Thermodynamically constrain Recon3D

Author: Ronan Fleming, Leiden University & National University of Ireland, Galway.

Reviewers:

INTRODUCTION

In flux balance analysis of genome scale stoichiometric models of metabolism, the principal constraints are uptake or secretion rates, the steady state mass conservation assumption and reaction directionality. Von Bertylanffy [1,4] is a set of methods for (i) quantitative estimation of thermochemical parameters for metabolites and reactions using the component contribution method [3], (ii) quantitative assignment of reaction directionality in a multi-compartmental genome scale model based on an application of the second law of thermodynamics to each reaction [2], (iii) analysis of thermochemical parameters in a network context, and (iv) thermodynamically constrained flux balance analysis. The theoretical basis for each of these methods is detailed within the cited papers.

PROCEDURE

Configure the environment

The default COBRA Toolbox paths are automatically changed here to work on the new version of vonBertalanffy

```
aPath = which('initVonBertalanffy');
basePath = strrep(aPath,['vonBertalanffy' filesep
'initVonBertalanffy.m'],'');
addpath(genpath(basePath))
folderPattern=[filesep 'old'];
method = 'remove';
editCobraToolboxPath(basePath,folderPattern,method)
```

```
removing: /home/rfleming/work/sbgCloud/code/fork-cobratoolbox/src/analysis/thermo/componentContribution/olcremoving: /home/rfleming/work/sbgCloud/code/fork-cobratoolbox/src/analysis/thermo/directionalityReport/olcremoving: /home/rfleming/work/sbgCloud/code/fork-cobratoolbox/src/analysis/thermo/groupContribution/oldremoving: /home/rfleming/work/sbgCloud/code/fork-cobratoolbox/src/analysis/thermo/inchi/oldremoving: /home/rfleming/work/sbgCloud/code/fork-cobratoolbox/src/analysis/thermo/molFiles/oldremoving: /home/rfleming/work/sbgCloud/code/fork-cobratoolbox/src/analysis/thermo/protons/oldremoving: /home/rfleming/work/sbgCloud/code/fork-cobratoolbox/src/analysis/thermo/trainingModel/old
```

```
aPath = which('initVonBertalanffy');
basePath = strrep(aPath,['vonBertalanffy' filesep
'initVonBertalanffy.m'],'');
addpath(genpath(basePath))
folderPattern=[filesep 'new'];
method = 'add';
editCobraToolboxPath(basePath,folderPattern,method)
```

```
adding: /home/rfleming/work/sbgCloud/code/fork-cobratoolbox/src/analysis/thermo/componentContribution/new adding: /home/rfleming/work/sbgCloud/code/fork-cobratoolbox/src/analysis/thermo/groupContribution/new adding: /home/rfleming/work/sbgCloud/code/fork-cobratoolbox/src/analysis/thermo/inchi/new
```

```
adding: /home/rfleming/work/sbgCloud/code/fork-cobratoolbox/src/analysis/thermo/molFiles/new adding: /home/rfleming/work/sbgCloud/code/fork-cobratoolbox/src/analysis/thermo/protons/new adding: /home/rfleming/work/sbgCloud/code/fork-cobratoolbox/src/analysis/thermo/trainingModel/new
```

All the installation instructions are in a separate .md file named vonBertalanffy.md in docs/source/installation

With all dependencies installed correctly, we configure our environment, verfy all dependencies, and add required fields and directories to the matlab path.

```
initVonBertalanffy
```

```
ChemAxon Marvin Beans is installed and working.

linux-vdso.so.1 (0x00007ffdddfcf000)

libc.so.6 => /lib/x86_64-linux-gnu/libc.so.6 (0x00007ffladeef000)

libopenbabel.so.5 => /usr/lib/libopenbabel.so.5 (0x00007fflade9f000)

libstdc++.so.6 => /usr/lib/x86_64-linux-gnu/libstdc++.so.6 (0x00007fflada85000)

libgcc_s.so.1 => /usr/local/bin/MATLAB/R2021a/sys/os/glnxa64/libgcc_s.so.1 (0x00007fflad86d000)

/lib64/ld-linux-x86-64.so.2 (0x00007fflae10f000)

libdl.so.2 => /lib/x86_64-linux-gnu/libdl.so.2 (0x00007fflad865000)

libz.so.1 => /lib/x86_64-linux-gnu/libz.so.1 (0x00007fflad849000)

libm.so.6 => /lib/x86_64-linux-gnu/libm.so.6 (0x00007fflad6fa000)

libgomp.so.1 => /usr/lib/x86_64-linux-gnu/libgomp.so.1 (0x00007fflad6b5000)

libpthread.so.0 => /lib/x86_64-linux-gnu/libpthread.so.0 (0x00007fflad692000)

babel must depend on the system libstdc++.so.6 not the one from MATLAB

Trying to edit the 'LD_LIBRARY_PATH' to make sure that it has the correct system path before the Matlab pages.
```

Select the model

The solution will be arch dependent

This tutorial is tested for the E. coli model iAF1260 and the human metabolic model Recon3Dmodel. However, only the data for the former is provided within the COBRA Toolbox as it is used for testing von Bertylanffy. However, the figures generated below are most suited to plotting results for Recon3D, so they may not be so useful for iAF1260. The Recon3D example uses values from literature for input variables where they are available.

```
%modelName = 'iAF1260';
%modelName='Ec_iAF1260_flux1';
% uncomment this line and comment the line below if you want to use the
other model- currently will not work without changes
modelName='Recon3DModel_301';
```

