

Supporting Online Material for

The Genetic Architecture of Maize Flowering Time

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Published 7 August 2009, *Science* **325**, 714 (2009)
DOI: 10.1126/science.1174276

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S1. Materials and Methods:

Germplasm: The germplasm selection process and the design of the population are elaborated in (S1). B73 was chosen as the reference line because it is an important founder line of the US maize crop, and it is currently being sequenced. We crossed 25 other lines that maximized world-wide diversity of maize on the basis of prior SSR marker surveys (S2). B73 was used as the maternal plant, and F1s were self-fertilized for 5 generations without intentional selection to make the lines. Each line descends from a unique F2 generation plant and also a single F5 generation plant, which was self-fertilized one generation and then sib-mated for one to three generations to create sufficient seed supplies to phenotype in replicated experiments.

Genotyping: The details of the genotyping can be found in (S3). Briefly, 7000 lines were scored for 1200 SNP markers. The markers were chosen to maximize information relative to B73. While this resulted in a segregating marker every 1.3cM on average within each family, there were substantial deviations in coverage between families and recombination was quite different between families (S3). This mosaic of genotyping results in limitations on the resolution of some of the QTL positions at this time. Each family is represented by 200 lines in the NAM panel, resulting in a final set of 5,000 mapping lines in total.

Genotyping was conducted to score polymorphisms at *Vgt1* and *ZmRap2.7* across the founder lines and lines with recombination events between *Vgt1* and *Vgt2*. Regions surrounding *Vgt1*, where (S4) detected SNPs in a set of 95 inbred lines, as well as the entire *ZmRap2.7* gene were PCR amplified and Sanger sequenced in the founders. In addition, a diverse association mapping panel of 282 lines (S5) was scored for variation at *Vgt1* and *ZmRap2.7* with sequencing and indel markers on an agarose gel. The associated marker data are presented in Table S5.

Primers:

Vgt1: GTACCAGCTCCCTCTGCTC; TGGAATGGATGTGAAGTGAG

Best *Rap2.7* Association Primers for sequencing: CCTAGGTGAGCAAGAGCAGA;
CTCTGCCATTGCATTGAT

Rap2.7 Insertion for Gel Scoring: CTAAAGGTCTTGGAACAGACAGG;
GAACTGAAGAAAACCAAGGCAAG

Phenotyping: The 5000 NAM lines were evaluated phenotypically in eight summer environments: Aurora, NY; Clayton, NC; Columbia, MO; and Urbana, IL across two years. Each experiment also included 200 randomly chosen intermated B73 x Mo17 (IBM) RILs plus an association mapping panel of 281 diverse inbred lines representing the global diversity of public maize lines (S5). Within each environment, one replicate of each of these 5481 unique lines was grown, but B73 and one other parental founder line were included in each sub-block of 22 plots to permit estimation of within-environment block effects and error variances. Experimental units consisted of plots of 12-25 plants each (depending on the environment), and randomized designs were used. DA and DS were measured as the number of days from planting to the initiation of pollen shed or silk emergence for half the plants within a plot ASI was computed as the difference between DS and DA.

Statistical Analyses:

Field Spatial Correction: Within each environment, mixed linear models were fit including family and line within family genotypic effects; effects of blocks, rows, and columns of the field designs; and spatially correlated residuals (S6). Likelihood ratio tests were used to determine which factors explained significant amounts of variation and were to be retained in the model. A combined mixed model across environments was then fitted, using the components of each individual environment model, adding family- and genotype-by-environment interaction terms and permitting heterogeneous spatially correlated error variance structures across environments. In addition, unique genetic variance components were fit for each of the 25 NAM cross families, the IBM, and the association panel. Best linear unbiased predictors (BLUPs) for each line were predicted from the combined analysis model with ASREML version 2.0 software (S7),

QTL Mapping approaches: Two approaches, composite interval mapping (CIM) and stepwise regression (SR) (S8), were used for QTL mapping. In SR, markers are added sequentially to the model, but can then be dropped if they no longer improve the fit of the model. CIM, using the markers from SR as cofactors to control for background genetic variance, uses maximum likelihood to test putative QTL and estimate their effects at regular intervals along the genome rather than testing only at marker positions. Inclusive CIM (ICIM; S9) is a variant of CIM.

In addition, we either mapped QTL on one population at a time or all the populations together. Mapping using all populations at once is referred to as joint mapping. In single population mapping, information on position of QTL is not shared between populations, while joint mapping uses QTL signal across all populations to identify QTL and position the QTL. Single population QTL mapping will not position QTL inaccurately, but it has far less power and precision than joint mapping. Joint mapping is most powerful when QTL are shared between populations.

Marker imputation: Since stepwise regression cannot use individuals with missing marker data, the first step in the analysis was to impute missing data. Since the genetic map is reasonably dense, the probability of double recombinants between any pair of adjacent markers is low. As a result, any interval flanked by markers originating from the same parent is assumed to have originated from that parent. To build the data set, markers within any interval whose nearest known flanking markers were both from B73 were assigned a value of 0. If the flanking markers both came from the non-B73 parent, markers in the intervals were assigned a value of 2. Markers in intervals flanked by a different parent on either side, i.e. recombinant intervals, were assigned a value of 1. Markers in intervals flanked by heterozygotes were assigned a value of 1. Markers in intervals flanked by a heterozygote and a homozygote were assigned a value of 0.5 or 1.5, depending on whether the homozygote carried B73 or non-B73 alleles.

Single Family Step-wise regression: SR was performed on an individual family by fitting markers as covariates rather than as classification variables. The analysis was performed using SAS GLMSELECT or as part of the ICIM software. Markers were allowed to enter

the model until the next marker added had a p-value greater than .001, at which point the analysis ended. At each step, markers were allowed to stay in the model as long as their marginal p-values were still less than .001. The significance level of .001 was chosen based on permutation analysis (S10).

Joint Stepwise regression: QTL were mapped in combined analysis of all 25 NAM families by joint stepwise regression of flowering time trait BLUPs on markers. In the joint stepwise regression, a family main effect was fit first, and then marker effects nested within families were selected to enter or leave the model based on the p-value calculated for the marginal F-test of that term. An appropriate p-value cutoff was determined to be 0.0001 from permutation tests (S10). After the model was fit with stepwise regression, each marker was dropped from the model one at a time and a single best marker was refit. This step checks the fit of each marker using the remaining QTL as background and improves the overall fit of the model to the data.

Inclusive composite interval mapping: QTL were mapped in individual NAM families using inclusive composite interval mapping (ICIM, S9), a modification of composite interval mapping (CIM). The method first determined a set of cofactors using stepwise regression to fit individual markers. It then scanned the entire genome at 0.1 cM intervals using maximum likelihood to test putative QTL at each point.

Joint inclusive composite interval mapping: We extended inclusive composite interval mapping (ICIM) to multiple population analysis and proposed a new statistical method. Two steps are included. In the first step, population and population by marker interactions were treated as fixed effects. Each marker has 26 levels (B73 and the other 25) in the consensus map. These parameters were included in a multiple regression model, that is,

$$\mathbf{Y} = b_0 + \boldsymbol{\alpha}\mathbf{u} + \mathbf{X}\boldsymbol{\beta} + \boldsymbol{\varepsilon} \quad (1)$$

where \mathbf{Y} is the vector of phenotypic values, b_0 is the intercept, $\mathbf{u}^T = (u_1, u_2, \dots, u_{25})$ is the effect vector indicating the cross effect of each founder with the common parent B73; $\boldsymbol{\alpha}$ is the $n \times 25$ incidence matrix relating each family mean u_f ($f=1, 2, \dots, 26$) to \mathbf{Y} ; $\boldsymbol{\beta}$ is the $26m \times 1$ effect vector of the $n \times 26m$ incidence matrix \mathbf{X} ; m is the number of markers; n is the total number of families; and $\boldsymbol{\varepsilon}$ is the vector of residual variance. To avoid over-fitting the model, we used stepwise regression to estimate the parameters in model (1). If the regression variable was not selected in the model, the corresponding coefficient was set to 0. On the basis of the coefficient estimation, one dimensional scanning was conducted in the second step. In total, the whole genome with the LOD threshold was set to 12.26 to guard against more than 2 false positive linkage results, i.e., genome-wise error rate GWER(2)=12.26 (S11).

Significant allele effects: Once significant loci were identified in the joint analysis, significant alleles were determined with a t-test comparison of their parental means versus the B73 allele. We used $\alpha=0.05$, which from an evaluation of the P -value distributions for all alleles at all loci is approximately an 8% FDR. We observed an elevated frequency of alleles with P -values between 0.05 and 0.125, so this is likely still

undercalling the total number of lines with significant effects. The $\alpha=0.05$ threshold was then used to determine the proportion of families with shared QTL, the number of significant QTL per family, and the significant alleles to predict the parental phenotypes.

QTL×Environment Interactions: QTL-by-environment (QxE) interactions were investigated by fitting the final QTL model based on the line BLUPs computed across all eight environments to a data set containing the BLUPs for each line from each environment analyzed separately. This model was augmented by the inclusion of environment, population-by-environment, and QTL by-environment effects for each QTL. Each QxE interaction was tested based on this general linear model. Any QxE interactions were then included in a similar mixed model that treated both QTL and QxE effects as random. Variance components for each QTL main effect and QxE interaction were estimated from the mixed model in ASREML version 2.0.

Epistasis: Epistasis was tested in two ways: within families separately and in a joint analysis combined across families. To test for epistatic interaction within individual families, we first selected an additive model by stepwise regression within each family separately (using $P < 0.001$ as a significance threshold). Then, we tested all possible pairwise marker interactions to identify the epistatic interaction with smallest p-value when combined with the selected additive model. Finally, permutation tests were used to test the null hypothesis of no epistasis for each family. Permuted data sets were formed by permuting residuals from the main effects only model and adding them back to the predicted values. If the minimum p-value from the original data was smaller than the fifth percentile of minimum p-values from 100 permutation analyses, we declared the presence of at least one significant epistatic interaction within that family.

To test for epistatic interactions across the entire NAM panel, we tested every possible marker pair for interactions (two main effects and one interaction nested within family) in a combined analysis across families and potentially significant interactions were identified as those exceeding the 5% false-discovery rate threshold estimated with the Benjamini-Hochberg procedure (S12). Selected marker pairs were then combined with the model developed from the NAM joint linkage additive QTL analysis. The epistatic interaction was deemed significant across families if $P < 0.0001$ in the combined model.

Pleiotropy: A central issue in evaluating pleiotropy in linkage populations is determining whether correlated effects are the product of linked loci or the same gene. So rather than focusing just on the co-localization of the QTL, we used correlations of effects estimates at each locus to evidence that the same QTL were responsible. With 25 families, we could correlate the effects at each locus for each trait against one another. Those with significantly correlated effects are likely to have the same genes and allele series that are producing the correlation. To remove the issues involved with fitting the QTL positions and estimating effects simultaneously, we produced effects estimates for all three traits (DS, DA, ASI) with the loci determined for each trait ($3 \times 3 = 9$ sets of estimates total). Then for each locus and each trait comparison, we calculated Pearson correlation between the effect estimates. Counts of significant correlation were determined with $P=0.05$, however, the significant loci were frequently much more significant. From this

method, for every pair of traits there are two sets of estimates of proportion of shared pleiotropic loci. For the DS-DA and DS-ASI the two proportions were virtually identical, for DA-ASI there is a slight spread from 14% versus 21%.

Population Structure: For each locus we correlated the effect estimates with Q matrix results from the program STRUCTURE, as reported (S5). These estimates reflect the relationships between tropical and temperate non-stiff stalks well, but they do a poor job in recognizing the Northern Flint-derived materials that are a small but important proportion of our germplasm. For these correlations, we used a modified Q vector representing tropicalness, so that the Northern lines (Il14H, P39, and Ms71) were set to values of -1; i.e., tropical lines had values near 1; mixed origin lines between 0 and 1, temperate lines had values near 0, and Northern lines were -1. Then for each trait and locus, a Pearson correlation between this population structure vector and the effect estimates for each QTL was calculated.

Table S1. Heritabilities for flowering Traits

Table S2. Summary Joint Linkage Stepwise Model and GxE ASREML Analysis for Days to Anthesis

Table S3. Summary Joint Linkage Stepwise Model and GxE ASREML Analysis for Days to Anthesis

Table S4. Summary Joint Linkage Stepwise Model and GxE ASREML Analysis for Days to Anthesis

Table S5. Associating polymorphisms at Vgt1 and Rap2.7 for the maize 282 association panel.

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Supporting Table 1. Heritabilities for flowering Traits

	ASI	Days to Anthesis	Days to Silk
Heritability of line BLUPs across all crosses (excluding association panel):			
Heritability of line BLUPs	0.78	0.94	0.94
Heritability of line BLUPs within each cross:			
B73xB97	0.64	0.77	0.77
B73xCML103	0.61	0.78	0.73
B73xCML228	0.68	0.90	0.89
B73xCML247	0.75	0.90	0.89
B73xCML277	0.71	0.92	0.91
B73xCML322	0.72	0.84	0.83
B73xCML333	0.61	0.84	0.82
B73xCML52	0.68	0.88	0.89
B73xCML69	0.67	0.82	0.80
B73xHp301	0.67	0.83	0.85
B73xI14H	0.61	0.84	0.86
B73xKi11	0.69	0.90	0.91
B73xKi3	0.67	0.84	0.85
B73xKy21	0.66	0.75	0.81
B73xM162W	0.69	0.82	0.83
B73xM37W	0.65	0.83	0.83
B73xMo17 (IBM)	0.67	0.84	0.84
B73xMo18W	0.81	0.88	0.90
B73xMS71	0.63	0.70	0.76
B73xNC350	0.71	0.83	0.84
B73xNC358	0.58	0.75	0.79
B73xOh43	0.71	0.71	0.81
B73xOh7B	0.71	0.81	0.80
B73xP39	0.62	0.87	0.86
B73xTx303	0.80	0.85	0.88
B73xTzi8	0.81	0.87	0.89
Association Panel	0.79	0.99	0.99
Average within-cross heritability (excluding association panel)	0.68	0.83	0.84
Cross Means (averaged across RILs within a cross)			
B73xB97	1.82	74.06	75.91
B73xCML103	0.26	78.13	78.39
B73xCML228	1.52	79.92	81.88
B73xCML247	1.73	80.58	82.27
B73xCML277	1.46	80.40	81.92
B73xCML322	1.25	77.78	79.16

B73xCML333	2.19	78.20	80.40
B73xCML52	1.53	82.79	84.52
B73xCML69	1.91	79.26	81.18
B73xHp301	1.68	74.53	76.11
B73xI14H	1.66	71.81	73.22
B73xKi11	2.22	79.47	82.12
B73xKi3	1.81	77.09	79.47
B73xKy21	1.92	76.28	78.29
B73xM162W	1.28	78.75	80.27
B73xM37W	1.65	77.54	79.20
B73xMo17 (IBM)	2.09	72.12	74.29
B73xMo18W	2.70	80.03	82.77
B73xMS71	1.06	71.78	72.36
B73xNC350	1.52	77.76	79.61
B73xNC358	1.53	75.57	77.01
B73xOh43	1.77	73.79	75.00
B73xOh7B	2.05	76.30	78.76
B73xP39	1.75	70.20	71.67
B73xTx303	2.60	78.11	80.77
B73xTzi8	2.61	78.47	81.39
Association Panel	1.88	78.41	80.06

Minimum and maximum RIL BLUPs across all crosses and association panel:

All Geno Max	6.65	91.20	94.57
All Geno Min	-1.53	66.05	66.83
All Geno Range	8.19	25.15	27.73

Minimum and maximum RIL BLUPs for each cross

B73xB97	(0.1, 3.9)	(70.7, 79.3)	(71.9, 81.2)
B73xCML103	(-1.5, 2)	(74.5, 83.5)	(75.1, 82.6)
B73xCML228	(-0.2, 3.9)	(74.1, 89.2)	(74.5, 91.2)
B73xCML247	(-1.0, 4.9)	(73.1, 89.4)	(75.6, 90.8)
B73xCML277	(-1.0, 4.1)	(75.0, 88.9)	(76.5, 90.2)
B73xCML322	(-1.1, 4.4)	(73.5, 84.1)	(74.3, 86.1)
B73xCML333	(0.6, 4)	(73.6, 84.2)	(76.4, 85.7)
B73xCML52	(-0.2, 3.7)	(78.3, 91.2)	(78.9, 94.6)
B73xCML69	(0.2, 4.3)	(75.2, 86.1)	(76.2, 85.4)
B73xHp301	(-0.3, 3.5)	(70.5, 80.0)	(71.3, 81.2)
B73xI14H	(0.1, 3.4)	(67.4, 78.9)	(68.0, 80.1)
B73xKi11	(0.3, 5.2)	(74.3, 87.6)	(75.8, 92.5)
B73xKi3	(0.1, 4.2)	(73.0, 82.2)	(74.9, 85.7)
B73xKy21	(-0.1, 3.7)	(72.5, 79.7)	(73.3, 83.0)
B73xM162W	(-0.5, 4.3)	(74.7, 85.6)	(75.4, 86.8)
B73xM37W	(-0.5, 4.4)	(73.2, 84.0)	(74.8, 85.1)
B73xMo17 (IBM)	(0.6, 4.7)	(68.0, 78.3)	(68.7, 80.0)
B73xMo18W	(0.3, 6.7)	(71.9, 86.0)	(73.0, 90.5)
B73xMS71	(-0.2, 3)	(69.2, 75.1)	(68.2, 76.8)
B73xNC350	(-1.0, 3.5)	(71.2, 82.6)	(72.1, 85.3)
B73xNC358	(0.2, 3.3)	(72.5, 80.6)	(72.9, 84.1)

B73xOh43	(-0.3, 4.3)	(70.9, 77.8)	(70.9, 80.8)
B73xOh7B	(0.1, 4.5)	(71.9, 80.6)	(73.6, 84.4)
B73xP39	(-0.7, 3.5)	(66.1, 78.5)	(66.8, 80.3)
B73xTx303	(0.1, 5.5)	(73.8, 85.6)	(75.7, 89.3)
B73xTzi8	(0.1, 5.8)	(72.6, 86.8)	(73.4, 91.1)
Association Panel	(-0.8, 7.0)	(58.4, 121.2)	(58.7, 125.8)

Founder line BLUPs

B73	1.05	73.96	75.01
B97	2.62	74.51	77.08
CML103	-0.76	82.64	81.87
CML228	2.10	92.48	94.08
CML247	1.48	93.46	95.00
CML277	2.42	91.10	93.29
CML322	1.81	82.97	85.01
CML333	2.29	84.70	86.96
CML52	1.99	93.87	98.70
CML69	1.31	88.29	89.34
Hp301	1.65	74.93	76.48
II14H	1.65	69.57	70.89
Ki11	2.71	88.38	91.04
Ki3	1.64	85.20	86.92
Ky21	2.88	80.92	83.87
M162W	-0.59	85.56	85.18
M37W	2.46	79.51	82.32
Mo18W	3.95	86.96	90.49
MS71	0.94	71.63	72.18
NC350	0.40	82.09	82.35
NC358	0.70	77.58	78.10
Oh43	1.24	70.73	71.47
Oh7B	1.90	83.02	85.15
P39	2.03	65.15	66.82
Tx303	2.86	82.64	85.68
Tzi8	2.78	88.72	91.85
Mo17	3.00	72.76	75.49
Founders' Max	3.95	93.87	98.70
Founders' Min	-0.76	65.15	66.82
Founders' Range	4.71	28.72	31.87
Founders' Var	1.14	61.53	68.76

