



Accurate estimation of rare cell type fractions from tissue omics data via hierarchical deconvolution

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Introduction

- The varying cellular fractions of tissue samples serve as the foundation for many downstream statistical analyses. Although biochemical methods can measure cell counts of samples, they are labor-intensive and costly.
- Cellular deconvolution methods have been developed to estimate cellular proportions. Supervised methods can be modeled as

$$\begin{matrix} \text{Bulk} & \text{Signature} & \text{Cell size} & \text{CT fractions} \\ X & = & ASP & + E \\ \begin{matrix} m \times n \\ [m \times n] \end{matrix} & & \begin{matrix} m \times K & K \times K & K \times n \\ [m \times K] & [K \times K] & [K \times n] \end{matrix} & & \begin{matrix} m \times n \\ [m \times n] \end{matrix} \end{matrix}$$

• m marker genes
• n tissue samples
• K cell types

- Cell type hierarchy has become important for understanding the topology of cell types across datasets^{1,2,3}.
- The increase of number of cell types begets co-linearity because of shared origins of cell differentiation and leads to rare cell types.
- Here we present Hierarchical Deconvolution (HiDecon), a penalized model pooling information across related cell types to tackle these challenges.

Estimation Model

- Given hierarchical tree, CT fractions of “parent” and “children” across layers have a summation relationship approximately.
- We introduce a cell type mapping matrix $B_{l,(l+1)} \in \mathbb{R}_+^{K_l \times K_{l+1}}$ modeling the relationship between layer l and $l+1$. e.g.,

$$p_{i1} \approx B_{1,2} p_{i2} = \begin{pmatrix} 1 & 0 & 0 & 0 \\ 0 & 1 & 1 & 1 \end{pmatrix} p_{i2}$$

- HiDecon model objective function (sample i) for an L layer tree:

$$f(p_i) = \sum_{l=1}^L m_l^{-1} \|x_{il} - A_l S_l p_{il}\|_2^2 + \lambda \sum_{l=1}^{L-1} K_l^{-1} \|p_{il} - B_{l,(l+1)} p_{i(l+1)}\|_2^2$$

$$p_{il} \geq 0, \quad \|p_{il}\|_1 = 1.$$

where p_i is a length $K = \sum_{l=1}^L K_l$ vector of cell proportions of all the nodes in the hierarchical tree, p_{il} denotes the layer l part of type fractions, x_{il} denotes bulk gene expression level of m_l markers in layer l , A_l and S_l are reference signature matrix and size factor matrix derived from single cell data respectively. Marker genes are selected for each layer.

- To estimate fractions for all layer simultaneously, we rewrite the model as:

$$f(p_i) = \|\tilde{x}_i - \tilde{A} p_i\|_2^2 + \lambda \|\tilde{B} p_i\|_2^2, \quad \text{such that } p_i \geq 0 \text{ and } \|p_i\|_1 = 1$$

$$\tilde{x} = \begin{pmatrix} m_1^{-1/2} x_{i1} \\ \vdots \\ m_L^{-1/2} x_{iL} \end{pmatrix}, \quad \tilde{A} = \begin{pmatrix} m_1^{-1/2} A_1 S_1 & \cdots & 0 \\ \vdots & \ddots & \vdots \\ 0 & \cdots & m_L^{-1/2} A_L S_L \end{pmatrix}, \quad \tilde{B} = \begin{pmatrix} K_1^{-1/2} I_{K_1} & \cdots & 0 & 0 \\ \vdots & \ddots & \vdots & \vdots \\ 0 & \cdots & K_{L-1}^{-1/2} I_{K_{L-1}} & 0 \end{pmatrix} - \begin{pmatrix} 0 & K_1^{-1/2} B_{1,2} & \cdots & 0 \\ \vdots & \vdots & \ddots & \vdots \\ 0 & 0 & \cdots & K_{L-1}^{-1/2} B_{(L-1),L} \end{pmatrix}$$

$$H(p_i) = \nabla_{p_i}^T f(p_i) = \tilde{A}^T \tilde{A} + \lambda \tilde{B}^T \tilde{B} \quad b_k = \tilde{A}^T \tilde{x}_i$$

