



A modular tool to aggregate results from bioinformatics analyses across many samples into a single report.

Report generated on 2024-01-28, 22:12 CET based on data in: /work_beegfs/sunam239/genom1.cs

General Statistics

Copy table

Configure Columns

Plot

Showing 5/5 rows and 13/13 columns.

Sample Name	N50 (Kbp)	Assembly Length (Mbp)	Organism	Contigs	CDS	% Duplication	M Reads After Filtering	GC content	% PF	% Adapter	% Dups	% GC	M Seqs
1_short_reads_qc 1													
1_short_reads_qc 241155E_R1						4.1%	3.2	45.1%	98.4%	0.1%	33.2%	45%	1.6
1_short_reads_qc 3											33.6%	45%	1.6
3_hybrid_assembly quast 3_hybrid_assembly	4 332.3Kbp	4.5Mbp											
4_annotated_genome strain			Genus species	7.0	3883								

QUAST

QUAST is a quality assessment tool for genome assemblies, written by the Center for Algorithmic Biotechnology. DOI: 10.1093/bioinformatics/btt086.

Assembly Statistics

Copy table

Configure Columns

Plot

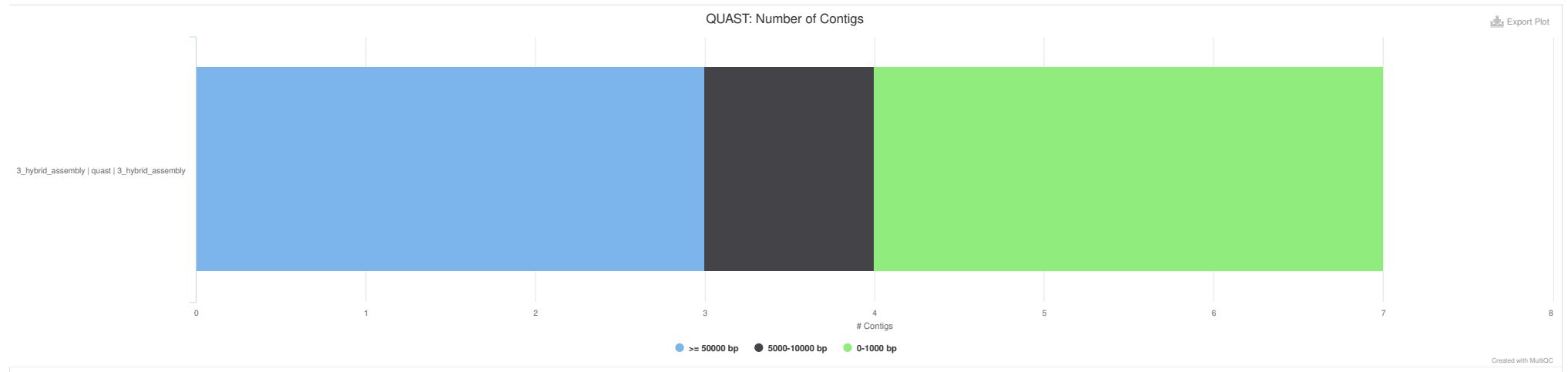
Showing 1/1 rows and 9/9 columns.

Sample Name	N50 (Kbp)	L50 (K)	Largest contig (Kbp)	Length (Mbp)	Misassemblies	Mismatches/100kbp	Indels/100kbp	Genes	Genome Fraction
3_hybrid_assembly quast 3_hybrid_assembly	4 332.3Kbp	0.0K	4 332.3Kbp	4.5Mbp	-	-	-	2 495	-%

Number of Contigs

This plot shows the number of contigs found for each assembly, broken down by length.

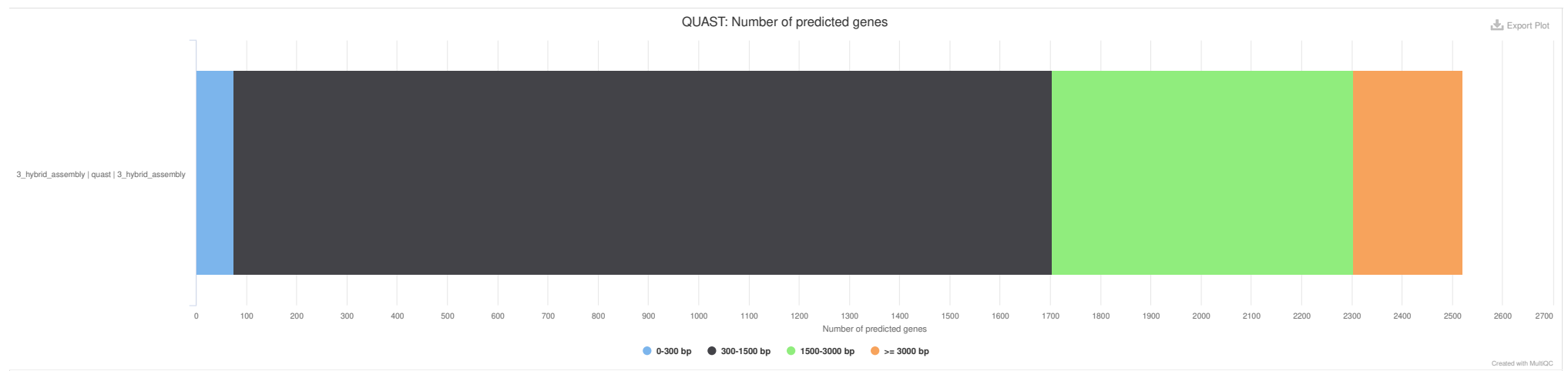
Counts Percentages



Number of Predicted Genes

This plot shows the number of predicted genes found for each assembly, broken down by length.