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Supporting Table 4. Summary Joint Linkage Stepwise Model and GxE ASREML Analysis for Days to Anthesis

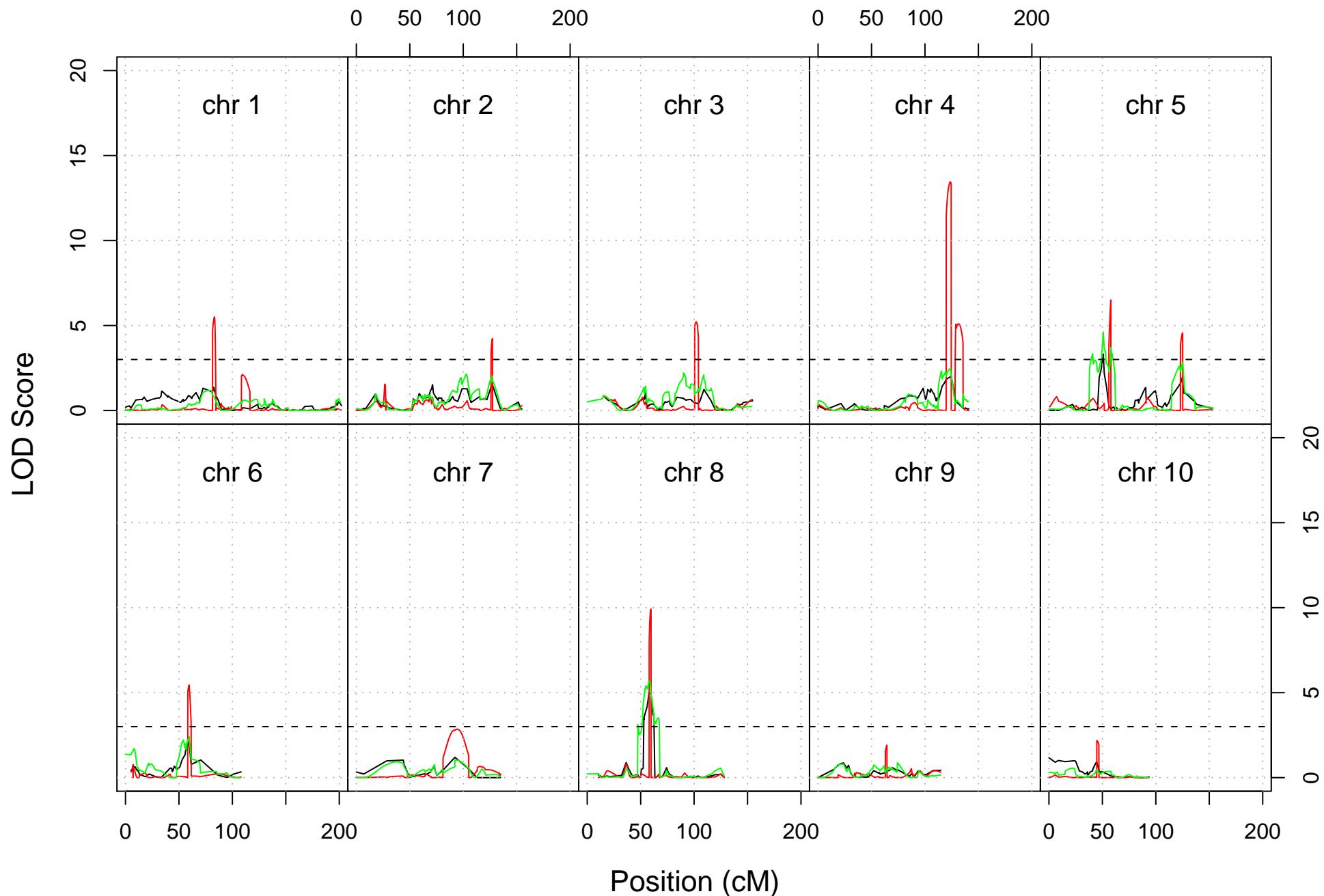
GLM							Map				ASREML							
Trait	Term	SS	df	MS	F	p	Rsq	marker_nu	marker	chr	pos	G_Gamma	G_Compoi	G_Comp/S	GxE_Gamr	GxE_Comp	GxE_Comp	G/GxE
asi	22	35.63	25	1.4252	3.377	2.9E-08	0.007798	m23	PZA02393	1	33.1	0.004682	0.002518	2.77	0.002014	0.001083	2.14	2.325041
asi	71	51.97	25	2.0788	4.925	1.16E-14	0.011375	m72	PZA02763	1	87.9	0.009572	0.005147	2.98	0.003482	0.001872	2.8	2.749129
asi	87	46.17	25	1.8468	4.375	2.51E-12	0.010105	m88	PZA00619	1	108.4	0.007906	0.004251	2.82	0.005246	0.002821	3.71	1.507172
asi	135	87.17	25	3.4868	8.262	1.32E-29	0.019079	m136	PZB00063	1	162.2	0.011424	0.006143	3.19	0.00162	0.000871	1.81	7.0508
asi	200	34.95	25	1.398	3.313	5.1E-08	0.007649	m201	PZA01753	2	40.6	0.005109	0.002747	2.76	0.002962	0.001593	2.84	1.724855
asi	247	58.16	25	2.3264	5.512	3.24E-17	0.012729	m248	PZA01321	2	82.8	0.009812	0.005276	3.06	0.00326	0.001753	2.98	3.010006
asi	282	35.42	25	1.4168	3.357	3.4E-08	0.007752	m283	PZA00804	2	123.9	0.006286	0.00338	2.86	0.002091	0.001125	2.21	3.005967
asi	297	40.36	25	1.6144	3.825	4.7E-10	0.008834	m298	PZA02170	2	151.6	0.008098	0.004354	3.05				
asi	311	74.71	25	2.9884	7.081	3.22E-24	0.016352	m312	PZB01944	3	28.9	0.013435	0.007224	3.1	0.005065	0.002724	3.85	2.652413
asi	324	44.25	25	1.77	4.193	1.45E-11	0.009685	m325	zb21.1	3	55.5	0.007303	0.003927	3	0.000948	0.00051	1.11	7.701247
asi	391	31.6	25	1.264	2.994	8.45E-07	0.006916	m392	PZA01228	3	100.1	0.005174	0.002782	2.96				
asi	461	42.82	25	1.7128	4.058	5.23E-11	0.009372	m462	PZA01759	4	55.4	0.008683	0.004669	3.13				
asi	496	34.63	25	1.3852	3.282	6.8E-08	0.007579	m497	PZA03275	4	85.2	0.005658	0.003042	2.86	0.001624	0.000873	1.83	3.484136
asi	613	48.22	25	1.9288	4.57	3.81E-13	0.010554	m614	PZA00273	5	68.7	0.006685	0.003595	2.87	0.003031	0.00163	2.86	2.205202
asi	648	43.12	25	1.7248	4.087	4E-11	0.009438	m649	PZA03172	5	98.2	0.007586	0.004079	3.04				
asi	672	35.01	25	1.4004	3.318	4.9E-08	0.007663	m673	PZA02015	5	124.9	0.005606	0.003015	2.88				
asi	683	30.79	25	1.2316	2.918	1.63E-06	0.006739	m684	PZA00158	6	5.1	0.005565	0.002993	2.91	0.001082	0.000582	1.31	5.143088
asi	744	41.47	25	1.6588	3.93	1.75E-10	0.009076	m745	PZA00223	6	75.8	0.01105	0.005942	3.03	0.001417	0.000762	1.36	7.800165
asi	751	31.26	25	1.2504	2.963	1.11E-06	0.006842	m752	PZA00889	6	94.1	0.006216	0.003343	2.77	0.001301	0.000699	1.36	4.77886
asi	787	37.9	25	1.516	3.592	4E-09	0.008295	m788	PZA02291	7	49.5	0.006237	0.003354	2.93	0.001193	0.000641	1.37	5.228876
asi	809	32.65	25	1.306	3.094	3.55E-07	0.007146	m810	PZA03728	7	75.3	0.006287	0.003381	2.77	0.001889	0.001016	1.85	3.328431
asi	818	35.57	25	1.4228	3.371	3E-08	0.007785	m819	PHM112.8	7	92.2	0.009467	0.005091	2.95	0.002937	0.001579	2.51	3.223118
asi	842	30.6	25	1.224	2.9	1.89E-06	0.006697	m843	PZA00368	8	17.1	0.005179	0.002785	2.93				
asi	892	55.72	25	2.2288	5.281	3.34E-16	0.012195	m893	PZA03637	8	64.1	0.010488	0.00564	3.06	0.003511	0.001888	3.14	2.987367
asi	924	27.5	25	1.1	2.606	2.2E-05	0.006019	m925	PZB00811	8	89.5	0.003843	0.002066	2.58	0.000462	0.000249	0.56	8.309895
asi	935	30.51	25	1.2204	2.892	2.04E-06	0.006678	m936	PHM14046	8	117.5	0.004777	0.002569	2.87				
asi	969	47.97	25	1.9188	4.546	4.77E-13	0.010499	m970	PZB00959	9	44.5	0.009939	0.005344	2.99	0.004136	0.002224	3.39	2.402681
asi	1002	44.4	25	1.776	4.208	1.26E-11	0.009718	m1003	PZA02397	9	64.5	0.008714	0.004686	3.02	0.001547	0.000832	1.58	5.632369
asi	1070	45.46	25	1.8184	4.308	4.79E-12	0.00995	m1071	PZA01919	10	44.7	0.007376	0.003966	3.01	0.002165	0.001164	2.28	3.406576
asi	Population	28.86	24	1.2025	2.849	4.41E-06	0.006317					Env	0.615168	0.330801	1.85			
asi	Model	2902.24	749	3.87482	9.18	0	0.635209					PopxEnv	0.162253	0.08725	7.12			
asi	ERROR	1666.71	3949	0.422059	.	.	.					Pop	0.178146	0.095797	2.83			

Supporting Table 5. Associating polymorphisms at Vgt1 and Rap2.7 for the maize 282 association panel.

LINE	Vgt1 MITE Rap2.7 GEL	Rap2.7 Seq 568-575	LINE	Vgt1 MITE Rap2.7 GEL	Rap2.7 Seq 568-575	LINE	Vgt1 MITE Rap2.7 GEL	Rap2.7 Seq 568-575	LINE	Vgt1 MITE Rap2.7 GEL	Rap2.7 Seq 568-575	LINE	Vgt1 MITE Rap2.7 GEL	Rap2.7 Seq 568-575	
4226	0	2	NS	C13A	0	2	NS	E2558W	0	0	NS	Mp339	0	0	NS
4722	2	0	NS	C164	0	0	NS	EP1	2	2	NS	MS1334	2	0	NS
33-16	0	2	NS	C166	0	2	NS	F2834T	0	2	NS	MS153	0	0	NS
38-11	2	0	NS	C190C	0	0	NS	F44	0	2	NS	MS71	2	0	0
A188	2	0	NS	C191B	0	2	NS	F6	0	0	NS	Mt42	0	0	NS
A214N	0	0	NS	CM105	2	0	NS	F7	2	2	NS	N192	0	0	NS
A239	0	0	NS	CM174	2	0	NS	GA209	0	2	NS	N28Ht	0	0	NS
A272	2	2	NS	CM37	2	0	NS	GT112	0	2	NS	N6	2	2	NS
A441-5	0	0	NS	CM7	2	0	NS	H105W	2	0	NS	N7A	0	0	NS
A554	0	0	NS	CML10	0	2	NS	H49	0	0	NS	NC222	0	0	NS
A556	2	0	NS	CML103	0	0	0	H84	0	2	NS	NC230	0	2	NS
A6	0	2	NS	CML108	2	2	NS	H91	0	0	NS	NC232	0	0	NS
A619	2	0	NS	CML11	0	2	NS	H95	0	0	NS	NC236	0	2	NS
A632	2	0	NS	CML14	2	2	NS	H99	2	0	NS	NC238	0	NS	SA24
A634	0	0	NS	CML154Q	0	0	NS	Hi27	0	2	NS	NC250	0	0	NS
A635	0	0	NS	CML157Q	2	2	NS	HP301	0	2	0	NC258	0	2	NS
A641	2	0	NS	CML158Q	0	2	NS	Hy	0	0	NS	NC260	0	0	NS
A654	2	0	NS	CML218	0	2	NS	I137TN	0	0	NS	NC262	2	2	NS
A659	0	0	NS	CML220	0	0	NS	I205	0	0	NS	NC264	2	0	NS
A661	0	0	NS	CML228	0	2	0	I29	2	2	NS	NC290A	2	2	NS
A679	2	0	NS	CML238	0	0	NS	IA2132	2	NS	NS	NC294	0	0	NS
A680	0	0	NS	CML247	0	0	0	Ia5125	2	0	NS	NC296	0	0	NS
A682	2	0	NS	CML254	0	0	NS	ID528	2	0	NS	NC296A	0	0	NS
Ab28A	2	0	NS	CML258	0	0	NS	ID569	0	2	NS	NC298	0	0	NS
B10	0	0	NS	CML261	0	0	NS	ID591	0	2	NS	NC300	0	0	NS
B103	0	0	NS	CML264	0	2	NS	II101	2	0	NS	NC302	0	0	NS
B104	0	0	NS	CML277	0	0	0	II14H	2	0	0	NC304	0	2	NS
B105	0	0	NS	CML281	2	0	NS	II677a	2	0	NS	NC306	0	0	NS
B109	0	0	NS	CML287	0	0	NS	K148	2	2	NS	NC310	0	0	NS
B115	0	2	NS	CML311	2	2	NS	K4	0	0	NS	NC314	0	0	NS
B14A	0	0	NS	CML314	0	0	NS	K55	0	2	NS	NC318	0	0	NS
B164	2	0	NS	CML321	0	0	NS	K64	0	0	NS	NC320	0	2	NS
B2-good	0	0	NS	CML322	0	2	2	Ki11	0	0	0	NC324	0	0	NS
B37	0	2	NS	CML323	0	2	NS	Ki14	0	0	NS	NC326	0	0	NS
B46	0	0	NS	CML328	0	2	NS	Ki2021	0	0	NS	NC328	0	0	NS
B52	0	2	NS	CML331	0	2	NS	Ki21	2	2	NS	NC33	0	2	NS
B57	0	2	NS	CML332	0	2	NS	Ki3	0	0	2	NC336	0	0	NS
B64	0	0	NS	CML333	0	2	2	Ki43	0	0	NS	NC338	0	2	NS
B68	0	0	NS	CML341	0	0	NS	Ki44	0	2	NS	NC340	0	0	NS
B73	0	0	0	CML38	0	0	NS	Ky21	0	0	0	NC342	0	2	NS
B73Htrhm	0	0	NS	CML45	0	2	NS	KY226	0	0	NS	NC344	0	2	NS
B75	0	0	NS	CML5	0	0	NS	KY228	0	0	NS	NC346	0	0	NS
B76	0	2	NS	CML52	0	2	2	L317	2	0	NS	NC348	0	0	NS
B77	0	2	NS	CML61	0	2	NS	L578	0	2	NS	NC350	0	0	NS
B79	0	0	NS	CML69	0	2	2	M14	0	0	NS	NC352	0	0	NS
B84	0	0	NS	CML77	2	2	NS	M162W	0	0	0	NC354	0	0	NS
B97	0	0	2	CML91	0	2	NS	M37W	0	0	0	NC356	0	0	NS
C103	2	0	NS	CML92	0	0	NS	MEF156-55-2	2	0	NS	NC358	0	2	0
C123	2	0	NS	CMV3	2	0	NS	Mo17	2	2	0	NC360	2	2	NS
C49A	0	0	NS	CO106	2	0	NS	Mo18W	0	2	2	NC362	2	2	NS
CH701-30	0	0	NS	CO125	2	0	NS	MO1W	0	0	NS	NC364	2	2	NS
CH9	0	0	NS	CO255	2	0	NS	Mo24W	0	2	NS	NC366	0	2	NS
Cl.7	0	2	NS	D940Y	0	0	NS	Mo44	0	0	NS	NC368	0	0	NS
Cl187-2	2	0	NS	DE_2	0	0	NS	Mo45	0	2	NS	ND246	2	2	NS
Cl21E	0	2	NS	DE_3	0	0	NS	Mo46	0	0	NS	Oh40B	2	0	NS
Cl28A	0	0	NS	DE1	0	0	NS	Mo47	0	0	NS	Oh43	0	0	0
Cl31A	0	2	NS	DE811	0	0	NS	MoG	0	2	NS	Oh43E	2	0	NS

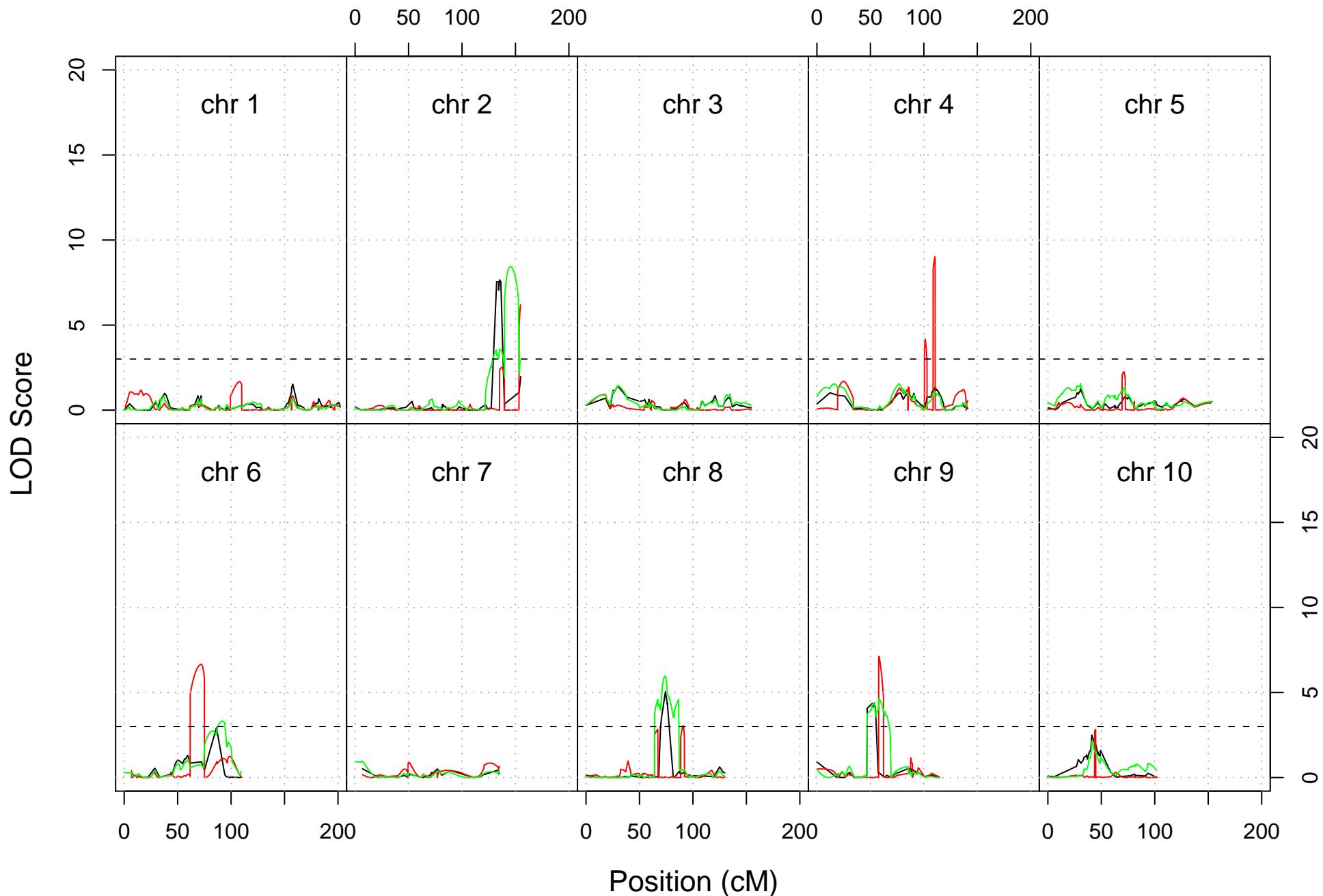
LOD Scores for B97 population, anthesis–silk interval

