

# Use of external protein standards to approach absolute quantification in yeast extracts

Aaron MILLAN-OROEPEZA  
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# Context of study: European project CHASSY

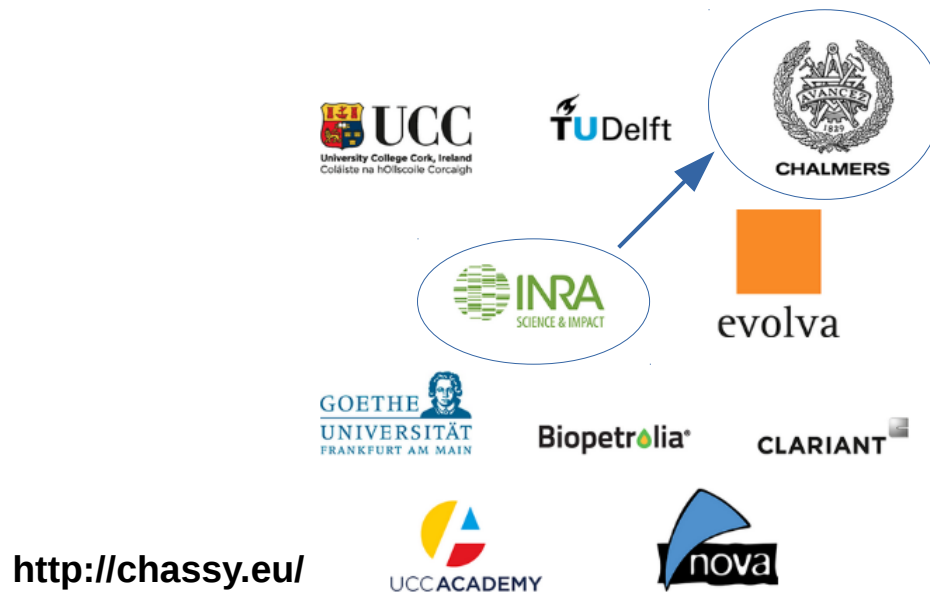
Consortium between nine academy and industry partners aiming to develop yeast as microbial factories using **systems and synthetic biology** (omics) to produce high-added value products of cosmetic and nutraceutic sectors.



<http://chassy.eu/>

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Consortium between nine academy and industry partners aiming to develop yeast as microbial factories using **systems and synthetic biology** (omics) to produce high-added value products of cosmetic and nutraceutic sectors.



Ivan Domenzain  
PhS student at Chalmer UT  
in charge of modeling

Need of absolute quantification in large proteomic datasets with reliable accuracy for **modeling at a genome-scale level**

# Relative and absolute quantification by MS

Quantification in proteomics relies on the ability to detect small changes in protein and peptide abundance in response to different conditions.

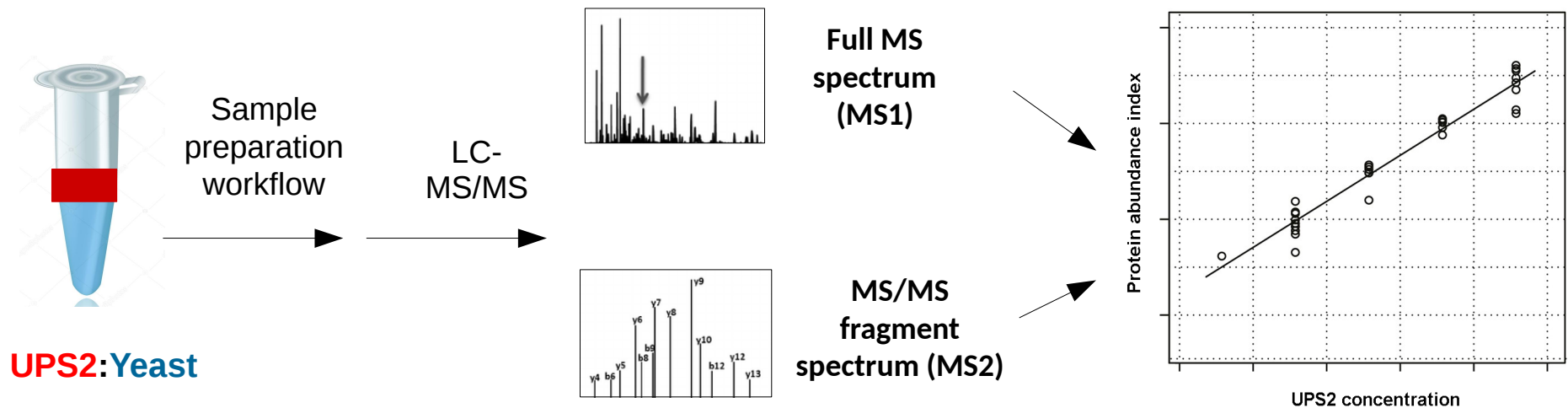
**Relative quantification** compares the levels of a specific protein in different conditions with results being expressed in relative units.

**Absolute quantification** is the determination of the exact amount or mass concentration of a protein (ex. fmol).

# Universal Proteomic Standards (UPS) for "absolute" quantification

**UPS2** is a mixture of 48 human proteins distributed in 6 orders of magnitude well-known to be difficult to analyse.

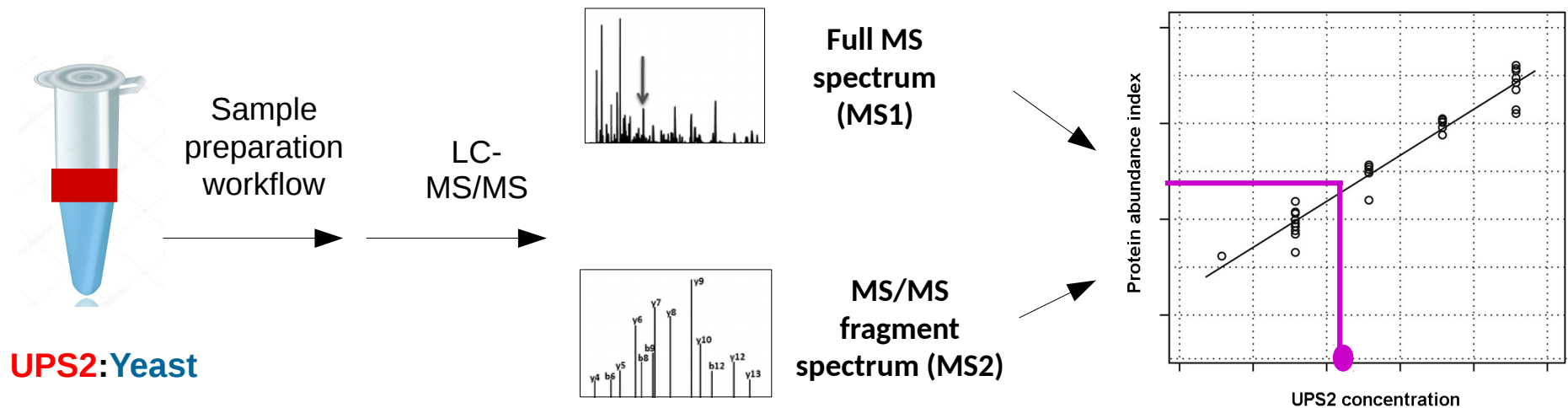
- 50 pmoles ( $10^{-12}$ ) – 500 amoles ( $10^{-18}$ )
- 6 groups (different quantity) of 8 proteins (different molecular weight)



