

A Compact Fiber Optic Eye Diagnostics System

Rafat R. Ansari and Kwang I. Suh
Lewis Research Center
Cleveland, Ohio

Michael A. DellaVecchia
Wills Eye Hospital
Philadelphia, Pennsylvania

Stephen Dubin
Drexel University
Philadelphia, Pennsylvania

J. Samuel Zigler, Jr.
National Eye Institute
Bethesda, Maryland

Prepared for
Technology 2005
cosponsored by NASA Headquarters and the Technology Utilization Foundation
Chicago, Illinois, October 24–26, 1995



National Aeronautics and
Space Administration

Trade names or manufacturers' names are used in this report for identification only. This usage does not constitute an official endorsement, either expressed or implied, by the National Aeronautics and Space Administration.

A COMPACT FIBER OPTIC EYE DIAGNOSTICS SYSTEM

Rafat R. Ansari and Kwang I. Suh*

NASA Lewis Research Center

Microgravity Fluids Branch, Space Experiments Division

Mail Stop 333-1, 21000 Brookpark Road, Cleveland, Ohio 44135

Phone: (216) 433-5008, Fax: (216) 977-7138, e-mail: ransari@lims01.lerc.nasa.gov

Michael A. DellaVecchia

(affiliated with Wills Eye Hospital, Philadelphia, Pennsylvania)

846 Farragut Road, Berwyn, Pennsylvania 19312

Phone: (610) 640-1578, Fax: (215) 726-6637, e-mail: dellavem@duvm.ocs.drexel.edu

Stephen Dubin

Biomedical Engineering and Science Institute

Drexel University

Philadelphia, Pennsylvania 19104

Phone: (215) 895-2219, Fax: (215) 895-4983, e-mail: dubinse@duvm.ocs.drexel.edu

J. Samuel Zigler Jr.

Laboratory for the Mechanisms of Ocular Disease

National Eye Institute, National Institutes of Health

9000 Rockville Pike, Bethesda, MD 20892

Phone: (301) 496-6669, Fax: (301) 496-1759, e-mail: szigler@helix.nih.gov

ABSTRACT

A new fiber optic probe developed for determining transport properties of sub-micron particles in fluids experiments in a microgravity environment has been applied to study different parts of the eye. The probe positioned in front of an eye, delivers a low power (\sim few μ W) light from a laser diode into the eye and guides the light which is back scattered by different components (aqueous humor, lens, and vitreous humor) of the eye through a receiving optical fiber to a photo detector. The probe provides rapid determination of macromolecular diffusivities and their respective size distributions in the eye lens and the gel-like material in the vitreous humor. For a clinical use, the probe is mounted on a standard slit-lamp apparatus simply using a Hruby lens holder. The capability of detecting cataracts, both nuclear and cortical, in their early stages of formation, in a non invasive and quantitative fashion, has the potential in patient monitoring and in developing and testing new drugs or diet therapies to “dissolve” or slow down the cataract formation before the surgery becomes necessary. The ability to detect biochemical and macromolecular changes in the vitreous structure can be very useful in identifying certain diseases of the posterior chamber and their complications, e.g., posterior vitreous detachment and diabetic retinopathy.

INTRODUCTION

The anterior chamber (space between the cornea and the lens) of a normal adult eye the eye is filled with a transparent water-like fluid known as aqueous humor. However, presence of elevated cholesterol and blood-sugar levels in patients with heart and diabetic related diseases can be identified in this fluid. An adult human lens is the size of an aspirin tablet. At birth the lens tissue is transparent. With age the lens gradually loses its transparency. The lens transparency can also be lost because of trauma, certain diseases (e.g. diabetes, uremia, chronic diarrhea), hereditary factors, radiation exposure, and continued use of certain medications over a long period of time including steroids, tranquilizers (phenothiazines), gout medications (allopurinol), psoriasis medications (psoralens), and antibiotics (tetracycline). A cataract is formed when the lens cloudiness hinders light transmission through the lens and the ability to focus a sharp

*National Research Council—NASA Research Associate at Lewis Research Center.

image on the retina. Cataracts remain the major cause of blindness affecting about 50 million people each year worldwide. It is estimated that over \$5 billion will be spent this year in treating cataract patients in the United States alone (Shulman, 1993). There is no medical treatment to prevent or halt the progression of a cataract; nor is there any way to reverse a cataract once it has formed. The only known treatment is surgical. However, a medical treatment could be possible if we understand how a cataract forms and what makes it grow (Shulman, 1993). The ability of detecting a cataract at the earliest stages of formation will be useful in patient monitoring and in the development and in screening possible “anticataract” drugs or “diet” therapies. Kupfer of the National Eye Institute (NEI) of the National Institutes of Health (NIH) predicts that by the year 2000 new drugs will slow the progression of cataracts which is the leading cause of blindness worldwide (Long, 1992). Most recently Sardi (1995) has strongly argued in favor of using multivitamins, antioxidants, and nutritional supplements in eradicating cataracts. In-between the lens and the retina a gel-like fluid known as vitreous humor fills the rest of the eye globe. This fluid comprises 80% of the total volume of the eye and is the least understood component of an eye. Vitreous diseases can be detected by optically examining this fluid.