Load a model

Load a model, and save it as the original model in the workspace, unless it is already loaded into the workspace.

```
clear model
global CBTDIR
modelFileName = [modelName '.mat']
```

```
modelFileName =
'Recon3DModel_301.mat'
```

```
modelDirectory = getDistributedModelFolder(modelFileName); %Look up the
folder for the distributed Models.
modelFileName= [modelDirectory filesep modelFileName]; % Get the full path.
Necessary to be sure, that the right model is loaded
switch modelName
    case 'Ec_iAF1260_flux1'
        modelFileName = [modelName '.xml']
        model = readCbModel(modelFileName);
        if model.S(952, 350) == 0
            model.S(952, 350)=1; % One reaction needing mass balancing in
iAF1260
        end
        model.metCharges(strcmp('asntrna[Cytosol]', model.mets))=0; % One
reaction needing charge balancing
    case 'iAF1260'
        model = readCbModel(modelFileName);
        model.mets = cellfun(@(mets)
strrep(mets,'_c','[c]'),model.mets,'UniformOutput',false);
        model.mets = cellfun(@(mets)
strrep(mets, '_e', '[e]'), model.mets, 'UniformOutput', false);
        model.mets = cellfun(@(mets)
strrep(mets, '_p', '[p]'), model.mets, 'UniformOutput', false);
        bool = strcmp(model.mets,'lipa[c]old[c]');
        model.mets{bool}='lipa_old_[c]';
        bool = strcmp(model.mets,'lipa[c]old[e]');
        model.mets{bool}='lipa_old_[e]';
        bool = strcmp(model.mets, 'lipa[c]old[p]');
        model.mets{bool}='lipa old [p]';
        if model.S(952, 350) == 0
            model.S(952, 350)=1; % One reaction needing mass balancing in
iAF1260
        end
        model.metCharges(strcmp('asntrna[c]', model.mets))=0; % One reaction
needing charge balancing
    case 'Recon3DModel Dec2017'
      model = readCbModel(modelFileName);
      model.csense(1:size(model.S,1),1)='E';
      %Hack for thermodynamics
      model.metFormulas{strcmp(model.mets, 'h[i]')}='H';
      model.metFormulas(cellfun('isempty',model.metFormulas)) = {'R'};
      if isfield(model, 'metCharge')
          model.metCharges = double(model.metCharge);
          model=rmfield(model, 'metCharge');
      modelOrig = model;
```

```
case 'Recon3DModel_301'
    model = readCbModel(modelFileName);
    %Hack for thermodynamics
    model.metFormulas(cellfun('isempty',model.metFormulas)) = {'R'};
    modelOrig = model;
    otherwise
        error('setup specific parameters for your model')
end
```

Each model.subSystems $\{x\}$ has been changed to a character array.

Set the directory containing the results

```
switch modelName
    case 'Ec_iAF1260_flux1'
        resultsPath=which('tutorial_vonBertalanffy.mlx');
        resultsPath=strrep(resultsPath,'/tutorial_vonBertalanffy.mlx','');
        resultsPath=[resultsPath filesep modelName ' results'];
        resultsBaseFileName=[resultsPath filesep modelName '_results'];
        resultsPath=which('tutorial_vonBertalanffy.mlx');
        resultsPath=strrep(resultsPath,'/tutorial_vonBertalanffy.mlx','');
        resultsPath=[resultsPath filesep modelName '_results'];
        resultsBaseFileName=[resultsPath filesep modelName ' results'];
    case 'Recon3DModel_Dec2017'
        basePath='~/work/sbqCloud';
        resultsPath=[basePath '/programReconstruction/projects/recon2models/
results/thermo/' modelName];
        resultsBaseFileName=[resultsPath filesep modelName ' '
datestr(now,30) ' '];
    case 'Recon3DModel 301'
        basePath=['~' filesep 'work' filesep 'sbgCloud'];
        resultsPath=which('tutorial vonBertalanffy.mlx');
        resultsPath=strrep(resultsPath,[filesep
'tutorial vonBertalanffy.mlx'],'');
        resultsPath=[resultsPath filesep modelName '_results'];
        resultsBaseFileName=[resultsPath filesep modelName '_results_'];
    otherwise
        error('setup specific parameters for your model')
end
```

Set the directory containing molfiles

```
switch modelName
   case 'Ec_iAF1260_flux1'
       molFileDir = 'iAF1260Molfiles';
   case 'iAF1260'
       molFileDir = 'iAF1260Molfiles';
   case 'Recon3DModel_Dec2017'
       molFileDir = [basePath '/data/metDatabase/explicit/molFiles'];
```

Set the thermochemical parameters for the model

```
switch modelName
    case 'Ec_iAF1260_flux1'
        T = 310.15; % Temperature in Kelvin
        compartments = {'Cytosol'; 'Extra_organism'; 'Periplasm'}; % Cell
compartment identifiers
        ph = [7.7; 7.7; 7.7]; % Compartment specific pH
        is = [0.25; 0.25; 0.25]; % Compartment specific ionic strength in
mol/L
        chi = [0; 90; 90]; % Compartment specific electrical potential
relative to cytosol in mV
    case 'iAF1260'
        T = 310.15; % Temperature in Kelvin
        compartments = ['c'; 'e'; 'p']; % Cell compartment identifiers
        ph = [7.7; 7.7; 7.7]; % Compartment specific pH
        is = [0.25; 0.25; 0.25]; % Compartment specific ionic strength in
mol/L
        chi = [0; 90; 90]; % Compartment specific electrical potential
relative to cytosol in mV
    case 'Recon3DModel_Dec2017'
        % Temperature in Kelvin
        T = 310.15;
        % Cell compartment identifiers
        compartments = ['c'; 'e'; 'g'; 'l'; 'm'; 'n'; 'r'; 'x';'i'];
        % Compartment specific pH
        ph = [7.2; 7.4; 6.35; 5.5; 8; 7.2; 7.2; 7; 7.2];
        % Compartment specific ionic strength in mol/L
        is = 0.15*ones(length(compartments),1);
        % Compartment specific electrical potential relative to cytosol in mV
        chi = [0; 30; 0; 19; -155; 0; 0;
-2.303*8.3144621e-3*T*(ph(compartments == 'x') - ph(compartments == 'c'))/
(96485.3365e-6); 0];
    case 'Recon3DModel_301'
        % Temperature in Kelvin
        T = 310.15;
        % Cell compartment identifiers
```

```
compartments = ['c'; 'e'; 'g'; 'l'; 'm'; 'n'; 'r'; 'x';'i'];
% Compartment specific pH
ph = [7.2; 7.4; 6.35; 5.5; 8; 7.2; 7.2; 7; 7.2];
% Compartment specific ionic strength in mol/L
is = 0.15*ones(length(compartments),1);
% Compartment specific electrical potential relative to cytosol in mV
chi = [0; 30; 0; 19; -155; 0; 0;
-2.303*8.3144621e-3*T*(ph(compartments == 'x') - ph(compartments == 'c'))/
(96485.3365e-6); 0];
otherwise
error('setup specific parameters for your model')
end
```

Set the default range of metabolite concentrations

```
switch modelName
    case 'Ec iAF1260 flux1'
        concMinDefault = 1e-5; % Lower bounds on metabolite concentrations
in mol/L
        concMaxDefault = 0.02; % Upper bounds on metabolite concentrations
in mol/L
        metBoundsFile=[];
    case 'iAF1260'
        concMinDefault = 1e-5; % Lower bounds on metabolite concentrations
in mol/L
        concMaxDefault = 0.02; % Upper bounds on metabolite concentrations
in mol/L
       metBoundsFile=[];
    case 'Recon3DModel_Dec2017'
        concMinDefault=1e-5; % Lower bounds on metabolite concentrations in
mol/L
        concMaxDefault=1e-2; % Upper bounds on metabolite concentrations in
mol/L
       metBoundsFile=which('HumanCofactorConcentrations.txt');%already in
the COBRA toolbox
    case 'Recon3DModel_301'
        concMinDefault=1e-5; % Lower bounds on metabolite concentrations in
mol/L
        concMaxDefault=1e-2; % Upper bounds on metabolite concentrations in
mol/L
       metBoundsFile=which('HumanCofactorConcentrations.txt');%already in
the COBRA toolbox
    otherwise
        error('setup specific parameters for your model')
end
```