ICIM = red, CIM = green, Stepwise = black



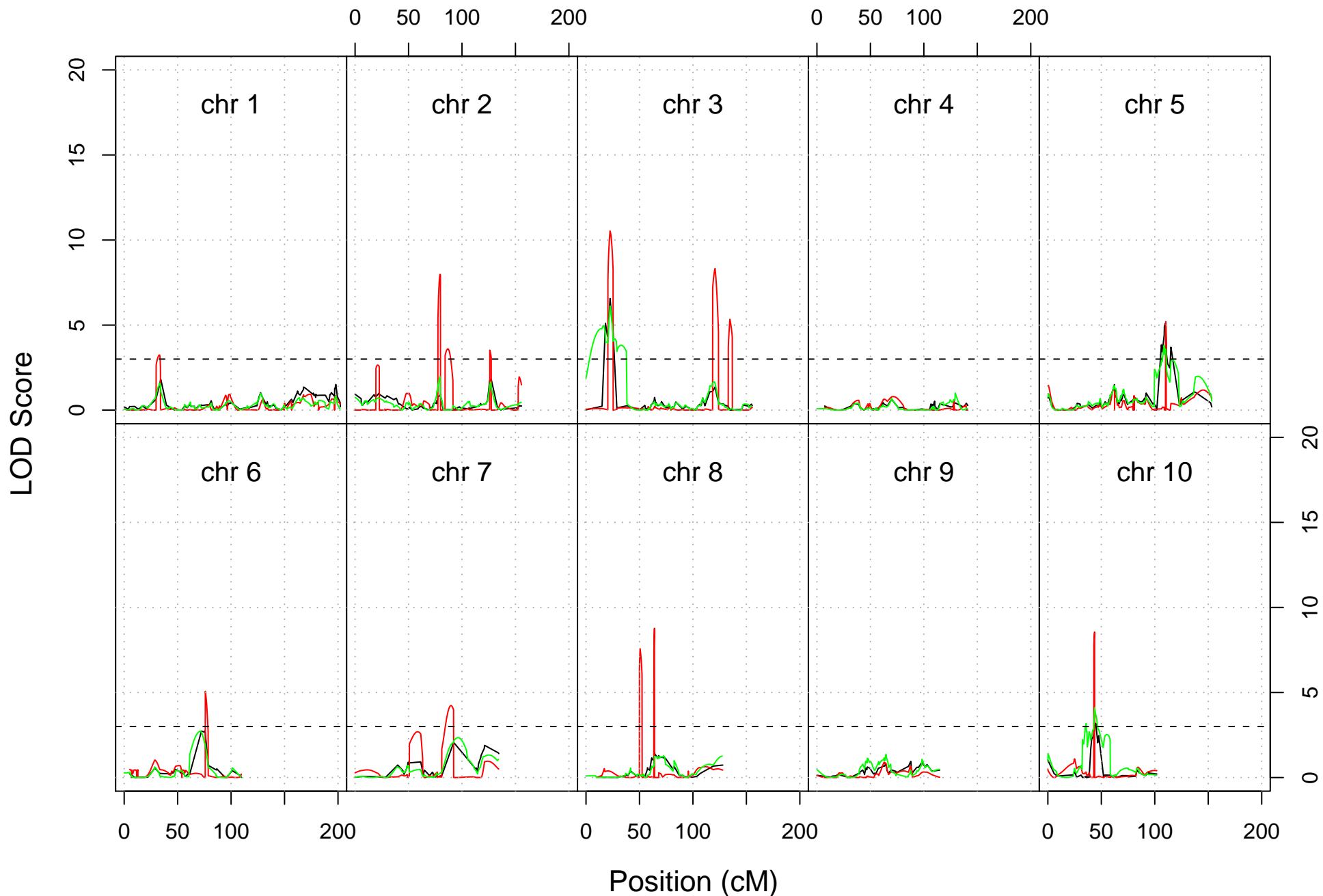
LOD Scores for CML103 population, anthesis–silk interval

ICIM = red, CIM = green, Stepwise = black



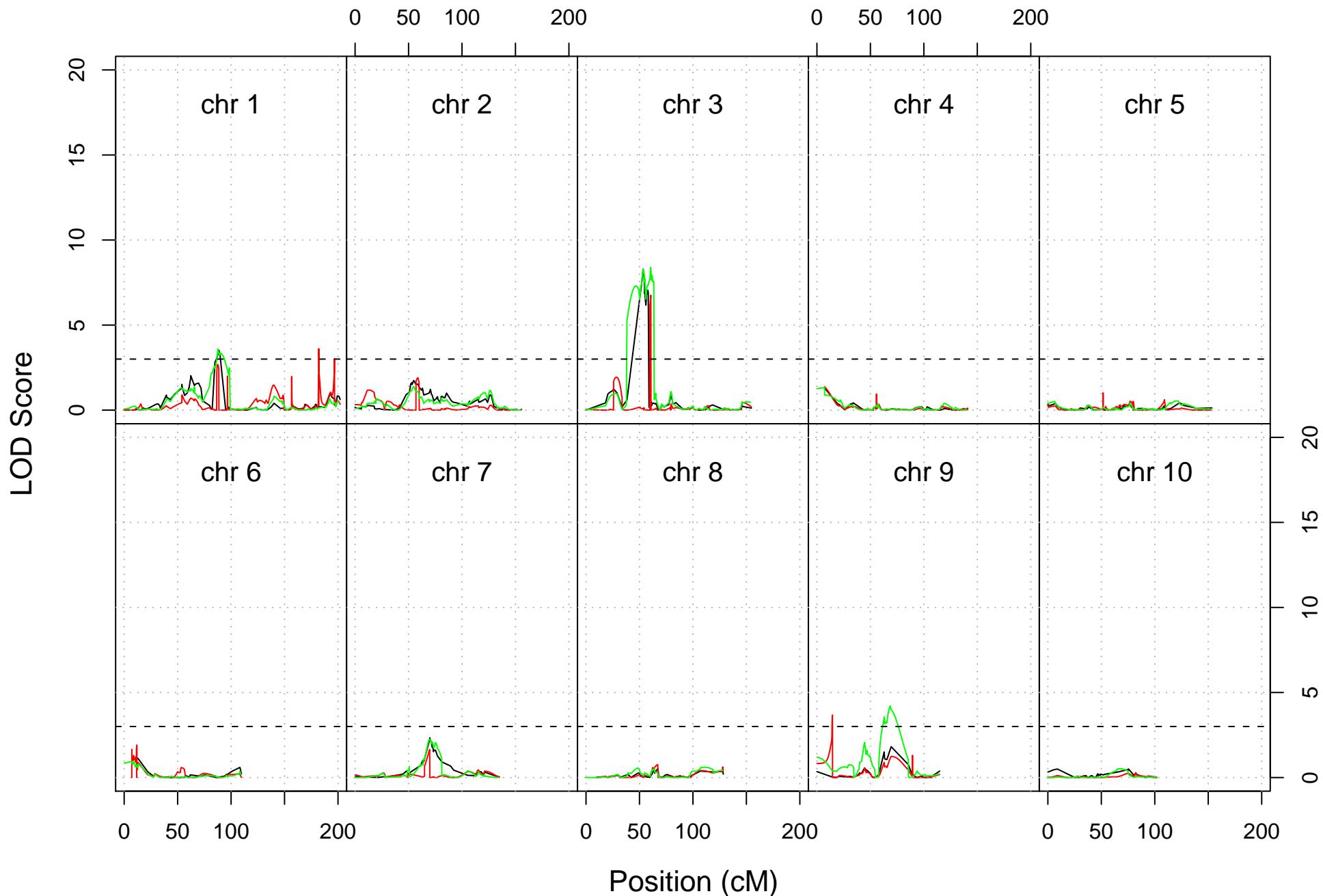
LOD Scores for CML228 population, anthesis–silk interval

ICIM = red, CIM = green, Stepwise = black



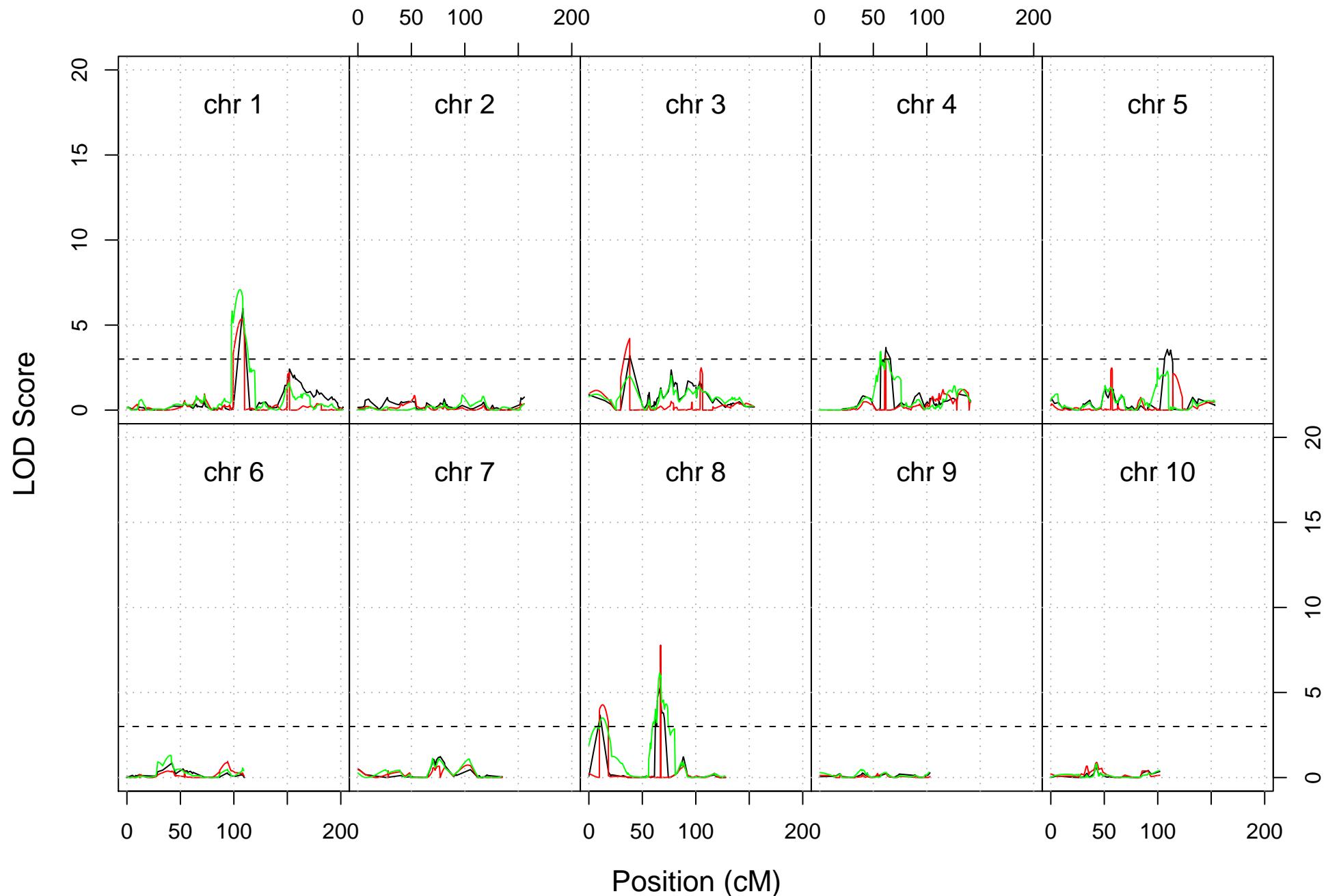
LOD Scores for CML247 population, anthesis–silk interval

ICIM = red, CIM = green, Stepwise = black



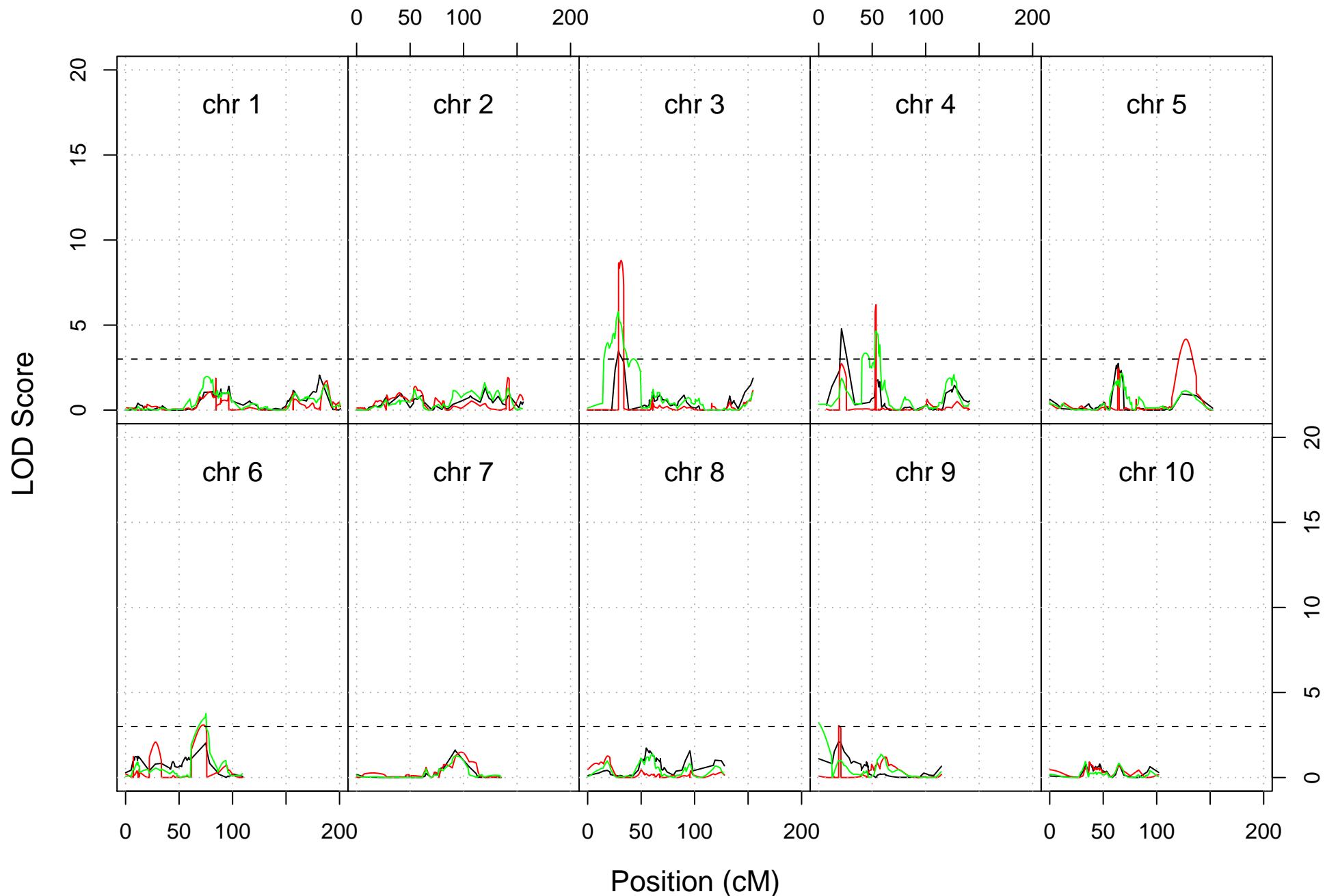
LOD Scores for CML277 population, anthesis–silk interval

ICIM = red, CIM = green, Stepwise = black



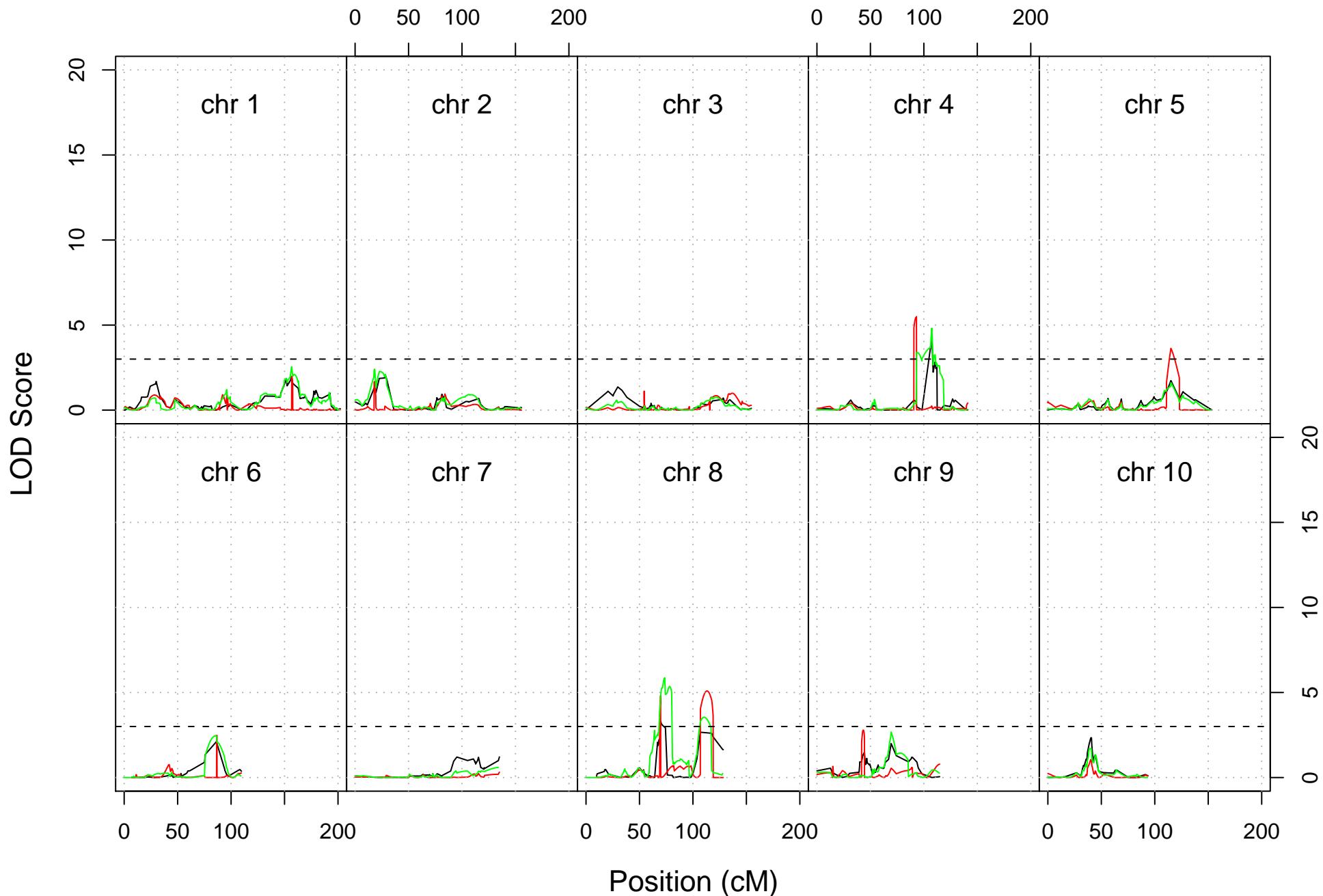
LOD Scores for CML322 population, anthesis–silk interval