The technique of dynamic light scattering (DLS) or quasi-elastic light scattering (QELS) was first applied to the study of cataractogenesis in the pioneering work of Tanaka and Benedek (1976). An extensive review of using QELS to study cataracts has been given by Bursell et al (1990). DLS/QELS being a non-invasive and quantitative technique, seem to hold promising potential in its use as a routine ophthalmic device. But, its commercial scope as an ophthalmic diagnostic tool in clinical settings hitherto not materialized. This is mainly due to bulky equipment, optical alignment problems, polydisperse and interacting nature of the scattering species, and radiation exposure time. Since the original application of QELS, some twenty years ago by Tanaka and Benedek (1976), every subsequent study focused on rather easily accessible part (anterior chamber) of the eye i.e. the aqueous humor and the lens. Most recently Ansari and et al. (1995a and 1995b) in the USA and Rovati et al. (1995) in Switzerland reported the first experiments directed to the studies of the vitreous.

In the past few years DLS instrumentation has embraced several new and innovative technological advances. The immediate impact of these have resulted in the miniaturization of DLS instrumentation (Brown et al. 1990). About four years ago a lens-less back-scatter fiber optic probe was developed by Dhadwal et al. (1991) to study concentrated particulate dispersions. Subsequently, Ansari et al. (1992) and Dhadwal et al. (1993) have shown the utility of this probe to the studies of cataractogenesis in excised bovine and cadaver eye lenses, and in live animals (Dhadwal, et al. 1995). Unfortunately these preliminary studies have certain limitations and pose severe constraints in probing different parts of the eye. Ansari and Suh (1995) recently developed a new DLS probe to study nucleation and aggregation phenomena during protein crystal growth in space (microgravity) experiments. Ansari et al. (1995) have also shown recently that this probe alleviates many major concerns discussed earlier when used as an ophthalmic diagnostics device and is much superior in performance when compared with the earlier reported work (Ansari, et al. 1992; Dhadwal, et al. 1993; 1995). The new probe provides accurate particle size determination of a variety of colloidal dispersions in 5 seconds at extremely low laser power levels. Furthermore, the major problem of probing different parts of the eye has been solved. In this report we briefly summarize some experimental in-vivo results on rabbits with naphtalene-induced cataract and on Philly mice, with hereditary cataract.

EXPERIMENTAL PROCEDURE AND SET UP

The laser light from a laser diode is transmitted by a compact backscatter fiber optic probe to the eye. Depending upon the position of the scattering volume, Brownian motion of the particles in the aqueous humor, protein crystallines inside the lens and the macromolecules in the vitreous humor are monitored. The incident laser radiation interacts with the submicron particles dispersed in different regions of the eye. The particles scatter light and the intensity of this scattered light fluctuates in time due to the Brownian or thermal motion of the dispersed particles in the suspending medium. In a dynamic medium such as eye, when illuminated by laser light, a rapidly changing interference pattern due to the differences in the optical density of the scattering medium can be coherently detected by a DLS spectrometer. The rapidly fluctuating interference pattern at a photodetector contains information about the dynamics of the scattering medium and can be extracted by constructing a correlation function. A correlation function is constructed after detecting the scattered light by a photodetector and appropriate electronic processing via a amplifier-discriminator stage and a digital correlator. In the simplest case of dilute dispersions of spherical particles the slope of the correlation function provides quick and accurate determination of the particle's translation diffusion coefficient. The diffusion coefficient data can

easily be transformed into average particle size, using Stokes-Einstein equation, provided the viscosity of the suspending medium, its temperature and refractive index are known.

A fiber optic probe comprising two monomode optical fibers and two GRIN micro lenses, as illustrated in Figure 1, provides a compact and remote means of studying the dynamical characteristics of the macromolecules in the eye. A 1.3 cm (diameter) x 2.3 cm (length) fiber optic probe contains the necessary optics to perform DLS measurements at a scattering angle of 161.5° . The probe is non-invasive and is conveniently positioned in front of the eye (cornea), but having no physical contact with any part of the eye. Two monomode optical fibers, each housed in a stainless steel ferrule, are mounted into a separate stainless steel housing. An air gap (0-0.5 mm) is intentionally left between the fiber housing and the lens housing in order to produce a tightly focused spot in the scattering volume. The two optical fibers in their housings are aligned and fixed into position off-axis with the GRIN lens. The two housings are placed inside a third (outer) housing made of stainless steel, and the back end of the housing is covered with a heat-shrink tubing. The two free ends of the optical fibers were terminated with FC/PC-type male connectors for easy mating with the laser diode and an avalanche photodiode detector (APD). The DLS data was analyzed using the commercial software provided by the Brookhaven Instruments Company of New York. The DLS probe is also equipped with a 2 mm fiberscope for imaging the eye. This eye imaging provides records of beam position in the eye for patient documentation.

