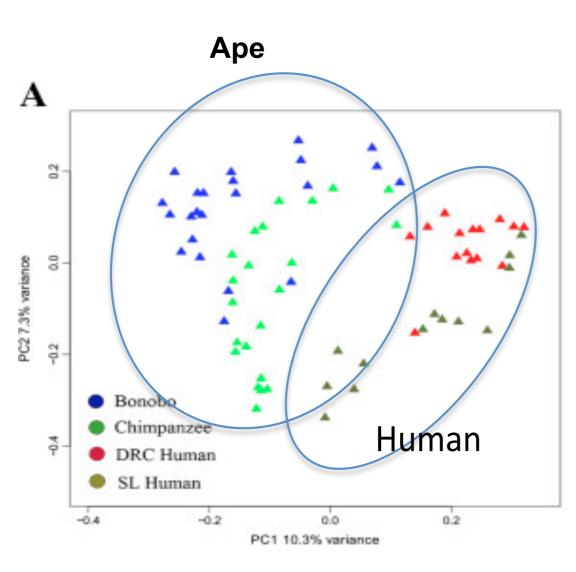
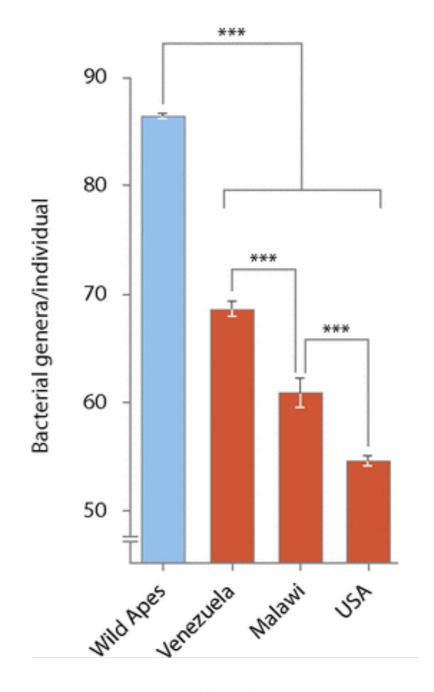
LESSONS FROM NEANDERTAL MICROBIOTA:
WORKING TOWARD ACCURATE METAGENOMIC
ANALYSIS FROM LOW-BIOMASS SAMPLES.



