

# CPIC Guideline Update on PharmGKB

**For: “Clinical Pharmacogenetics Implementation Consortium Guidelines for Dihydropyrimidine Dehydrogenase Genotype and Fluoropyrimidine Dosing”**

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**URL:**

<http://www.pharmgkb.org/guideline/PA166109594>

<http://www.pharmgkb.org/guideline/PA166122686>

<http://www.pharmgkb.org/guideline/PA166122687>

## **Description:**

There was an issue with the stranding for nucleotide changes for DPYD\*3 and DPYD\*7 in Table S1. DPYD\*3 is a G>del change on the positive chromosomal strand, while DPYD\*7 is an insertion of ATGA on the positive chromosomal strand. Changes to table S1 are in red.

Additionally, The CPIC authors recommend that the *DPYD*\*4, \*5, \*6 and \*9A alleles be categorized as "normal" activity, in part based upon the recent publication [Comparative Functional Analysis of DPYD Variants of Potential Clinical Relevance to Dihydropyrimidine Dehydrogenase Activity](#). Changes to table S2 are in red.

Please see the updated guideline at:

<http://www.pharmgkb.org/guideline/PA166109594>

<http://www.pharmgkb.org/guideline/PA166122686>

## **Supplemental Material**

### **Clinical Pharmacogenetics Implementation Consortium Guidelines for Dihydropyrimidine Dehydrogenase Genotype and Fluoropyrimidine Dosing**

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## Literature Review

A literature search of the PubMed database (1966 to March 2013) using the keywords ((DPD OR DPYD OR Dihydropyrimidine Dehydrogenase) AND (fluorouracil OR 5-FU OR fluoropyrimidines OR capecitabine OR tegafur) AND genotype) was performed and results were limited to those available in English. Further articles were found via the reference sections of reviews.

## Available Genetic Test Options

Commercially available genetic testing options change over time. An updated and fully linked table is available at <http://www.pharmgkb.org/gene/PA145>. Furthermore, the Genetic Testing Registry (GTR) provides a central location for voluntary submission of genetic test information by providers and is available at <http://www.ncbi.nlm.nih.gov/gtr/conditions/C2720286/>.

## Genetic Test Interpretation

The haplotype, or star (\*) allele name, is determined by a specific SNP or a combination of SNPs that are interrogated in the genotyping analysis. The genotypes that constitute the haplotype, or star (\*) alleles for *DPYD*, and the rs# for each of the specific genomic nucleotide alterations that define the alleles, are described in Supplemental Table S1. However, at least one SNP, rs67376798, is associated with low activity but has not been given a \* allele designation.

The genotype results are generally reported as a diplotype, which include one maternal and one paternal star allele (e.g. \*1/\*2A). The functional status associated with each of the common \* alleles is summarized in Supplemental Table S2.

DPD function associated with the known *DPYD* allelic variants is summarized in Supplemental Table S2. The dosing recommendations in this guideline are specific for variant alleles in which there are clear data linking the *DPYD* genotype to fluoropyrimidine toxicity (\*2A, \*13, and rs67376798) (Supplemental Table S5). However, several other variants have been reported to be associated with reduced enzyme activity and/or linked to fluoropyrimidine toxicity, albeit with somewhat weaker evidence. These variants have been categorized as “probable reduced-function” or “unknown/unclear/contradictory” based on the lack of evidence linking these genotypes to fluoropyrimidine toxicity or conflicting evidence, respectively. All of the variants listed in the “probable reduced-function” category (\*3, \*8, \*9B, \*10, \*11, and \*12) are very rare (0-0.15% allele frequency in populations studied) and thus, reports describing DPD activity and the effects on fluoropyrimidine toxicity for these variants are rare. For example, *DPYD*\*7, \*8, and \*10 have only been reported in one study analyzing *DPYD* in 17 families (22 individuals) with complete dihydropyrimidine dehydrogenase deficiency (no detectable DPD activity in fibroblasts, <0.2% of controls).(1) To date, there are no studies linking these variant alleles (\*7, \*8, and \*10) to toxicities related to fluoropyrimidines. Although the variants listed in the “unknown/unclear/contradictory” category have been observed in patients experiencing fluoropyrimidine toxicities, there is a lack of studies making a clear association between these variants and fluoropyrimidine

toxicity and/or decreased DPD activity. These alleles would therefore be informative in any *DPYD* testing.