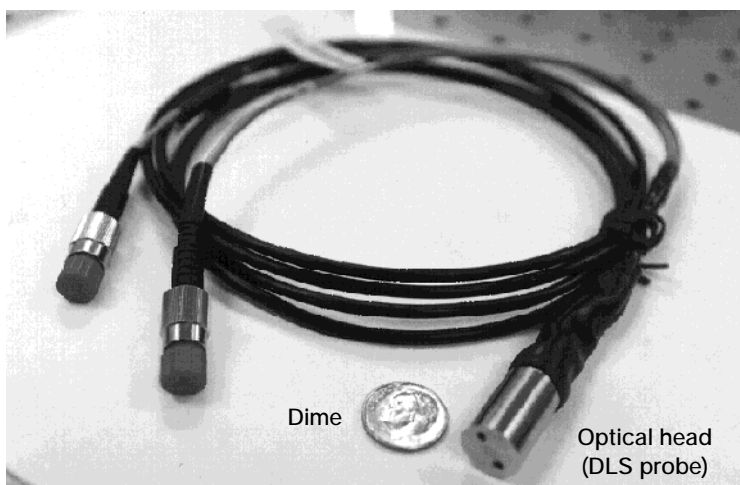
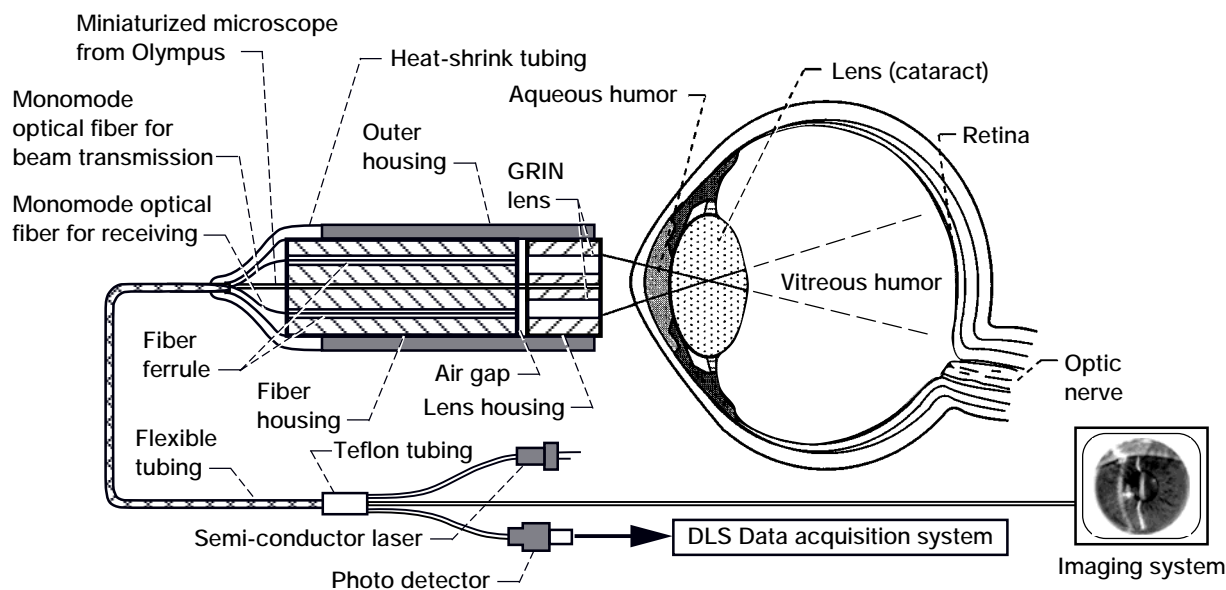


Figure 1.—A compact device for eye diagnostics.

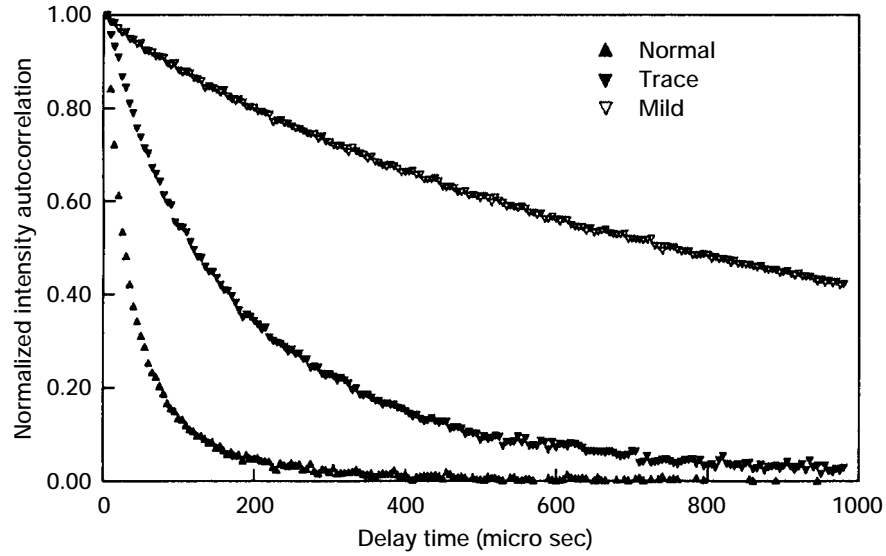


Figure 2.—In-vivo DLS eye diagnostic measurements in live Philly mice.