# Universal Proteomic Standards (UPS) for "absolute" quantification






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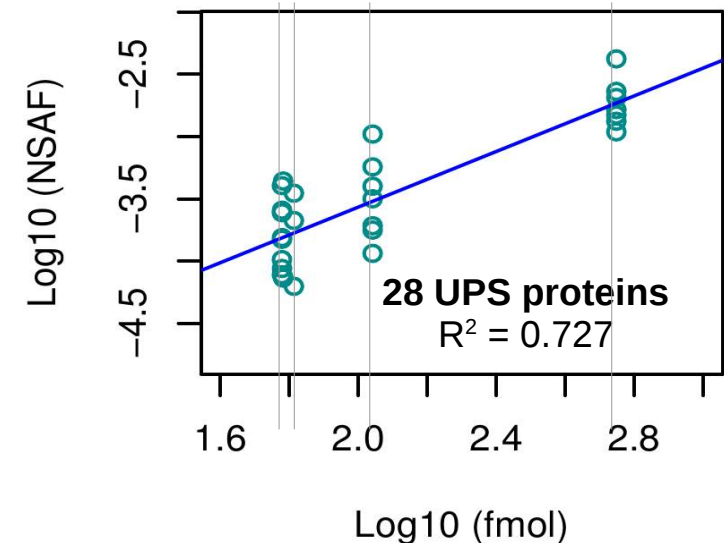
The regression curve between protein **quantification indexes** (MS1 or MS2) and **UPS proteins** concentrations will be used to calculate the **absolute values** of yeast to generate suitable datasets for genome-scale modeling

# Not all the UPS2 proteins can be detected

-  **High costs** of standard 10 µg / 580-750 €
-  Limited to a small number of samples per experiment due to high amounts of UPS2 spiked (**1-4 µg UPS2** per sample)
-  A maximum of **4 orders of magnitude (32 proteins)** have been detected  
(Soufi et al. 2015, Front Microbiol)
-  Ambiguity of details concerning the spiking amounts
-  First tests showed difficulty to detect UPS proteins in complex mixtures (14 proteins, 2 orders of magnitude)

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By adding **UPS1**, standard with the **same 48 proteins** at equimolar concentrations, we were able to generate 2 new orders of magnitude in a complex sample



# Sample preparation : spiked samples

UPS1 (6  $\mu\text{g}$ )

UPS2 (10.6  $\mu\text{g}$ )

Yeast sample x 15 samples  
(5 conditions x 3 replicates)



Reduction, Alkylation, Digestion (Lys-C and  
Trypsin, 1:30)



**127 ng UPS1** → 1 : 6 (UPS:yeast)  
**382 ng UPS2** → 1 : 2 (UPS:yeast)  
**750 ng Yeast**

**1 : 1.5 (UPS:yeast)**

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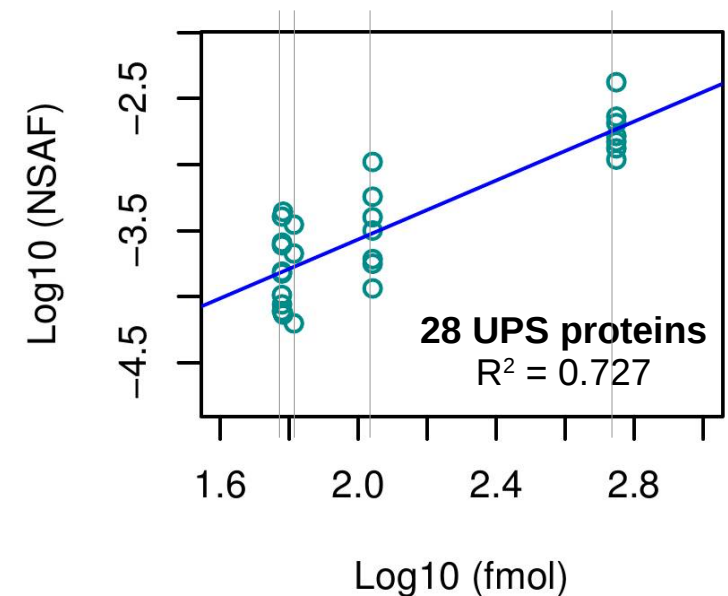
LC-MS/MS

Orbitrap Fusion™ Lumos™ Tribrid™  
50-cm column  
216 min run

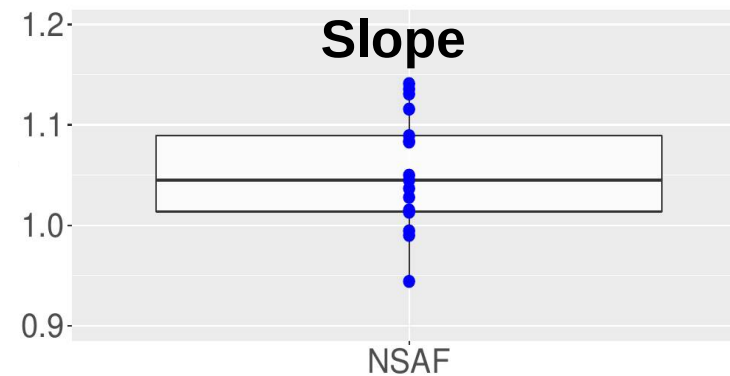
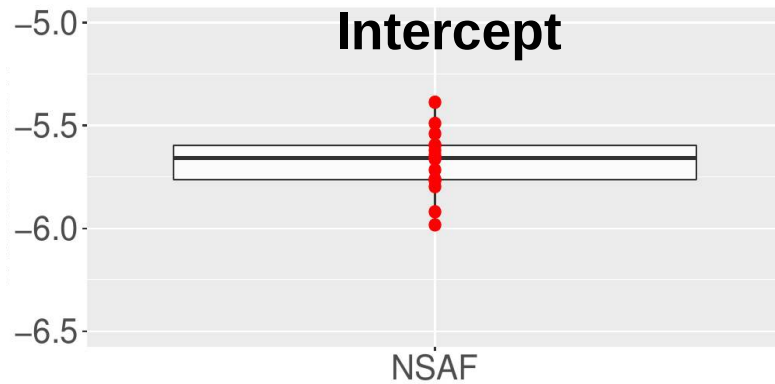
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**1 : 1.5 (UPS:yeast)**