### **Other considerations**

Several other genes may influence responses to 5-fluorouracil (2, 3). The most well-studied of these are *ABCB1*, *MTHFR* and *TYMS*, although to date results have been inconsistent and predictive dosing strategies have yet to be successfully applied. Some of the testing options for 5-fluorouracil toxicity and *DPYD* also include testing for other gene variants in *TYMS* and *MTHFR*. Furthermore, Fernandez-Rozadilla *et al* performed a genome-wide association study on 221 colorectal cancer patients (including a validation set of 791 patients) that had been treated with a 5-fluorouracil-based regimen. Seven SNPs (rs16857540 (*NLGN1*), rs2465403 (*COLEC10*), rs10876844 (*OR10AE3P*, *PSMB2P*), rs10784749, rs17626122 (*PARD3B*), rs7325568 and rs4243761) showed evidence of association with adverse drug reactions. However, they also evaluated the association signals for seven SNP variants that had been linked to 5-fluorouracil-related toxicity in the literature (*DPYD*\*5 and \*9A, rs18010919 (*UMPS*), rs1801133 (*MTHFR*), rs34743033, rs34489327 (*TYMS*), rs1695 (*GSTP1*)). Four of these variants had good proxy SNPs in the study, although none of them showed a statistically significant association. These associations underscore the potential importance of other genes that may contribute increased risk of toxicity of 5-fluorouracil (4).

### **Level of Evidence**

The evidence summarized in Supplemental Table S5 is graded using a scaled modified slightly from Valdes *et al* (5).

High: Evidence includes consistent results from well-designed, well-conducted studies.

Moderate: Evidence is sufficient to determine effects, but the strength of the evidence is limited by the number, quality, or consistency of the individual studies; generalizability to routine practice; or indirect nature of the evidence.

Weak: Evidence is insufficient to assess the effects on health outcomes because of limited number or power of studies, important flaws in their design or conduct, gaps in the chain of evidence, or lack of information

### **Strength of Recommendation**

CPIC's dosing recommendations (Table 2, main manuscript) are based weighting the evidence from a combination of preclinical functional and clinical data, as well as on some existing disease-specific consensus guidelines. Some of the factors that are taken into account include in vivo clinical outcome for reference drug, in vivo PK/PD for reference drug, and in vitro enzyme activity with probe substrate only.

Overall, the dosing recommendations are simplified to allow rapid interpretation by clinicians. We chose to use a slight modification of a transparent and simple system for just three categories for recommendations adopted from the rating scale for evidence-based recommendations on the use of retroviral agents

(<http://aidsinfo.nih.gov/contentfiles/AdultandAdolescentGL.pdf>): strong, where “the evidence is high quality and the desirable effects clearly outweigh the undesirable effects”; moderate, in which “there is a close or uncertain balance” as to whether the evidence is high quality and the desirable clearly outweigh the undesirable effects; and optional, in which the desirable effects are closely balanced with undesirable effects and there is room for differences in opinion as to the need for the recommended course of action.

Strong recommendation for the statement  
Moderate recommendation for the statement  
Optional recommendation for the statement

The strength of the 5-fluorouracil dosing recommendations (Table 2; main manuscript) is based on the fact that some variants (*DPYD*\*2A, \*13, and rs67376798) clearly affect DPD activity, and DPD activity is clearly related to 5-fluorouracil clearance, and 5-fluorouracil exposure is associated with its toxic effects. Therefore, reduction of 5-fluorouracil dosage in patients with these variants may prevent severe and possibly life-threatening toxicities. The strength of the capecitabine and tegafur dosing recommendations is based on the fact that both of these drugs are prodrugs of 5-fluorouracil and metabolized by DPD in the same manner.