ICIM = red, CIM = green, Stepwise = black



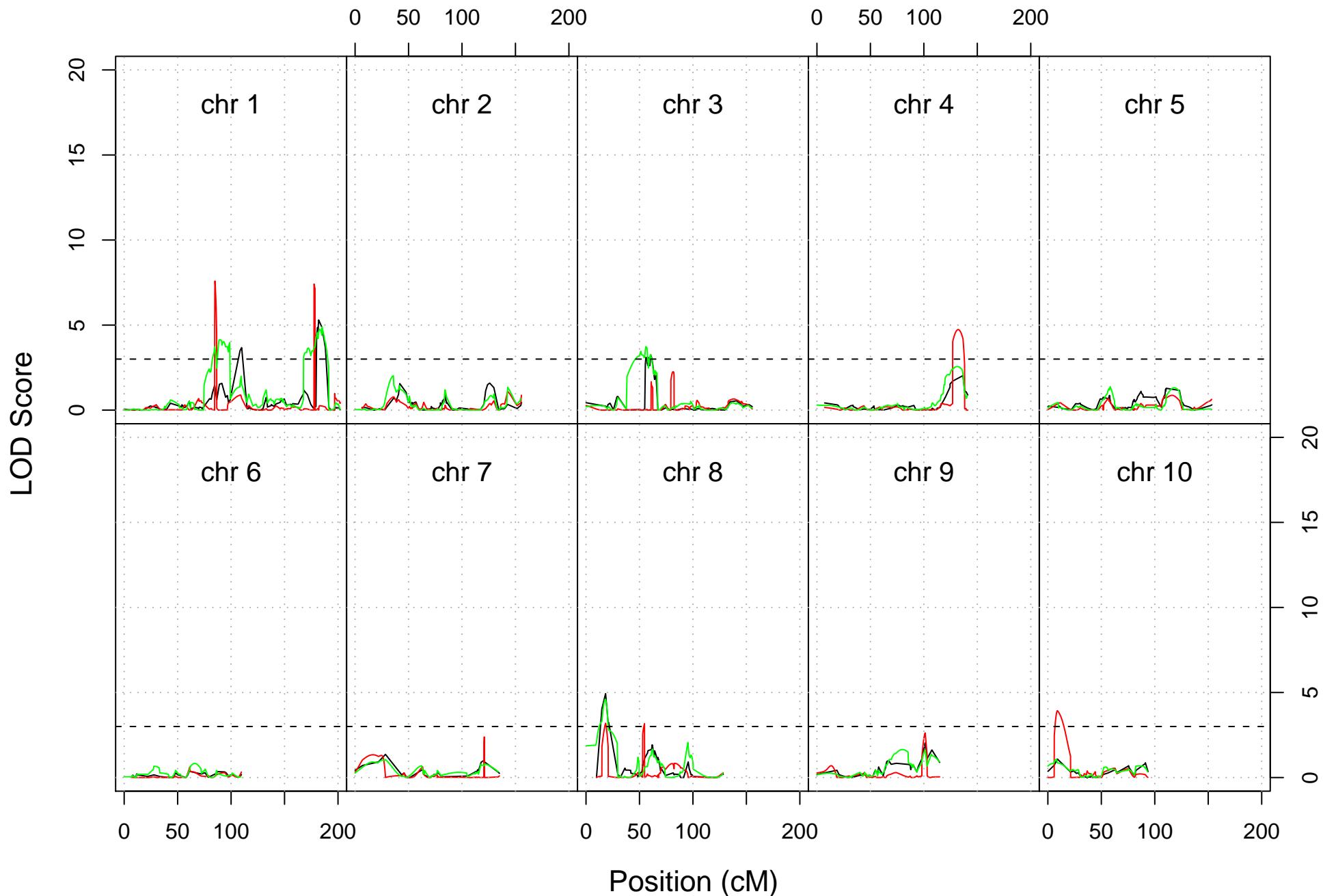
LOD Scores for CML333 population, anthesis–silk interval

ICIM = red, CIM = green, Stepwise = black



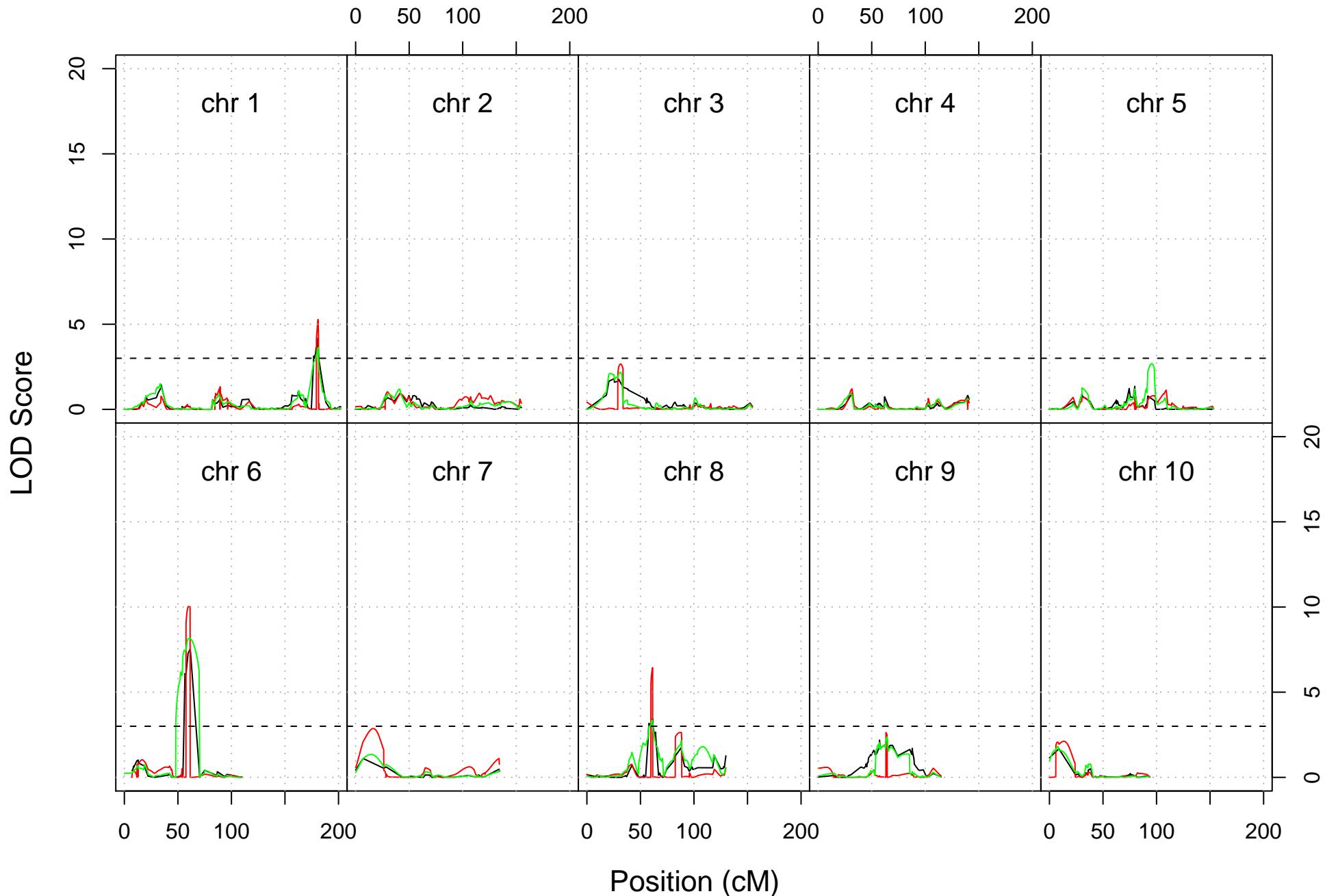
LOD Scores for CML52 population, anthesis–silk interval

ICIM = red, CIM = green, Stepwise = black



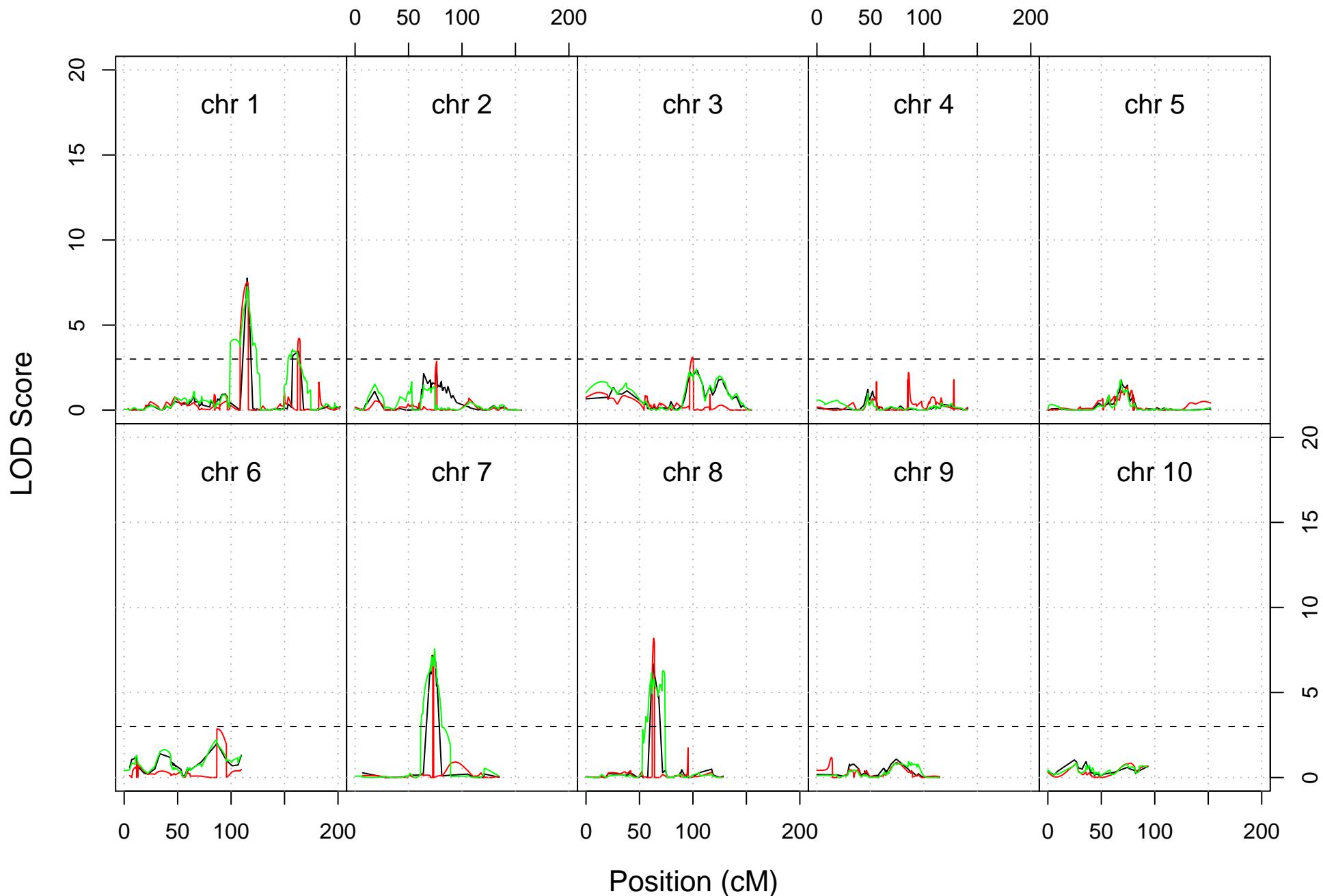
LOD Scores for CML69 population, anthesis–silk interval

ICIM = red, CIM = green, Stepwise = black



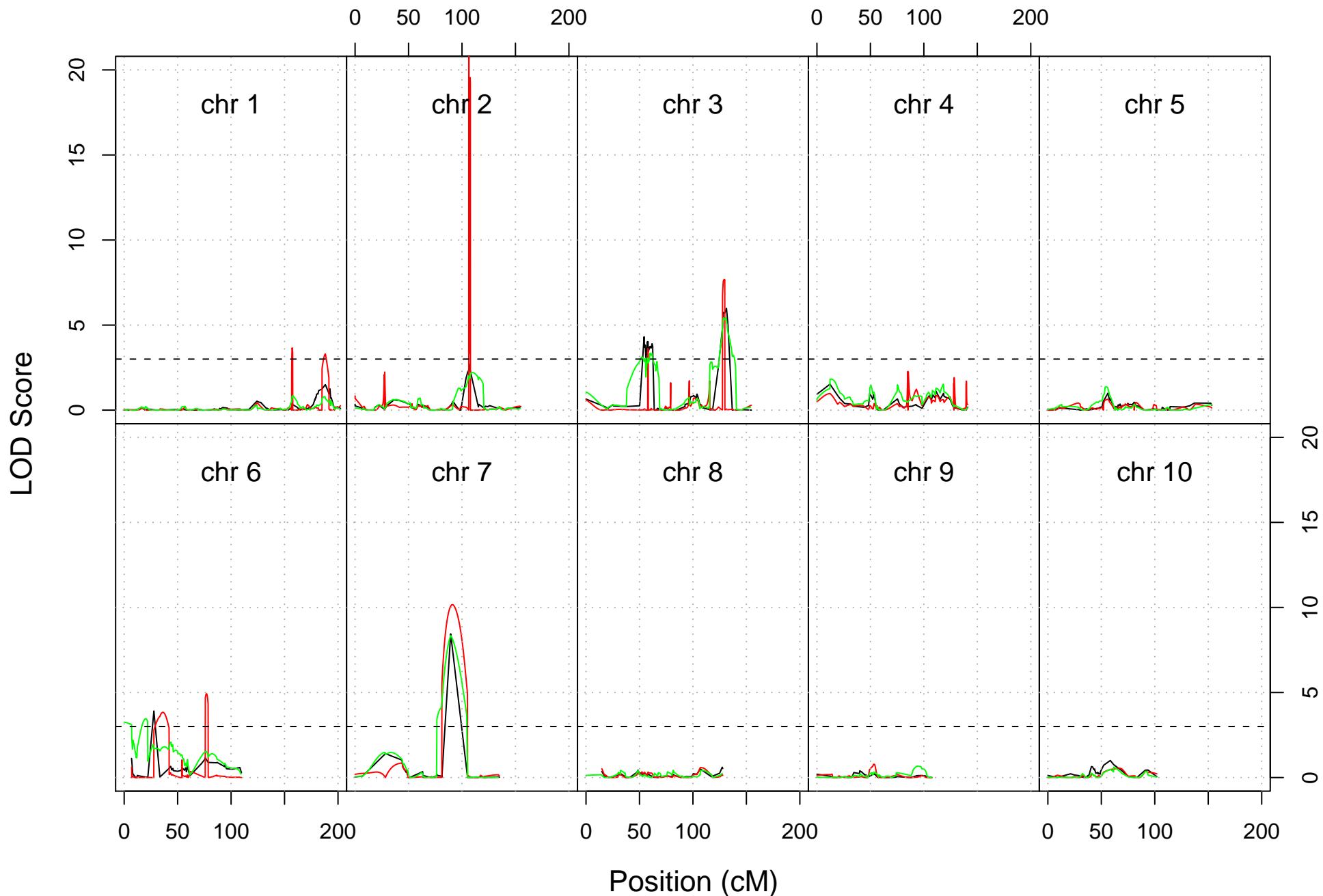
LOD Scores for HP301 population, anthesis–silk interval

ICIM = red, CIM = green, Stepwise = black



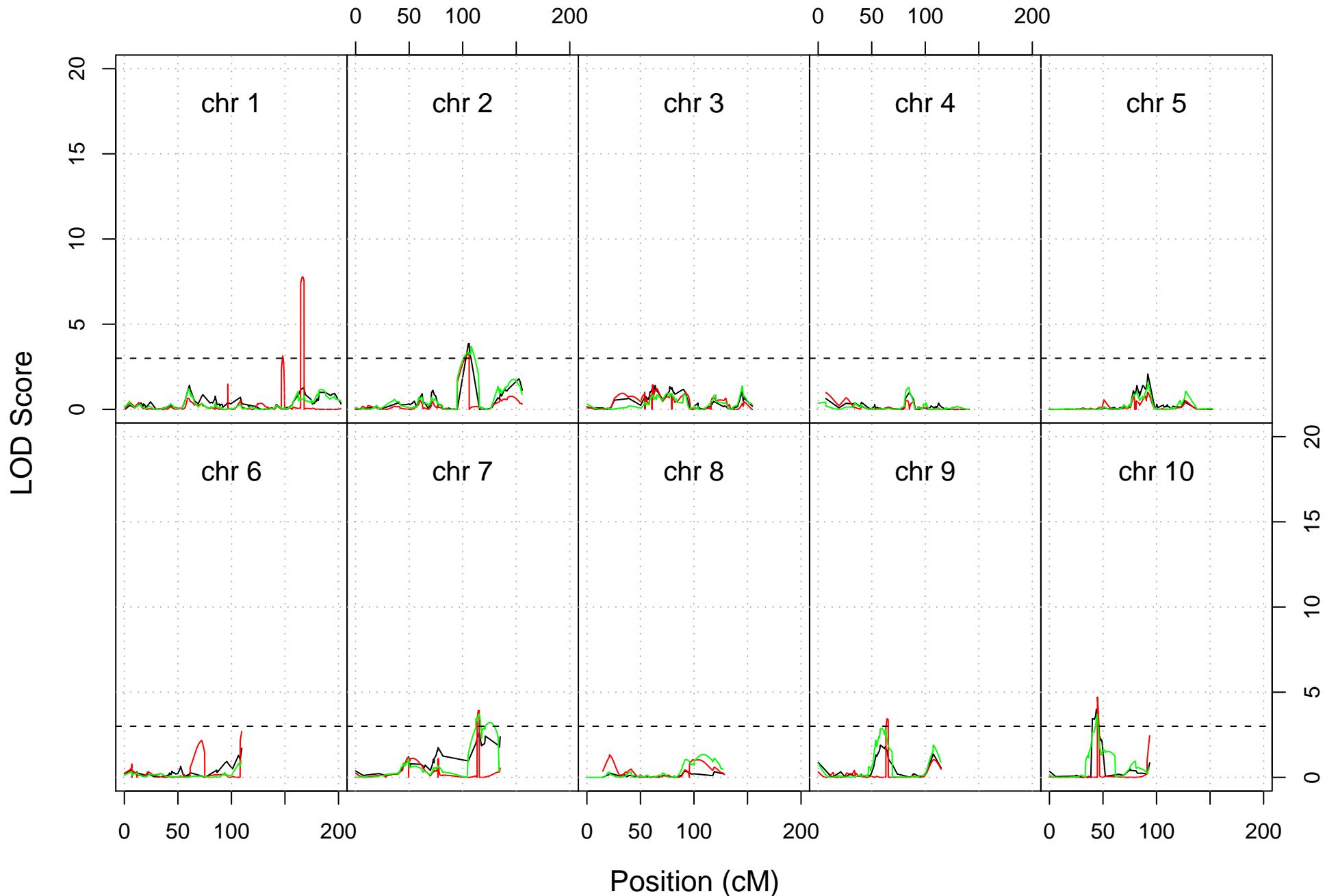
LOD Scores for IL14H population, anthesis–silk interval