Each yeast sample was **spiked**  
with UPS1 and UPS2 after  
digestion

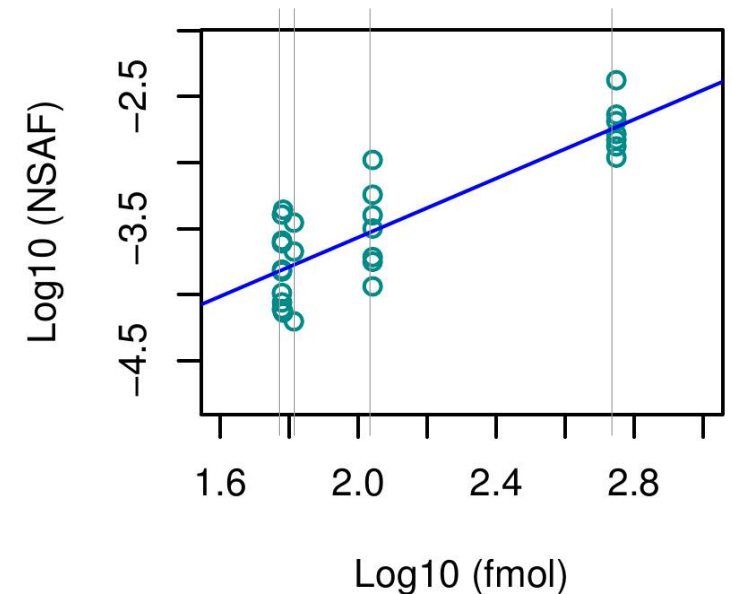


# Spiked samples showed similar linear regressions

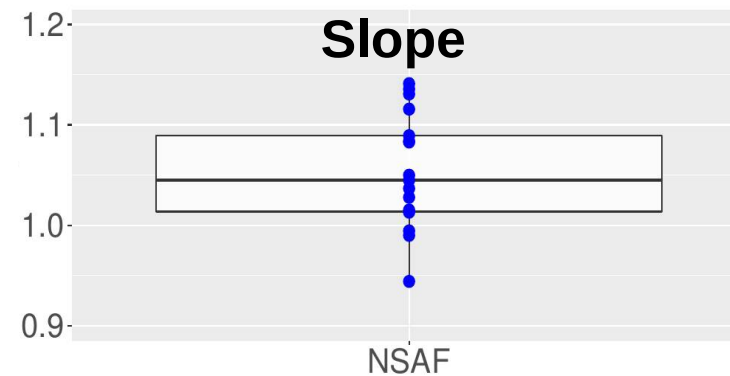
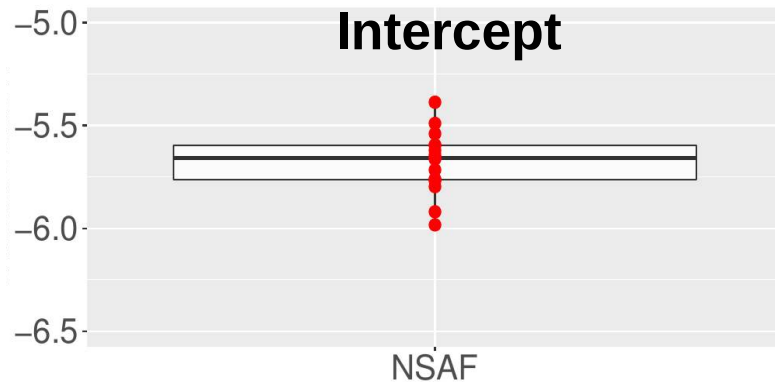


n = 15 samples

Despite the dispersion of proteins on each order of magnitude, the slopes and intercepts were consistent in the 15 samples

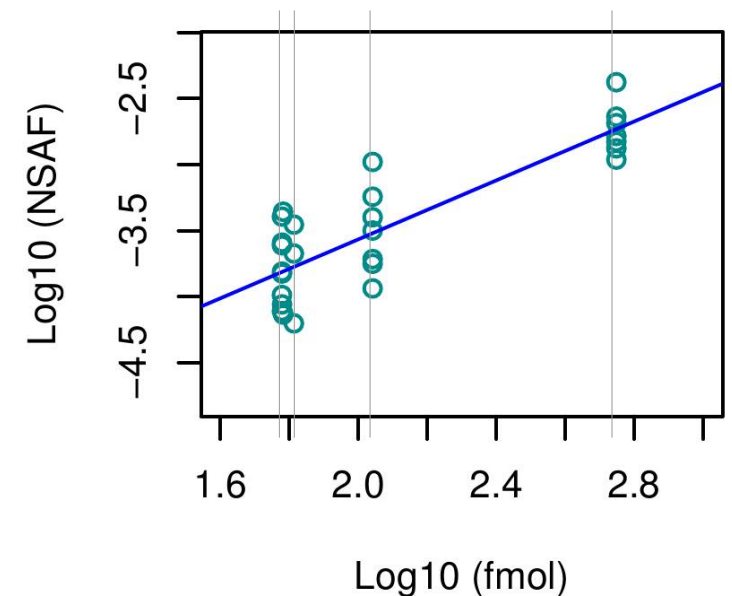


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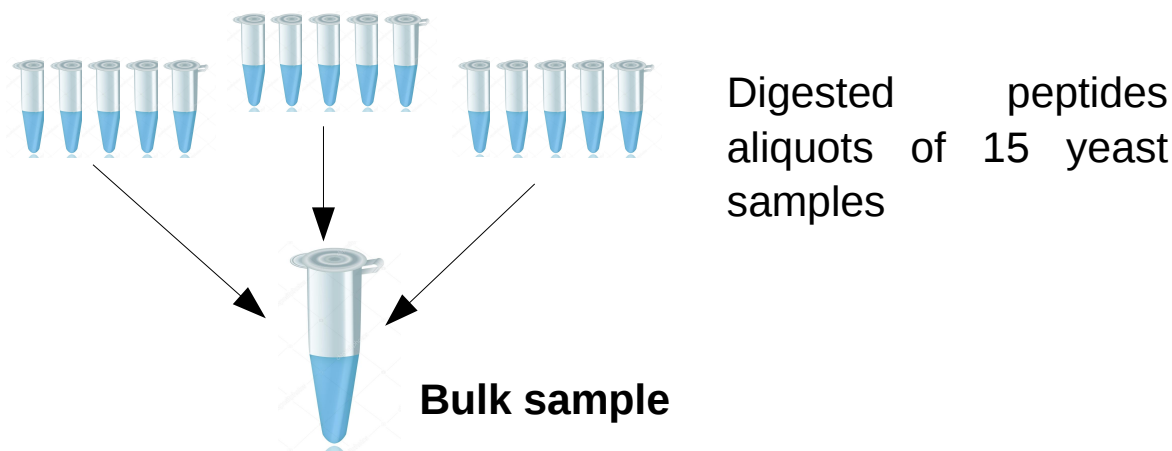
n = 15 samples

Despite the dispersion of proteins on each order of magnitude, the slopes and intercepts were consistent in the 15 samples



Can we use a **representative sample** during the proteomics analysis ?