<b>Supplemental Table S1. Genotypes<sup>1</sup> that constitute the * alleles for <i>DPYD</i></b>			
<b>Allele</b>	<b>Constituted by genotypes at:</b>	<b>Position on NM_000110.3</b>	<b>Amino acid change</b>
*1			
*2A	rs3918290 C>T	c.1905+1G>A	n/a
*2B	rs3918290 C>T and rs1801159 T>C	c.1905+1G>A and c.1627A>G	n/a and Ile543Val
*3	rs72549303 C>del	c.1898delC	Pro633fx
*4	rs1801158 C>T	c.1601G>A	Ser534Asn
*5	rs1801159 T>C	c.1627A>G	Ile543Val
*6	rs1801160 C>T	c.2194G>A	Val732Ile
*7	rs72549309 TCAT>del	c.302delTinsTCAT	Ile101fx
*8	rs1801266 G>A	c.703C>T	Arg235Trp
*9A	rs1801265 A>G	c.85T>C	Cys29Arg
*9B	rs1801265 A>G and rs1801267 C>T	c.85T>C and c.2657G>A	Cys29Arg and Arg886His
*10	rs1801268 C>A	c.2983G>T	Val995Phe
*11	rs72549306 C>A	c.1003G>T	Val335Leu
*12	rs80081766 C>T and rs78060119 C>A	c.62G>A and c.1156G>T	Arg21Gln and Glu386Ter
*13 <sup>§</sup>	rs55886062 A>C	c.1679T>G	Ile560Ser
None <sup>¶</sup>	rs67376798 T>A	c.2846A>T	Asp949Val

1: Bases reported on the positive chromosomal strand from Golden Path. See [www.pharmgkb.org](http://www.pharmgkb.org) for updates on *DPYD* gene alleles and nomenclature

Nomenclature from \*1-10 from McLeod et al (6), \*11 and \*12 from Mattison et al, (7), \*13 from Johnson et al, (8).

<b>Supplemental Table S2. Association between allelic variants and DPD function</b>		
<b>Functional Status</b>	<b>Alleles</b>	<b>Phenotype</b>
Functional / normal activity/ wild-type <sup>1</sup>	*1	
Non-functional, variant, or mutant / no activity	*2A	Associated with toxicity in most (3, 9-12), but not all (13, 14). Observed in patients with low DPD activity (11, 14-21) and DPD deficiency (1). Observed in patients with severe or fatal toxicity (15, 22-27). Associated with reduced 5-fluorouracil clearance (12, 17, 21) and inactive catalytic activity (28).
	*13	Associated with toxicity (12). Observed in patients with low DPD activity (8, 29). Observed in patients with severe or fatal toxicity (20, 27). Associated with reduced 5-fluorouracil clearance (12).
	rs67376798	Observed in individuals with low DPD activity (20, 30). Associated with toxicity (3, 10-12, 23, 27). Associated with reduced 5-fluorouracil clearance (12, 21).
Probable Reduced-function / decreased activity (these alleles are mostly very rare and so reports have been rare)	*3	Observed in individuals with DPD deficiency (1, 31).
	*7	Observed in individuals with DPD deficiency (1).
	*8	Observed in individuals with low DPD activity (1).
	*9B	Observed in individuals with low DPD activity (31, 32).
	*10	Observed in individuals with DPD deficiency (1).
	*11	Observed in individuals with low DPD activity (33).
	*12	Observed in individuals with low DPD activity (33).



Unknown/unclear/contradictory evidence	*4 <sup>†</sup>	Observed in individuals with low DPD activity (29, 34), but not in another study (35). Associated with toxicity (27, 36, 37).
	*5 <sup>†</sup>	Associated with toxicity in some (38) but not other studies (39). Not associated with low DPD activity (35). Not associated with DPD protein levels (40).
	*6 <sup>†</sup>	Associated with toxicity (10, 41). Not associated with low DPD activity (1)
	*9A <sup>†</sup>	Associated with reduced DPD activity (31) but not associated with reduced DPD in other study (29). Associated with toxicity (42)
1: an important caveat for all genotyping tests is that the decision to assign an allele a “wild-type” status is based upon a genotyping test that interrogates only the most common and already-proven sites of functional variation. In human DNA, it is always possible that a new, previously undiscovered (and therefore un-interrogated) site of variation may confer loss-of-function in an individual, and thus lead to the rare possibility of a non-functional allele being erroneously called as “wild-type.”		