RESULTS

The Philly mouse is a good animal model of hereditary cataract (Kador et al. 1980). Philly mice develop cataracts spontaneously between day 26 and 33 after their birth. We applied our new fiber optic probe to study the progression of cataractogenesis in this mouse model and to show the feasibility and safe use of our device in laboratory animals. Figure 2 shows DLS autocorrelation data on three Philly mice. The data includes a 45 day old normal mouse of the control FVB/N strain and two Philly mice roughly 26-29 days old. The slope of the correlation function gives a translational diffusion coefficient which can be converted into particle size and particle size distributions (see Ansari et al. 1995). One of us (MAD—an ophthalmologist) conducted eye examinations of these mice with a slit-lamp apparatus and concluded a normal (transparent) and two other eyes having trace and mild cataracts. These examinations were

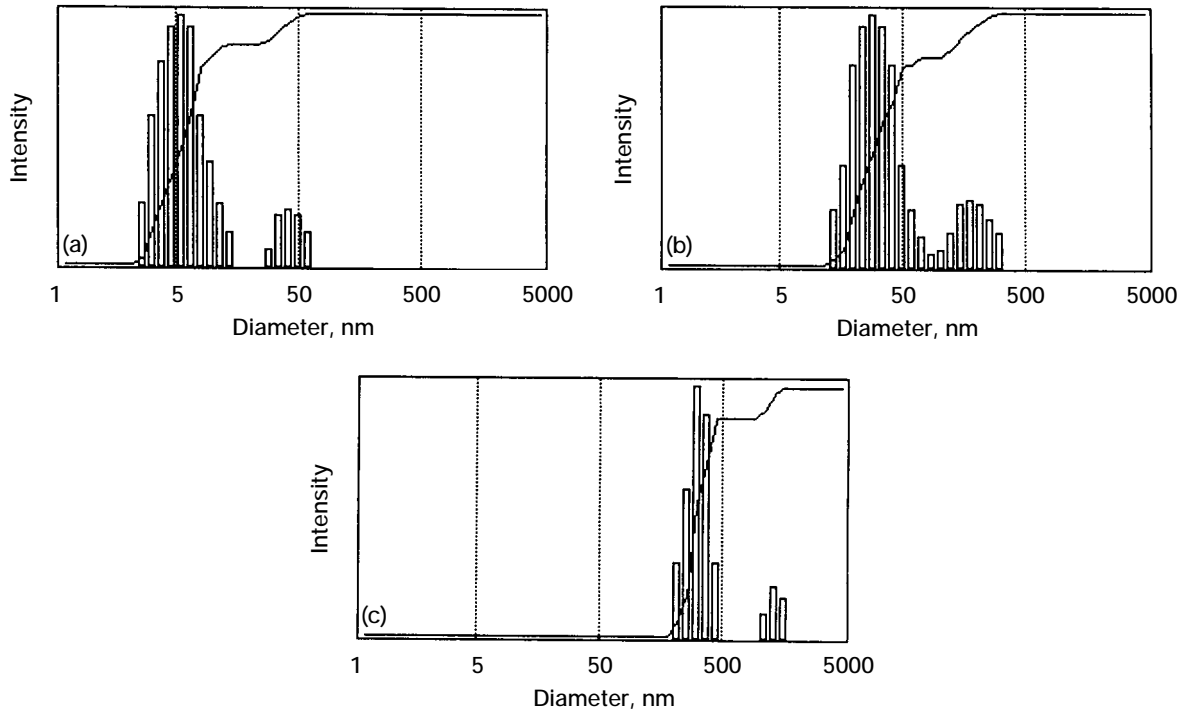


Figure 3(a-c).— Size distributions.

conducted soon (within few minutes) after the DLS measurements were completed. Each measurement only took 5 seconds at a laser power of 50 μ W. The changing slope of the autocorrelation functions is an indication of cataractogenesis. The DLS autocorrelation data is converted into particle size distribution using an exponential sampling program and is shown in Figure 3 (a-c). Although conversion of the DLS data into particle size distributions require certain assumptions regarding the viscosity of the lens fluid these size values do indicate a trend as the cataract progress. These measurements suggest that we can quantitatively monitor cataractogenesis with reasonable reliability, reproducibility (5%-10%), and accuracy.