# Preparation of representative sample: bulk



**127 ng UPS1** → 1 : 6 (UPS:yeast)  
**382 ng UPS2** → 1 : 2 (UPS:yeast)  
**750 ng Bulk**

**1 : 1.5 (UPS:yeast)**



LC-MS/MS

Orbitrap Fusion™ Lumos™ Tribrid™  
50-cm column  
216 min run



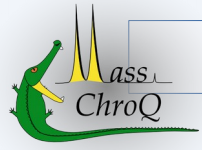
The costs of UPS standards are reduced using a bulk sample

# Bioinformatic workflow



X!Tandempipeline

2739 yeast proteins  
31 UPS proteins



MassChroQ

Data cleaning

Select **UPS** proteins

(5) IBAQ, (6) Top3

Linear regression

**XIC**

Select **UPS** proteins

(1) PAI, (2) emPAI,  
(3) Spectral Count,  
(4) NSAF

Linear regression

Abundance inference on  
yeast proteins

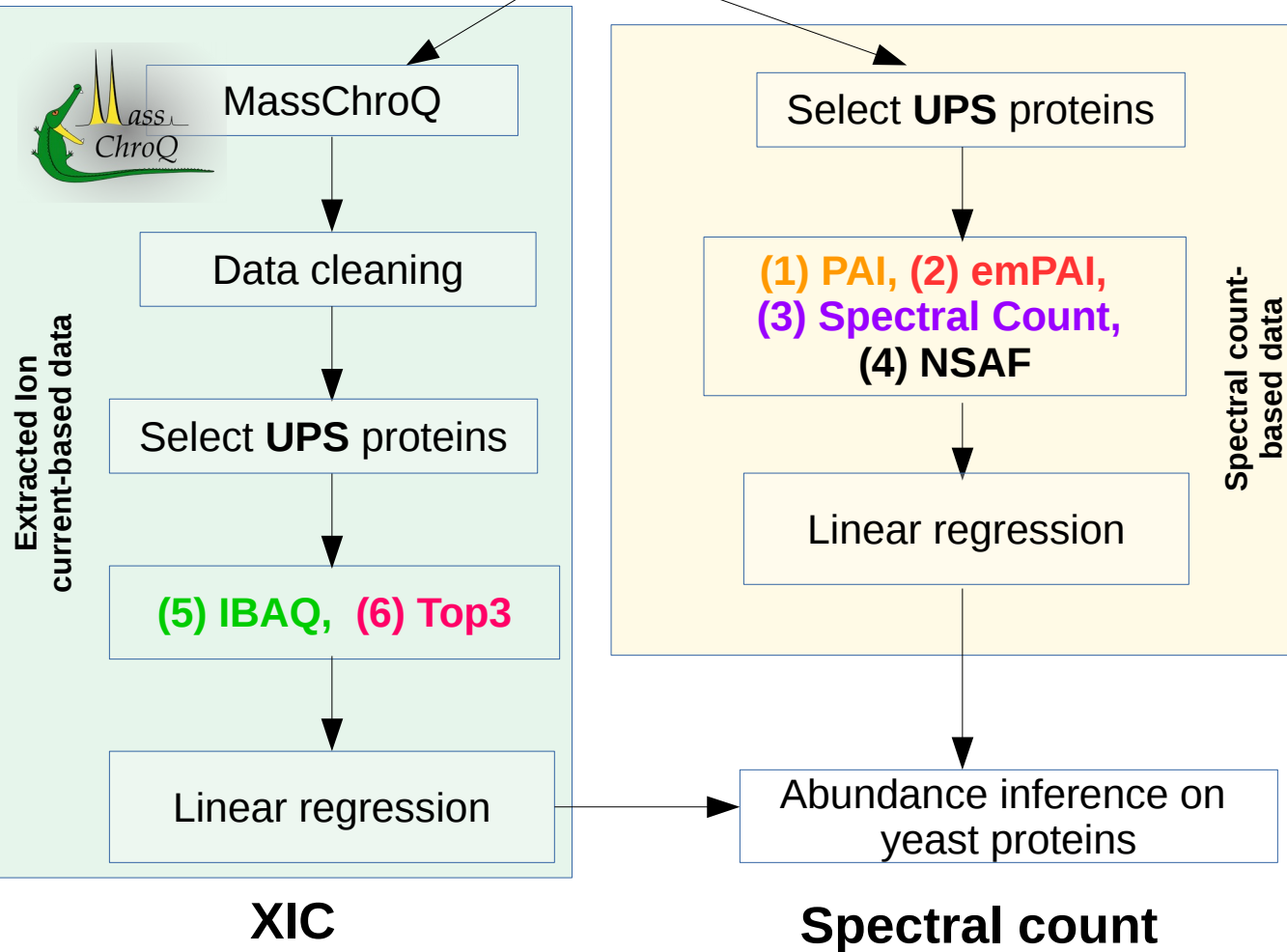
**Spectral count**

# Bioinformatic workflow



**X!Tandempipeline**

2739 yeast proteins  