Counts Percentages



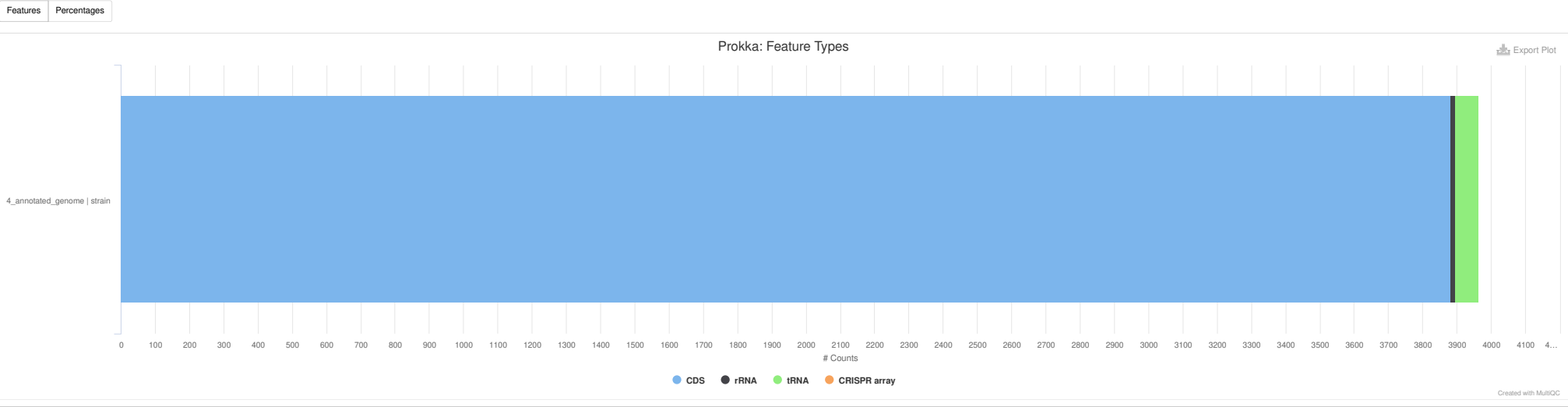
Number of Partially Predicted Genes

This plot shows the number of partially predicted genes found for each assembly, broken down by length.

Error - was not able to plot data.

Prokka

Prokka is a software tool for the rapid annotation of prokaryotic genomes. DOI: 10.1093/bioinformatics/btu153.
This barplot shows the distribution of different types of features found in each contig.



NanoStat

NanoStat reports various statistics from a long read sequencing dataset in FASTQ, BAM or sequencing summary format. DOI: 10.1093/bioinformatics/bty149.

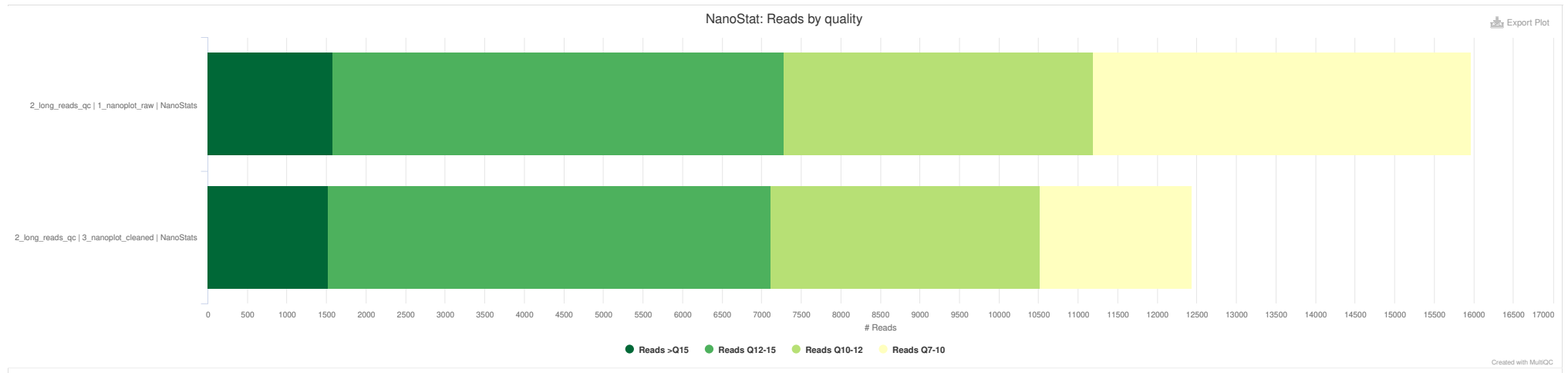
Summary Statistics

Copy tableConfigure ColumnsPlotShowing $2\frac{1}{2}$ rows and $5\frac{1}{7}$ columns.