Set the desired confidence level for estimation of thermochemical parameters

The confidence level for estimated standard transformed reaction Gibbs energies is used to quantitatively assign reaction directionality.

```
switch modelName
    case 'Ec_iAF1260_flux1'
        confidenceLevel = 0.95;
        DrGt0_Uncertainty_Cutoff = 20; %KJ/KMol
    case 'iAF1260'
        confidenceLevel = 0.95;
        DrGt0_Uncertainty_Cutoff = 20; %KJ/KMol
    case 'Recon3DModel_Dec2017'
        confidenceLevel = 0.95;
        DrGt0_Uncertainty_Cutoff = 20; %KJ/KMol
    otherwise
        confidenceLevel = -1;%bypass addition of uncertainty temporarily
        %confidenceLevel = 0.95;
        DrGt0_Uncertainty_Cutoff = 20; %KJ/KMol
end
```

Prepare folder for results

```
if ~exist(resultsPath,'dir')
    mkdir(resultsPath)
end
cd(resultsPath)
```

Set the print level and decide to record a diary or not (helpful for debugging)

```
printLevel=2;
diary([resultsPath filesep 'diary.txt'])
```

Setup a thermodynamically constrained model

Read in the metabolite bounds

```
setDefaultConc=1;
setDefaultFlux=0;
rxnBoundsFile=[];
model=readMetRxnBoundsFiles(model,setDefaultConc,setDefaultFlux,concMinDefault,concMaxDefault,metBoundsFile,rxnBoundsFile,printLevel);
```

```
Reading metabolite conc bounds from: /home/rfleming/work/sbgCloud/code/fork-cobratoolbox/src/analysis/ther
             adp[c]
                          1e-07
                                        0.0019
                                        0.0094
             adp[m]
                          0.0026
             amp[c]
                          1e-07
                                        0.0012
             atp[c]
                        0.00129
                                        0.0049
                         0.0028
                                        0.0204
             atp[m]
                                   0.0001168
             coa[c]
                       2.92e-05
             coa[m]
                         0.0022
                                        0.0039
```

na1[c]	1e-07	0.025
na1[e]	0.1326	0.1554
nad[c]	0.00010546	0.0007572
nad[m]	0.0005	0.0075
nadh[c]	9.2574e-07	0.00038294
nadh[m]	1e-07	0.0011
nadp[c]	1e-07	5.8284e-06
nadp[m]	1e-07	0.0015
nadph[c]	1e-07	0.00037523
nadph[m]	1e-07	0.0042
nh4[c]	0.0007	0.0009
pi[c]	0.001	0.0063
ppi[c]	0.0021	0.0076
udp[g]	1.4e-06	0.00014

Check inputs

```
model =
configureSetupThermoModelInputs(model,T,compartments,ph,is,chi,concMinDefault
,concMaxDefault,confidenceLevel);
```

Field metCompartments is missing from model structure. Attempting to create it. Attempt to create field metCompartments successful.

Warning: Setting temperature to a value other than 298.15 K may introduce error, since enthalpies and heat

Add InChI to model

```
%[model, pKaErrorMets] = setupComponentContribution(model,molFileDir);
model = addInchiToModel(model, molFileDir, 'sdf', printLevel);
```

```
Creating MetStructures.sdf from molfiles.
Percentage of metabolites without mol files: 9.1%
Converting SDF to InChI strings.
5835 = number of model metabolites
5835 ... with mol files
0 ... without mol files
4949 ... with nonstandard inchi
886 ... without nonstandard inchi
108 ... compositie inchi removed
```

Add pseudoisomers to model

```
[model, nonphysicalMetBool, pKaErrorMetBool] =
addPseudoisomersToModel(model, printLevel);
```

Estimating metabolite pKa values.

```
ChemAxon's pKa calculator plugin returned an error for 2 metabolites:
                         { 'InChI=1/C5H3N4O3/c10-3-1-2(7-4(11)6-1)8-5(12)9-3/h(H3,6,7...' }
    {'pchol2ste_hs'}
                         { 'InChI=1/C26H55N07P/c1-5-6-7-8-9-10-11-12-13-14-15-16-17-1...' }
```

Assuming that metabolite species in model.metFormulas are representative for metabolites where pKa could r 5835 = number of model metabolites

217 = number of nonphysical model metabolites

3 = number of model metabolites with pKa error

Check elemental balancing of metabolic reactions

```
ignoreBalancingOfSpecifiedInternalReactions=1;
if ~exist('massImbalance','var')
    if isfield(model,'Srecon')
        model.S=model.Srecon;
    end
    % Check for imbalanced reactions
    fprintf('\nChecking mass and charge balance.\n');
    %Heuristically identify exchange reactions and metabolites exclusively
involved in exchange reactions
    if ~isfield(model,'SIntMetBool') || ~isfield(model,'SIntRxnBool') ||
ignoreBalancingOfSpecifiedInternalReactions
        %finds the reactions in the model which export/import from the model
        %boundary i.e. mass unbalanced reactions
        %e.g. Exchange reactions
             Demand reactions
              Sink reactions
        model = findSExRxnInd(model,[],printLevel);
    end
    if ignoreBalancingOfSpecifiedInternalReactions
        [nMet,nRxn]=size(model.S);
        ignoreBalancingMetBool=false(nMet,1);
        for m=1:nMet
              if strcmp(model.mets{m}, 'Rtotal3coa[m]')
응
                  pause(0.1);
응
              end
            if ~isempty(model.metFormulas{m})
ignoreBalancingMetBool(m,1)=numAtomsOfElementInFormula(model.metFormulas{m}, '
FULLR');
            end
        end
ignoreBalancingRxnBool=getCorrespondingCols(model.S,ignoreBalancingMetBool,mo
del.SIntRxnBool, 'inclusive');
        SIntRxnBool=model.SIntRxnBool;
        model.SIntRxnBool=model.SIntRxnBool & ~ignoreBalancingRxnBool;
    end
    printLevelcheckMassChargeBalance=-1; % -1; % print problem reactions to
a file
    %mass and charge balance can be checked by looking at formulas
[massImbalance,imBalancedMass,imBalancedCharge,imBalancedRxnBool,Elements,mis
singFormulaeBool,balancedMetBool]...
checkMassChargeBalance(model,printLevelcheckMassChargeBalance,resultsBaseFile
Name);
```

```
model.balancedRxnBool=~imBalancedRxnBool;
model.balancedMetBool=balancedMetBool;
model.Elements=Elements;
model.missingFormulaeBool=missingFormulaeBool;

%reset original boolean vector
if ignoreBalancingOfSpecifiedInternalReactions
    model.SIntRxnBool=SIntRxnBool;
end
```

```
Checking mass and charge balance.

Assuming biomass reaction is: biomass_maintenance

ATP demand reaction is not considered an exchange reaction by default. It should be mass balanced:

DM_atp_c_ h2o[c] + atp[c] -> h[c] + adp[c] + pi[c]

There are mass imbalanced reactions, see /home/rfleming/work/sbgCloud/code/fork-COBRA.tutorials/analysis/v

There are mass balanced, but charge imbalanced reactions, see /home/rfleming/work/sbgCloud/code/fork-COBRA.
```