ICIM = red, CIM = green, Stepwise = black



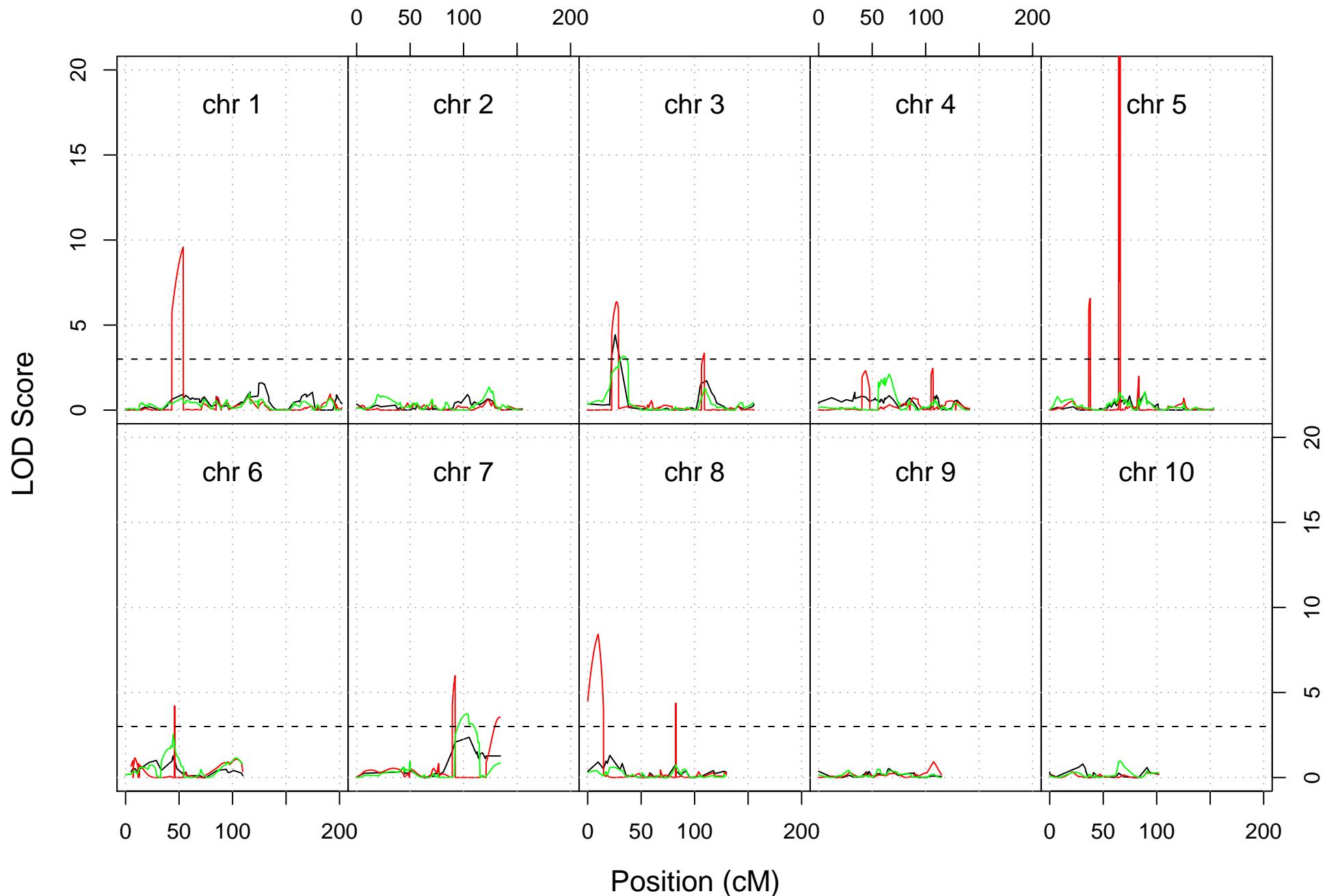
LOD Scores for KI11 population, anthesis–silk interval

ICIM = red, CIM = green, Stepwise = black



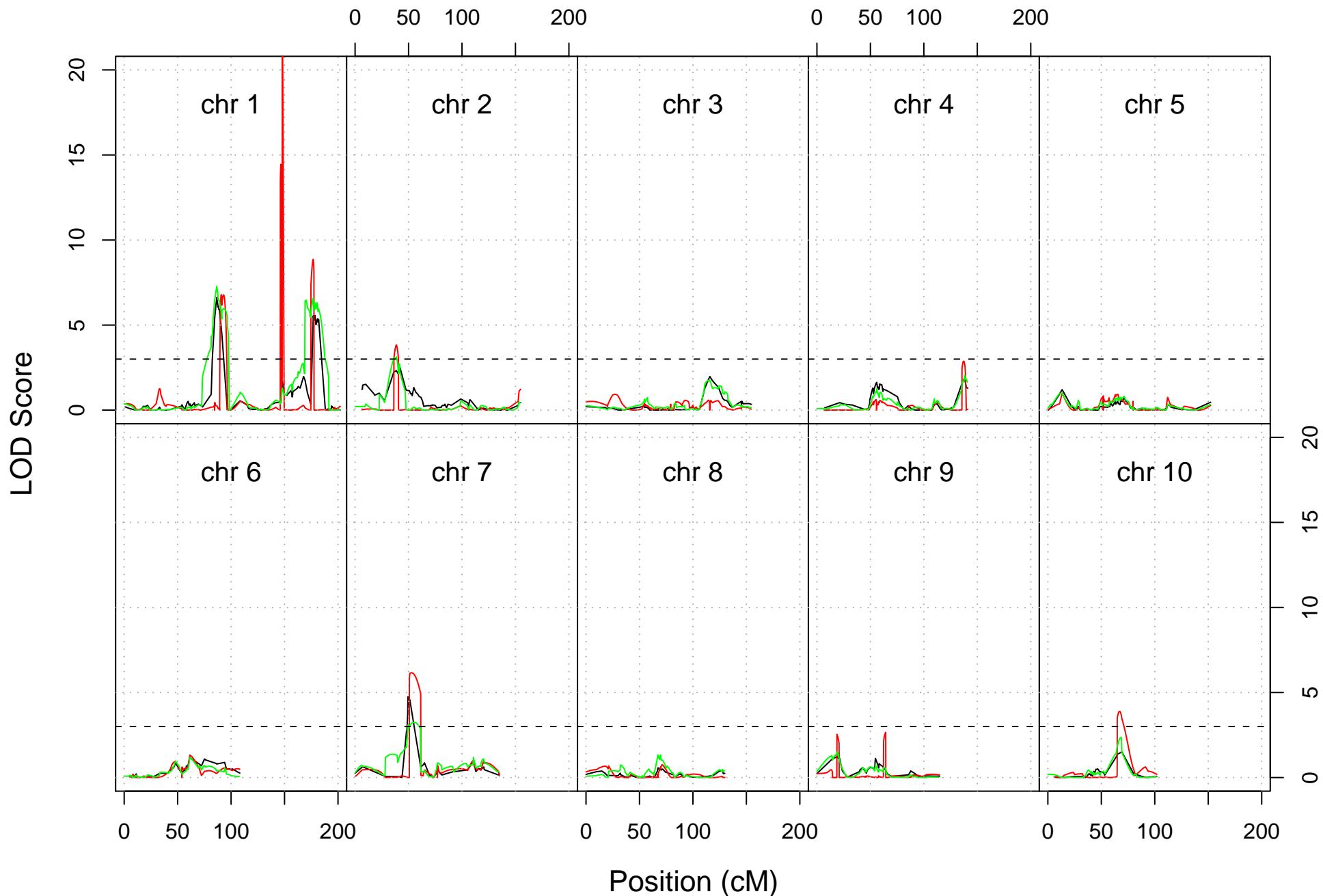
LOD Scores for KI3 population, anthesis–silk interval

ICIM = red, CIM = green, Stepwise = black



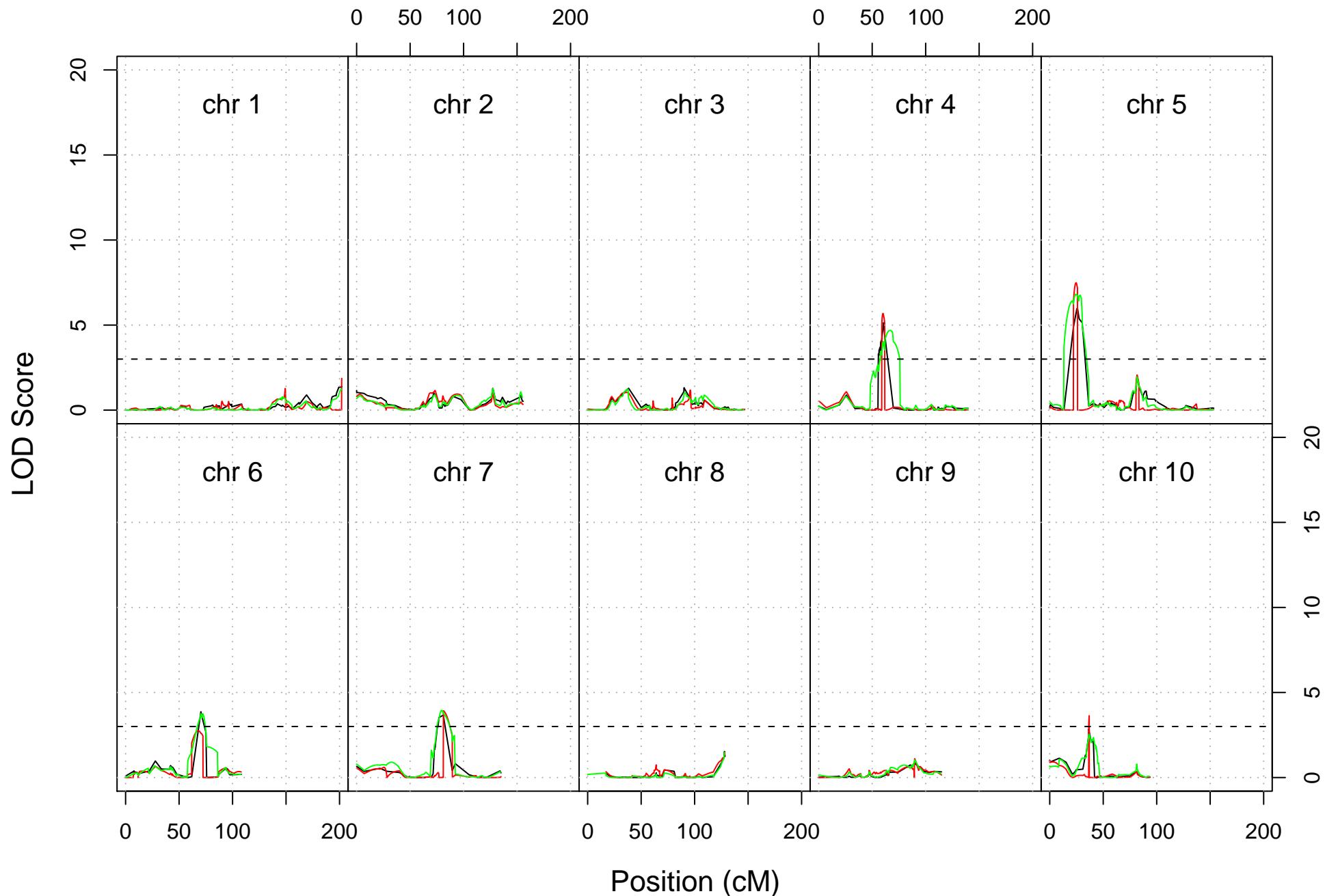
LOD Scores for KY21 population, anthesis–silk interval

ICIM = red, CIM = green, Stepwise = black



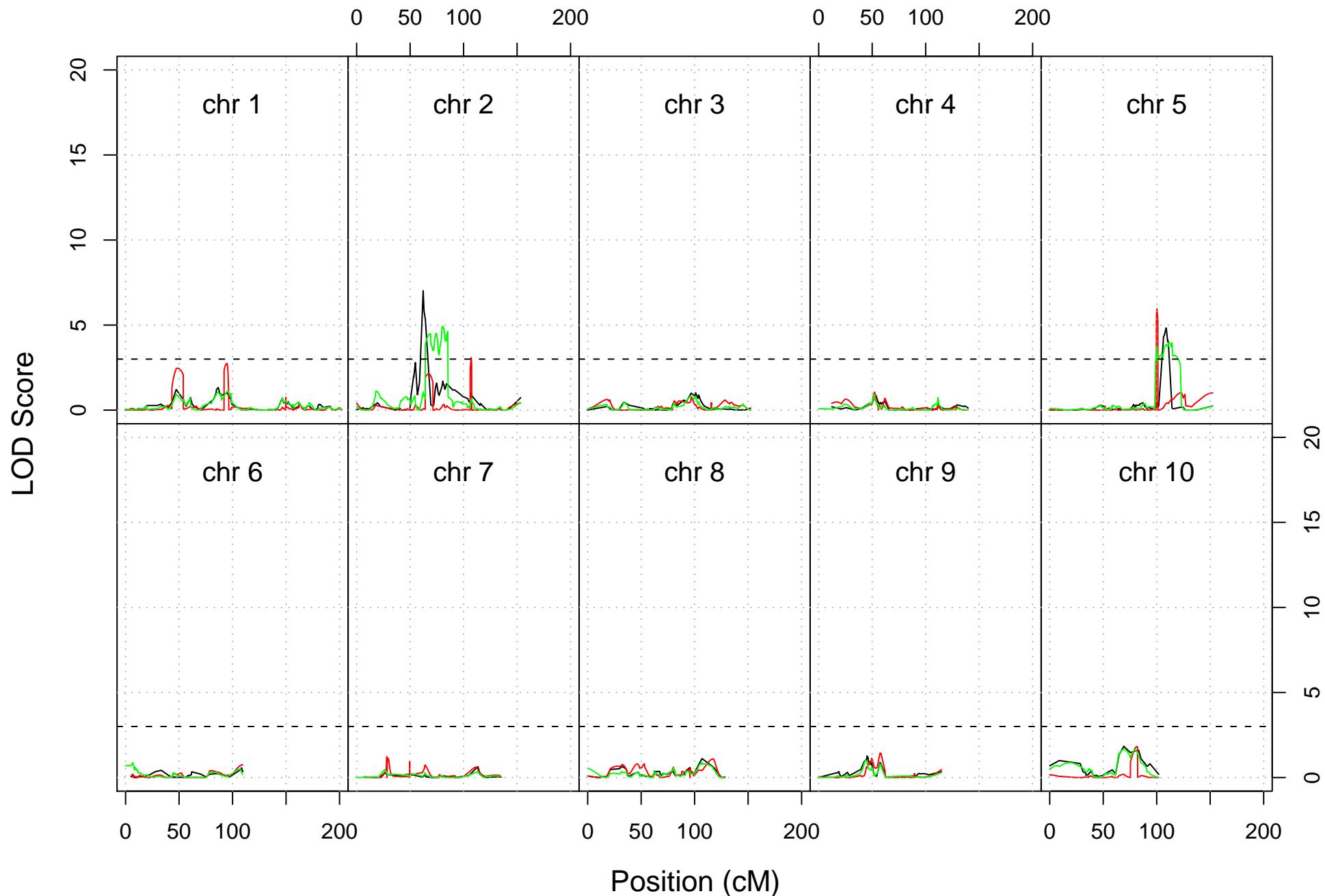
LOD Scores for M162W population, anthesis–silk interval

ICIM = red, CIM = green, Stepwise = black



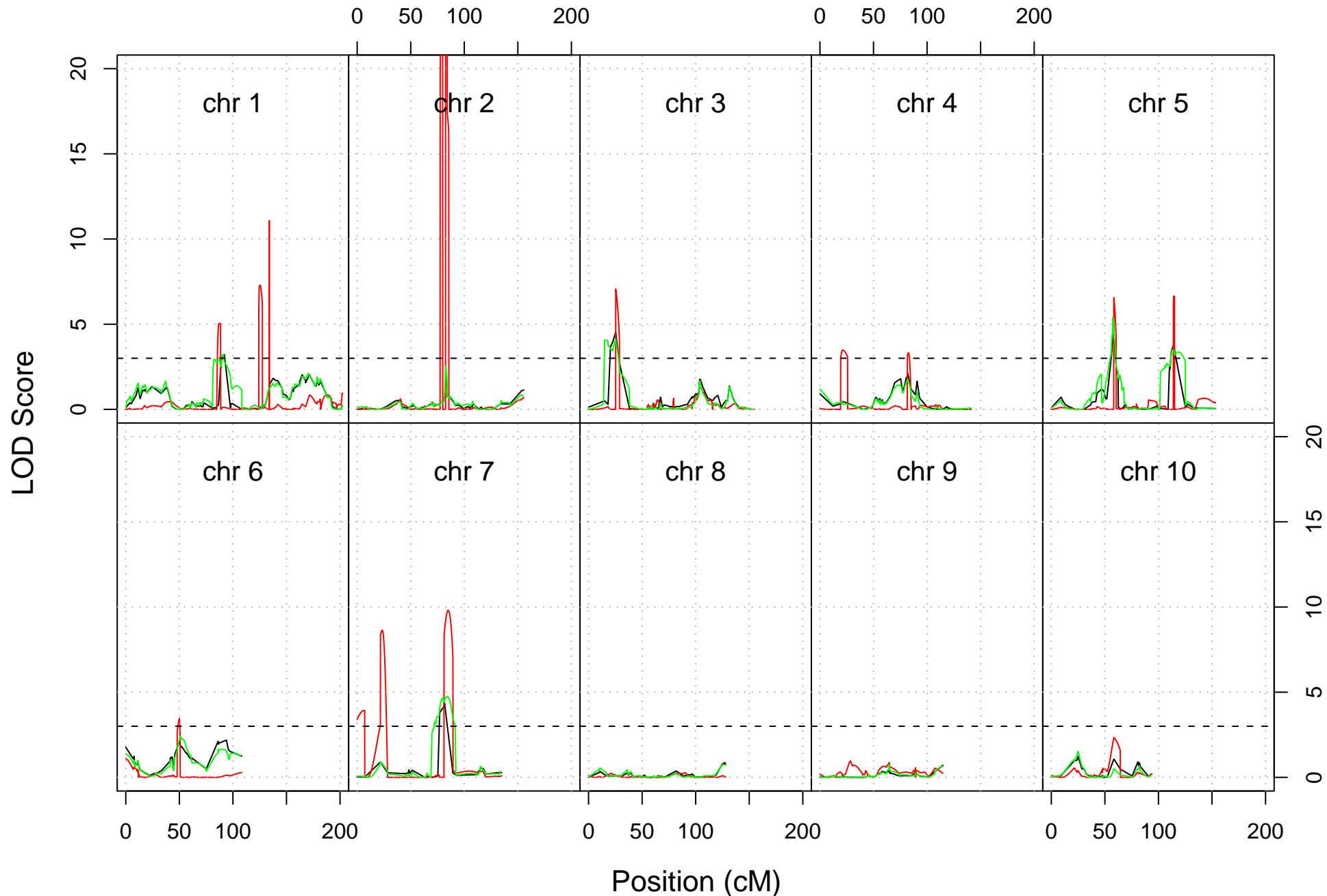
LOD Scores for M37W population, anthesis–silk interval