Sample Name	Median length	Read N50	Median Qual	# Reads (K)	Total Bases (Mb)
2_long_reads_qc 1_nanoplot_raw NanoStats	3 270 bp	21 971 bp	11.7	16.0	146.3
2_long_reads_qc 3_nanoplot_cleaned NanoStats	4 477 bp	22 747 bp	12.5	12.4	131.7

Reads by quality

Read counts categorised by read quality (Phred score).

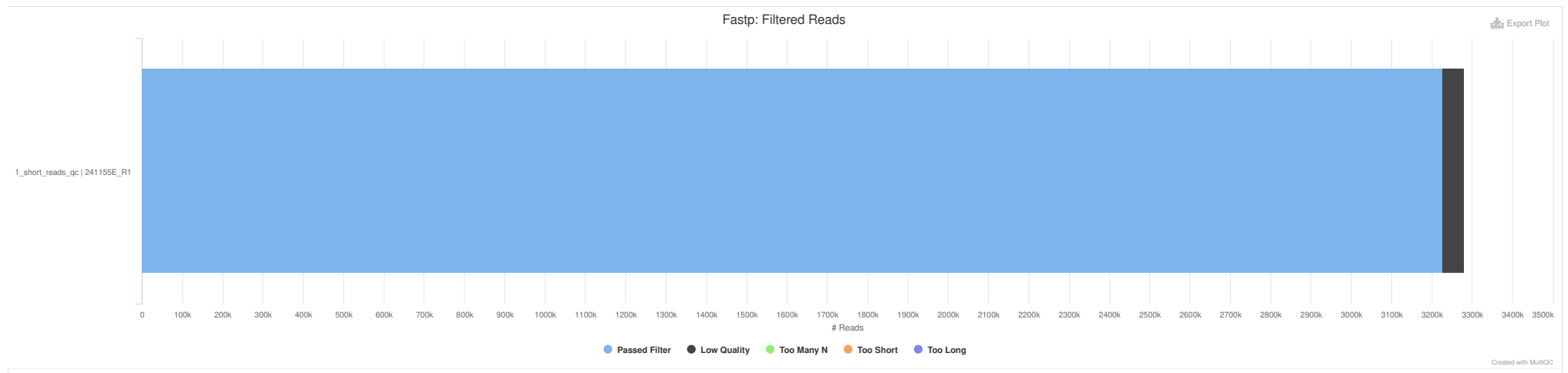
fastp

Version: 0.23.4

fastp An ultra-fast all-in-one FASTQ preprocessor (QC, adapters, trimming, filtering, splitting...). DOI: 10.1093/bioinformatics/bty560.

Filtered Reads

Filtering statistics of sampled reads.

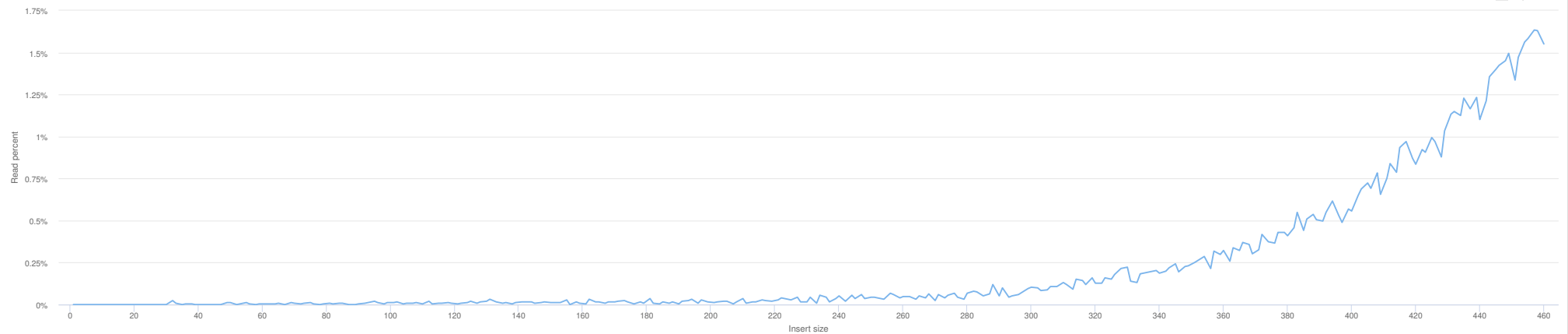
Insert Sizes

Insert size estimation of sampled reads.

Y-Limits:

Fastp: Insert Size Distribution

 Export Plot



Sequence Quality

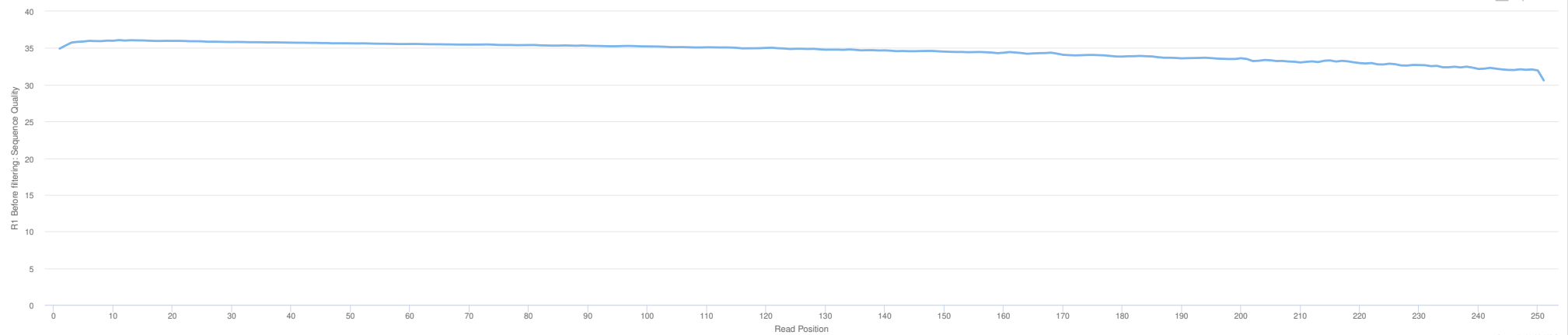
Average sequencing quality over each base of all reads.