Create the thermodynamic training model

removing: /home/rfleming/work/sbgCloud/code/fork-cobratoolbox/src/analysis/thermo/componentContribution/ol

```
removing: /home/rfleming/work/sbgCloud/code/fork-cobratoolbox/src/analysis/thermo/directionalityReport/old
removing: /home/rfleming/work/sbgCloud/code/fork-cobratoolbox/src/analysis/thermo/groupContribution/old
removing: /home/rfleming/work/sbgCloud/code/fork-cobratoolbox/src/analysis/thermo/inchi/old
removing: /home/rfleming/work/sbgCloud/code/fork-cobratoolbox/src/analysis/thermo/molFiles/old
removing: /home/rfleming/work/sbgCloud/code/fork-cobratoolbox/src/analysis/thermo/protons/old
removing: /home/rfleming/work/sbgCloud/code/fork-cobratoolbox/src/analysis/thermo/trainingModel/old
adding: /home/rfleming/work/sbgCloud/code/fork-cobratoolbox/src/analysis/thermo/componentContribution/new
adding: /home/rfleming/work/sbgCloud/code/fork-cobratoolbox/src/analysis/thermo/groupContribution/new
adding: /home/rfleming/work/sbgCloud/code/fork-cobratoolbox/src/analysis/thermo/inchi/new
adding: /home/rfleming/work/sbgCloud/code/fork-cobratoolbox/src/analysis/thermo/molFiles/new
adding: /home/rfleming/work/sbqCloud/code/fork-cobratoolbox/src/analysis/thermo/protons/new
adding: /home/rfleming/work/sbgCloud/code/fork-cobratoolbox/src/analysis/thermo/trainingModel/new
ChemAxon Marvin Beans is installed and working.
   linux-vdso.so.1 (0x00007ffe937e7000)
    libc.so.6 => /lib/x86_64-linux-gnu/libc.so.6 (0x00007f25ad526000)
   libopenbabel.so.5 => /usr/lib/libopenbabel.so.5 (0x00007f25ad2d6000)
    libstdc++.so.6 => /usr/lib/x86_64-linux-gnu/libstdc++.so.6 (0x00007f25ad0bc000)
   libgcc_s.so.1 => /usr/local/bin/MATLAB/R2021a/sys/os/glnxa64/libgcc_s.so.1 (0x00007f25acea4000)
    /lib64/ld-linux-x86-64.so.2 (0x00007f25ad746000)
   libdl.so.2 => /lib/x86_64-linux-gnu/libdl.so.2 (0x00007f25ace9c000)
   libz.so.1 => /lib/x86_64-linux-gnu/libz.so.1 (0x00007f25ace80000)
   libm.so.6 => /lib/x86_64-linux-gnu/libm.so.6 (0x00007f25acd31000)
```

```
libgomp.so.1 => /usr/lib/x86_64-linux-gnu/libgomp.so.1 (0x00007f25accec000)
libpthread.so.0 => /lib/x86_64-linux-gnu/libpthread.so.0 (0x00007f25accc9000)
```

babel must depend on the system libstdc++.so.6 not the one from MATLAB

Trying to edit the 'LD_LIBRARY_PATH' to make sure that it has the correct system path before the Matlab pa The solution will be arch dependent Successfully added 3914 values from TECRDB Successfully added 223 formation energies Successfully added 13 redox potentials mol2inchi: could not generate inchi for C00080 0 molecules converted 2 audit log messages createInChIStruct: no molecule identifier in C00080 mol2inchi: could not generate inchi for C00080 0 molecules converted 2 audit log messages mol2inchi: could not generate inchi for C00080 0 molecules converted 2 audit log messages mol2inchi: could not generate inchi for C00080 0 molecules converted 2 audit log messages mol2inchi: could not generate inchi for C00125 babel: Alias R was not chemically interpreted createInChIStruct: no molecule identifier in C00125 mol2inchi: could not generate inchi for C00125 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C00125 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C00125 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C00126 babel: Alias R was not chemically interpreted createInChIStruct: no molecule identifier in C00126 mol2inchi: could not generate inchi for C00126 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C00126 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C00126 babel: Alias R was not chemically interpreted mol2inchi: no annotation in C00225 createInChIStruct: no molecule identifier in C00225 mol2inchi: no annotation in C00225 mol2inchi: no annotation in C00225 mol2inchi: no annotation in C00225 mol2inchi: could not generate inchi for C00229 babel: Alias R was not chemically interpreted createInChIStruct: no molecule identifier in C00229 mol2inchi: could not generate inchi for C00229 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C00229 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C00229 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C00342 babel: Alias R was not chemically interpreted createInChIStruct: no molecule identifier in C00342 mol2inchi: could not generate inchi for C00342 babel: Alias R was not chemically interpreted

mol2inchi: could not generate inchi for C00342

babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C00342 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C00343 babel: Alias R was not chemically interpreted createInChIStruct: no molecule identifier in C00343 mol2inchi: could not generate inchi for C00343 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C00343 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C00343 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C01003 babel: Alias R was not chemically interpreted createInChIStruct: no molecule identifier in C01003 mol2inchi: could not generate inchi for C01003 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C01003 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C01003 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C01194 babel: Alias R was not chemically interpreted createInChIStruct: no molecule identifier in C01194 mol2inchi: could not generate inchi for C01194 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C01194 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C01194 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C01209 babel: Alias R was not chemically interpreted createInChIStruct: no molecule identifier in C01209 mol2inchi: could not generate inchi for C01209 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C01209 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C01209 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C01277 babel: Alias R was not chemically interpreted createInChIStruct: no molecule identifier in C01277 mol2inchi: could not generate inchi for C01277 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C01277 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C01277 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C01281 babel: Alias R was not chemically interpreted createInChIStruct: no molecule identifier in C01281 mol2inchi: could not generate inchi for C01281 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C01281 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C01281 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C01299 babel: Alias R was not chemically interpreted createInChIStruct: no molecule identifier in C01299 mol2inchi: could not generate inchi for C01299 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C01299 babel: Alias R was not chemically interpreted