HiDecon estimation algorithm:

- Initialize $p_i = H^{-1}b$. (Coordinate-wise descend algorithm⁴)
- If $p_i \geq 0$, return p_i . If not, proceed to step 3.
- Let $p_{ik} = \max\{p_{ik}, 0\}$, for all $k \in \{1, \dots, K\}$.
- $p_{ik} = \max\left\{0, p_{ik} + \frac{b_k - p_{ik}^T H_{*k}}{H_{kk}}\right\}$, where H_{*k} is the k th column of H .
- Check the KKT conditions

$$(|H_{*k}^T p_i - b_k| \leq \epsilon) \text{ OR } (H_{*k}^T p_i - b_k \geq 0 \text{ AND } p_{ik} \leq \epsilon), \forall k \in \{1, \dots, K\}.$$

- If KKT are satisfied, return p_i . Otherwise, repeat step 4-5.

Parameter Selection

- For tuning parameter λ , we employ a resampling method shown as below:

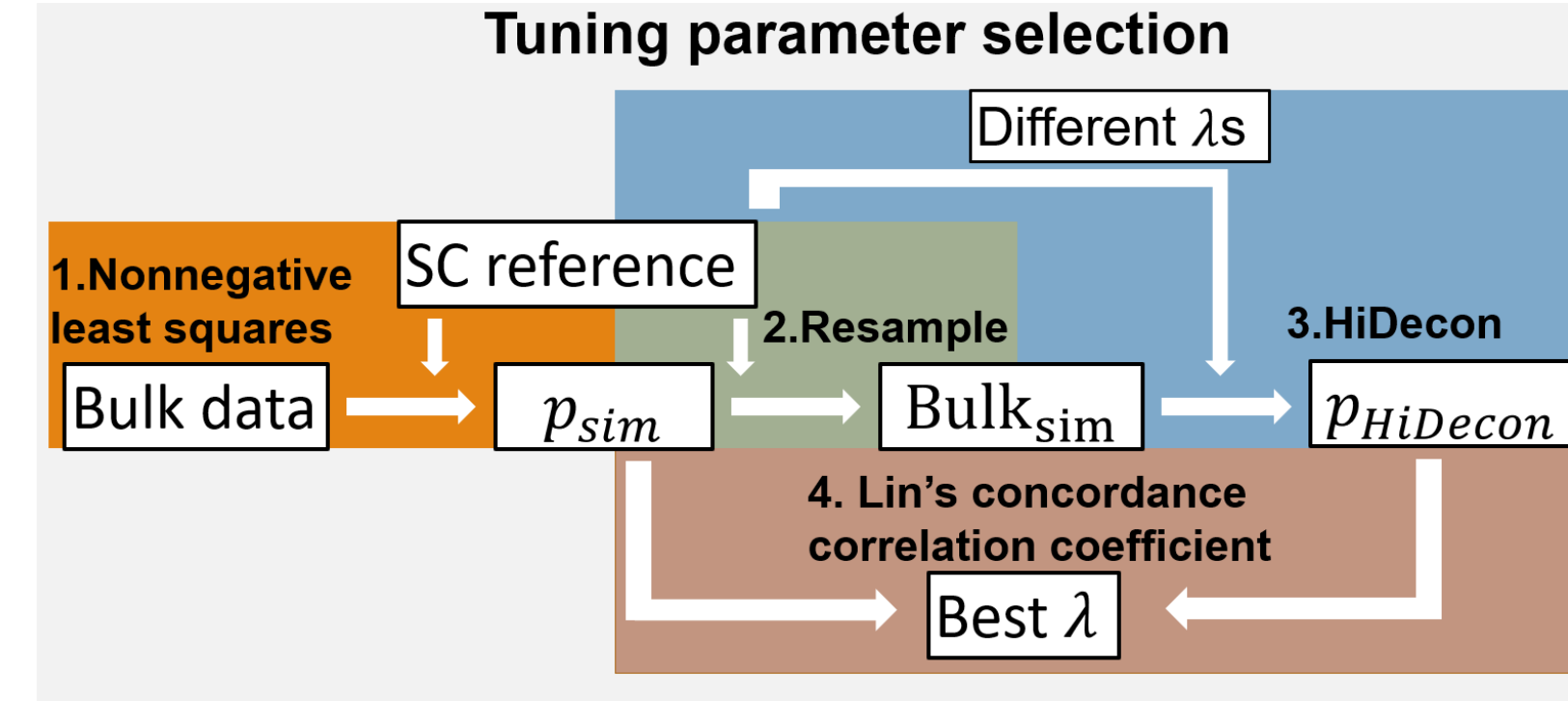


Figure 1: Flow chart for HiDecon tuning parameter selection.

Simulation Studies

Metrics

- MAE: mean absolute error compared with true fraction $\text{avg}(|P - \hat{P}|)$
- CCC (Lin's Concordance Correlation Coefficient⁵): $\frac{2\text{cov}(P_{k*}, \hat{P}_{k*})}{\sigma_{P_{k*}}^2 + \sigma_{\hat{P}_{k*}}^2 + (\text{avg}(P_{k*} - \hat{P}_{k*}))^2}$

Data: real large scale COVID-19 individual level scRNA-seq dataset⁶ of PBMC (peripheral blood mononuclear cells) with subtypes:

- Simulated bulk data: averaged across single cells within sample (126 samples).
- Reference: all individual level single cell data used for simulating bulk data.