Y-Limits:

Read 1: Before filtering Read 1: After filtering Read 2: Before filtering Read 2: After filtering

Fastp: Sequence Quality

 Export Plot

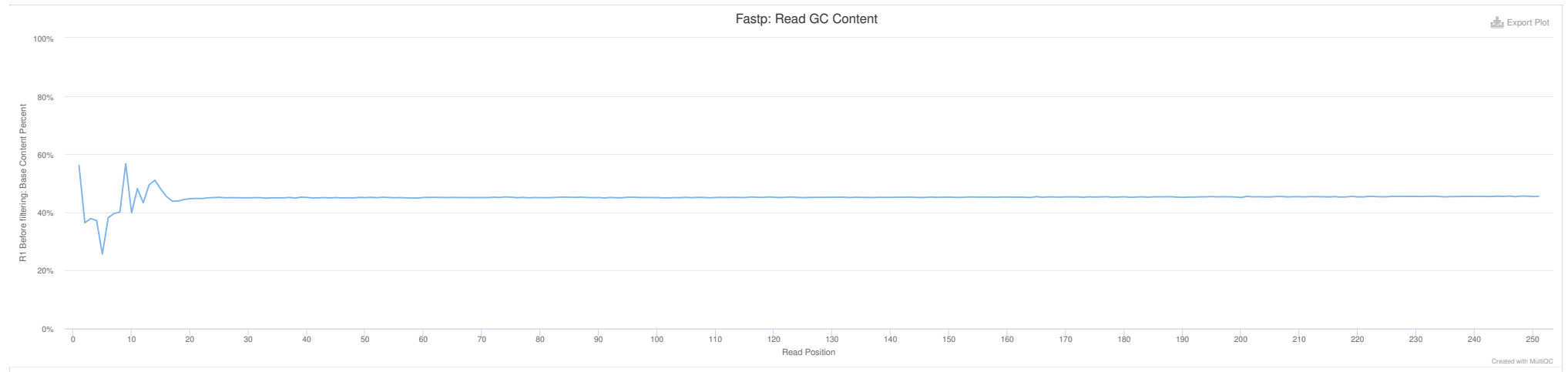


GC Content

Average GC content over each base of all reads.

Y-Limits:

Read 1: Before filtering Read 1: After filtering Read 2: Before filtering Read 2: After filtering

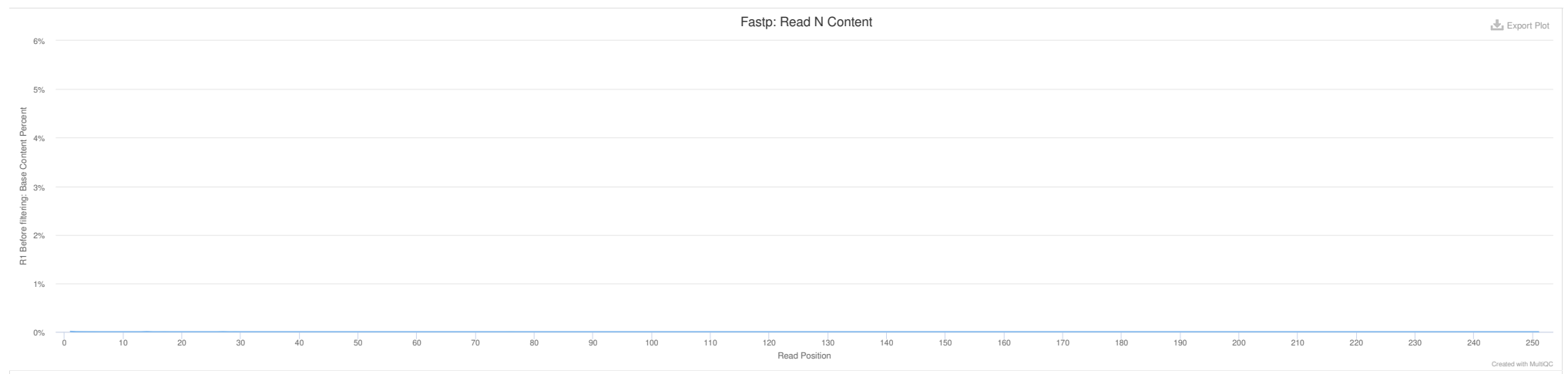


N content

Average N content over each base of all reads.

