```
model = lm(data = immune_test, VNT ~ IgG29)
# Confidence Intervals
confint(model, level = .99)
##
                    0.5 %
                             99.5 %
## (Intercept) -1.9320169 -1.092745
                0.8447733 1.071036
## IgG29
x = seq(min(immune_test$IgG29), max(immune_test$IgG29), .01)
mypoints = data.frame(IgG29 = x)
plot(immune_test$IgG29, immune_test$VNT, pch = 16, ylab = "Virus Neutralizing Titers",
     xlab = "Log Immunoglobulin G 29 Days After Inoculation",
     main = "VNT vs IgG29 With 95% Confidence Intervals")
myCI = predict(model, newdata = mypoints, interval = "confidence")
lines(x, myCI[, 2], col = "purple")
lines(x, myCI[, 3], col = "purple")
lines(x, myCI[, 1], col = "blue", lwd = 2)
```

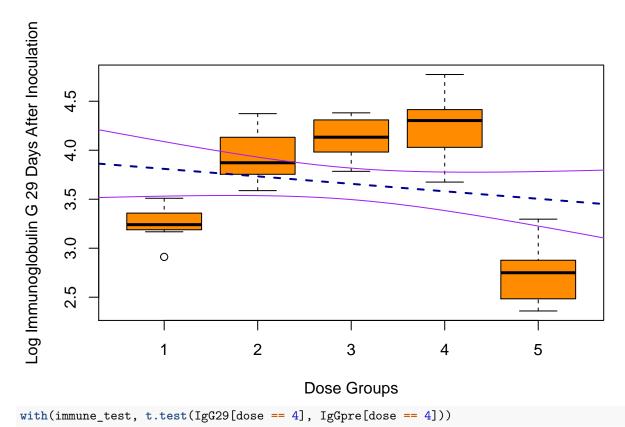
## VNT vs IgG29 With 95% Confidence Intervals



Log Immunoglobulin G 29 Days After Inoculation

```
m = aov(IgG29 ~ as.factor(dose), data = immune_test)
summary(m)
##
                   Df Sum Sq Mean Sq F value Pr(>F)
## as.factor(dose) 4 19.951
                               4.988
                                      87.43 <2e-16 ***
## Residuals
                     3.138
                               0.057
                   55
## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
# We can conclude from the analysis of variance test that the coefficents of effects for different dose
# on Virus Neutralizing Titers, are jointly significant. In other words, the means of VNT per each dose
# group are signficantly different from each other. We are told this information through our F Value,
# which is essentially the ratio of the between-group variance against the within-group variance
# in our model.
confint(m, level = .95)
##
                        2.5 %
                                   97.5 %
## (Intercept)
                     3.1210186 3.3973796
## as.factor(dose)2 0.4801546 0.8709881
## as.factor(dose)3 0.6718084 1.0626419
## as.factor(dose)4 0.7847145 1.1755480
## as.factor(dose)5 -0.7291440 -0.3383105
library(sqldf)
## Loading required package: gsubfn
## Loading required package: proto
## Warning in fun(libname, pkgname): couldn't connect to display ":0"
## Loading required package: RSQLite
```

```
ans = sqldf("
      SELECT dose, avg(IgG29)
      FROM immune_test
      GROUP BY dose
      ")
print(ans)
    dose avg(IgG29)
##
      1 3.259199
## 1
       2 3.934771
## 2
## 3
       3 4.126424
## 4
       4
          4.239330
          2.725472
## 5
       5
# The greatest log concentration seems to belong to dose group 4, at 4.239330.
# It's 95% Confindence Interval Values are (0.7847145, 1.1755480), with regards to its effect on
# Log Immunoglobulin G 29 Days After Inoculation.
# Running a boxplot regression with Confindence Intervals required that I converted "dose"
# to a numeric value.
immune_test$dose = as.numeric(immune_test$dose)
x = seq(0, 6, .01)
mypoints = data.frame(dose = x)
m = lm(IgG29 ~ dose, data = immune_test)
boxplot(IgG29 ~ dose, data = immune_test, col = "darkorange", xlab = "Dose Groups",
        ylab = "Log Immunoglobulin G 29 Days After Inoculation")
myCI = predict(m, newdata = mypoints, interval = "confidence")
abline(m, lty = 2, col = "darkblue", lwd = 2)
lines(x, myCI[, 2], col = "purple")
lines(x, myCI[, 3], col = "purple")
```



```
##
##
   Welch Two Sample t-test
## data: IgG29[dose == 4] and IgGpre[dose == 4]
## t = 22.965, df = 19.172, p-value = 2.065e-15
\#\# alternative hypothesis: true difference in means is not equal to 0
## 95 percent confidence interval:
## 3.244909 3.895281
## sample estimates:
## mean of x mean of y
## 4.2393304 0.6692356
# Yes it is signficantly different.
diff = sqldf("SELECT IgG29 - IgGpre
      FROM immune_test
      WHERE dose = 4")
# Subtracting the baseline data from the IgPGpre data in the group 4 dose condition.
diff
##
      IgG29 - IgGpre
```