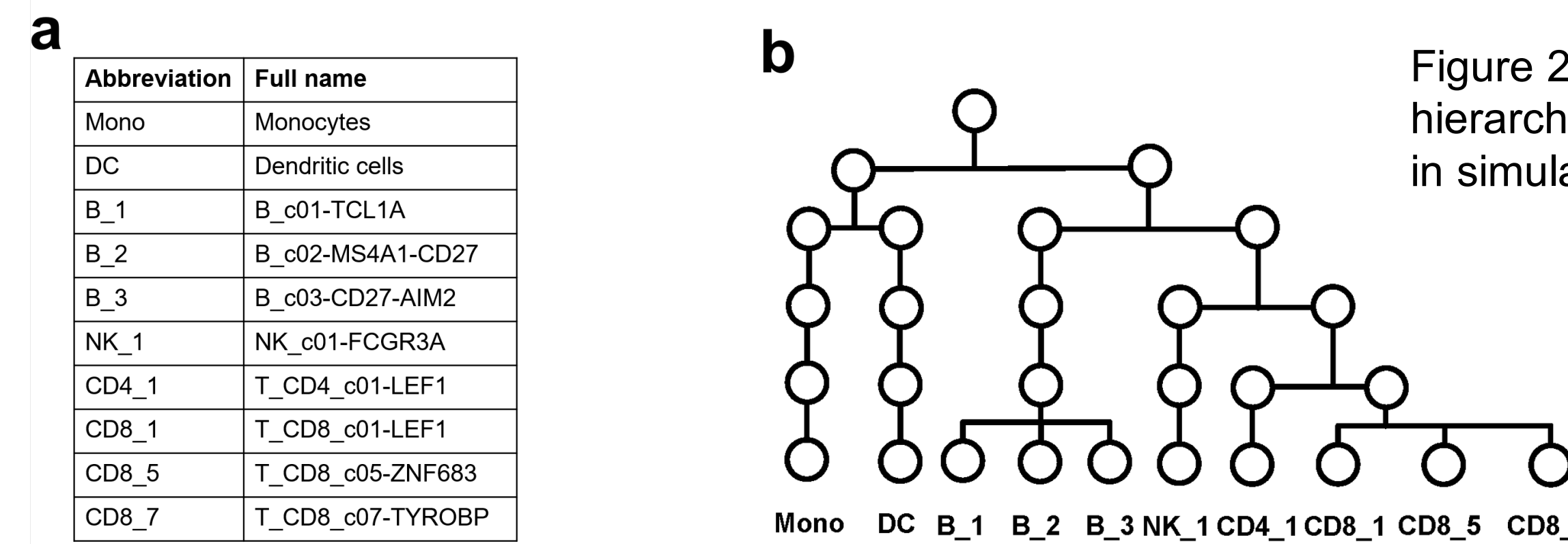


Figure 2: Cell types and hierarchical tree we use in simulation.

	Mono	DC	B_1	B_2	B_3	NK_1	CD4_1	CD8_1	CD8_5	CD8_7	Mean CCC	MAE
HiDecon	(0.38)	(0.02)	(0.07)	(0.03)	(0.03)	(0.07)	(0.13)	(0.09)	(0.09)	(0.08)	0.57	0.05
CIBERSORT	0.88	0.31	0.66	0.31	0.37	0.01	0.42	0.76	0.25	0.25	0.42	0.07