mol2inchi: could not generate inchi for C01299 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C01931 babel: Alias R was not chemically interpreted createInChIStruct: no molecule identifier in C01931 mol2inchi: could not generate inchi for C01931 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C01931 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C01931 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C02163 babel: Alias R was not chemically interpreted createInChIStruct: no molecule identifier in C02163 mol2inchi: could not generate inchi for C02163 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C02163 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C02163 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C02553 babel: Alias R was not chemically interpreted createInChIStruct: no molecule identifier in C02553 mol2inchi: could not generate inchi for C02553 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C02553 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C02553 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C02554 babel: Alias R was not chemically interpreted createInChIStruct: no molecule identifier in C02554 mol2inchi: could not generate inchi for C02554 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C02554 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C02554 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C02780 0 molecules converted 2 audit log messages

createInChIStruct: no molecule identifier in C02780
mol2inchi: could not generate inchi for C02780
0 molecules converted
2 audit log messages

mol2inchi: could not generate inchi for C02780
0 molecules converted
2 audit log messages

mol2inchi: could not generate inchi for C02780
0 molecules converted

0 molecules converted
2 audit log messages

mol2inchi: could not generate inchi for C02839 babel: Alias R was not chemically interpreted createInChIStruct: no molecule identifier in C02839 mol2inchi: could not generate inchi for C02839 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C02839 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C02839 babel: Alias R was not chemically interpreted babel: Alias R was not chemically interpreted

mol2inchi: could not generate inchi for C02988 babel: Alias R was not chemically interpreted createInChIStruct: no molecule identifier in C02988 mol2inchi: could not generate inchi for C02988 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C02988 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C02988 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C02992 babel: Alias R was not chemically interpreted createInChIStruct: no molecule identifier in C02992 mol2inchi: could not generate inchi for C02992 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C02992 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C02992 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C03127 babel: Alias R was not chemically interpreted createInChIStruct: no molecule identifier in C03127 mol2inchi: could not generate inchi for C03127 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C03127 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C03127 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C03511 babel: Alias R was not chemically interpreted createInChIStruct: no molecule identifier in C03511 mol2inchi: could not generate inchi for C03511 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C03511 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C03511 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C03875 babel: Alias R was not chemically interpreted createInChIStruct: no molecule identifier in C03875 mol2inchi: could not generate inchi for C03875 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C03875 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C03875 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C03939 babel: Alias R was not chemically interpreted createInChIStruct: no molecule identifier in C03939 mol2inchi: could not generate inchi for C03939 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C03939 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C03939 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C04246 babel: Alias R was not chemically interpreted createInChIStruct: no molecule identifier in C04246 mol2inchi: could not generate inchi for C04246 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C04246 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C04246 babel: Alias R was not chemically interpreted mol2inchi: could not generate inchi for C04618

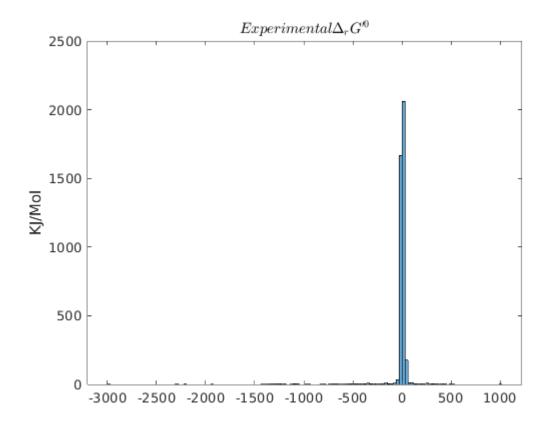
```
babel: Alias R was not chemically interpreted
createInChIStruct: no molecule identifier in C04618
mol2inchi: could not generate inchi for C04618
babel: Alias R was not chemically interpreted
mol2inchi: could not generate inchi for C04618
babel: Alias R was not chemically interpreted
mol2inchi: could not generate inchi for C04618
babel: Alias R was not chemically interpreted
mol2inchi: could not generate inchi for C04688
babel: Alias R was not chemically interpreted
createInChIStruct: no molecule identifier in C04688
mol2inchi: could not generate inchi for C04688
babel: Alias R was not chemically interpreted
mol2inchi: could not generate inchi for C04688
babel: Alias R was not chemically interpreted
mol2inchi: could not generate inchi for C04688
babel: Alias R was not chemically interpreted
mol2inchi: could not generate inchi for C06020
babel: Alias R was not chemically interpreted
createInChIStruct: no molecule identifier in C06020
mol2inchi: could not generate inchi for C06020
babel: Alias R was not chemically interpreted
mol2inchi: could not generate inchi for C06020
babel: Alias R was not chemically interpreted
mol2inchi: could not generate inchi for C06020
babel: Alias R was not chemically interpreted
mol2inchi: could not generate inchi for C06021
babel: Alias R was not chemically interpreted
createInChIStruct: no molecule identifier in C06021
mol2inchi: could not generate inchi for C06021
babel: Alias R was not chemically interpreted
mol2inchi: could not generate inchi for C06021
babel: Alias R was not chemically interpreted
mol2inchi: could not generate inchi for C06021
babel: Alias R was not chemically interpreted
mol2inchi: could not generate inchi for C06567
babel: Alias R was not chemically interpreted
createInChIStruct: no molecule identifier in C06567
mol2inchi: could not generate inchi for C06567
babel: Alias R was not chemically interpreted
mol2inchi: could not generate inchi for C06567
babel: Alias R was not chemically interpreted
mol2inchi: could not generate inchi for C06567
babel: Alias R was not chemically interpreted
672 = number of model metabolites
657 ... with mol files
15 ... without mol files
627 ... with nonstandard inchi
45 ... without nonstandard inchi
0 ... compositie inchi removed
```

Estimating metabolite pKa values for training trainingModel...

```
...done.
```

There are mass imbalanced reactions, see /home/rfleming/work/sbgCloud/code/fork-COBRA.tutorials/analysis/vPerforming reverse Legendre transform

```
figure
histogram(trainingModel.DrGt0)
title('$Experimental \smallskip \Delta_{r}
G^{\prime0}$','Interpreter','latex')
```



```
fprintf('%u%s\n',nnz(trainingModel.DrGt0==0),' = number of zero DrGt0, i.e.
experimental apparent equilibrium constant equal to one')
```