<sup>†</sup>Designated as fully functional by the Dutch Pharmacogenetics Working Group based on the lack of an association to toxicity reported in studies and/or decreased clearance or activity (43).

<b>Supplemental Table S3. Frequencies<sup>1</sup> of alleles in major race<sup>2</sup> groups</b>				
Allele	Caucasian	Asian	African-American or Black	Middle Eastern
*2A	0.00862	0.0015	0	0
*3	0	0	0	0
*4	0.0194	0.001	0.00237	0.0293
*5	0.147	0.268	0.177	0.119
*6	0.0412	0.015	0.0451	0.092
*7	0.00122	0	n/a	n/a
*9A	0.182	0.0315	0.137	n/a
*11	n/a	0.0015	n/a	n/a
*12	0	0	n/a	n/a
*13	0.001	0	n/a	n/a
IVS10-15T>C	n/a	0.018	0.042	n/a
rs75017182	0.0155	n/a	n/a	n/a
rs67376798	0.0111	n/a	n/a	n/a
<sup>1</sup> Average frequencies are reported based on the average from the actual numbers of subjects with each allele reported in multiple studies. See Supplemental Table S4 for details and references.				
<sup>2</sup> Race/ethnic group designations correspond to those indicated in Supplemental Table S4				

<b>Supplemental Table S4. <i>DPYD</i> minor allele frequency</b>															
<b>Pooled grouping</b>	<b>Ethnicity</b>	<b><i>DPYD</i> minor allele frequency (%)</b>													<b>Total Alleles</b>
		<b>*2A</b>	<b>*3</b>	<b>*4</b>	<b>*5</b>	<b>*6</b>	<b>*7</b>	<b>*9A</b>	<b>*11</b>	<b>*12</b>	<b>*13</b>	<b>IVS10 - 15T&gt; C</b>	<b>rs7501718 2</b>	<b>rs6737679 8</b>	
Caucasian	Finnish (15)	2.2	-	-	-	-	-	-	-	-	-	-	-	-	90
Caucasian	Finnish (34)	-	-	3.3	7.2	6.7	-	-	-	-	-	-	-	-	180
Caucasian	Scottish (44)	0	0	0.8	28	5.8	-	-	-	-	-	-	-	-	120
Caucasian	Dutch (45)	0.9 1	-	-	-	-	-	-	-	-	-	-	-	-	2714
Caucasian	German (22)	0.5	-	-	-	-	-	-	-	-	-	-	-	-	1702
Caucasian	German (30)	-	-	1.6	14	2.0	0.3	19	-	-	-	-	-	0.6	314
Caucasian	French (21)	0.6	-	-	-	-	0	17. 7	-	0	-	-	-	-	504
Caucasian	French (12)	1.1	-	-	-	-	-	-	-	0	0.1	-	-	1.0	974
Caucasian	Swedish (46)	1.6	-	-	-	-	-	-	-	-	-	-	-	-	1284
Caucasian	Bosnian (47)	2.0	-	-	-	-	-	-	-	-	-	-	-	-	100
Caucasian	Dutch (48)	-	-	-	-	-	-	-	-	-	-	-	1.3	-	382
Caucasian	German (48)	-	-	-	-	-	-	-	-	-	-	-	1.7	-	906
Caucasian	Turkish (49)	0	-	-	-	-	-	-	-	-	-	-	-	-	500