31 UPS proteins



**PAI:** Observed peptides divided by the number of theoretically observable peptides

**emPAI:**  $10^{\text{PAI}} - 1$

**Spectral Count (SC):** Number of MS2 spectra assigned to a protein

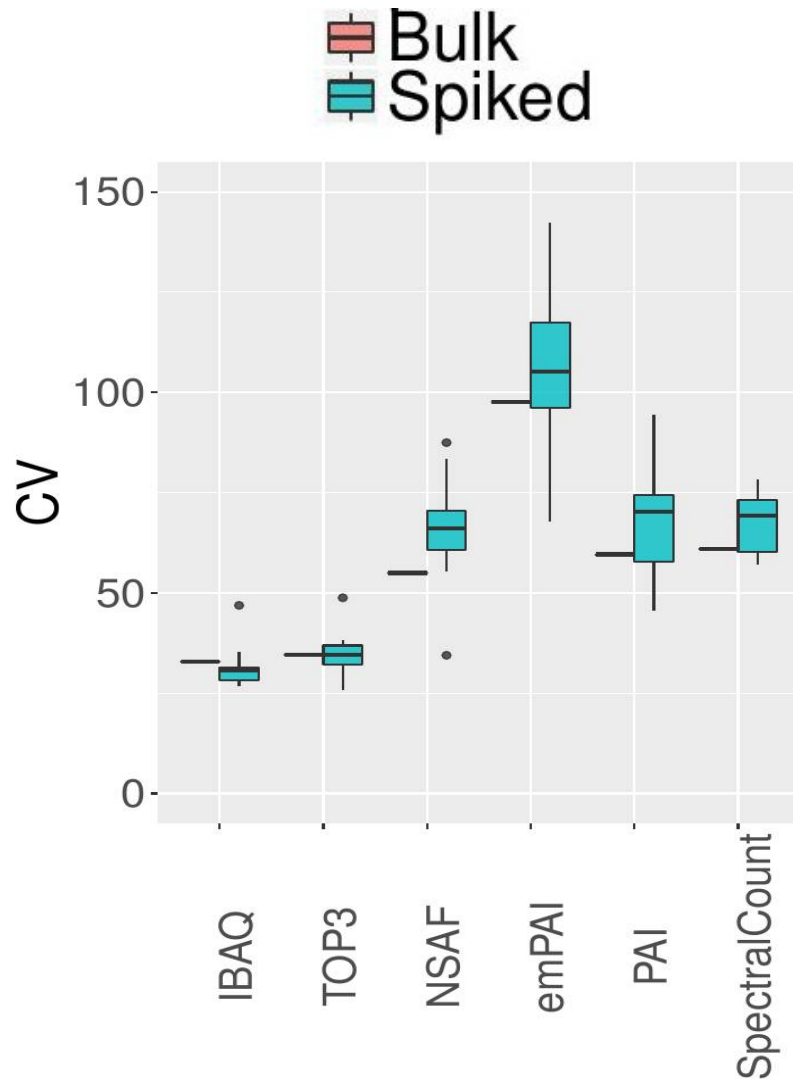
**NSAF:** 
$$\frac{SC/L}{\sum (SC/L)}$$
  
L = length of protein in aa

**IBAQ:** Sum of MS1 intensities of a protein divided by the number of theoretically observable peptides

**Top3:** Average of the 3 most intense MS1 intensities of a protein

Aiming to find the best protein abundance index

# Variation of different abundance indexes

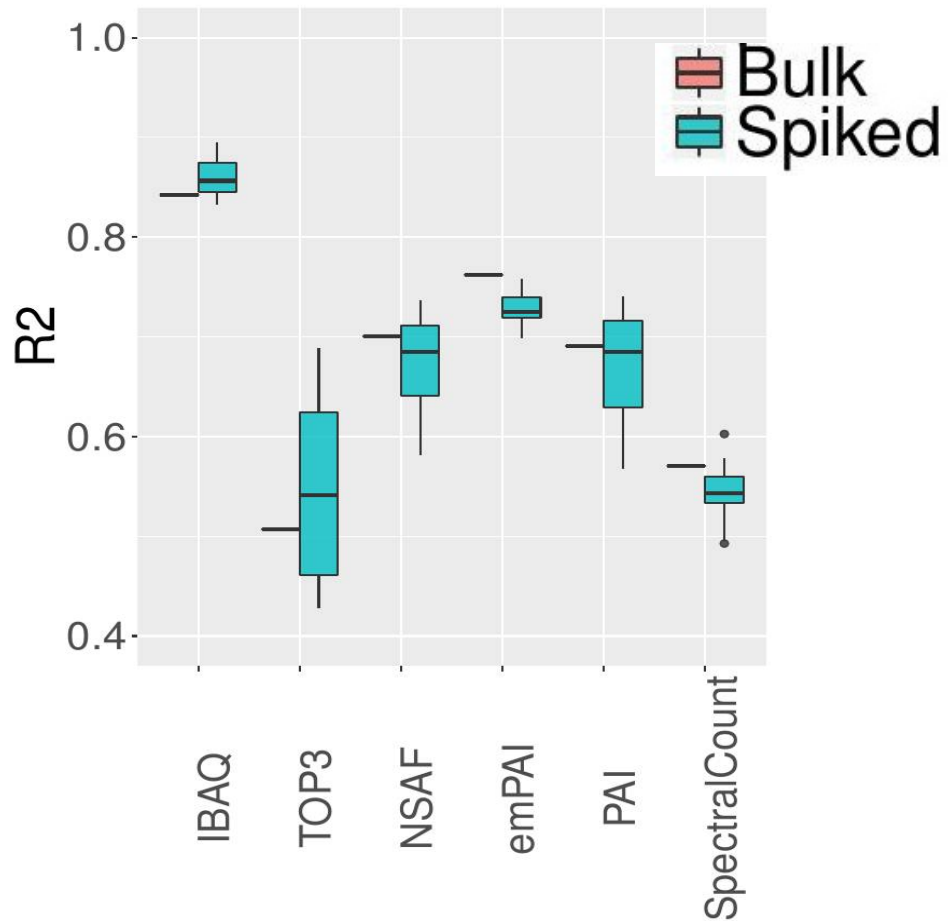


- Overall, the method **IBAQ** showed **the best coefficient of variation (CV)**.
- On every index **bulk** sampling showed a good coefficient of variation compared to spiked samples.