Y-Limits:

Read 1: Before filtering Read 1: After filtering Read 2: Before filtering Read 2: After filtering



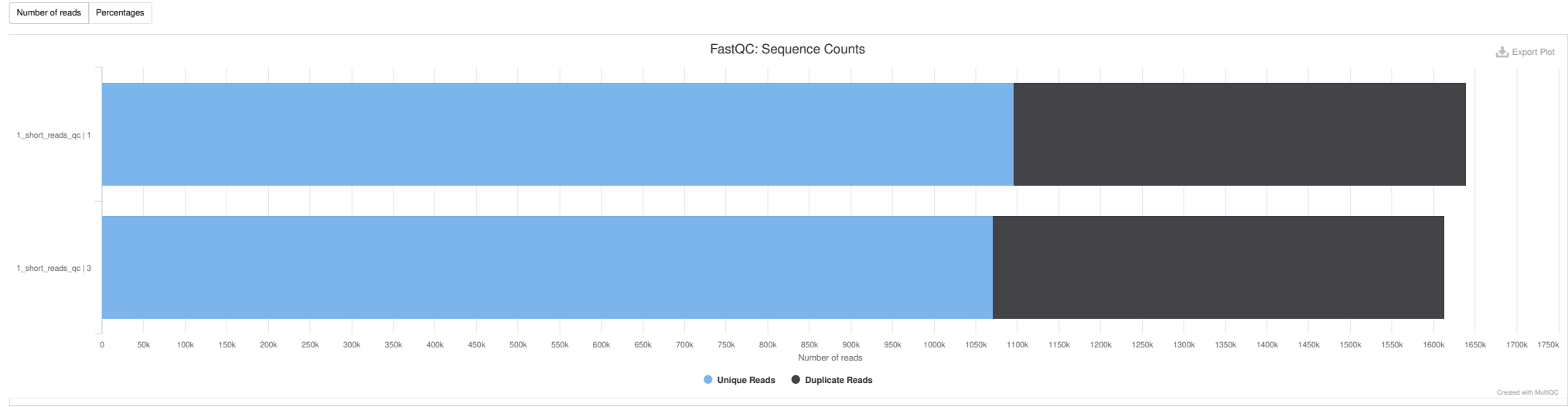
FastQC

Version: 0.12.1

FastQC is a quality control tool for high throughput sequence data, written by Simon Andrews at the Babraham Institute in Cambridge.

Sequence Counts

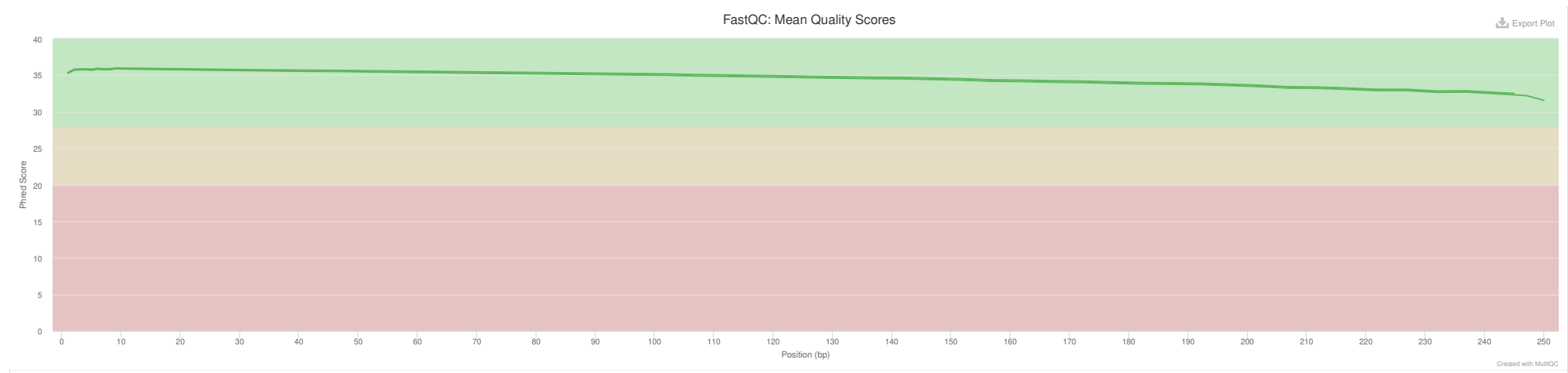
Sequence counts for each sample. Duplicate read counts are an estimate only.



Sequence Quality Histograms

2

The mean quality value across each base position in the read.