<b>Supplemental Table S4. <i>DPYD</i> minor allele frequency</b>															
<b>Pooled grouping</b>	<b>Ethnicity</b>	<b><i>DPYD</i> minor allele frequency (%)</b>													<b>Total Alleles</b>
		<b>*2A</b>	<b>*3</b>	<b>*4</b>	<b>*5</b>	<b>*6</b>	<b>*7</b>	<b>*9A</b>	<b>*11</b>	<b>*12</b>	<b>*13</b>	<b>IVS10 - 15T&gt; C</b>	<b>rs7501718 2</b>	<b>rs6737679 8</b>	
Caucasian	Turkish (50)	0.46	-	-	-	-	-	-	-	-	-	-	-	-	436
African	Afro-American (15)	0	-	-	-	-	-	-	-	-	-	-	-	-	40
African	Afro-American (34)	-	-	0.5	22.7	1.9	-	-	-	-	-	-	-	-	210
African	Tunisian (51)	0	0	0	12.7	7.1	-	13.7	-	-	-	4.2	-	-	212
Asian	Japanese (15)	0	-	-	-	-	-	-	-	-	-	-	-	-	70
Asian	Taiwanese (15)	2.7	-	-	-	-	-	-	-	-	-	-	-	-	72
Asian	Taiwanese (34)	-	-	0	21	1.4	-	-	-	-	-	-	-	-	262
Asian	Japanese (34)	-	-	1.1	35.2	4.4	-	-	-	-	-	-	-	-	100
Asian	Taiwanese (40)	-	0	28.3	1.2	-	-	2.2	-	-	-	-	-	-	600
Asian	Japanese (52)	-	-	-	-	-	-	2.9	-	-	-	-	-	-	2692

<b>Supplemental Table S4. <i>DPYD</i> minor allele frequency</b>															
<b>Pooled grouping</b>	<b>Ethnicity</b>	<b><i>DPYD</i> minor allele frequency (%)</b>													<b>Total Alleles</b>
		<b>*2A</b>	<b>*3</b>	<b>*4</b>	<b>*5</b>	<b>*6</b>	<b>*7</b>	<b>*9A</b>	<b>*11</b>	<b>*12</b>	<b>*13</b>	<b>IVS10 - 15T&gt; C</b>	<b>rs7501718 2</b>	<b>rs6737679 8</b>	
Asian	Japanese (53)	0	-	-	28.3	1.5	0	2.9	0.15	0	0	1.8	-	-	682
Asian	Chinese (38)	0	0	0	30	-	-	4.7	-	0	-	-	-	-	150
Asian	Chinese (54)	0	0	0	20.8	0.7	-	7.0	-	-	0	-	-	-	284
Middle Eastern	Egyptian (55))	0	0	2.9	11.9	9.2	-	-	-	-	-	-	-	-	478

Supplemental Table S5. Evidence linking <i>DPYD</i> genotype with DPD phenotype and dihydropyrimidine toxicity			
Type of experimental model (in vitro, in vivo preclinical, or clinical)	Major findings	References	Level of evidence <sup>a,b</sup>
In vitro	<i>DPYD*2A</i> was expressed in mammalian cells and the DPD enzymatic activity was determined relative to wild-type. The <i>DPYD*2A</i> was found to be catalytically inactive.	Offer et al. (2013) (28)	High
Clinical	Severe neurotoxicity due to 5-fluorouracil and complete DPD deficiency in a patient with familial pyrimidinemia was observed. A markedly prolonged 5-fluorouracil elimination half-life was observed after administration of a test dose of 5-fluorouracil in this patient.	Diasio et al. (1988) (56)	Moderate
Clinical	In a study of patients with severe 5-fluorouracil-related toxicity and low DPD activity in peripheral blood mononuclear cells isolated from patients (n=14), six were heterozygous for the <i>DPYD*2A</i> variant and one heterozygous for the <b>rs67376798</b> variant.	van Kuilenburg et al. (2000) (16)	Moderate
Clinical	In a study of cancer patients with reduced (n=23) or normal (n=14) DPD activity, 1 patient with reduced activity and 5-fluorouracil toxicity was heterozygous for <i>DPYD*13</i> and one patient with normal activity was heterozygous for <i>DPYD*2A</i> .	Collie-Duguid et al. (2000) (29)	Moderate
Clinical	In a study of patients with severe 5-FU-related toxicity (n=25), 5 were heterozygous and 1 was homozygous for the <i>DPYD*2A</i> variant. Lethal outcome was seen in the homozygous and two of the heterozygous cases.	Raida et al. (2001)(22)	Moderate
Clinical	Severe toxicity observed in a single patient with colorectal cancer treated with 5-fluorouracil heterozygous for the <i>DPYD*2A</i> . Further analysis revealed low DPD activity in peripheral blood mononuclear cells and reduced clearance in this patient as compared to control patients with normal 5-fluorouracil symptoms.	Maring et al. (2002)(17)	Moderate