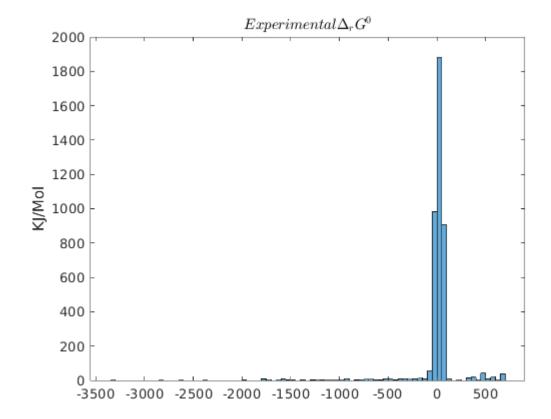
35 = number of zero DrGtO, i.e. experimental apparent equilibrium constant equal to one

formulas =
printRxnFormula(trainingModel,trainingModel.rxns(trainingModel.DrGt0==0));

```
TECRDB_79
           C01101
                      <=>
                             C00231
TECRDB_244
            C00041
                      <=>
                            C00133
TECRDB_580
             C00002 + C00065 + C01650
                                              C00013 + C00020 + C02553
                                        <=>
TECRDB_698
            C00003 + C00644
                                      C00004 + C00085
                              <=>
TECRDB_733
            C01101
                     <=>
                             C00231
            C00002 + C00055
                                      C00008 + C00112
TECRDB_815
                              <=>
TECRDB_1090
           C00001 + C00006 + C00311
                                       <=>
                                               C00005 + C00026 + C00288
TECRDB_1272
           C00001 + C06322
                                       C06749
                             <=>
             C00041
                              C00133
TECRDB_2030
                       <=>
TECRDB_2202
             C00003 + C00579
                              <=>
                                      C00004 + C00248
TECRDB_2339
             C00063 + C00103
                                      C00013 + C00501
                               <=>
TECRDB_2392
             C01213
                       <=>
                              C00683
TECRDB_2584
             C00041
                        <=>
                              C00133
TECRDB_2620
             C00026 + C00041
                               <=>
                                      C00022 + C00025
TECRDB_2621
             C00002 + C01107
                               <=>
                                      C00008 + C01143
TECRDB_2629
                              C00217
             C00025
                        <=>
TECRDB 2639
             C00002 + C00104
                               <=>
                                      C00008 + C00081
TECRDB 2746
             C00031
                       <=>
                              C00095
             C00010 + C00042 + C00044
TECRDB_2791
                                               C00009 + C00035 + C00091
                                        <=>
TECRDB_2841
             2 C00008 <=>
                               C00002 + C00020
TECRDB_2894
             C00041
                              C00133
                       <=>
TECRDB_3608
                              C00095
             C00031
                        <=>
```

```
C00047
TECRDB_3640
                      <=>
                             C00739
TECRDB_3803 C00636
                      <=>
                             C00275
TECRDB_3808
TECRDB_4052
            C00075 + C00103
                             <=>
                                    C00013 + C00029
            C00041
                      <=>
                             C00133
TECRDB_4271
            C00935
                             C00190
                      <=>
TECRDB_4375 C00123
                            C01570
                      <=>
TECRDB_4377
                                    C00106 + C00620
            C00009 + C00299
                             <=>
C00022 + C00025
                              <=>
TECRDB_4536 C00031
                            C00095
                      <=>
TECRDB_4537 C00031
                             C00095
                       <=>
FORM_C00023
                      C00023
               <=>
FORM_C00034
                <=>
                      C00034
FORM_C00080
                <=>
                      C00080
```

```
figure
histogram(trainingModel.DrG0)
title('$Experimental \medskip \Delta_{r} G^{0}$','Interpreter','latex')
ylabel('KJ/Mol')
```



```
fprintf('%u%s\n',nnz(trainingModel.DrG0==0),' = number of zero DrG0. i.e.
equilibrium constant equal to one and same number of hydrogens on both
sides')
```

16 = number of zero DrGO. i.e. equilibrium constant equal to one and same number of hydrogens on both side

```
formulas =
printRxnFormula(trainingModel,trainingModel.rxns(trainingModel.DrG0==0));
```

```
C01101
TECRDB_733
                <=> C00231
C06749
C00133
                 <=>
                 <=>
                      C00683
                 <=>
                      C00133
                      C00217
                 <=>
TECRDB_2894 C00041
                      C00133
                 <=>
                     C00739
TECRDB_3640 C00047
                 <=>
TECRDB_4052 C00041
                     C00133
                 <=>
TECRDB_4271 C00935
                     C00190
                 <=>
TECRDB_4375 C00123
                      C01570
                 <=>
FORM_C00023
          <=> C00023
FORM_C00034
            <=> C00034
FORM_C00080
            <=>
                 C00080
```

Create Group Incidence Matrix

Create the group incidence matrix (G) for the combined set of all metabolites.

```
save('data_prior_to_createGroupIncidenceMatrix')
```

```
%param.fragmentationMethod='manual';
param.fragmentationMethod='abinito';
param.printLevel=0;
param.modelCache=['autoFragment_' modelName];
param.debug=1;
param.radius=2;
```

```
combinedModel = createGroupIncidenceMatrix(model, trainingModel, param);

Creating group incidence matrix
There are 574 fragments unique to the training model.
There are 914 fragments in common between the training and test models.
There are 2659 fragments unique to the test model.

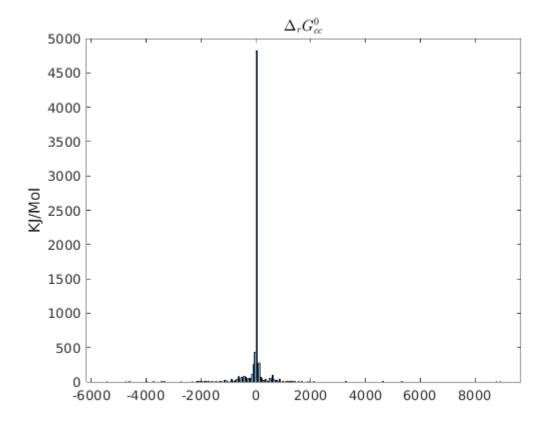
save('data_prior_to_componentContribution','model','combinedModel')
```

Apply component contribution method

```
if ~isfield(model,'DfG0')
    [model,solution] = componentContribution(model,combinedModel);
end
```

Running Component Contribution method

```
figure
histogram(model.DrG0(~model.unconstrainedDrG0_cc))
title('$\Delta_{r} G^{0}_{cc}$','Interpreter','latex')
ylabel('KJ/Mol')
```