#### EVOLUTIONARY HISTORY OF THE HOMINID MICROBIOTA

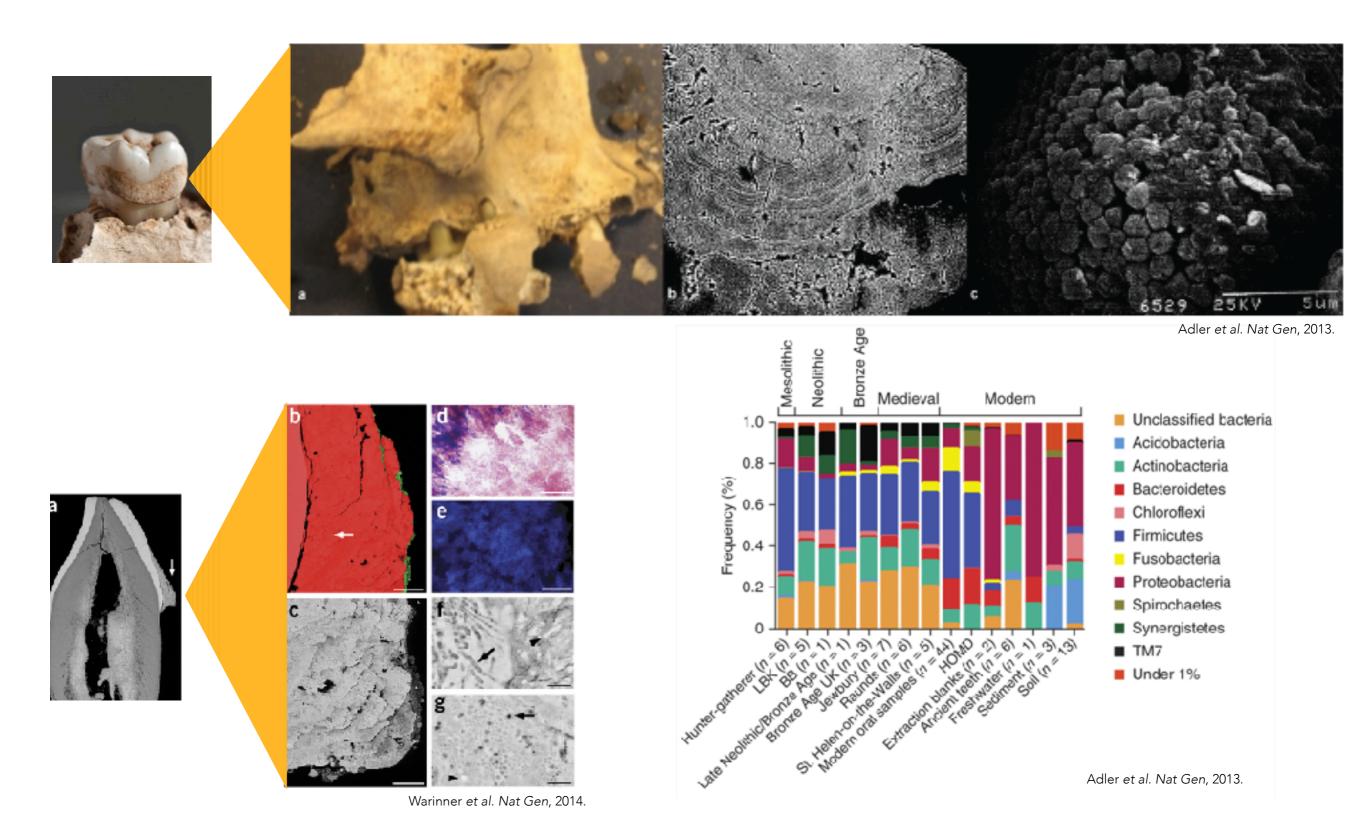


Li et al. BMC Microbiology 2013

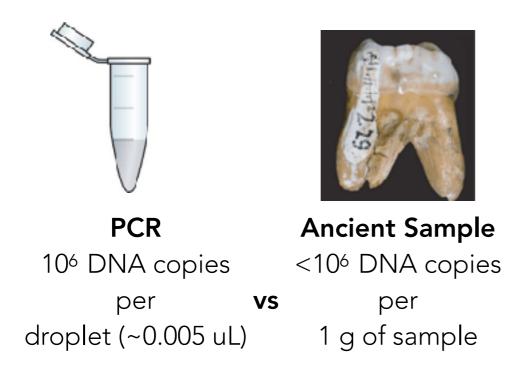


Moeller, et al. PNAS, 2014.

# ANCIENT DENTAL CALCULUS IS A FOSSILISED BACTERIAL RECORD



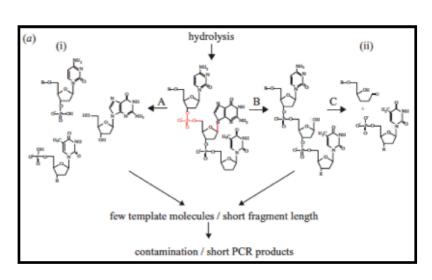
#### WHY ARE ANCIENT SAMPLES PROBLEMATIC?