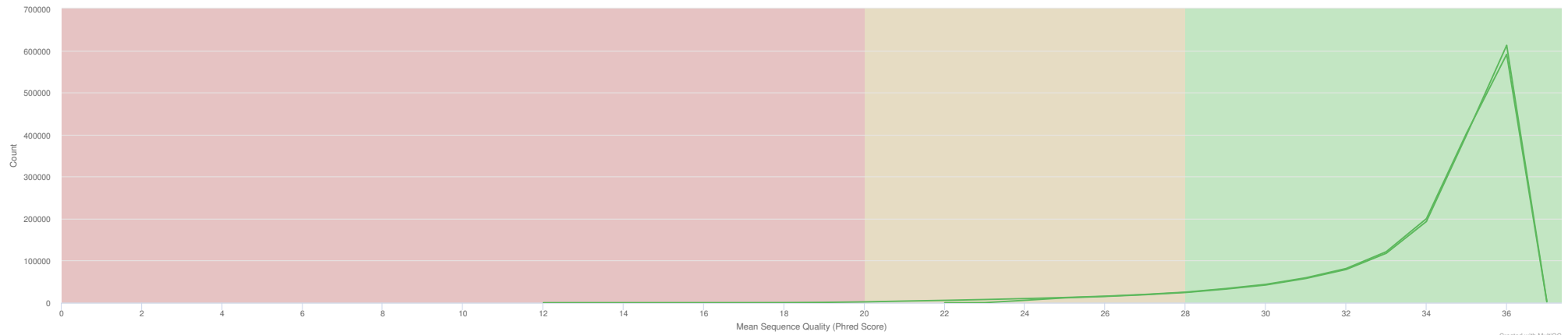
Per Sequence Quality Scores

2

The number of reads with average quality scores. Shows if a subset of reads has poor quality.

Y-Limits: on

FastQC: Per Sequence Quality Scores

[Export Plot](#)

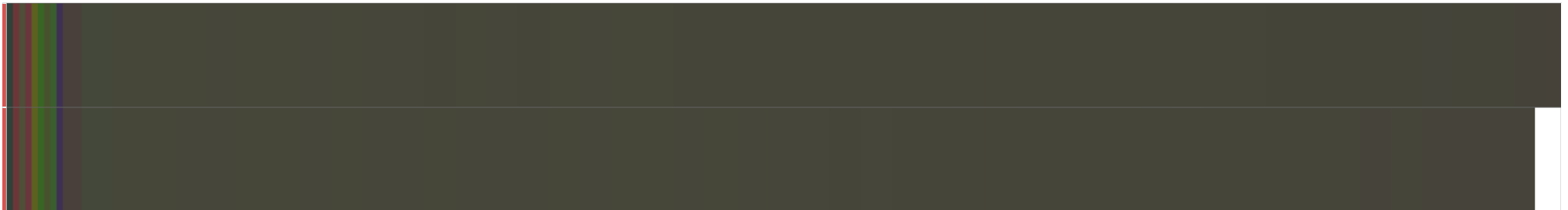
Per Base Sequence Content

The proportion of each base position for which each of the four normal DNA bases has been called.

[Click a sample row to see a line plot for that dataset.](#)

i Rollover for sample name

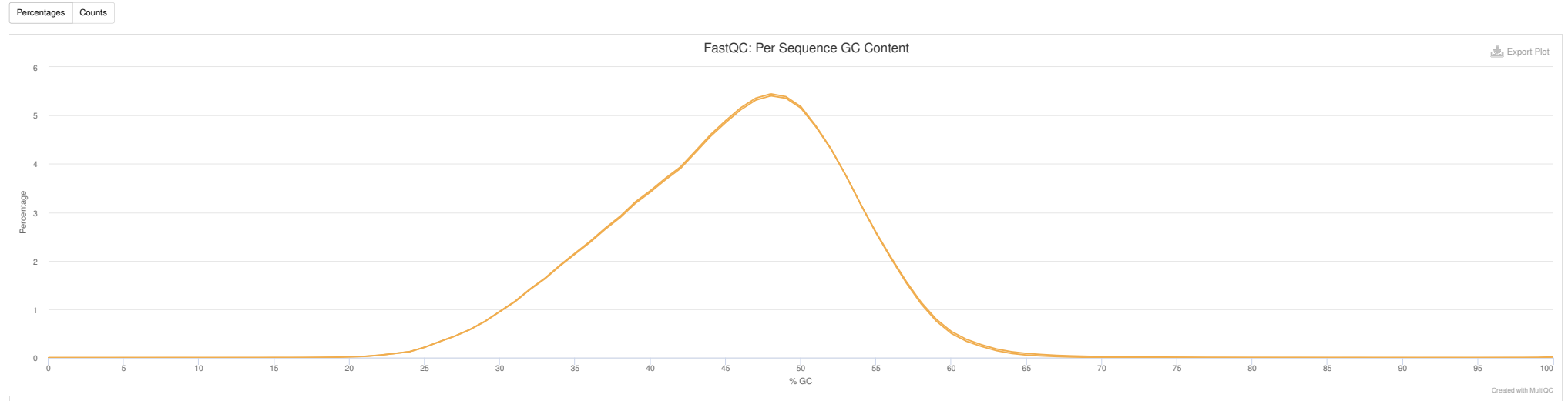
Position: - %T: - %C: - %A: - %G: -

[Export Plot](#)