Figure 4 and 5 shows autocorrelation function profiles of two Belgian-belted rabbits. Both rabbits were about six months in age. One rabbit was kept on a normal diet while the other rabbit was fed 1 gm of naphthalene per 1 Kg of body weight for 3 days per week for a period of three weeks. One of us (MAD) regularly monitored the progression of cataract in these rabbits as a function of time using a slit-lamp apparatus until a mild cataract was noticed. The DLS

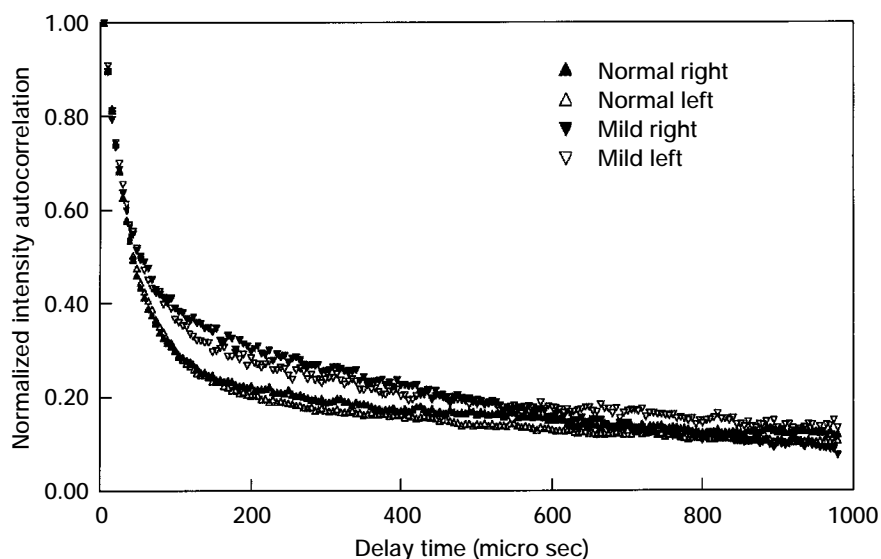


Figure 4.—In-vivo DLS eye diagnostic measurements in live rabbits.

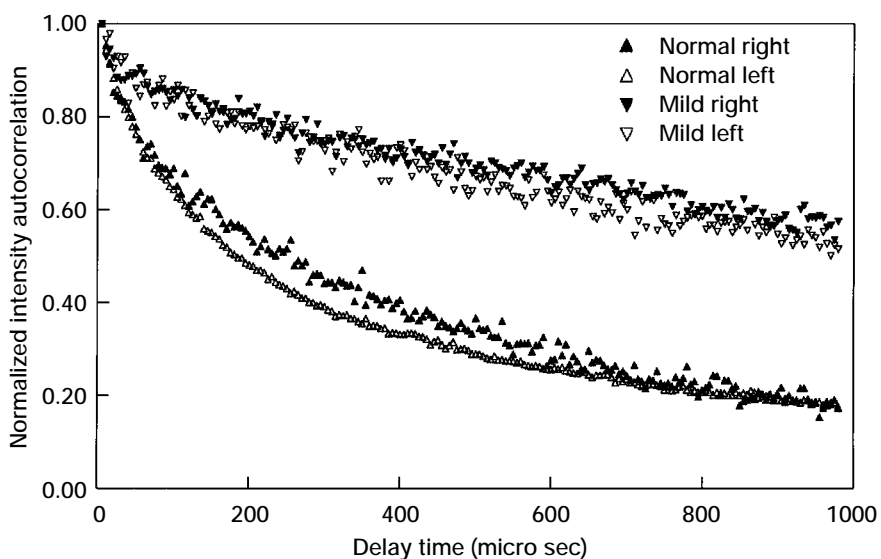


Figure 5.—In-vivo DLS eye diagnostic measurements in live rabbits. Naphthalene induced cataract on rabbits.

measurements were performed in the anterior cortex (see Figure 4) and in the nucleus (Figure 5). In the anterior cortex we do not see any appreciable change in the slope of the correlation functions in either eyes of either the controlled or the naphthalene-fed rabbit. The change in slopes in the correlation functions of the cortex and the lens nucleus in the normal rabbits is consistent with earlier observations (Ansari et al. 1995, and Libondi et al. 1986). In the nuclear region (Figure 4) we see appreciable change in the slope of the correlation functions due to mild cataracts in the naphthalene-fed rabbit. These preliminary results are now guiding us into a research effort to study a group of several controlled and naphthalene-induced cataract rabbits to get a detailed understanding of cataractogenesis.

In Figure 6 we present preliminary results identifying detailed crystalline structure of a transparent excised bovine eye lens. A 3-D plot is constructed after scanning the lens by the new fiber optic probe. This plot shows average protein size (α -crystalline+aggregates) as one moves from the anterior cortex to nucleus to posterior cortex inside the lens capsule.

The vitreous humor of the eye described by Sebag (1989) is the least understood part of an eye. We present preliminary DLS measurements made *in vivo*. The vitreous body exhibits a two-exponential behavior, i.e. a fast and a slowly diffusing component, consistent with its gel-like properties. We see almost constant value for the fast component in this region. We ascribe this fast component due to the diffusion of hyaluronate molecular coils in water and the slow diffusion component due to the collagen-fibril network. A size distribution of the vitreous is presented in Figure 7. We calculated the molecular weight for the fast component to be 3.78×10^6 daltons. This is consistent with the range ($2-4.5 \times 10^6$) of values given by Balazs and Delinger (1984). More *in-vivo* experimental work is being performed at this time in our laboratory to fully understand the structure of the vitreous humor.