# Fragmented and damaged DNA crosslinking/hydrolysis/oxidation



## Low 'endogenous' DNA yield



Willerslev and Cooper. Proc Biol Sci. 2005 Jan 7; 272(1558): 3-16

Contamination (modern or environmental) sample collection/handling & lab or reagent contamination

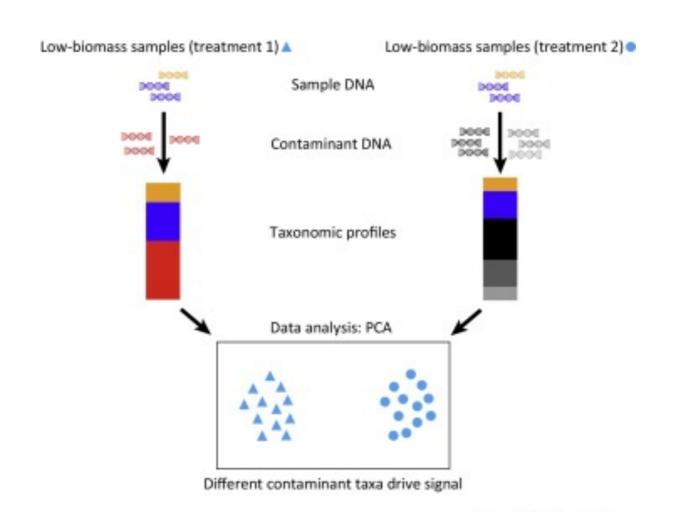
## Modern samples can also be similarly problematic



Skin swabs
Placenta and Pre-term infants
Respiratory biopsies
Blue Ice
Ancient/Historic Specimens
and more...



### CONCEPTS IN CONTAMINATION



- 1. Dedicated DNA extraction/processing facilities
- 2. Decontamination, if possible
- 3. Monitoring laboratory & environmental contamination
- 4. Efficient DNA extractions and library preparation
- 5. Accurate analysis and verification tools

# 1. REDUCING CONTAMINATION WITH CLEAN FACILITIES





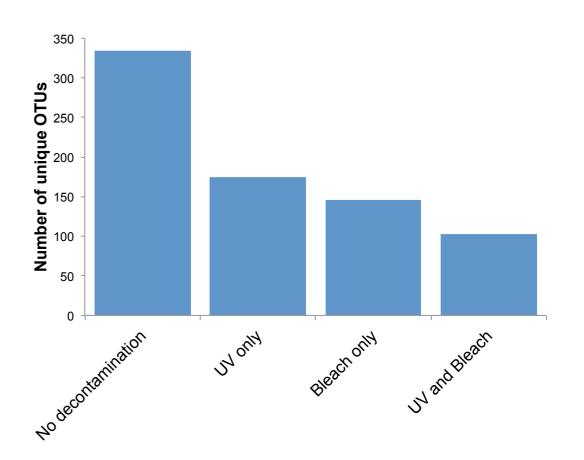
Isolated pre-PCR facility Isolated ventilation Glove boxes Nightly UV irradiation Clean -> Dirty workflow Bleach treatment of stock Reagents tested for DNA Appropriate dress

### 2. DECONTAMINATION MATTERS!

#### **Decontamination Procedures**

- A. Remove surface
- B. Decontaminate sample
  - -UV or washing, if able
- C. Include controls every time
  - -PCR negatives
  - -Extraction blank controls
- D. Duplicate samples
- E. Remove contaminants with bioinformatic methods