Clinical	Severe toxicity and profound DPD deficiency (in peripheral blood mononuclear cells) in a breast cancer patient heterozygous for <b>DPYD*2A</b> and <b>DPYD*13</b> treated with a 5-fluorouracil-based regimen.	Johnson <i>et al.</i> (2002) (8)	Moderate
Clinical	In a study of cancer patients (n=60) suffering from severe grade 3-4 toxicity after the administration of 5-fluorouracil, 17 patients were heterozygous and 1 patient was homozygous for <b>DPYD*2A</b> variant (28%). All patients but one with the <b>DPYD*2A</b> variant had reduced DPD activity in their peripheral blood mononuclear cells.	van Kuilenburg <i>et al.</i> (2002)(18)	Moderate
Clinical	In a study of cancer patients (n=25) suffering from severe grade 4 toxicity after the administration of 5-fluorouracil, 9 patients were heterozygous and 1 patient was homozygous for <b>DPYD*2A</b> variant and decreased DPD activity could be detected in peripheral blood mononuclear cells.	Van Kuilenburg <i>et al.</i> (2002) (19)	Moderate
Clinical	In a study of Portuguese cancer patients receiving fluorouracil-based chemotherapy (n=73), 8 had grade 3-4 toxicity, one of which was heterozygous for the <b>DPYD*2A</b> variant.	Salgueiro <i>et al.</i> (2004) (57)	Moderate
Clinical	In a large study of colorectal cancer patients (n=252), where ten patients were heterozygous for either the <b>DPYD*2A</b> variant (n=3) or the <b>DPYD rs67376798</b> (n=8) variant, seven had toxicity (grade 3-4), one of which was fatal (heterozygous for both variants). Two patients had grade I toxicity and the dose was immediately reduced. Only one patient with the <b>rs67376798</b> had no toxic side effects. <b>DPYD*2A</b> and <b>rs67376798</b> resulted in statistically significant reduced clearance of 5-fluorouracil and lower DPD activity (as determined by dihydrouracil/uracil ratio) as compared to patients with wild-type <b>DPYD</b> .	Boisdron-Cellier <i>et al.</i> (2007) (21)	High
Clinical	In a large study of cancer patients treated with various fluorouracil-based regimens (n=487) 60% of patients with either the <b>DPYD*2A</b> (n=6 of 10) and <b>rs67376798</b> (n=6 of 10) variants experienced grade 3 or 4 toxicity. Four of those with no toxicity were receiving reduced doses. One patient with <b>DPYD*13</b> was identified and experienced grade 3 or 4 toxicity. Patients with these variants had significantly reduced 5-fluorouracil plasma clearance.	Morel <i>et al.</i> (2006) (12)	High