## 1

## 2

## 3

## 4

## 5

## 6 ## 7

## 8

4.150838

3.867206

3.731578

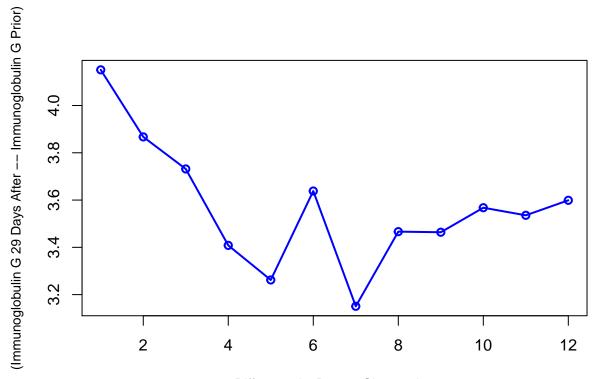
3.408268

3.262033 3.638150

3.150635 3.466389

```
## 9
            3.463865
## 10
            3.567617
            3.535515
## 11
## 12
            3.599043
mean(diff$`IgG29 - IgGpre`)
## [1] 3.570095
# The mean of the difference is 3.570095, the 95\% CI, in the above t-test, is (3.244909,\ 3.895281).
names(diff) <- c("difference")</pre>
# Attaching Dose Value to the Data Frame, and using a rudimentry method to attach '4' to every row valu
diff = sqldf("
      SELECT *,
      (CASE difference
      WHEN 100000 THEN 'No Comment'
      ELSE '4'
      END)
      AS 'Dose'
      FROM diff
      ")
diff
##
      difference Dose
## 1
       4.150838
## 2
        3.867206
                    4
## 3
        3.731578
## 4
        3.408268
## 5
        3.262033
                    4
## 6
        3.638150
                    4
## 7
        3.150635
                    4
## 8
        3.466389
                    4
## 9
        3.463865
                    4
## 10
        3.567617
                    4
## 11
        3.535515
## 12
        3.599043
                    4
# Here is the data frame, corresponding to all of the dose values and there respective 'IgG29 - IgGpre'
# values.
plot(diff$difference, type = "o", xlab = "Difference by Dose 4 Observation",
     ylab = "(Immunoglobulin G 29 Days After -- Immunoglobulin G Prior)", cex.lab = .8,
```

col = "blue", lwd = 2)



Difference by Dose 4 Observation

```
## Expected_VNT
## 1 2.575645
```

```
mean(diff$difference)
```