DNA Contamination



A. Farrer, in prep. Check out the kitome: Salter, et al., 2014, BMC Biology, 12, 87.





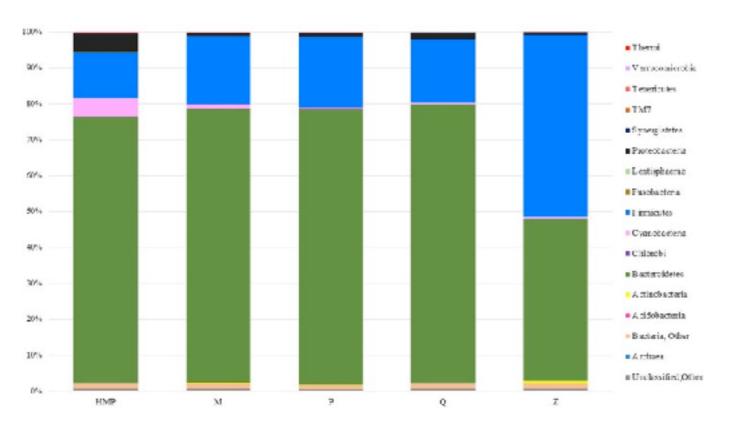








# 3. DNA EXTRACTION BIASES AND EFFICIENCY MUST BE CONSIDERED



1800 1600 1400 1200 1000 800 400 200 QG - Silica Kit

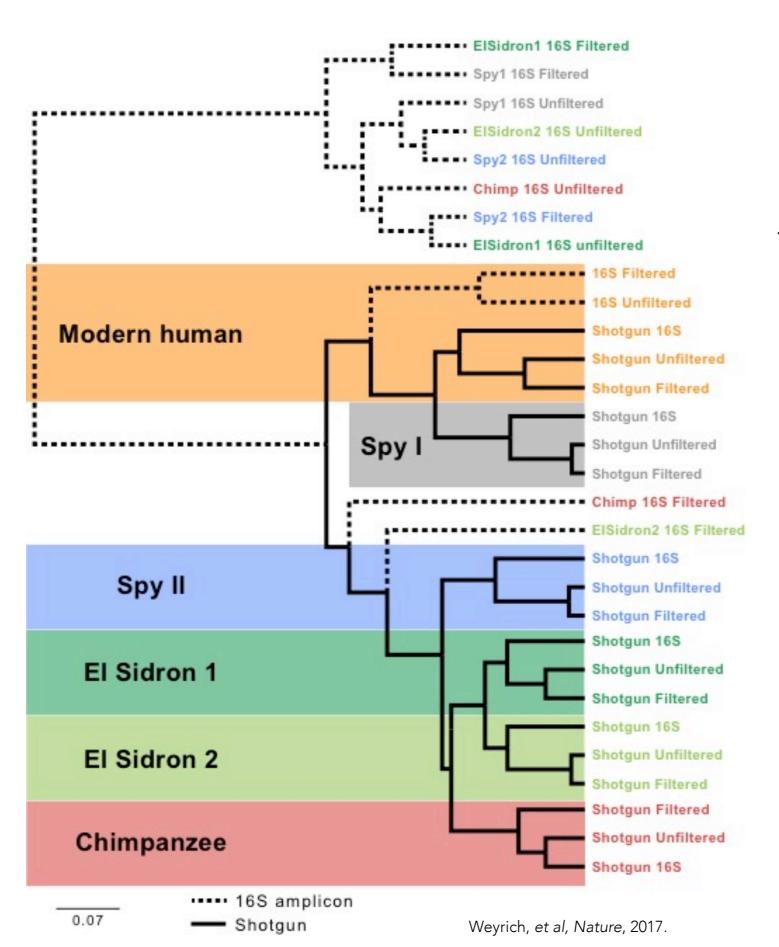
Mackenzie, et al. Front Microbiol. 2015; 6: 130.

Extraction methods bias can results, and limit downstream comparisons.

QIITA!

Extraction efficiency matters!