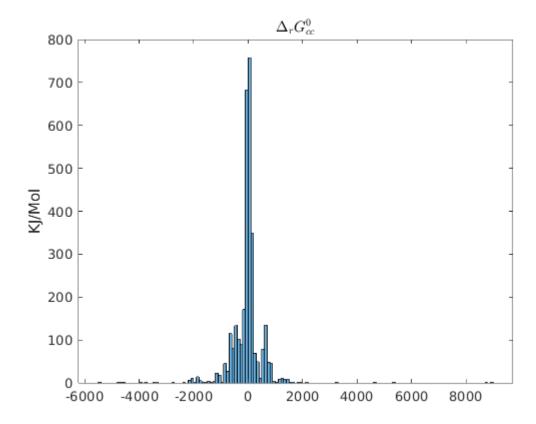
```
fprintf('%u%s\n',length(model.DrG0),' model reactions')
```

10600 model reactions

```
fprintf('%u%s\n',nnz(model.unconstrainedDrG0_cc),' of which have partially
unconstrained groups in DrG0_cc')
```

3147 of which have partially unconstrained groups in DrGO_cc

```
figure
model.transportRxnBool = transportReactionBool(model);
bool = model.SIntRxnBool & ~model.transportRxnBool &
  ~model.unconstrainedDrG0_cc;
histogram(model.DrG0(bool))
title('$\Delta_{r} G^{0}_{cc}$','Interpreter','latex')
ylabel('KJ/Mol')
```



```
fprintf('%u%s\n',length(model.DrG0),' model reactions')
10600 model reactions
fprintf('%u%s\n',nnz(model.unconstrainedDrG0_cc),' of which have partially
unconstrained groups in DrGO_cc')
3147 of which have partially unconstrained groups in DrGO_cc
ind=find(model.unconstrainedDrG0_cc);
formulas = printRxnFormula(model.model.rxns(ind(1:10)));
2AMACSULT
          2amac[c] + nadph[c] + paps[c]
                                        ->
                                              nadp[c] + Lcyst[c] + pap[c]
2DR1PP h2o[c] + 2dr1p[c] -> pi[c] + drib[c]
34DHPLACOX h2o[c] + nad[c] + 34dhpac[c] -> 2h[c] + nadh[c] + 34dhpha[c]
34DHPLACOX_NADP_ h2o[c] + nadp[c] + 34dhpac[c]
                                              <=> 2 h[c] + nadph[c] + 34dhpha[c]
34DHXMANDACOX
            h2o[c] + nad[c] + 34dhmald[c] ->
                                                2 h[c] + nadh[c] + 34dhoxmand[c]
                                                        2 h[c] + nadph[c] + 34dhoxmand[c]
34DHXMANDACOX_NADP_ h2o[c] + nadp[c] + 34dhmald[c]
                                                  <=>
3AIBTm 2mop[m] + glu_L[m]
                                 akg[m] + 3aib[m]
                           <=>
3HAO o2[c] + 3hanthrn[c]
                           -> h[c] + cmusa[c]
      h2o[m] + b2coa[m]
3HBCDm
                            <=>
                                 3hbcoa_R[m]
       h[c] + 34dhphe[c]
                                 co2[c] + dopa[c]
3HLYTCL
                            ->
```

Setup a thermodynamically constrained model

```
save('debug_prior_to_setupThermoModel')
```

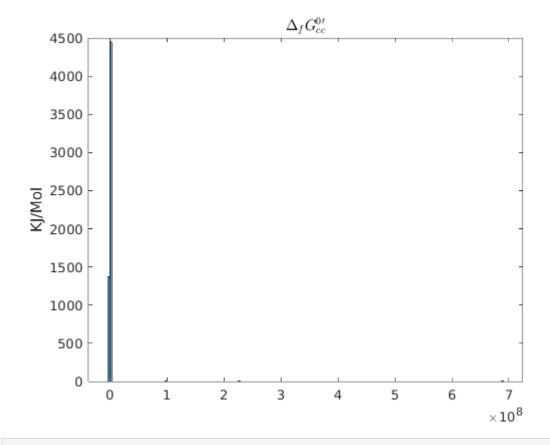
```
if ~isfield(model,'DfGt0')
  model = setupThermoModel(model,confidenceLevel);
```

Estimating standard transformed Gibbs energies of formation.

Estimating bounds on transformed Gibbs energies.

Additional effect due to possible change in chemical potential of Hydrogen ions for transport reactions. Additional effect due to possible change in electrical potential for transport reactions.

```
figure
histogram(model.DfGt0)
title('$\Delta_{f} G^{0\circ}_{cc}$','Interpreter','latex')
ylabel('KJ/Mol')
```



Generate a model with reactants instead of major microspecies

```
if ~isfield(model,'Srecon')
    printLevel_pHbalanceProtons=-1;

model=pHbalanceProtons(model,massImbalance,printLevel_pHbalanceProtons,result
sBaseFileName);
end
```

Warning: vonBertalanffy:pHbalanceProtons 'Hydrogen unbalanced reconstruction reactions exist!

Determine quantitative directionality assignments

```
if ~exist('directions','var') | 1
    fprintf('Quantitatively assigning reaction directionality.\n');
    [model, directions] =
    thermoConstrainFluxBounds(model,confidenceLevel,DrGt0_Uncertainty_Cutoff,printLevel);
end
```

```
Quantitatively assigning reaction directionality.
9/10600 reactions with DrGtMin=DrGtMax~=0
4/10600 reactions with DrGtMin=DrGtMax=0
The following reactions have DrGtMax=DrGtMin=0:
H2Oter
       h2o[c] <=>
                      h2o[r]
      h2o[n]
H2Otn
               <=>
                     h2o[c]
<=>
HMR_1095 h[c]
                      h[n]
ACYP
```

Analyse thermodynamically constrained model

Choose the cutoff for probablity that reaction is reversible

```
cumNormProbCutoff=0.2;
```