Test conducted on the 31 UPS proteins

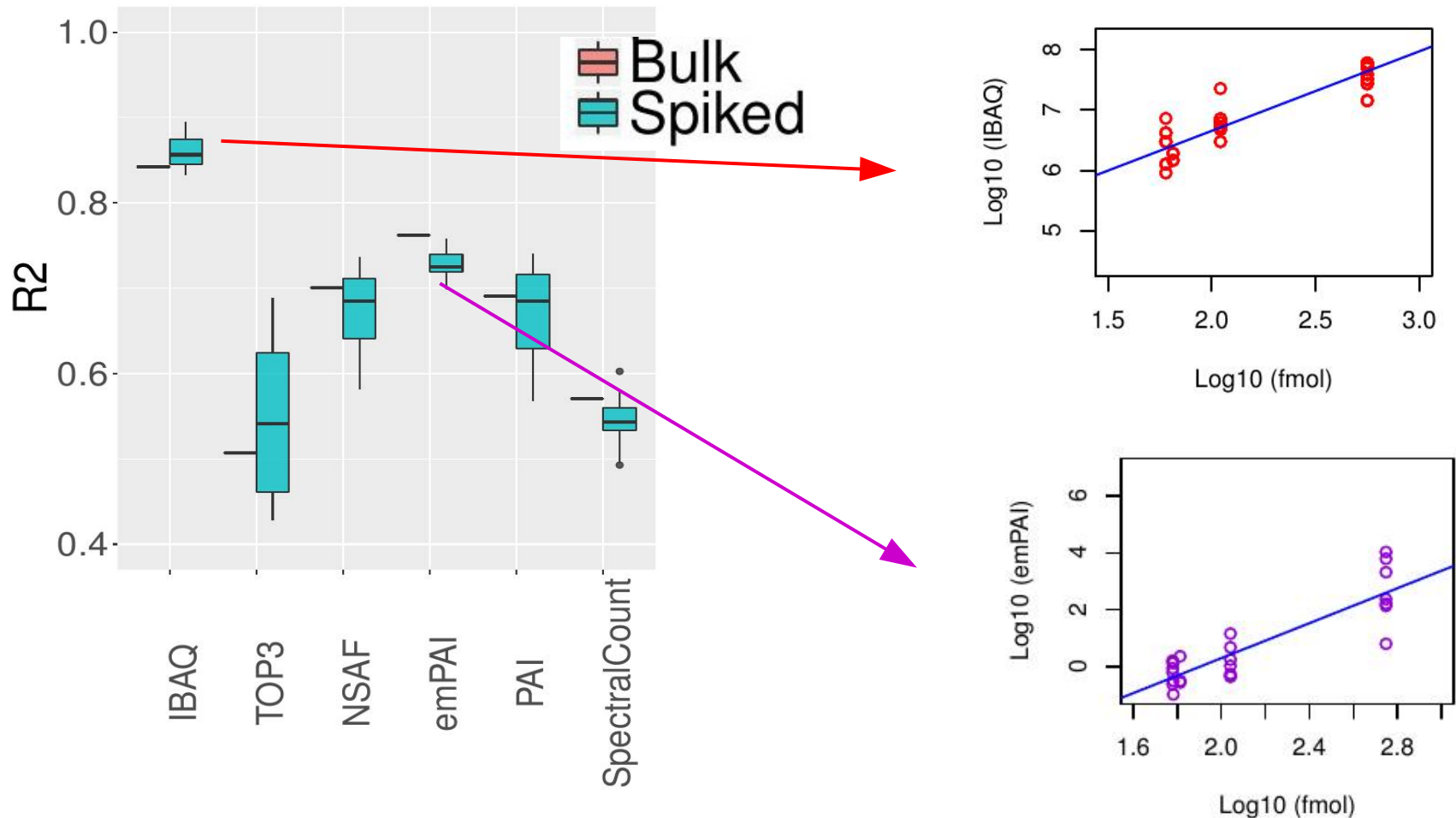


# Coefficient of determination between the different abundance indexes



Test conducted on the 31 UPS proteins

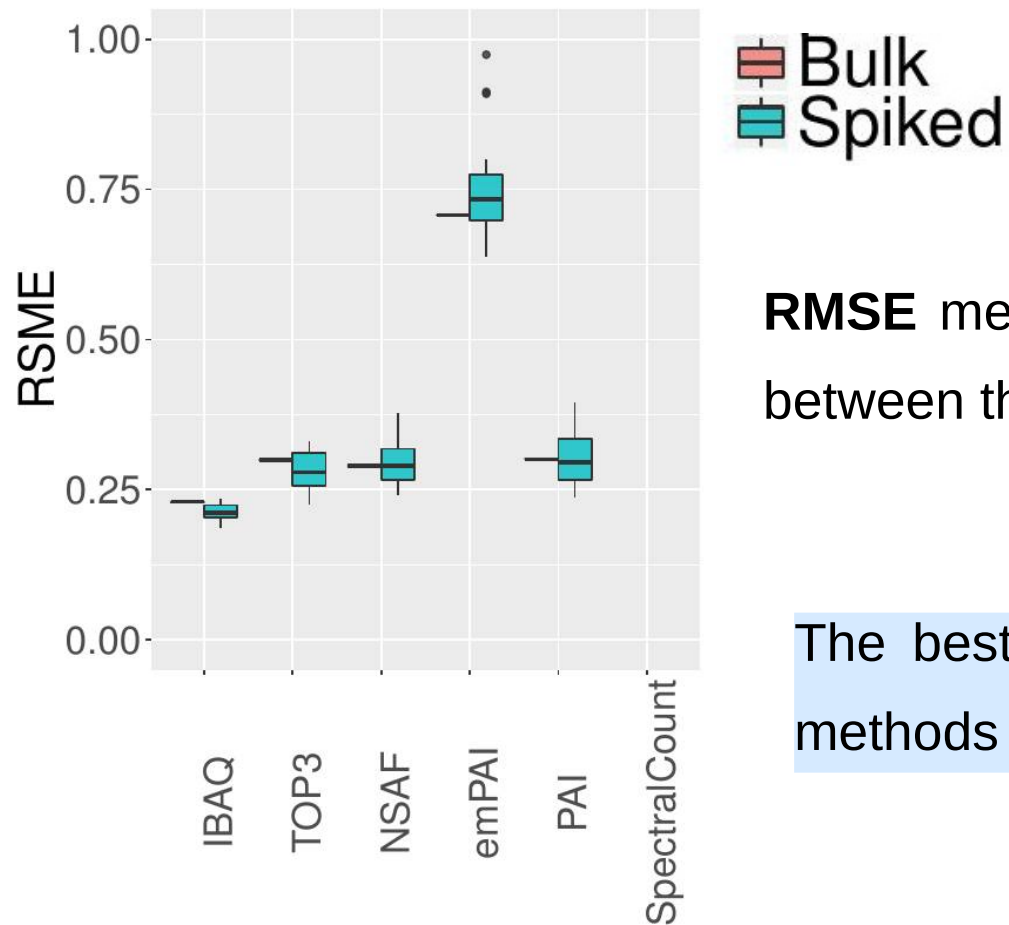
# Coefficient of determination between the different abundance indexes



Test conducted on the 31 UPS proteins

- The method **IBAQ** showed the best linearity  $R^2 = 0.89$

# Goodness of the fit between the different abundance indexes



**RMSE** measures the quality of the fit between the actual data and the predicted model.

The best goodness of fit were obtained by the methods **TOP3**, **NSAF** and **IBAQ**.