CONCLUSION

Our newly developed DLS probe for eye diagnostics offers several unique features. These include use of extremely low laser power levels ($\sim 10-50 \mu\text{W}$), sufficient penetration depth to diagnose aqueous, lens, and the vitreous body, touch free operation (no physical contact with the eye), fast data acquisition time ($\sim 2-10$ seconds), high spatial coherence (β) or signal/noise values, free of optical alignment, no need for vibration isolation devices, 100% solid state (laser/detector) operation, and a modular approach which uses a Hruby lens holder on a regular slit-lamp apparatus providing quantitative monitoring of the entire eye (see Figure 8). The viewing area inside the eye can be controlled by using the joy stick of the slit-lamp apparatus. In addition to routine eye examination with a slit-lamp apparatus, our method provides early detection of cataracts, the characterization of hyaluronic acid chains and the collagen protein

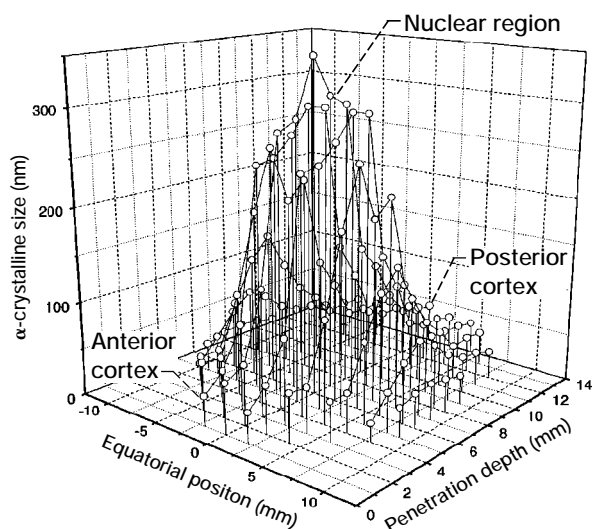


Figure 6.—3-D Scan of protein population in a excised bovine lens.

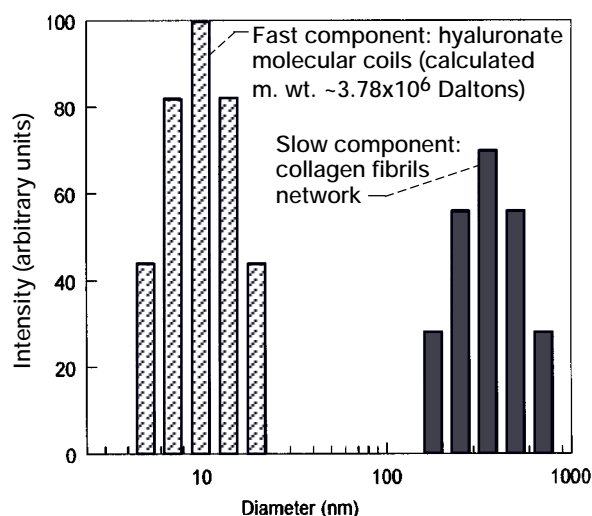


Figure 7.—Six month old rabbit right eye posterior chamber; vitreous size distribution (CONTIN analysis) penetration depth from the corneal surface 11.0 mm.

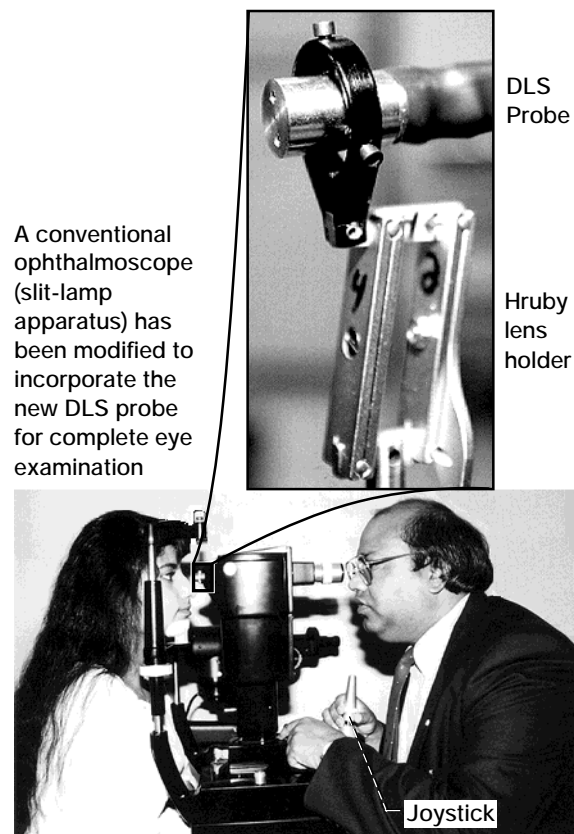


Figure 8.—A compact device for eye diagnostics.

fibrils in the vitreous humor, and though not demonstrated here, the characterization of protein molecules and the metabolic waste product in the aqueous fluid in the anterior chamber of the eye can also be achieved. The patient correlation functions and the corresponding size distribution data can be integrated into OPIS[®] (Ophthalmic Patient Information System) commercial computer software program, facilitating patient record keeping and follow up visits.