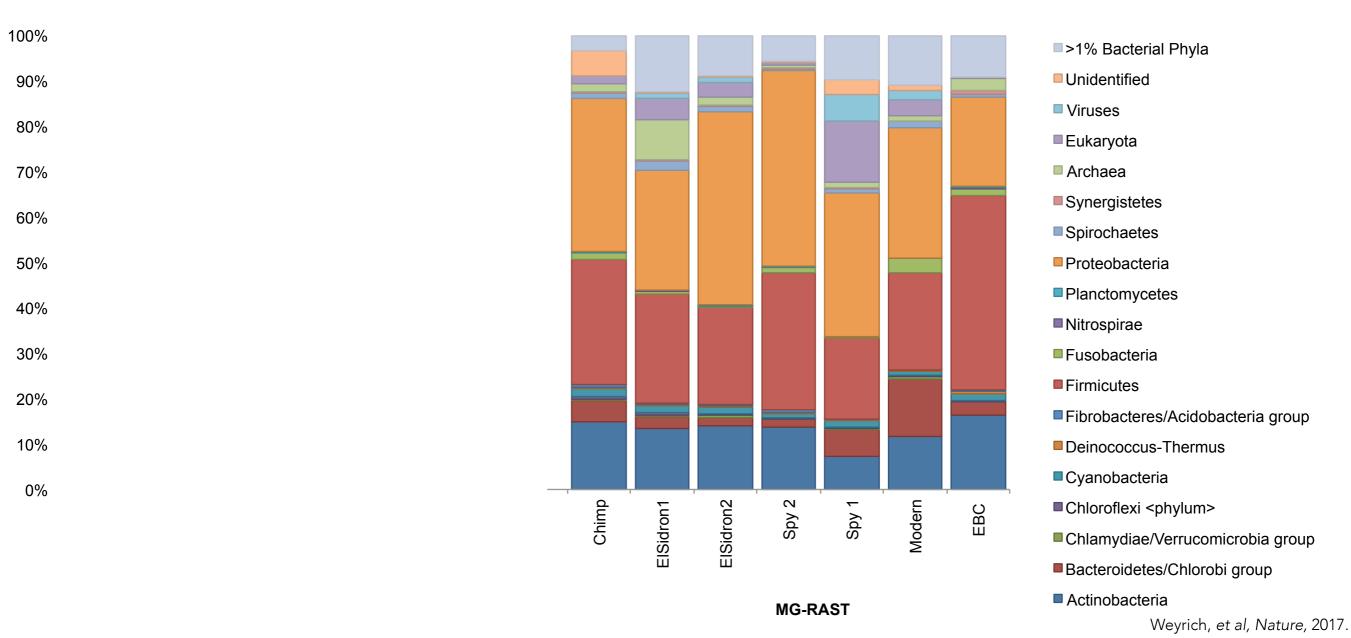
#### 3. LIBRARY PREPARATION ALSO MATTERS



16S rRNA gene seq will likely not work for highly degraded samples.

Filtering helps (sometimes!)

## 4. BIOINFORMATIC METHODS TO IDENTIFY SPECIES FROM SAMPLES



Satisfies input requirements; accurate; rapid nucleotide vs protein

Current methods applied: DIAMOND, MALTX

# 4. BIOINFORMATIC METHODS TO ACCOUNT FOR CONTAMINATION

1. Decontam

https://github.com/benjjneb/decontam

2. MEGAN6CE

https://github.com/husonlab/megan-ce

3. Direct Filtering

QIIME2, etc.

4. ContaminantAssessment

Comparison to known lists in: Salter, et al., *BMC Biol*, 2014 or Weyrich, et al., MER, 2019

# RIDE CHECKLIST FOR PERFORMING/REVIEWING LOW MICROBIAL BIOMASS MICROBIOME STUDIES

**Report** the experimental design and approaches used to reduce and assess the contributions of contamination.

**Include** controls to assess contaminant DNA. One of each type of negative control (sampling blanks, DNA extraction blanks, and no-template amplification) must be included per sampling, extraction, or amplification batch.

**Determine** the level of contamination by comparing biological samples to controls.

**Explore** contaminant taxa within each study and report their impact on the interpretation of biological samples.

# WHAT FACTORS SHOULD BE CONSIDERED BEFORE WORKING WITH LOW BIOMASS SAMPLES?

Before you start...

Consider your working environment.

Assess contamination.

Decide on the best extraction method.

## During analysis...

Use clean environments for processing.

Include extraction blank and PCR controls.

Don't let bioinformatics be a black box.

Scrutinise and test your results!

### And enjoy!