ICIM = red, CIM = green, Stepwise = black



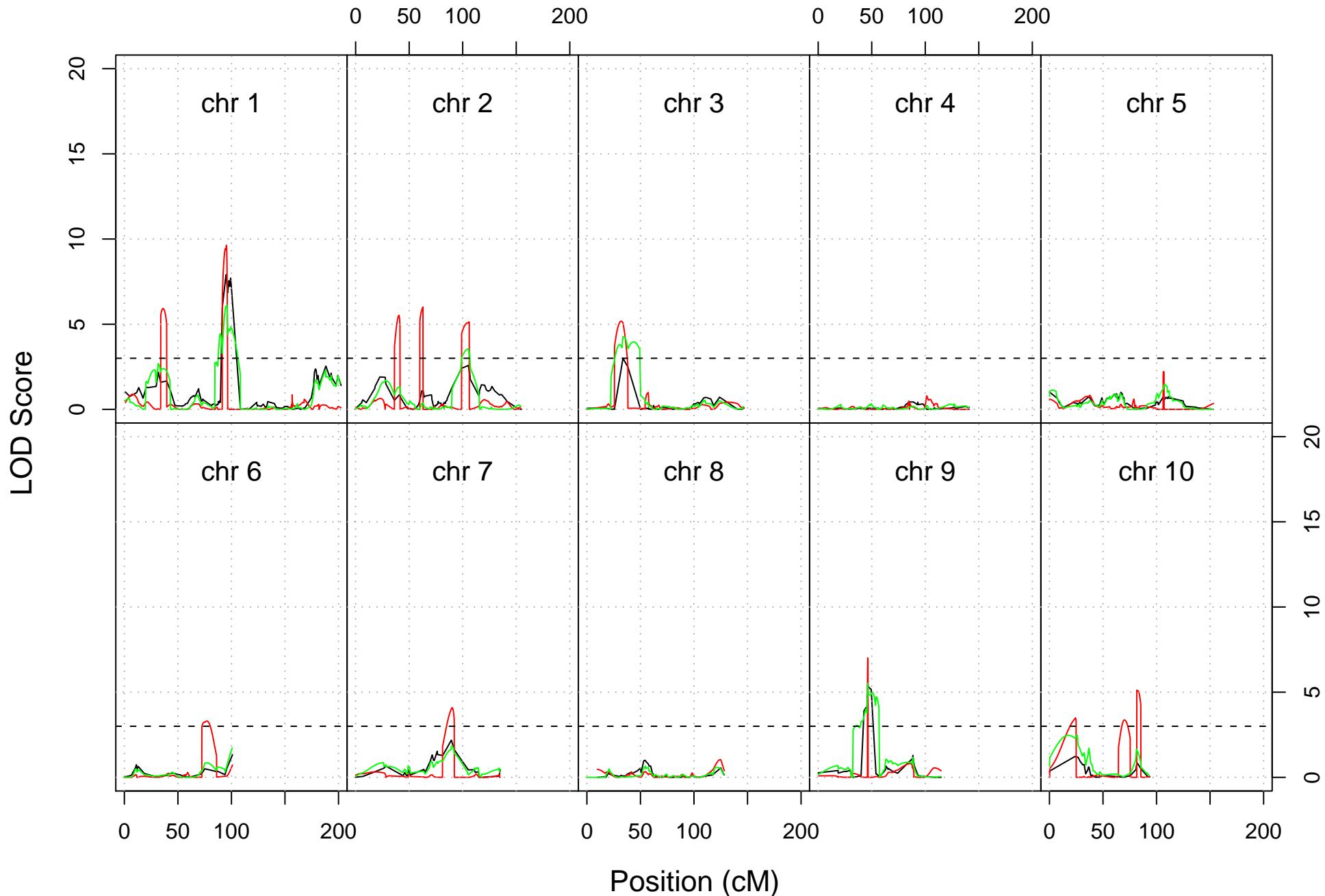
LOD Scores for MO18W population, anthesis–silk interval

ICIM = red, CIM = green, Stepwise = black



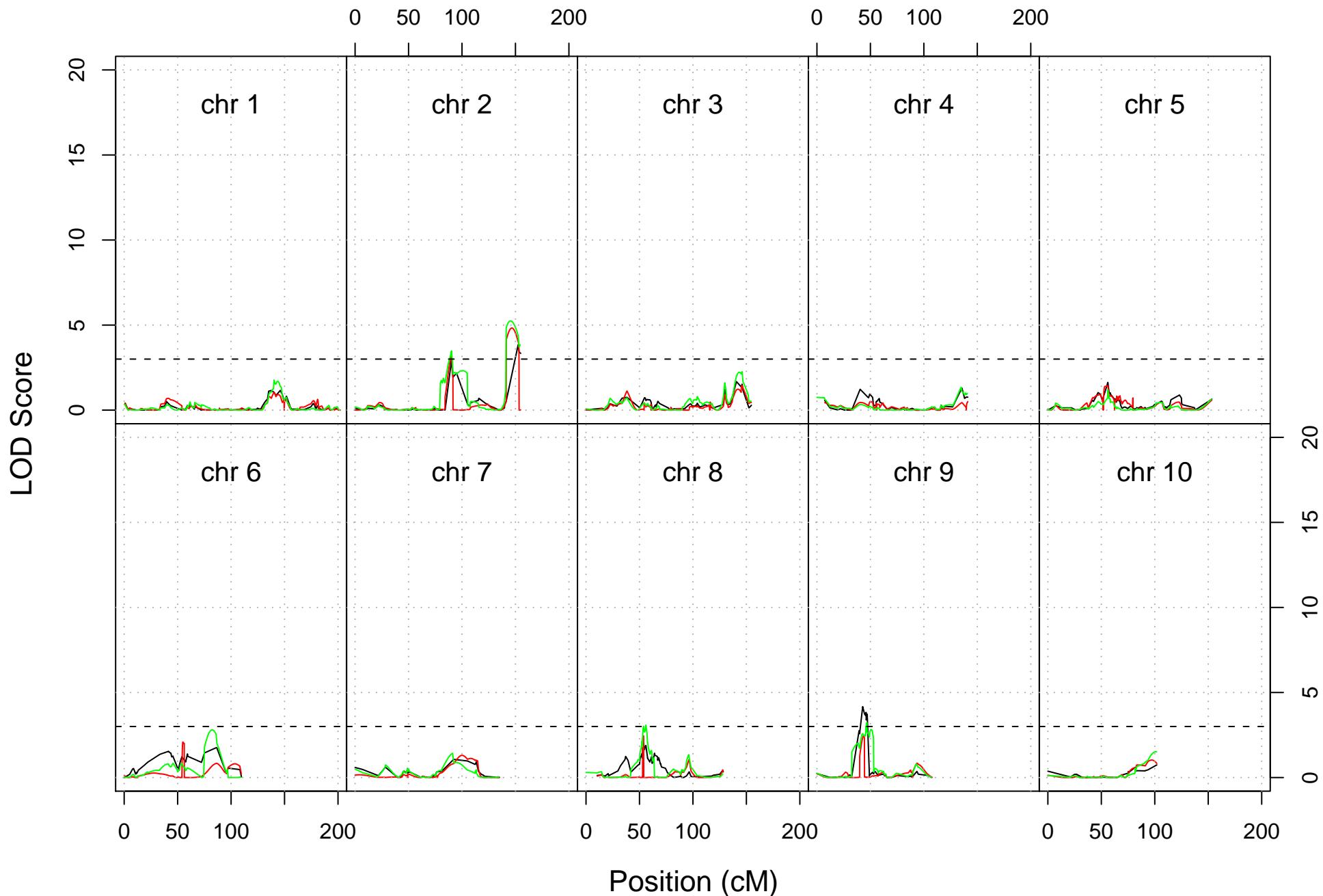
LOD Scores for MS71 population, anthesis–silk interval

ICIM = red, CIM = green, Stepwise = black



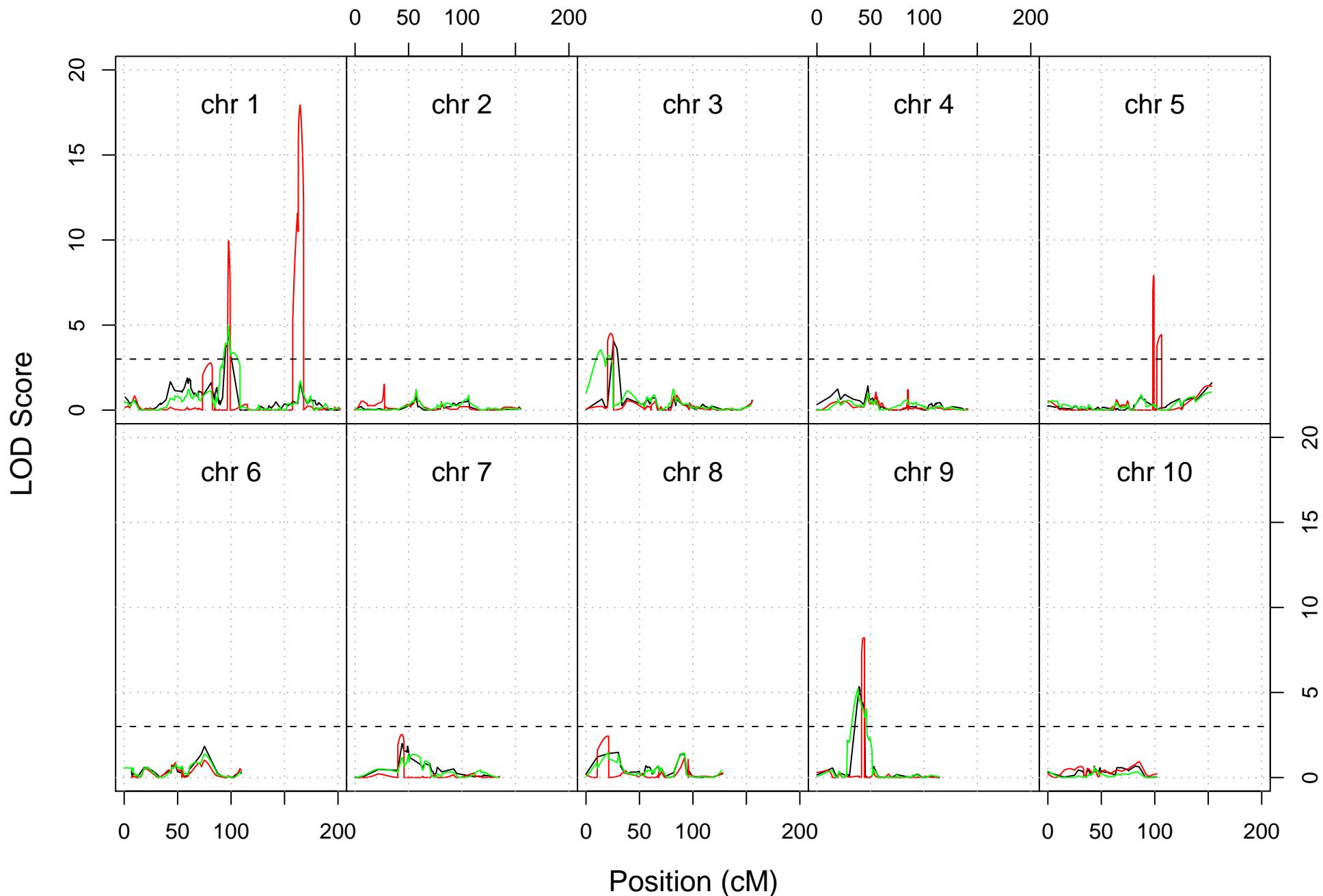
LOD Scores for NC350 population, anthesis–silk interval

ICIM = red, CIM = green, Stepwise = black



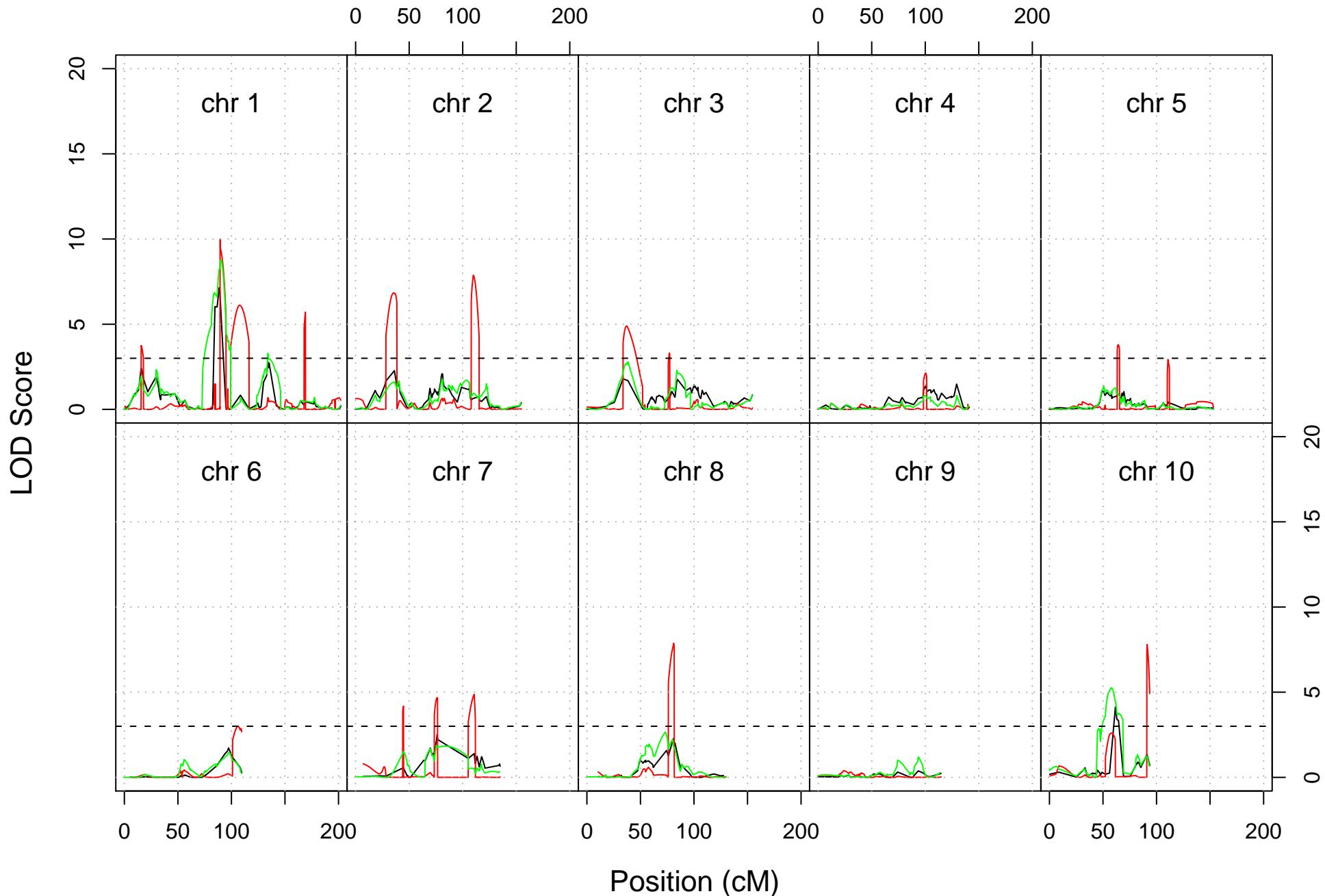
LOD Scores for NC358 population, anthesis–silk interval

ICIM = red, CIM = green, Stepwise = black



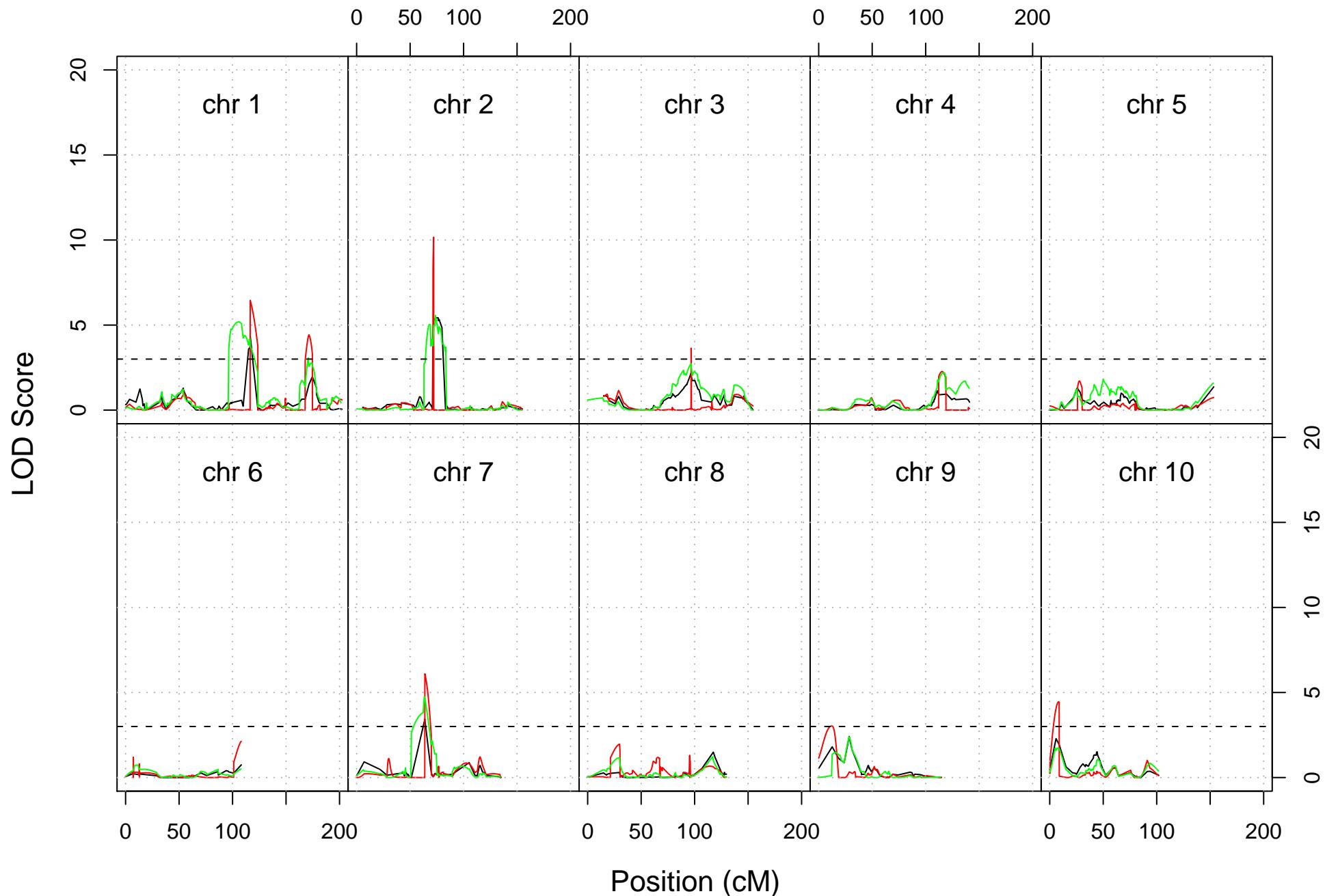
LOD Scores for OH43 population, anthesis–silk interval