In terms of 1991 Medicare dollars, according to Kolberg (1995) \$3.4 billion were paid for cataract surgery than for any other single procedure. We therefore conclude that our new technology described in this paper opens up enormous opportunity in eye care and in finding medical cures for the eradication of cataracts.

ACKNOWLEDGEMENTS

The authors are grateful to the Microgravity Science and Applications Division (MSAD), code UG of the NASA headquarters for supporting this research. This work was completed under a NASA grant NCC 166-3/CWRU. Kwang Suh wishes to thank NASA Lewis Research Center and the National Research Council for the award of a postdoctoral research fellowship.

REFERENCES

- Ansari, R.R., Dhadwal, H.S., Campbell, M.C.W. and DellaVecchia, M.A. (1992) "A Fiber Optic Sensor for Ophthalmic Refractive Diagnostics," Proc. of Fiber Optic Medical and Fluorescent Sensors and Applications, January 23–24, 1992, Los Angeles, CA., SPIE Vol. 1648.
- Ansari, R.R., Suh, K.I., DellaVecchia, M.A. and Dubin, S. (1995a) "Early Diagnosis of Cataracts Using a Fiber Optic System," in audio/video Proc. Cataract and Refractive Surgery: Current Issues, Session 2-B, ASCRS Symposium on Cataract, IOL and Refractive Surgery, April 1–5, 1995, San Diego, CA.

- Ansari, R.R., Suh, K.I., DellaVecchia, M.A. and Dubin, S. (1995b) "Ophthalmic Diagnostics Using a New Dynamic Light Scattering Fiber Optic Probe," Proc. Medical Application, Conference on lasers in Ophthalmology III, SPIE, Bios Europe' 95, Barcelona, Spain, September 12–16, 1995, **2632**(18).
- Ansari, R.R. and Suh, K.I. (1995) "Sizing of Colloidal Particles and Protein Molecules in a Hanging Fluid Drop", Proc. Biomedical Optoelectronics in Clinical Chemistry and Bio-technology, SPIE, Bios Europe' 95, Barcelona, Spain, September 12–16, 1995. **2629**(23).
- Balazs, E.A. and Denlinger, J.L. (1984) "The Vitreous," in: The Eye, vol. IA, editor, H. Davson, Academic Press, 533–589.
- Brown, J.G., Burnett, J., Mansbridge, C.I., Moir, C. (1990) "Miniature Laser Light Scattering Instrumentation for Particle Size Analysis," Appl. Opt. **29**, 4159–4169.
- Bursell, S.E., Magnante, P.C. and Chylack Jr., L.T. (1990) "In vivo Uses of Quasielastic Light Scattering Spectroscopy as a Molecular Probe in the Anterior Segment of the Eye," in Noninvasive Diagnostic Techniques in Ophthalmology, editor Barry R. Masters, Springer-Verlag, New York.
- Carper, D., Smith-Gill, S.J. and Kinoshita, J.H., "Immunocytochemical Localization of the 27K b-Crystallin Polypeptide in the Mouse Lens during Development Using a Specific Monoclonal Antibody: Implications for Cataract Formation in the Philly Mouse", Developmental Biology 113, 104-109 (1986).
- Dhadwal, H.S., Ansari, R.R., and Meyer, W.V. (1991) "A Fiber Optic Probe for Particle Sizing in Concentrated Suspensions," Rev. Sci. Instrum. **62**(12), December 1991.
- Dhadwal, H.S., Ansari, R.R. and DellaVecchia, M.A. (1993) "Coherent Fiber Optic Sensor for Early Detection of Cataractogenesis in a Human Eye Lens," J. Opt. Engineering, **32**(2), February 1993.
- Dhadwal, H.S., Ansari, R.R., DellaVecchia, M.A. and Dubin, S. (1995) "Fiber Optic System for In-vivo Sizing of Proteins in Animal Eye Lenses," Proc. Ophthalmic Technologies V, SPIE, **2393**, 227–236, February 4–5, 1995, San Jose, CA.
- Kador, P.F., Fukui, H.N., Fukushi, S., Jernigan Jr., H.M., and Kinoshita, J.H., "Philly Mouse: A New Model of Hereditary Cataract", Exp. Eye Res. 30, 59-68, (1980).
- Kolberg, R., "Intersecting Orbits: NIH and NASA's Joint Explorations" in "*The NIH Catalyst*", vol.3, issue 5, September-October 1995.
- Libondi, T., Magnante, P., Chylack L.T., and Benedek, G.B., "In Vivo Measurements of the Aging Rabbit Lens Using Quasi Elastic Light Scattering", Current Eye Research **5**, 6, 411-419, (1986).
- Long, M.E. (1992) "The Sense of Sight," in National Geographic, **182**(5), Nov. 1992.
- Rovati, L., Fankhauser, F. II, and Ricka, J. (1995) "Dynamic Light Scattering Spectroscopy of In-vivo Human Vitreous," Proc. Medical Application, Conference on Lasers in Ophthalmology III, SPIE, Bios Europe' 95, Barcelona, Spain, September 12–16, 1995, **2632**(19).
- Sardi, B. (1995) "Eradicating Cataracts," Townsend letter for doctors, June 1995.
- Sebag, J. (1989) "*The Vitreous: Structure, Function, and Pathobiology*," Springer-Verlag, New York.
- Shulman, J. (1993) "*Cataracts: The Complete Guide from Diagnostics to Recovery for Patients and Families*," St. Martins Press, New York.
- Tanaka, T. and Benedek, G.B. (1976) "Observation of Protein Diffusivity in Intact Human and Bovine Lenses With Application to Cataract," Investigative Ophthalmology, **14**(6), 449–456.