Build Boolean vectors with reaction directionality statistics

```
[model,directions]=directionalityStats(model,directions,cumNormProbCutoff,pri
ntLevel);
```

```
9/10600 reactions with DrGtMin=DrGtMax~=0
4/10600 reactions with DrGtMin=DrGtMax=0
Qualitative internal reaction directionality:
             internal reconstruction reaction directions.
             forward reconstruction assignment.
     5208
             reverse reconstruction assignment.
        4
     3579
             reversible reconstruction assignment.
Quantitative internal reaction directionality:
     internal reconstruction reaction directions.
     8036
             of which have a thermodynamic assignment.
      751
            of which have no thermodynamic assignment.
     1636
            forward thermodynamic only assignment.
     1512 reverse thermodynamic only assignment.
     4888
             reversible thermodynamic only assignment.
Qualitiative vs Quantitative:
             Reversible -> Reversible
     2525
      347
             Reversible -> Forward
      583
             Reversible -> Reverse
      120
              Reversible -> Uncertain
              Forward -> Forward
     1286
      929
              Forward -> Reverse
     2362
              Forward -> Reversible
      631
              Forward -> Uncertain
              Reverse -> Reverse
        3
              Reverse -> Forward
        1
              Reverse -> Reversible
              Reversible -> Uncertain
```

```
Breakdown of relaxation of reaction directionality, Qualitiative vs Quantitative:
             qualitatively forward reactions that are quantitatively reversible (total).
     1183
             of which are quantitatively reversible by range of dGt0. P(\Delta_{r}G^{\infty}) > 0.7
             of which are quantitatively reversible by range of dGt0. 0.3< P(\Delta_{r})^{0} < 0
             of which are quantitatively reversible by range of dGt0. P(\Delta_{r}G^{\sigma}) < 0.3
     1179
       56
             of which are quantitatively forward by fixed dGr0t, but reversible by concentration alone (
             of which are quantitatively reverse by dGr0t, but reversible by concentration (negative fix
             of which are quantitatively forward by dGr0t, but reversible by concentration (positve fixed)
        0
        0
             of which are quantitatively reverse by dGr0t, but reversible by concentration (uncertain ne
             of which are quantitatively forward by dGr0t, but reversible by concentration (uncertain po
% directions
                  a structue of boolean vectors with different directionality
응
                  assignments where some vectors contain subsets of others
응
  qualtiative -> quantiative changed reaction directions
응
응
    .forward2Forward
    .forward2Reverse
응
응
    .forward2Reversible
응
    .forward2Uncertain
응
    .reversible2Forward
응
    .reversible2Reverse
응
    .reversible2Reversible
응
    .reversible2Uncertain
응
    .reverse2Forward
응
    .reverse2Reverse
응
    .reverse2Reversible
    .reverse2Uncertain
응
    .tightened
응
응
% subsets of qualtiatively forward -> quantiatively reversible
응
    .forward2Reversible bydGt0
응
    .forward2Reversible_bydGt0LHS
응
    .forward2Reversible bydGt0Mid
응
    .forward2Reversible bydGt0RHS
응
응
    .forward2Reversible byConc zero fixed DrG0
    .forward2Reversible_byConc_negative_fixed_DrG0
응
    .forward2Reversible_byConc_positive_fixed_DrG0
응
```

Write out reports on directionality changes for individual reactions to the results folder.

.forward2Reversible_byConc_negative_uncertain_DrG0
.forward2Reversible_byConc_positive_uncertain_DrG0

응

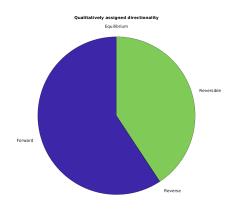
```
fprintf('%s\n','directionalityChangeReport...');
directionalityChangeReport...
directionalityChangeReport(model,directions,cumNormProbCutoff,printLevel,resultsBaseFileName)
```

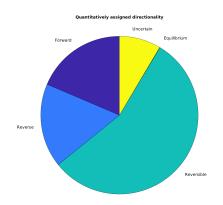
Generate pie charts with proportions of reaction directionalities and changes in directionality

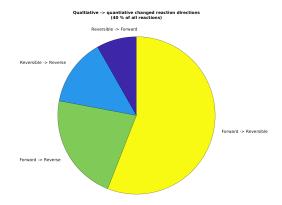
fprintf('%s\n','directionalityStatFigures...');

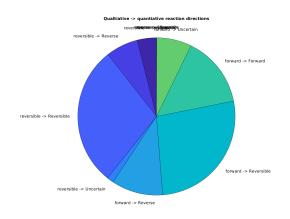
directionalityStatFigures...

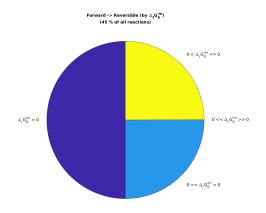
directionalityStatsFigures(directions,resultsBaseFileName)

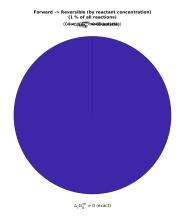








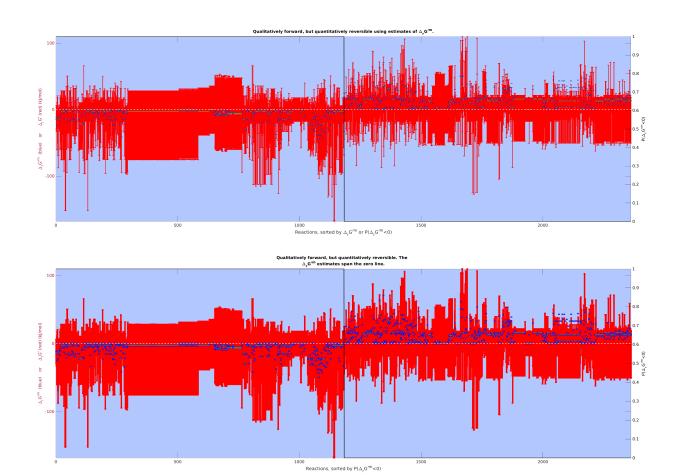




Generate figures to interpret the overall reasons for reaction directionality changes for the qualitatively forward now quantiatiavely reversible reactions

```
if any(directions.forward2Reversible)
    fprintf('%s\n','forwardReversibleFigures...');
    forwardReversibleFigures(model,directions,confidenceLevel)
end
```

forwardReversibleFigures...



Write out tables of experimental and estimated thermochemical parameters for the model

```
generateThermodynamicTables(model,resultsBaseFileName);
save([datestr(now,30) '_' modelName '_thermo'],'model')
save([datestr(now,30) '_vonB_tutorial_complete'])
```

REFERENCES

- [1] Fleming, R. M. T. & Thiele, I. von Bertalanffy 1.0: a COBRA toolbox extension to thermodynamically constrain metabolic models. Bioinformatics 27, 142–143 (2011).
- [2] Haraldsdóttir, H. S., Thiele, I. & Fleming, R. M. T. Quantitative assignment of reaction directionality in a multicompartmental human metabolic reconstruction. Biophysical Journal 102, 1703–1711 (2012).
- [3] Noor, E., Haraldsdóttir, H. S., Milo, R. & Fleming, R. M. T. Consistent Estimation of Gibbs Energy Using Component Contributions. PLoS Comput Biol 9, e1003098 (2013).
- [4] Fleming, R. M. T., Predicat, G., Haraldsdóttir, H. S., Thiele, I. von Bertalanffy 2.0 (in preparation).