Test conducted on the 31 UPS proteins

# Selection the abundance index to compute absolute quantification on yeast

Parameter	Indexes ordered by performance
CV median	Top3, <b>IBAQ</b> , <b>NSAF</b> , PAI, Spectral count, emPAI
R <sup>2</sup>	<b>IBAQ</b> , emPAI, <b>NSAF</b> , PAI, Top3, Spectral count
RMSE	Top3, <b>IBAQ</b> , <b>NSAF</b> , PAI, emPAI, Spectral count

**NSAF : 2739 proteins (ALL !)**

**IBAQ :1776 proteins**

**Top3 : 709 proteins**

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**IBAQ:** Sum of MS1 intensities of a protein divided by the number of theoretically observable peptides

**NSAF:** 
$$\frac{SC/L}{\sum (SC/L)}$$

L= length of protein in aa

# Validation of absolute values (in progress)



**Yeast proteins**  
**8 lab proteins**  
(known concentrations)

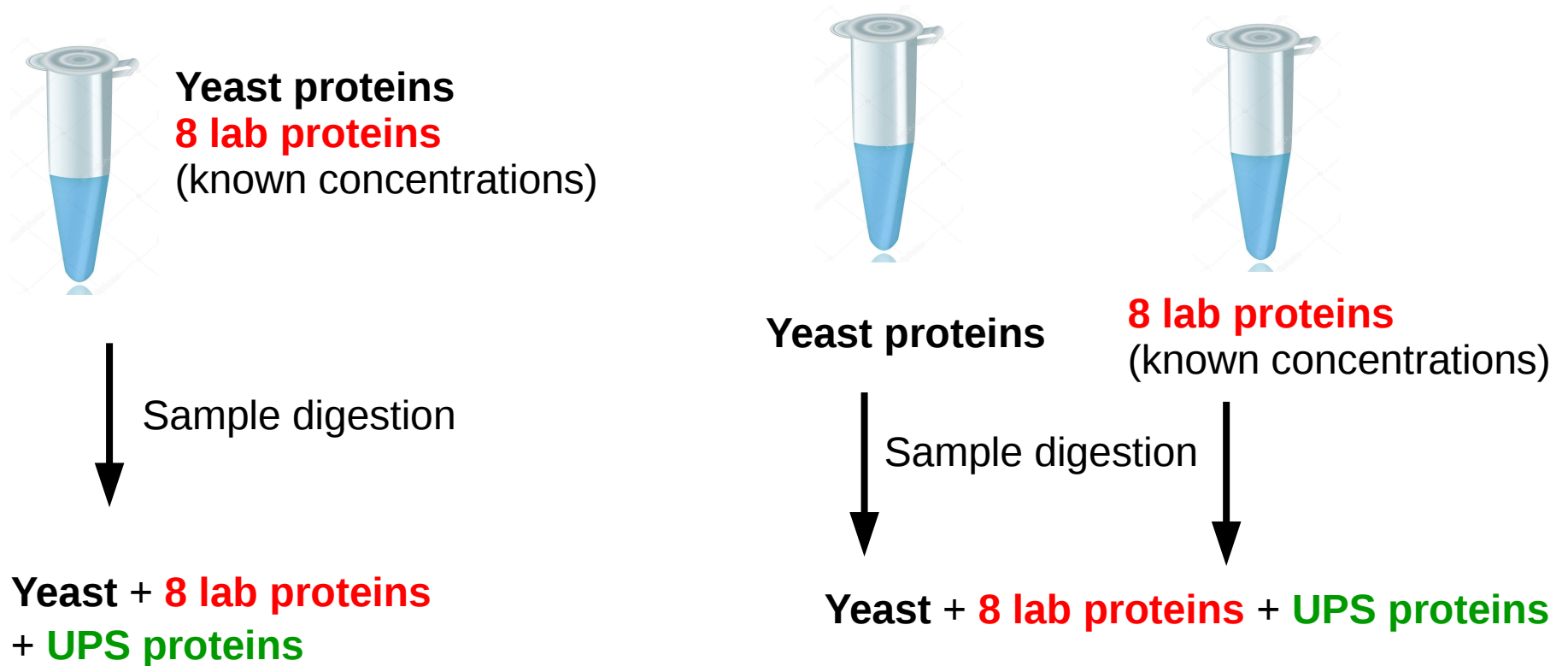


Sample digestion

**Yeast + 8 lab proteins**  
**+ UPS proteins**

Aiming to compare the estimated values against the theoretical values

# Validation of absolute values (in progress)



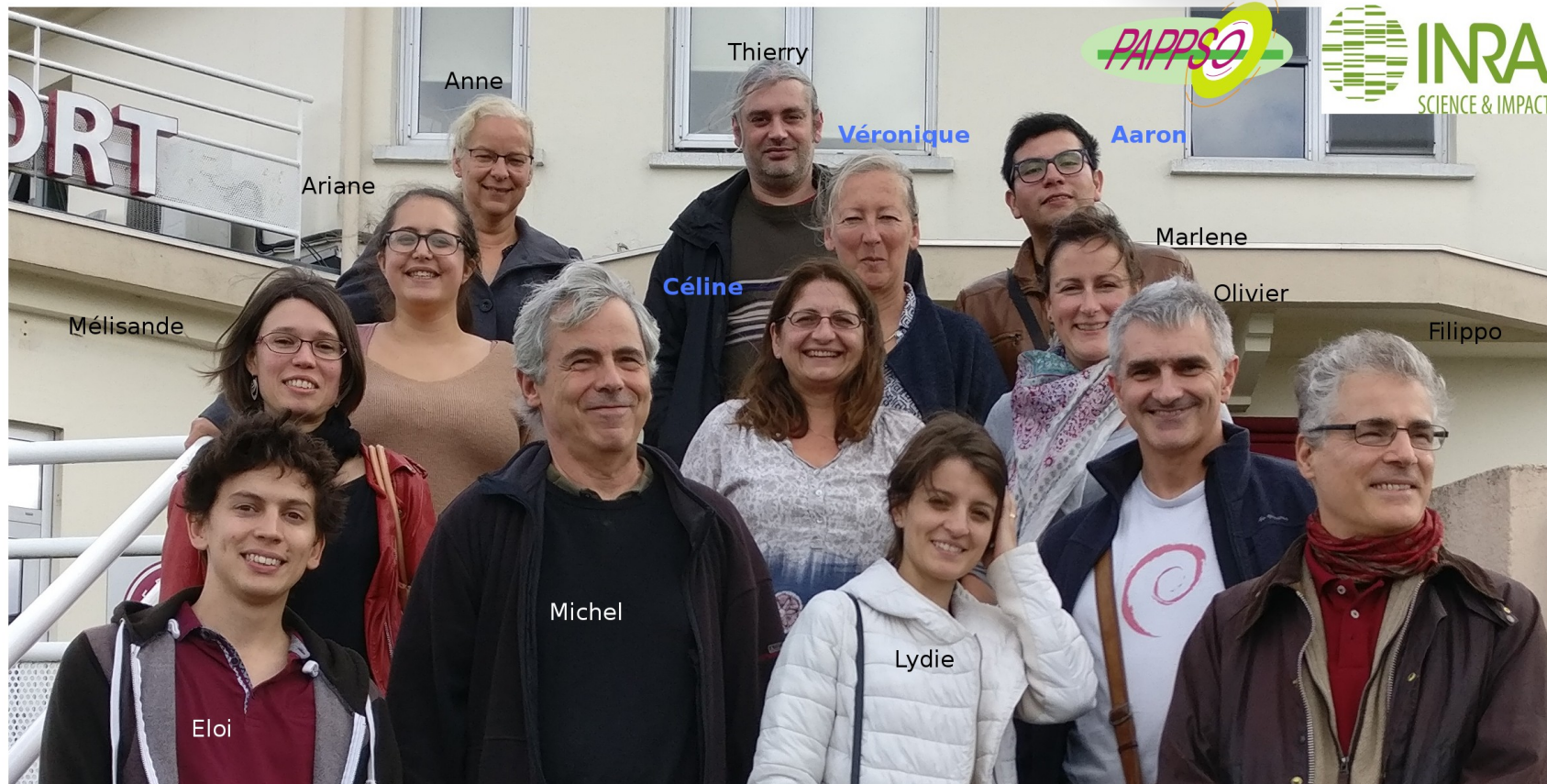
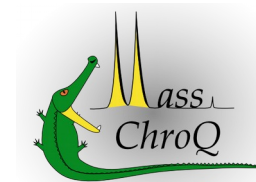
Aiming to compare the estimated values against the theoretical values