dtangle	0.35	0.08	0.56	0.18	0.30	0.47	0.51	0.51	0.60	0.69	0.43	0.06
MuSiC	0.89	0.01	NA	NA	0.16	NA	NA	0.35	0.22	0.39	0.20	0.08

Table 1: CCC and MAE in the simulation study for different deconvolution methods.

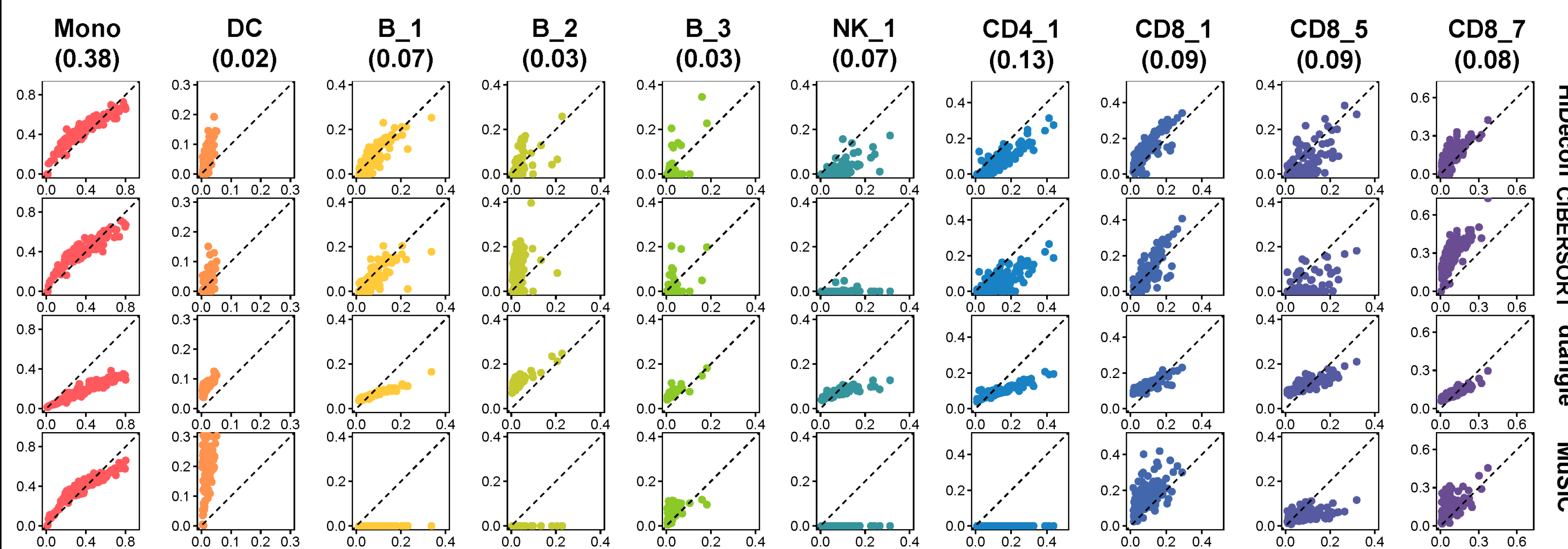


Figure 3: Scatter plots of cellular fractions in simulation study for different methods