#### ## [1] 3.570095

```
\# Using a lgG concentration of 3.570095: From the Excel Sheet 34 in the control and 2 in
# the Treatment got the disease.
agrestiCaffo <- function(x1,n1,x2,n2,alpha=.05) {</pre>
  # The Agresti-Coull proportion estimate with alpha = .05
              <- qnorm(1-alpha/2);
  zCrit
 pHat1
              <- (x1)/(n1);
              <- (x1+1)/(n1+2);
 pPrime1
              <- (pPrime1*(1-pPrime1)/(n1+2));
 pPrimeV1
 pHat2
              <- (x2)/(n2);
 pPrime2
              <- (x2+1)/(n2+2);
              <- (pPrime2*(1-pPrime2)/(n2+2));
 pPrimeV2
           <- sqrt(pPrimeV1+pPrimeV2);
 pPrimeSD
```

```
1CI
              <- pPrime1-pPrime2 - zCrit*pPrimeSD;
  uCI
              <- pPrime1-pPrime2 + zCrit*pPrimeSD;
  zScore
              <- (pPrime1-pPrime2)/pPrimeSD;
 pValue
              <- (1-pnorm(abs(zScore)))*2;
  pValueGgtP <- (1-pnorm(zScore))</pre>
                   <- list()
 myParms
  myParms$zCrit
                   <- zCrit;
  myParms$pHat1
                   <- pHat1;
  myParms$pPrime1 <- pPrime1;</pre>
 myParms$pHat2
                   <- pHat2;
 myParms$pPrime2 <- pPrime2;</pre>
 myParms$deltaP
                   <- pHat1-pHat2;
 myParms$lower
                   <- 1CI;
 myParms$upper
                   <- uCI;
 myParms$pValue
                   <- pValue;
 myParms$pValueGgtP <- pValueGgtP;</pre>
 return(myParms)
agrestiCaffo(34, 36, 2, 36)
## $zCrit
## [1] 1.959964
## $pHat1
## [1] 0.9444444
##
## $pPrime1
## [1] 0.9210526
## $pHat2
## [1] 0.0555556
##
## $pPrime2
## [1] 0.07894737
##
## $deltaP
## [1] 0.8888889
## $lower
## [1] 0.7208551
##
## $upper
## [1] 0.9633554
## $pValue
## [1] 0
##
## $pValueGgtP
## [1] 0
```

```
agrestiCoull <- function(x,n,alpha = .05) {</pre>
  \# The Agresti-Coull proportion estimate with alpha = .05
  zCrit
              <- qnorm(1-alpha/2)
 pHat
              <- (x)/(n)
 pPrime
              <- (x+2)/(n+4)
 pPrimeSD
             <- sqrt(pPrime*(1-pPrime)/(n+4))
  lCI
             <- pPrime - zCrit*pPrimeSD;</pre>
              <- pPrime + zCrit*pPrimeSD;
  uCI
  1CI[1CI<0] <-0;</pre>
  uCI[uCI>1] <-1;
  myParms
                   <- list()
  myParms$zCrit
                   <- zCrit;
  myParms$pHat
                   <- pHat;
  myParms$pPrime
                   <- pPrime;
  myParms$lower
                   <- 1CI;
 myParms$upper
                   <- uCI;
 return(myParms)
}
dose_4_data = sqldf("SELECT IgG29
                    FROM immune_test
                    WHERE dose = 4")
not_dose_4_data = sqldf("SELECT IgGpre
                        FROM immune_test
                        WHERE dose != 4")
mean(dose_4_data$IgG29)
## [1] 4.23933
mean(not_dose_4_data$IgGpre)
## [1] 0.674661
# Yes there is a significant difference between the post 29 day data in dose 4 and the
# pre inoculation data in the rest of the dose categories.
# Second bonus question solved through k-clustering means.
immune_cluster <- kmeans(immune_test[, 1:3], centers = 5, nstart = 20)</pre>
immune_cluster = data.frame(immune_test, cluster = factor(immune_cluster$cluster))
ggplot(immune_cluster, aes(x = IgG29, y = VNT, color = cluster,
                           shape = as.factor(dose))) + geom_point()
```