ICIM = red, CIM = green, Stepwise = black



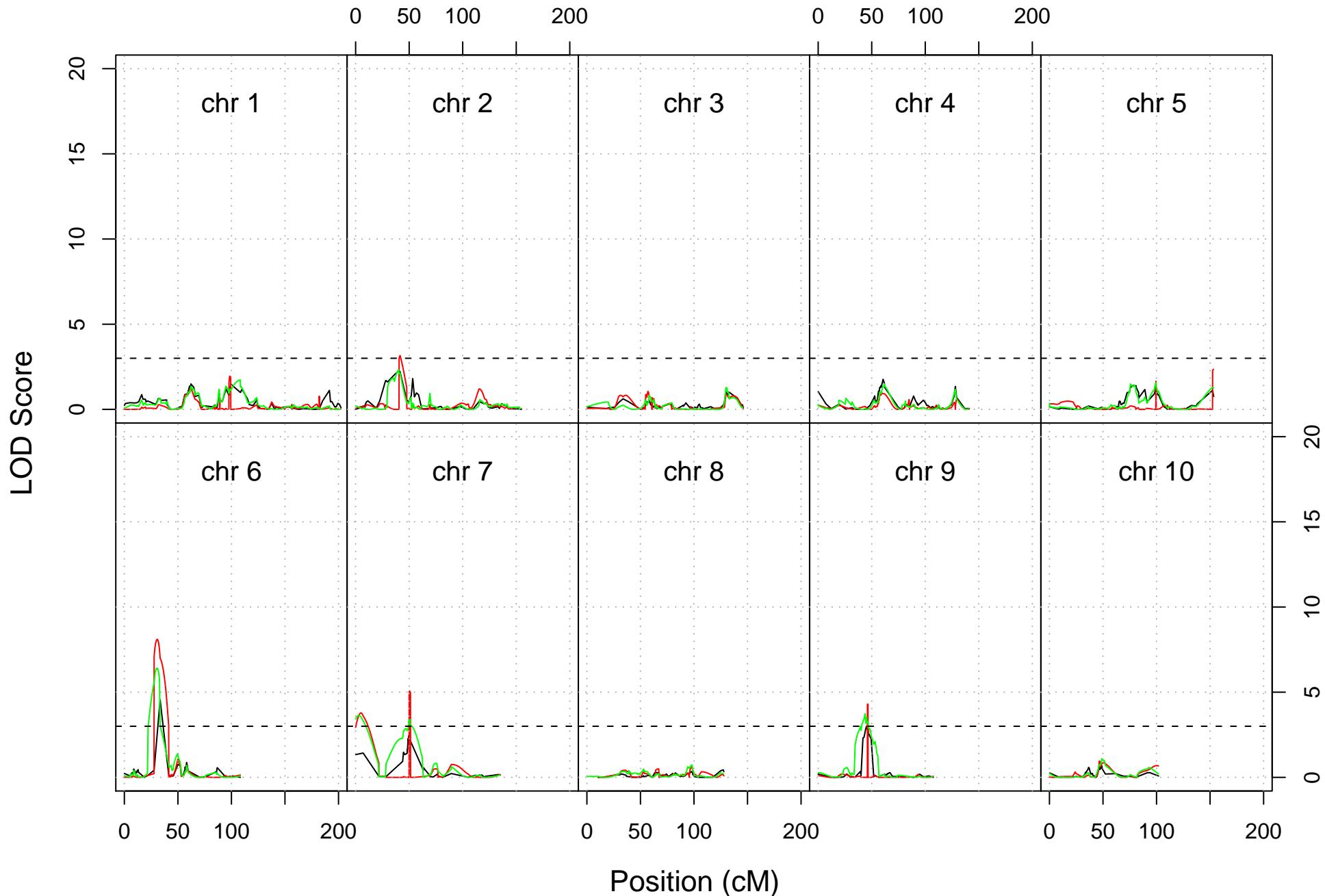
LOD Scores for OH7B population, anthesis–silk interval

ICIM = red, CIM = green, Stepwise = black



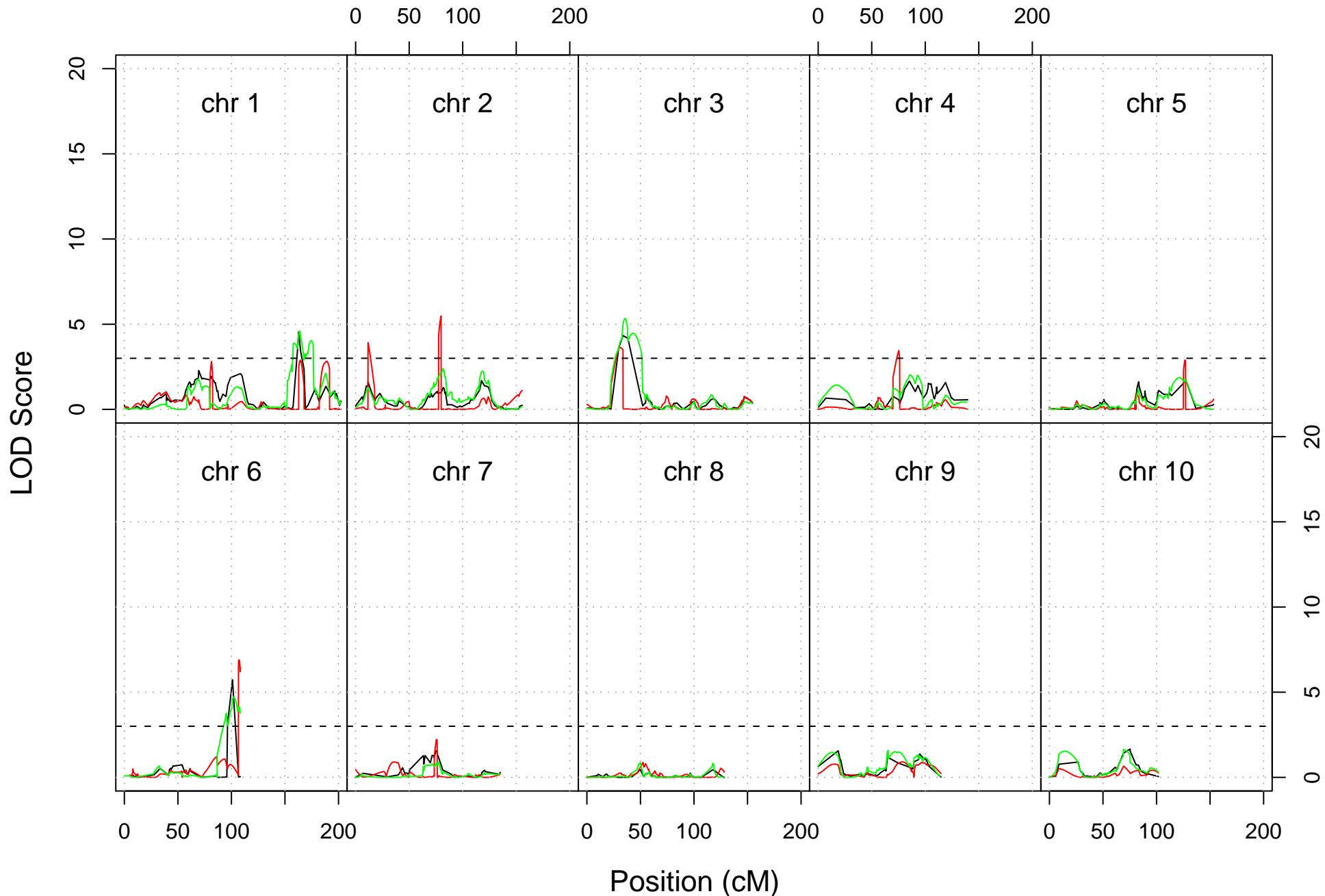
LOD Scores for P39 population, anthesis–silk interval

ICIM = red, CIM = green, Stepwise = black



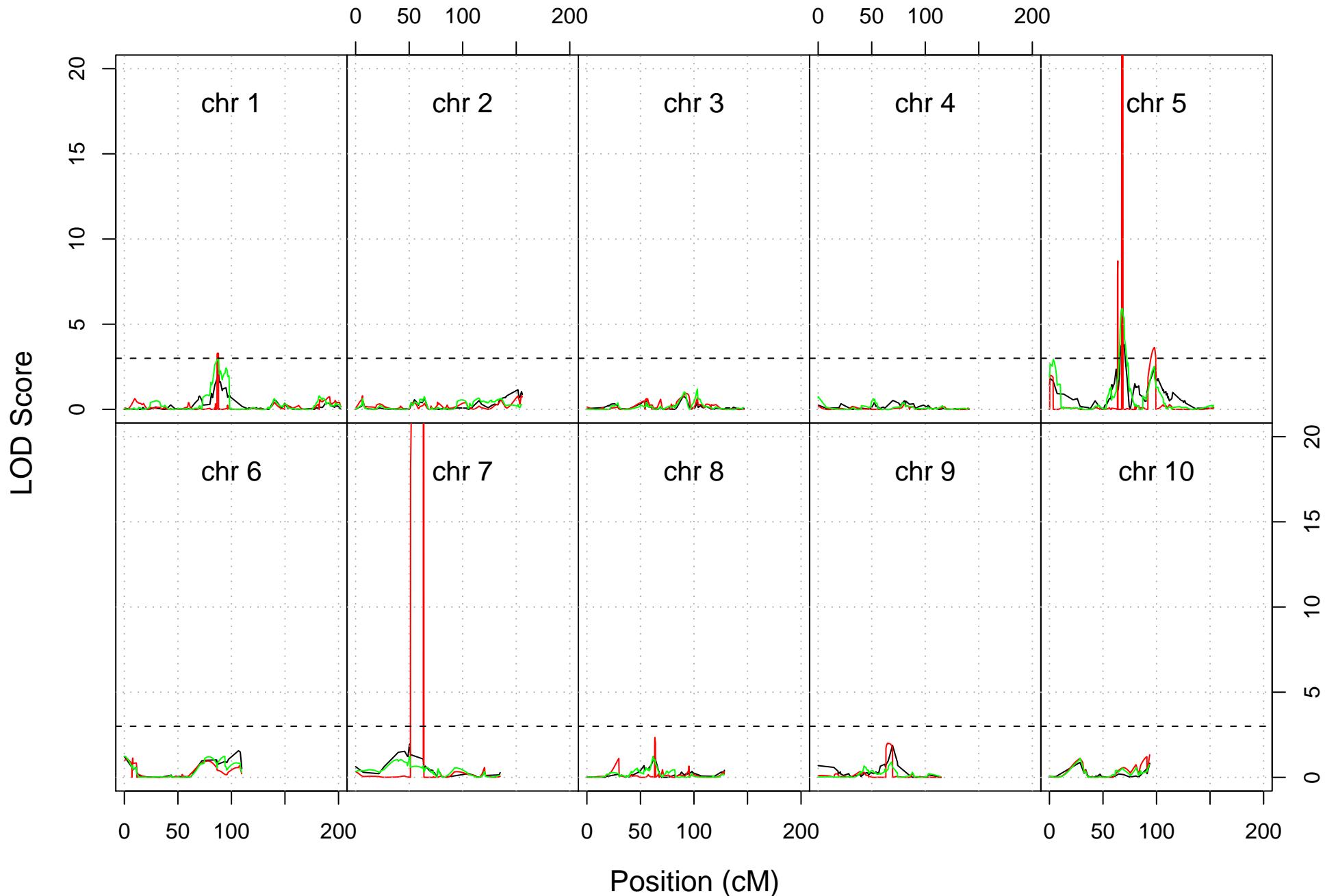
LOD Scores for TX303 population, anthesis–silk interval

ICIM = red, CIM = green, Stepwise = black



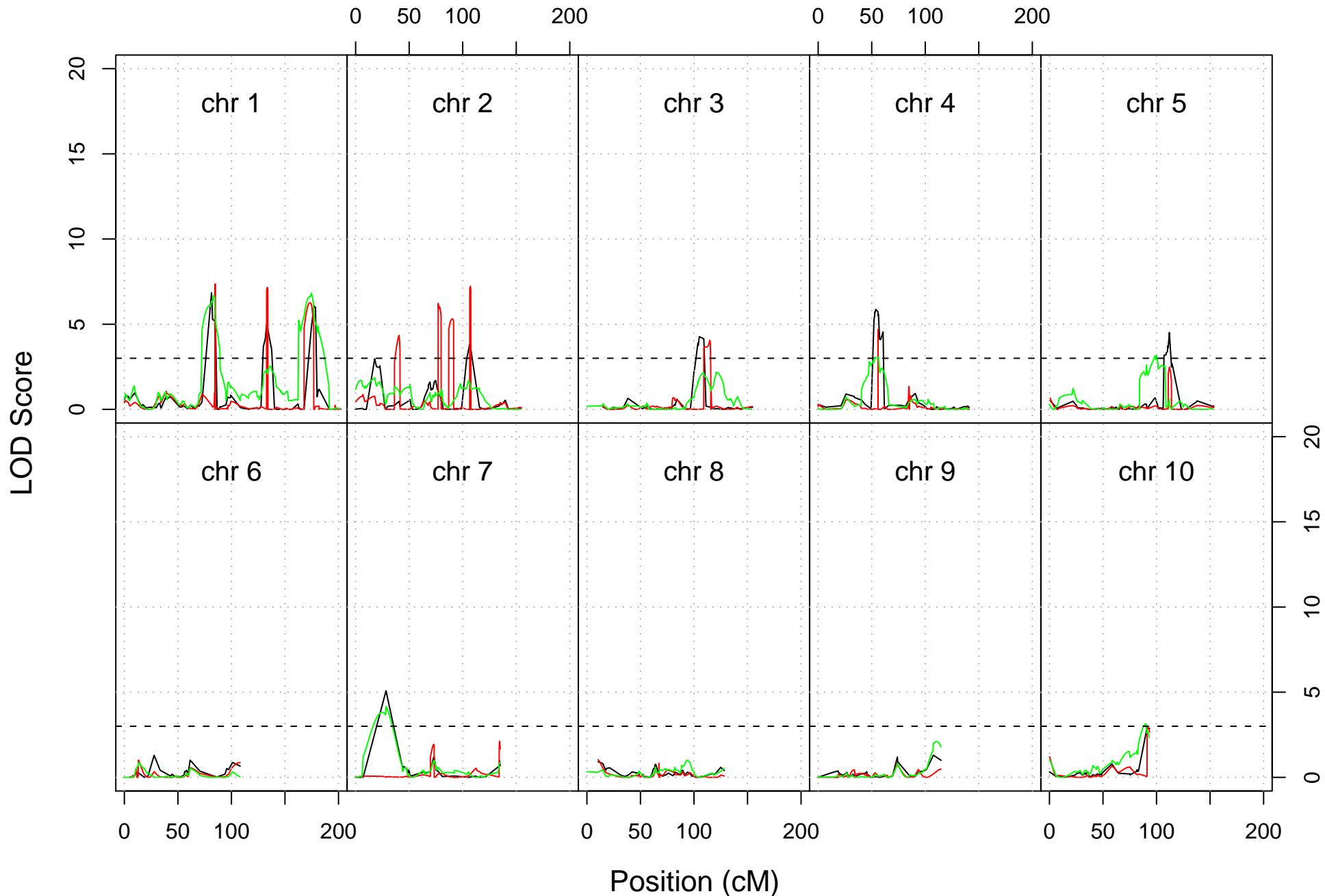
LOD Scores for TZI8 population, anthesis–silk interval

ICIM = red, CIM = green, Stepwise = black



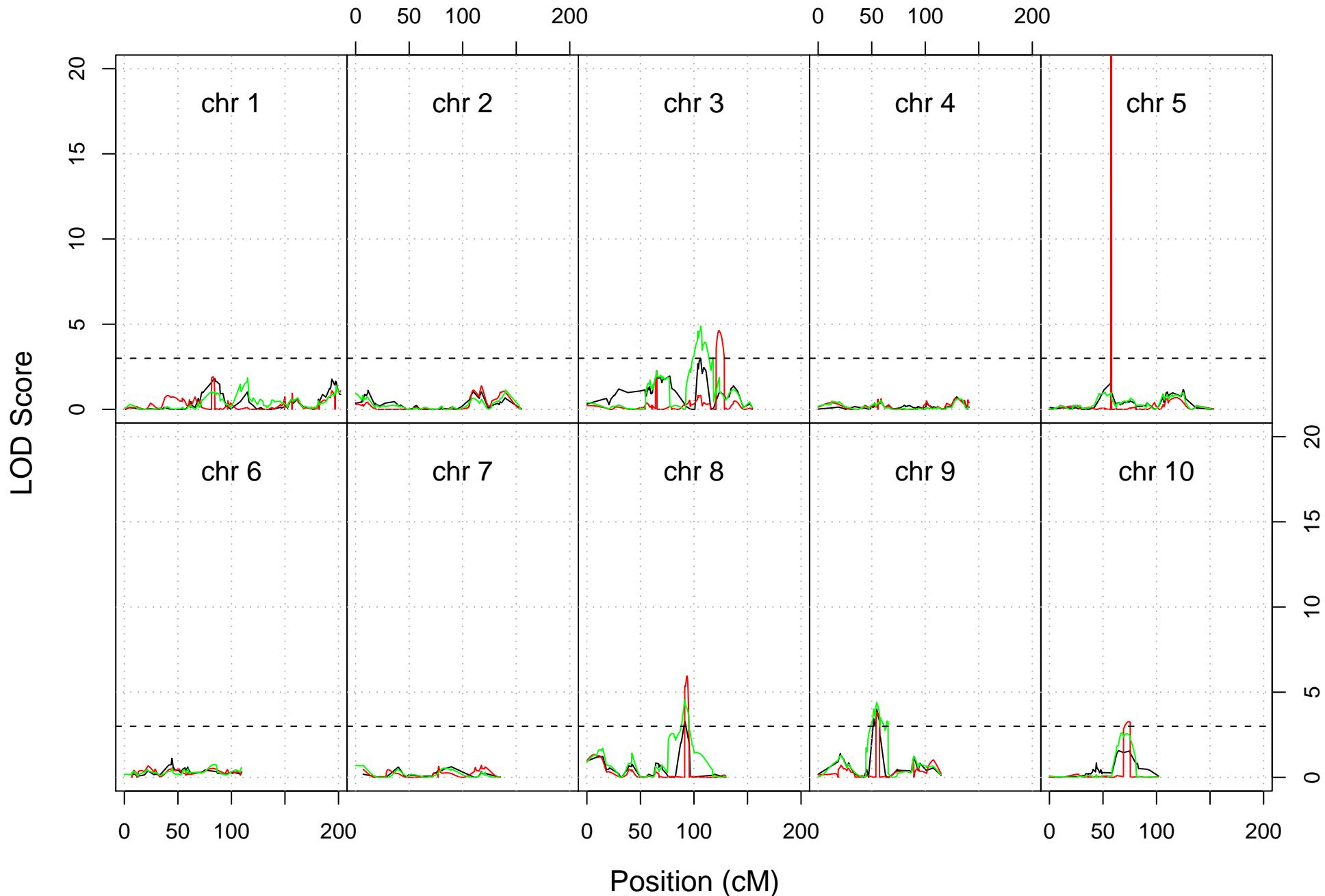
LOD Scores for B97 population, days to anthesis