Clinical	Fatal toxicity in a single patient heterozygous for the <b>DPYD*2A</b> variant with metastatic pancreatic adenocarcinoma treated with bolus fluorouracil.	Saif <i>et al.</i> (2007) (24)	Moderate
Clinical	In a retrospective study of case reports of patients with various cancers treated with fluorouracil-based regimens who experienced severe toxicity (n=93), two heterozygotes for <b>DPYD*2A</b> were found. Both patients had low DPD activity in their peripheral blood mononuclear cells.	Magne <i>et al.</i> (2007) (14)	Weak
Clinical	In a study of colorectal cancer patients treated with fluorouracil regimens (n=76), nine patients had reduced clearance and increased toxicity, one of whom was heterozygous for the <b>DPYD*2A</b> variant and two of whom were heterozygous for the <b>rs67376798</b> variant.	Capitain <i>et al.</i> (2008) (11)	Moderate
Clinical	In a large prospective study of fluorouracil monotherapy for various cancers (n=683), <b>DPYD*2A</b> was associated with increased risk of mucositis and leukopenia. Six out of thirteen patients with the <b>DPYD*2A</b> variant and 3 out of the 5 patients with the <b>rs67376798</b> had grade 3-4 toxicity.	Schwab <i>et al.</i> (2008) (3)	High
Clinical	In a large study of Polish colorectal cancer patients treated with fluorouracil (n=252), four patients had grade 3-4 neutropenia one of whom was heterozygous for <b>DPYD*2A</b> .	Sulzyc-Bielicka <i>et al.</i> (2008) (58)	Moderate
Clinical	In a study of patients with various cancers treated with either fluorouracil or capecitabine (n=181), five patients with <b>DPYD*2</b> variant and one patient with <b>rs67376798</b> variant all experienced grade 3-4 toxicity.	Gross <i>et al.</i> (2008)(59)	Moderate
Clinical	In a study of colorectal cancer patients treated with 5-fluorouracil- or capecitabine-based regimens (n=76 grade 3-4 toxicity; n=48 tolerant), <b>DPYD*2A</b> was associated with increased risk of mucositis. Five out of five patients with <b>DPYD*2A</b> variant had grade 3-4 toxicity (no patients with this variant were detected in the tolerant group).	Kleibl <i>et al.</i> (2009) (41)	Moderate
Clinical	In a study of patients with various cancers treated with fluorouracil-based regimens (n=111), the <b>DPYD*2A</b> variant was not significantly associated with increased risk of toxicity however only a single heterozygote was observed; this patient experienced grade 0-2 toxicity. One patient with the <b>DPYD*13</b> variant experienced grade 0-2 toxicity.	Amstutz <i>et al.</i> (2009) (13)	Weak



Clinical	In a large prospective clinical trial of various fluorouracil-based regimens for colorectal cancer (n=750), the <b>DPYD*2A</b> was not significantly associated with increased risk of toxicity. Seven patients were heterozygous for <b>DPYD*2A</b> of whom 4 experienced grade 3-4 toxicity.	Braun <i>et al.</i> (2009) (42)	Moderate
Clinical	In a study of patients with various cancers treated with a fluorouracil-based regimen and experiencing grade 3 or 4 toxicities (n=47), 4 patients were heterozygous for the <b>DPYD*2A</b> variant, 4 were heterozygous for the <b>rs67376798</b> variant and one was heterozygous for the <b>DPYD*13</b> variant accounting for 19% of all toxicity cases.	Loganayagam <i>et al.</i> (2010) (20)	Moderate
Clinical	In a large study of metastatic colorectal cancer patients treated with fluorouracil regimens (n=349), only two heterozygotes for <b>DPYD*2A</b> were observed, both of whom had grade 4 neutropenia.	Boige <i>et al.</i> (2010) (60)	Moderate
Clinical	In a study of cancer patients receiving capecitabine or fluorouracil (n=50), one patient was heterozygous for <b>DPYD*2A</b> and experienced grade 4 toxicity which was fatal.	Ceric <i>et al.</i> (2010) (47)	Moderate
Clinical	In a study of patients with gastrointestinal cancers who experienced severe fluorouracil toxicity (n=45), two patients were heterozygous for the <b>DPYD*2A</b> variant.	Savva-Bordalo <i>et al.</i> , (2010) (26)	Moderate
Clinical	In a study of patients with colorectal cancer treated with fluorouracil or capecitabine regimens (n=68), <b>DPYD*2A</b> was associated with increased risk of toxicity. Two <b>DPYD*2A</b> heterozygotes were observed in this cohort. One <b>rs67376798</b> heterozygote was observed but did not experience toxicity.	Kristensen <i>et al.</i> (2010) (9)	Moderate
Clinical	In a study of French breast cancer patients receiving capecitabine (n=105), a single patient who was heterozygous for the <b>DPYD*2A</b> variant had fatal toxicity.	Largillier <i>et al.</i> (2006) (61)	Moderate
Clinical	In a study of patients with colorectal cancer treated with capecitabine regimens (n=568), <b>DPYD*2A</b> and <b>rs67376798</b> were associated with increased risk of severe toxicity. Seven out of seven patients heterozygous for <b>DPYD*2A</b> and seven out of eight patients heterozygous for <b>DPYD rs67376798</b> had grade 3-4 toxicity.	Deenen <i>et al.</i> , (2011) (10)	High