Robustness analysis: We use simulated data and add noises $N(0, sd^2)$ to bulk data with maximum sd equals the sd of bulk data. Experiments are repeated 50 times using different random seeds and we averaged metrics among repetitions.

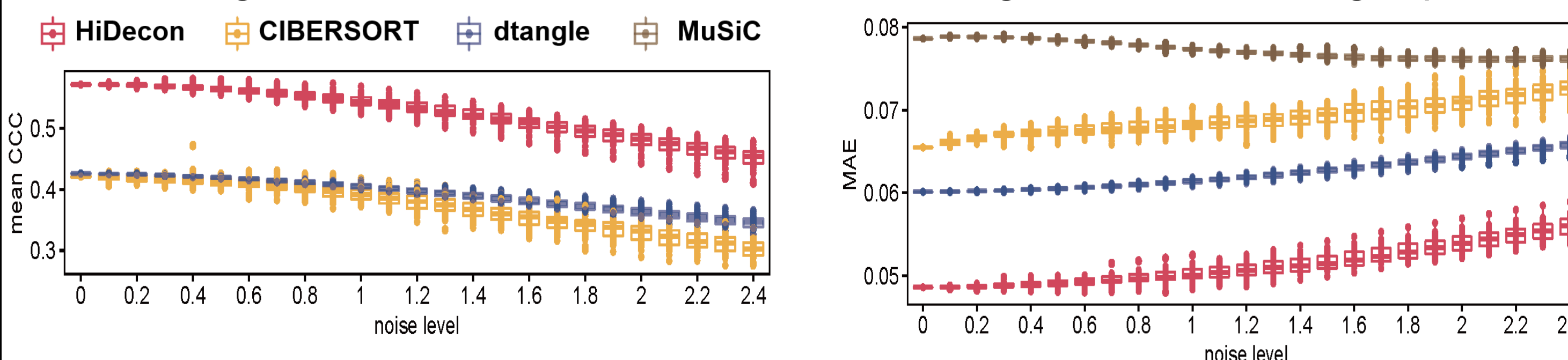


Figure 4: CCC (upper panel) and MAE in robustness analysis. (MuSiC CCC missing due to all 0 estimates).

Real Data Applications

Real data application: FHS data (Framingham Heart Study)^{7,8,9}. Human blood data.

- Bulk data: 4,110 blood samples with measured cell type fractions.
- Reference: LM22 data (microarray data).
- Ground truth: measured blood cell counts in FHS.

	Neutrophil (0.60)	Lymphocyte (0.28)	Monocyte (0.09)	Eosinophil (0.03)	Mean CCC	MAE
HiDecon	0.13	0.57	0.04	0.28	0.26	0.10
CIBERSORT	0.15	0.31	0.02	0.06	0.13	0.15
dtangle	0.02	0.17	0.01	0.01	0.05	0.17
HEpiDISH	0.12	0.25	0.03	0.04	0.11	0.17
MuSiC	NA	0.08	0.00	NA	0.02	0.32

Table 2: CCC and MAE in FHS data study for methods.