REPORT DOCUMENTATION PAGE			Form Approved OMB No. 0704-0188	
Public reporting burden for this collection of information is estimated to average 1 hour per response, including the time for reviewing instructions, searching existing data sources, gathering and maintaining the data needed, and completing and reviewing the collection of information. Send comments regarding this burden estimate or any other aspect of this collection of information, including suggestions for reducing this burden, to Washington Headquarters Services, Directorate for Information Operations and Reports, 1215 Jefferson Davis Highway, Suite 1204, Arlington, VA 22202-4302, and to the Office of Management and Budget, Paperwork Reduction Project (0704-0188), Washington, DC 20503.				
1. AGENCY USE ONLY (Leave blank)	2. REPORT DATE November 1995	3. REPORT TYPE AND DATES COVERED Technical Memorandum		
4. TITLE AND SUBTITLE A Compact Fiber Optic Eye Diagnostics System		5. FUNDING NUMBERS WU-963-45-0A NCC3-166		
6. AUTHOR(S) Rafat R. Ansari, Kwang I. Suh, Michael A. DellaVecchia, Stephen Dubin and J. Samuel Zigler, Jr.				
7. PERFORMING ORGANIZATION NAME(S) AND ADDRESS(ES) National Aeronautics and Space Administration Lewis Research Center Cleveland, Ohio 44135-3191		8. PERFORMING ORGANIZATION REPORT NUMBER E-10007		
9. SPONSORING/MONITORING AGENCY NAME(S) AND ADDRESS(ES) National Aeronautics and Space Administration Washington, D.C. 20546-0001		10. SPONSORING/MONITORING AGENCY REPORT NUMBER NASA TM-107119		
11. SUPPLEMENTARY NOTES Prepared for Technology 2005 cosponsored by NASA Headquarters and the Technology Utilization Foundation, Chicago, Illinois, October 24-26, 1995. Rafat R. Ansari, NASA Lewis Research Center; Kwang I. Suh, National Research Council—NASA Research Associate at Lewis Research Center; Michael A. DellaVecchia (affiliated with Wills Eye Hospital, Philadelphia, Pennsylvania) 846 Farragut Road, Berwyn, Pennsylvania 19312; Stephen Dubin, Drexel University, Bio-medical Engineering and Science Institute, Philadelphia, Pennsylvania 19104; and J. Samuel Zigler, Jr., Laboratory for the Mechanisms of Ocular Disease, National Eye Institute, National Institutes of Health, 9000 Rockville Pike, Bethesda, Maryland 20892. Responsible person, Rafat R. Ansari, organization code 6712, (216) 433-5008.				
12a. DISTRIBUTION/AVAILABILITY STATEMENT Unclassified - Unlimited Subject Categories 35, 51, and 74 This publication is available from the NASA Center for Aerospace Information, (301) 621-0390.		12b. DISTRIBUTION CODE		
13. ABSTRACT (Maximum 200 words) A new fiber optic probe developed for determining transport properties of sub-micron particles in fluids experiments in a microgravity environment has been applied to study different parts of the eye. The probe positioned in front of an eye, delivers a low power (~ few μ W) light from a laser diode into the eye and guides the light which is back scattered by different components (aqueous humor, lens, and vitreous humor) of the eye through a receiving optical fiber to a photo detector. The probe provides rapid determination of macromolecular diffusivities and their respective size distributions in the eye lens and the gel-like material in the vitreous humor. For a clinical use, the probe is mounted on a standard slit-lamp apparatus simply using a Hruby lens holder. The capability of detecting cataracts, both nuclear and cortical, in their early stages of formation, in a non invasive and quantitative fashion, has the potential in patient monitoring and in developing and testing new drugs or diet therapies to "dissolve" or slow down the cataract formation before the surgery becomes necessary. The ability to detect biochemical and macromolecular changes in the vitreous structure can be very useful in identifying certain diseases of the posterior chamber and their complications, e.g., posterior vitreous detachment and diabetic retinopathy.				
14. SUBJECT TERMS Patient monitoring; Ophthalmic technologies; Gels; Dynamic light scattering; Cataracts		15. NUMBER OF PAGES 11		
		16. PRICE CODE A03		
17. SECURITY CLASSIFICATION OF REPORT Unclassified	18. SECURITY CLASSIFICATION OF THIS PAGE Unclassified	19. SECURITY CLASSIFICATION OF ABSTRACT Unclassified	20. LIMITATION OF ABSTRACT	