# Conclusion

- Considering the performance of the 6 indexes we propose the use of **NSAF** and **IBAQ**
- The use of **bulk** for linear regression represent a solution to the high costs of UPS standards.
- The UPS standards are a good choice to perform absolute quantification suitable for modeling (4 orders of magnitude, 31/48 UPS proteins in a complex mixture)



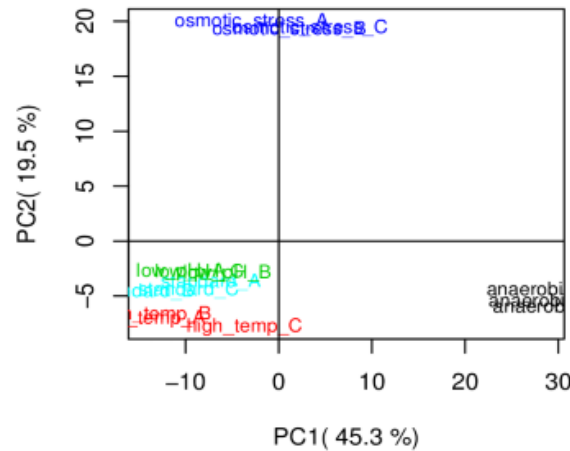
# Thank you



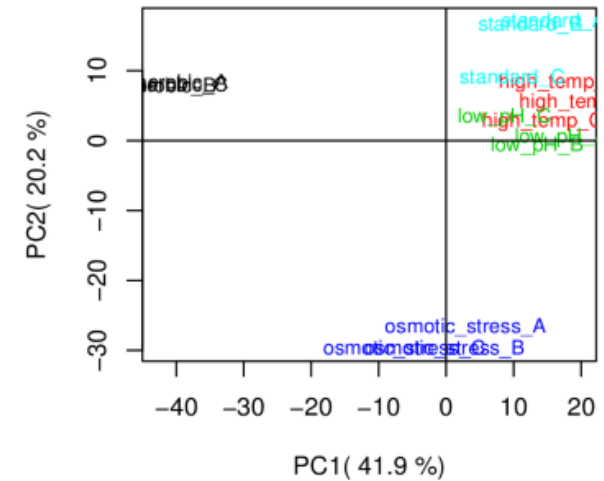
# PCA of “absolute” and relative quantification on yeast extracts

## RELATIVE QUANTIFICATION

Spectral Count: 2316 proteins  
XIC: 1575 proteins



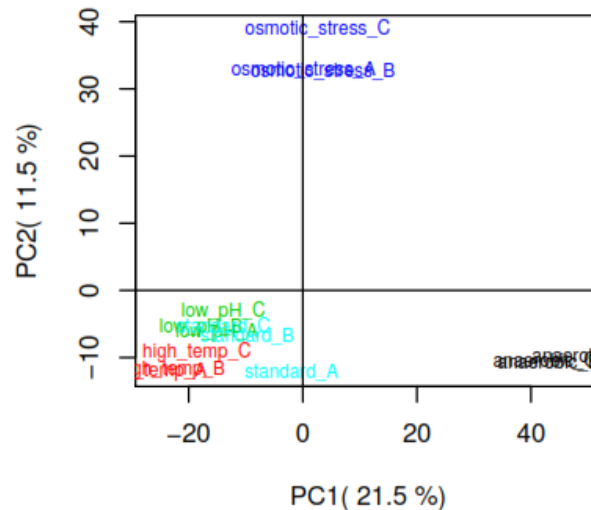
PCA based on Spectral Counts



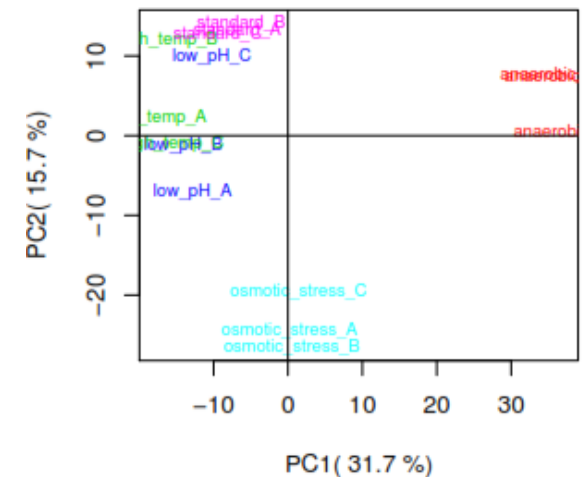
PCA based on XIC

## ABSOLUTE QUANTIFICATION

NSAF : 2739 proteins  
IBAQ : 1776 proteins



PCA based on NSAF -bulk



PCA based on IBAQ -bulk