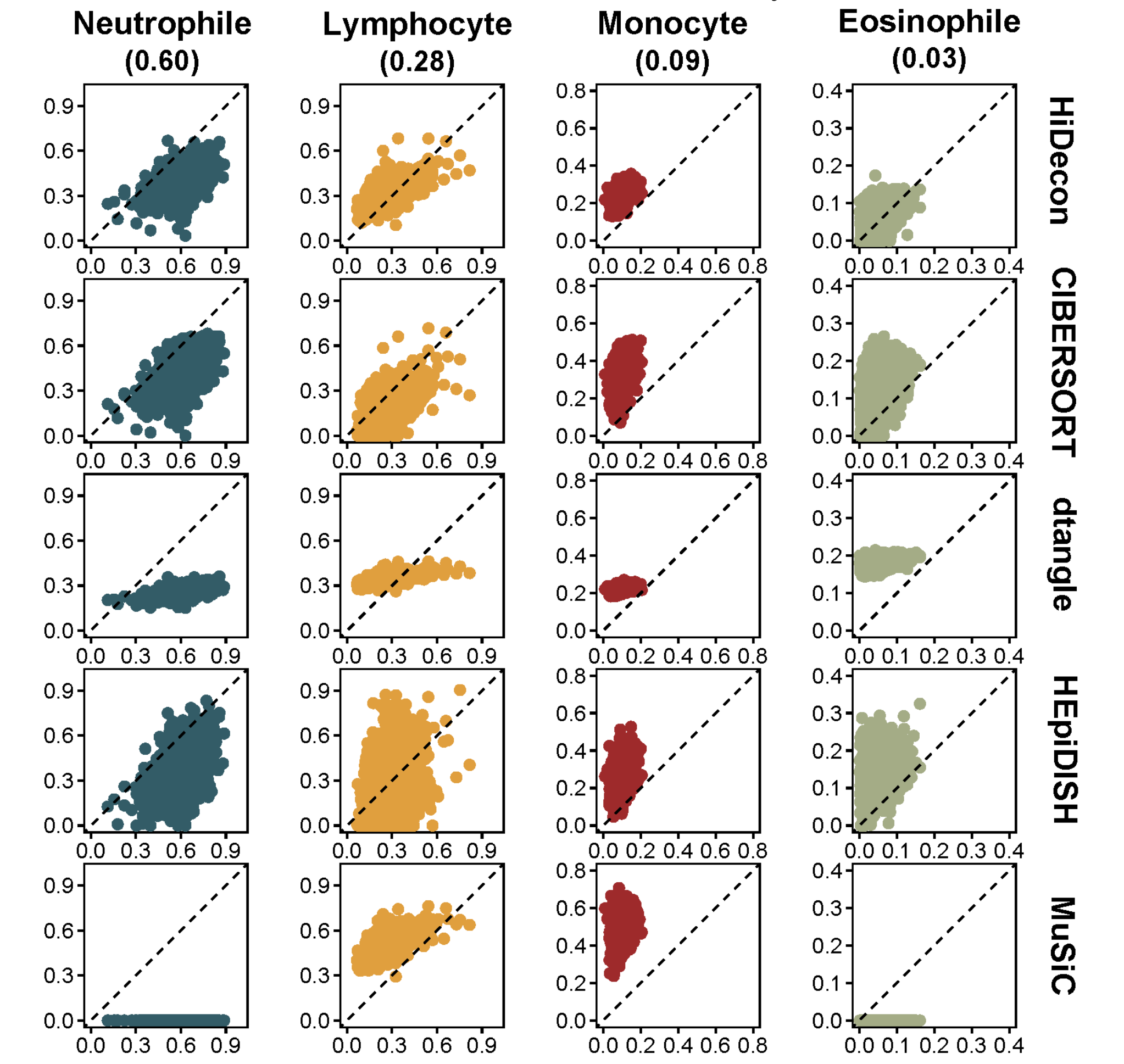


Figure 5: Scatter plots in the FHS real data study for different deconvolution methods.

Conclusions

- We developed HiDecon to incorporate a hierarchical cell type tree to facilitate the estimation of related cell types.
- HiDecon can incorporate complex tree structure with more flexibility.
- HiDecon can provide accurate estimates especially for rare cell types.
- We offer a user-friendly R package along with a brief tutorial hosted on <https://github.com/randell/HiDecon>.

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