Per Sequence GC Content

2

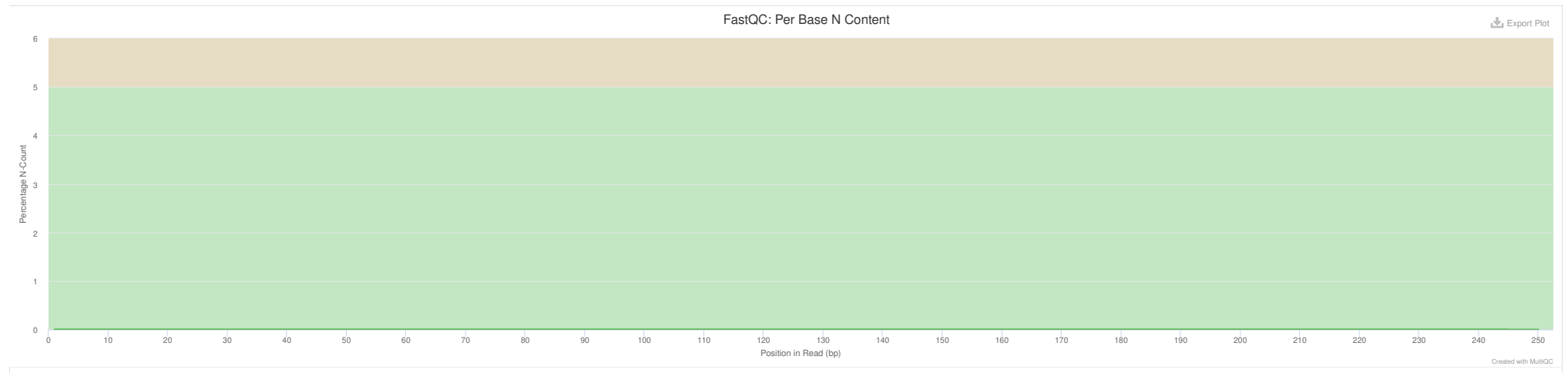
The average GC content of reads. Normal random library typically have a roughly normal distribution of GC content.

Y-Limits: 

Per Base N Content

2

The percentage of base calls at each position for which an N was called.

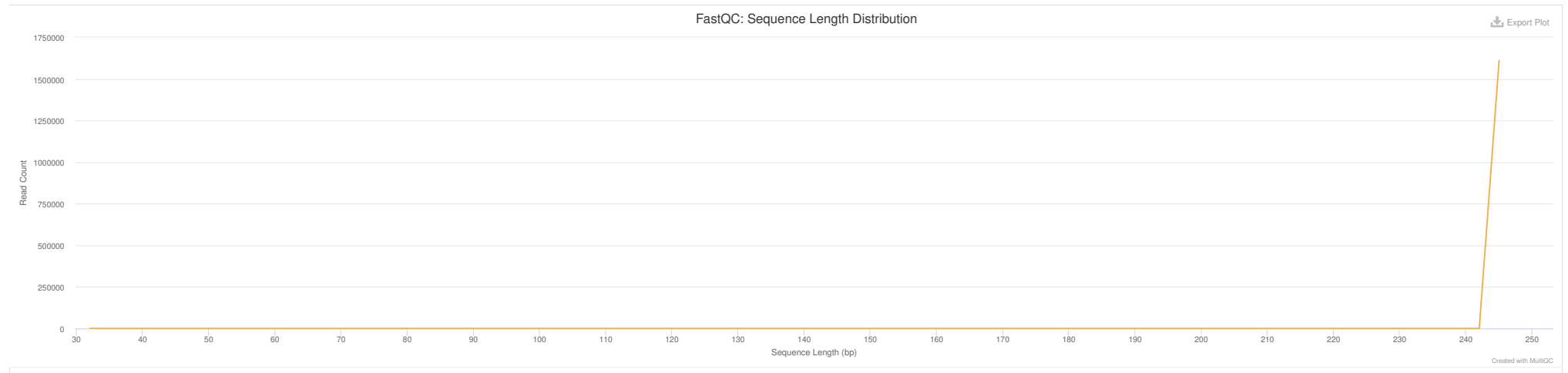
Y-Limits: 

Sequence Length Distribution

1 1

The distribution of fragment sizes (read lengths) found. See the FastQC help

Y-Limits: on

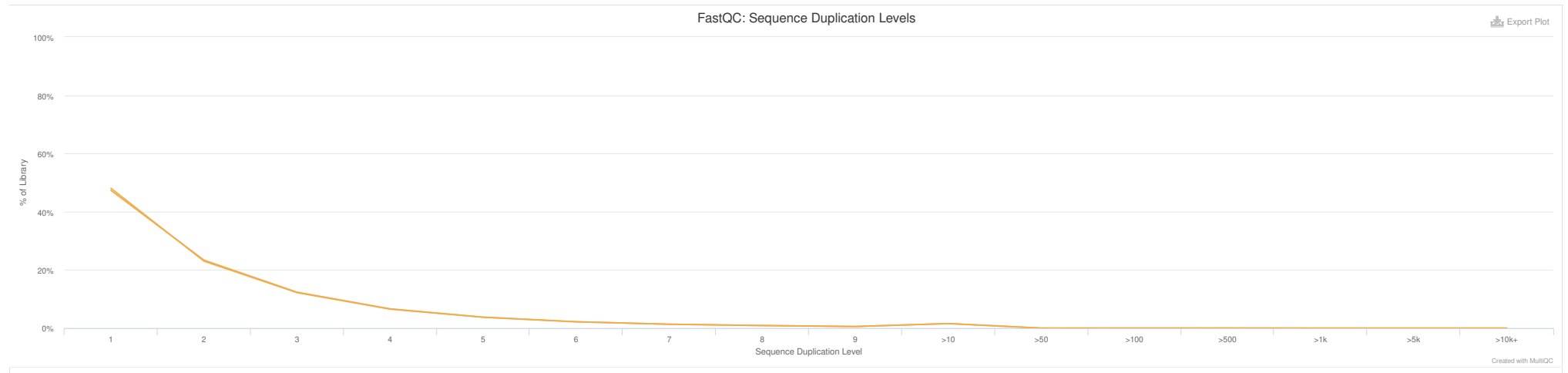


Sequence Duplication Levels

2

The relative level of duplication found for every sequence.