Clinical	In a study of patients with rectal cancer receiving tegafur (n=63), a single patient who was heterozygous for variant <b>DPYD*2A</b> experienced very early grade 4 neutropenia and a patient who was heterozygous for variant <b>rs67376798</b> experienced grade four diarrhea.	Cellier <i>et al</i> , (2011) (25)	Moderate
Clinical	In a study of cancer patients receiving 5-fluorouracil, patients who were heterozygous for variant <b>DPYD*2A</b> (n=3) were treated with a dose reduction of 50% and still experienced severe toxicities that resulted in hospitalization of patient and premature discontinuation of treatment.	Magnani <i>et al</i> , (2013) (62)	High

<sup>a</sup>Rating Scheme for Quality of Evidence as per (5)

<sup>b</sup>Some of the small case series, although not strong individually, collectively do support a strong recommendation.

**Supplemental Table S6. Summary of the effects of *DPYD* variants on 5-fluorouracil clearance**

Reference	<i>DPYD</i> *2A	rs67376798	<i>DPYD</i> *13	Wild-type or no <i>DPYD</i> variant noted
Deenen <i>et al.</i> , (2011) (10)	Mean dose intensity after 6 cycles = 56% (n=5)	Mean dose intensity after 6 cycles = 76% (n=5)	No included	Mean dose intensity after 6 cycles =90% (n=410)
Morel <i>et al.</i> (2006) (12)	5-FU clearance NCI grade 0-2 tox = 72.11 L h <sup>-1</sup> m <sup>-2</sup> (n=4)  5-FU clearance for pts with NCI grade 3-4 = 54.32 L h <sup>-1</sup> m <sup>-2</sup> (n=6)	5-FU clearance for pts with NCI grade 0-2 tox = 89.68 L h <sup>-1</sup> m <sup>-2</sup> (n=4)  5-FU clearance for pts with NCI grade 3-4 = 60.76 L h <sup>-1</sup> m <sup>-2</sup> (n=6)	5-FU clearance for pts with NCI grade 3-4 = 41.06 L h <sup>-1</sup> m <sup>-2</sup> (n=1)	5-FU clearance for pts with NCI grade 0-2 tox = 136.33 L h <sup>-1</sup> m <sup>-2</sup> (n=264)  5-FU clearance for pts with NCI grade 3-4 = 78.67 L h <sup>-1</sup> m <sup>-2</sup> (n=19)
Boisdron-Cellier <i>et al.</i> (2007) (21)	5-FU clearance FuFol 21.22 L h <sup>-1</sup> m <sup>-2</sup> (n=2)  UH2/U mean (n=2) 5.16±0.07 µg/L	5-FU clearance FuFol 43.9 L h <sup>-1</sup> m <sup>-2</sup> (n=7)  UH2/U mean (n=7) 4.4±1.6 µg/L	Not included	5-FU clearance FuFol 104.7 L h <sup>-1</sup> m <sup>-2</sup> (n=163)  UH2/U mean (n=163) 8.1±2.5 µg/L

5-FU, 5-fluorouracil; FuFol, Weekly 5-FU 1200 mg/m<sup>2</sup>, 4 h infusion plus 200 mg/m<sup>2</sup> folinic acid

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