Y-Limits: on



Overrepresented sequences by sample


2

The total amount of overrepresented sequences found in each library.

2 samples had less than 1% of reads made up of overrepresented sequences

Top overrepresented sequences

Top overrepresented sequences across all samples. The table shows 20 most overrepresented sequences across all samples, ranked by the number of samples they occur in.

 Copy table Showing 0% rows.

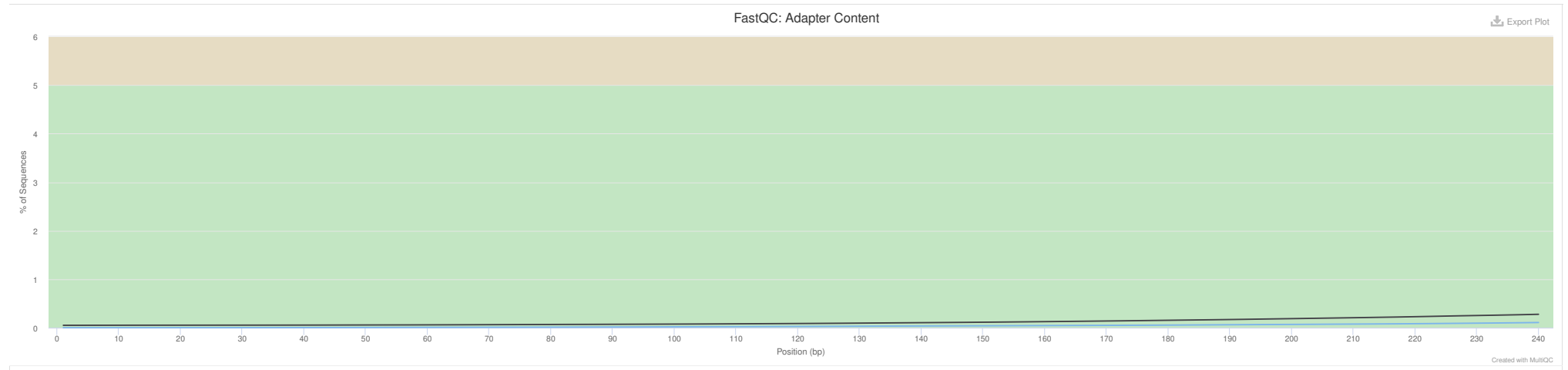
Overrepresented sequence

Adapter Content

2

The cumulative percentage count of the proportion of your library which has seen each of the adapter sequences at each position.

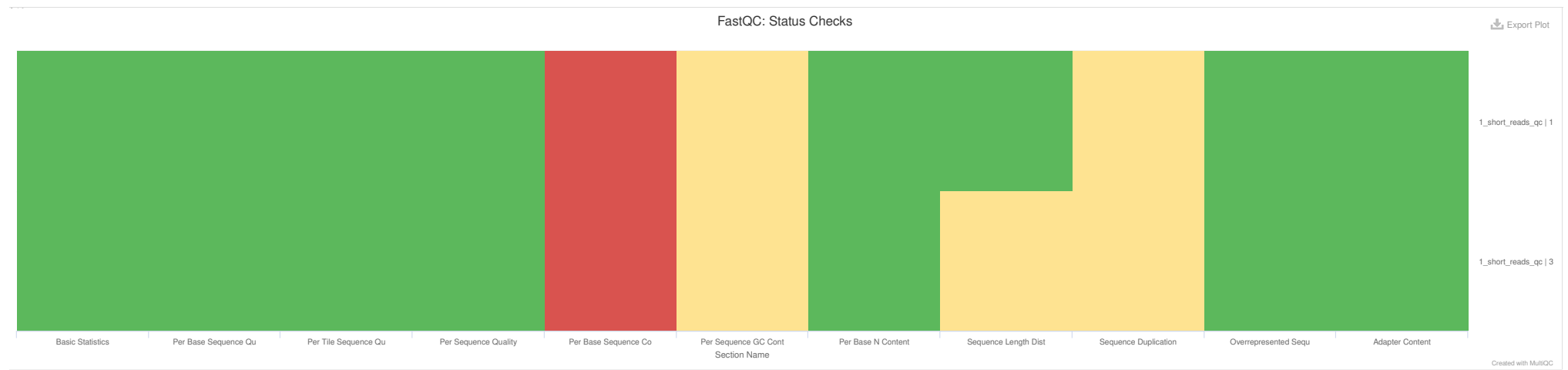
Y-Limits:



Status Checks

Status for each FastQC section showing whether results seem entirely normal (green), slightly abnormal (orange) or very unusual (red).

 Sort by highlight **Min:** 0  **Max:** 1 



Software Versions

Software Versions lists versions of software tools extracted from file contents.

 Copy table

Software	Version
FastQC	0.12.1
fastp	0.23.4

MultiQC v1.19 - Written by [Phil Ewels](#), available on [GitHub](#).
This report uses [HighCharts](#), [jQuery](#), [jQuery UI](#), [Bootstrap](#), [FileSaver.js](#) and [clipboard.js](#).