ICIM = red, CIM = green, Stepwise = black



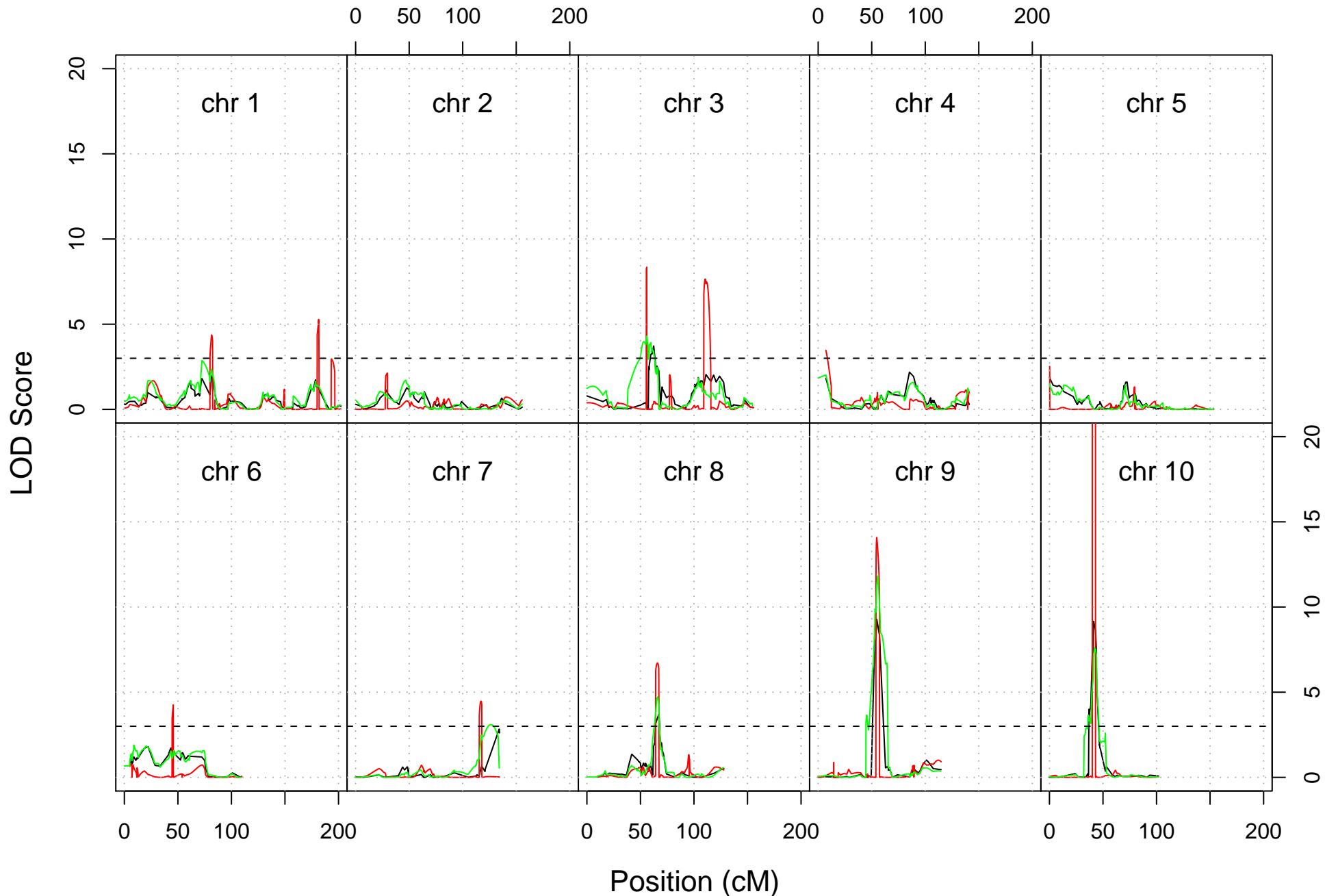
LOD Scores for CML103 population, days to anthesis

ICIM = red, CIM = green, Stepwise = black



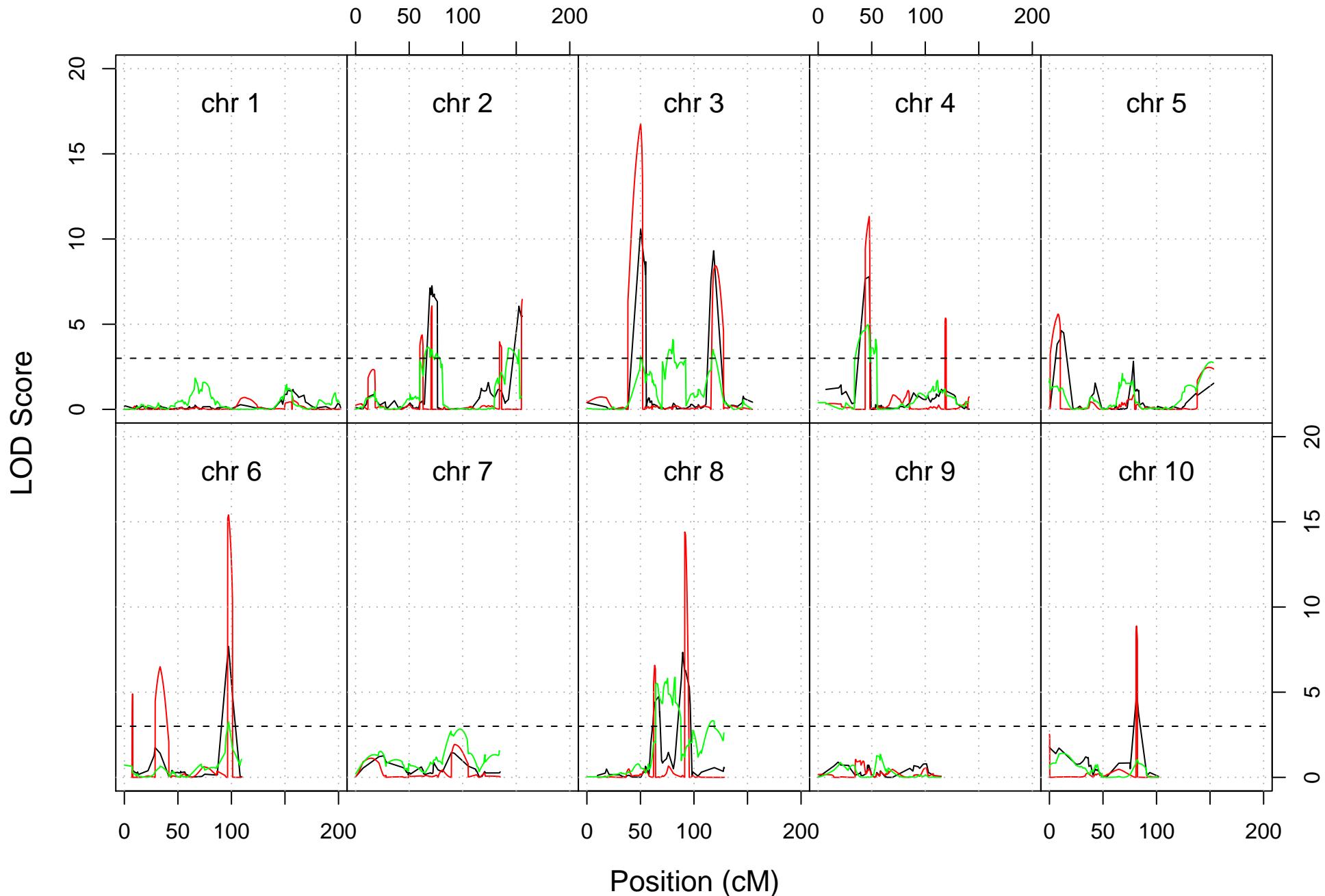
LOD Scores for CML228 population, days to anthesis

ICIM = red, CIM = green, Stepwise = black



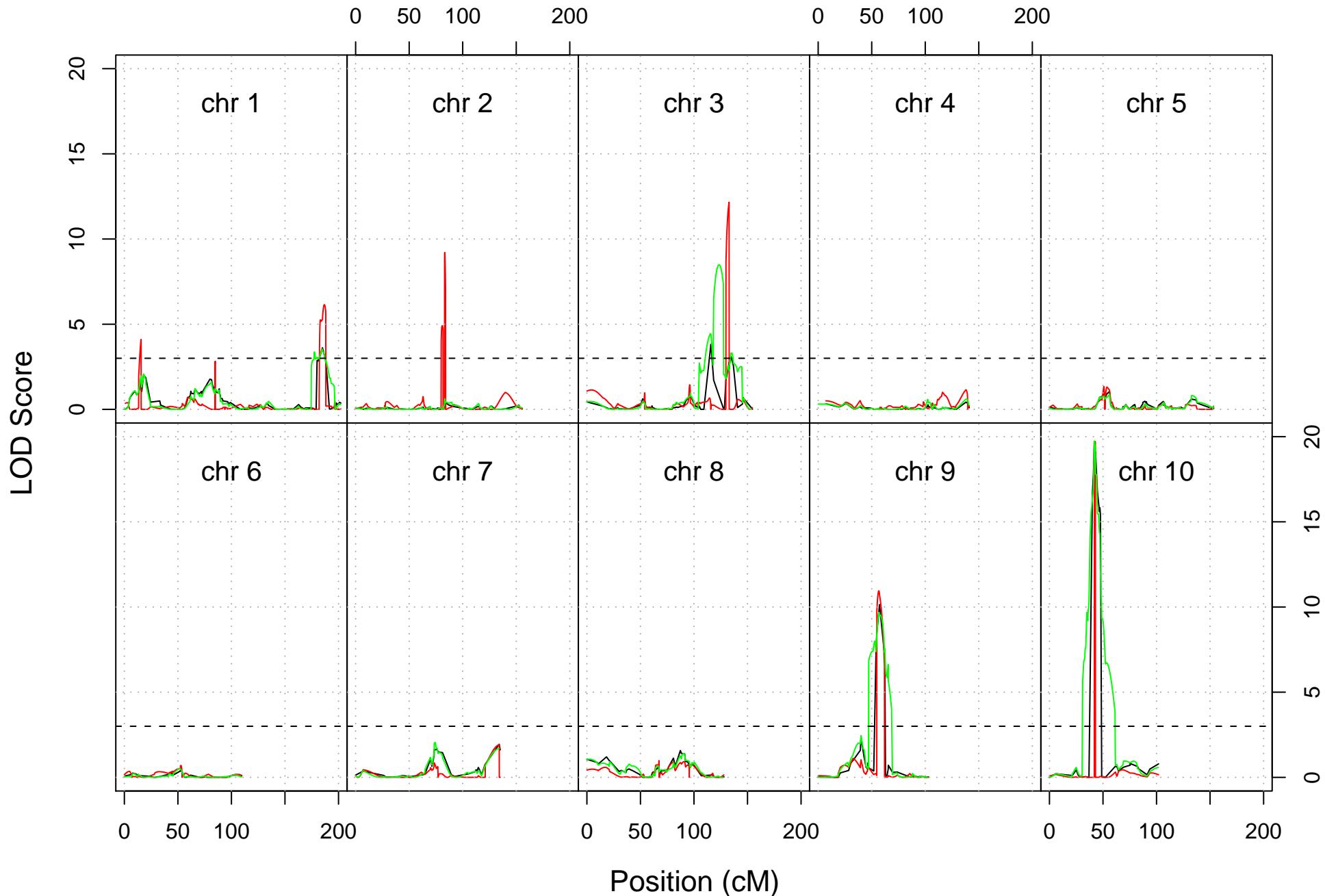
LOD Scores for CML247 population, days to anthesis

ICIM = red, CIM = green, Stepwise = black



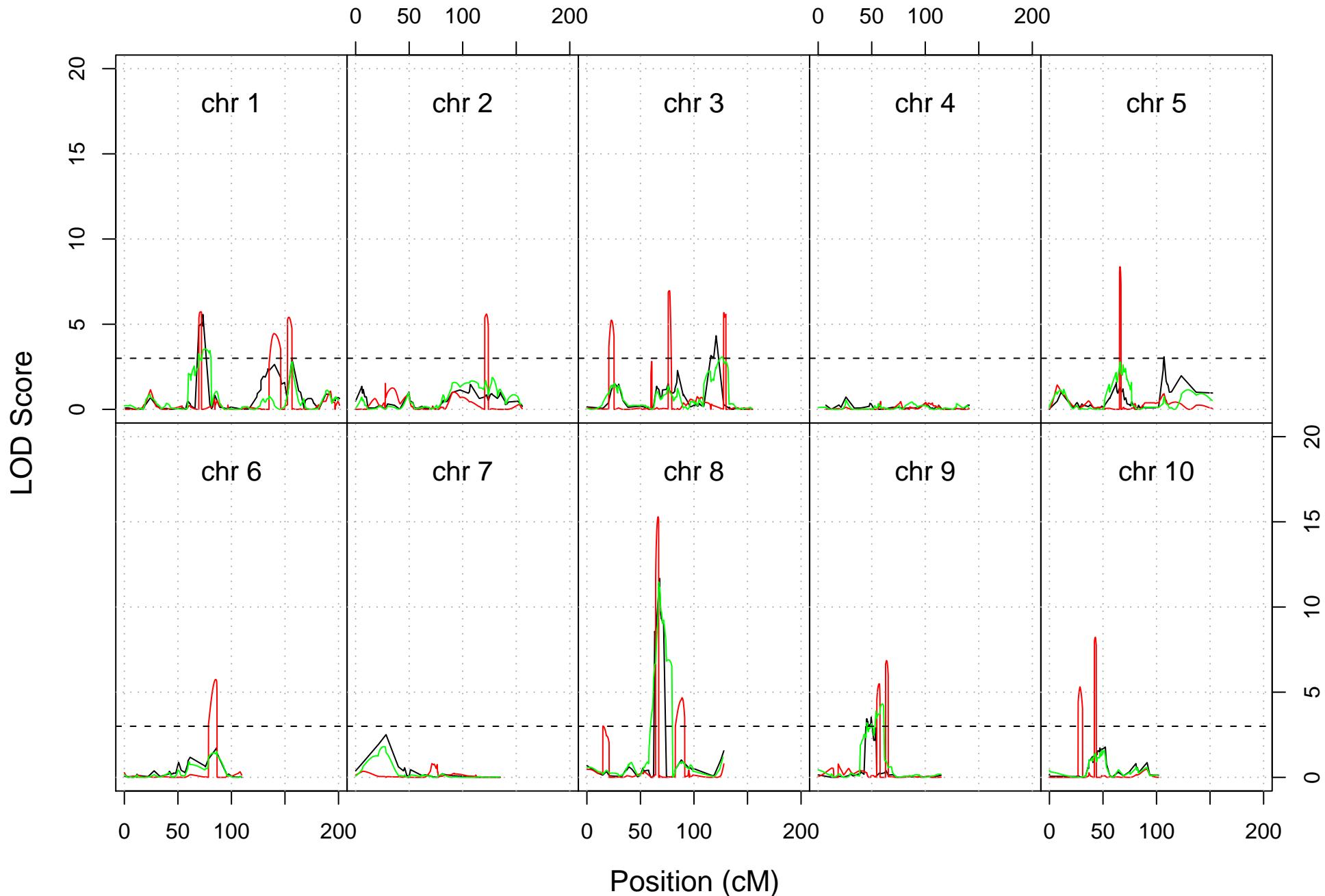
LOD Scores for CML277 population, days to anthesis

ICIM = red, CIM = green, Stepwise = black



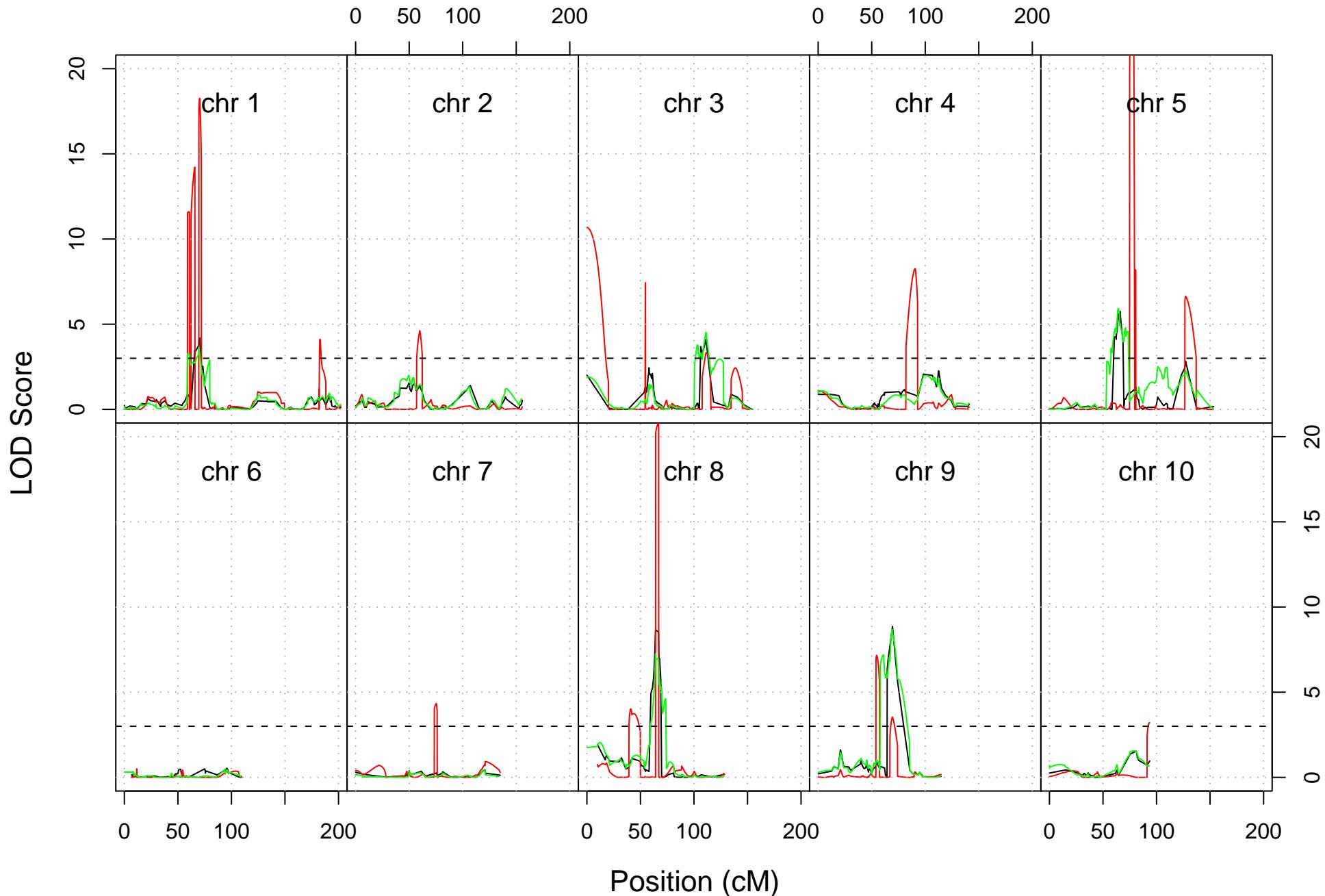
LOD Scores for CML322 population, days to anthesis

ICIM = red, CIM = green, Stepwise = black



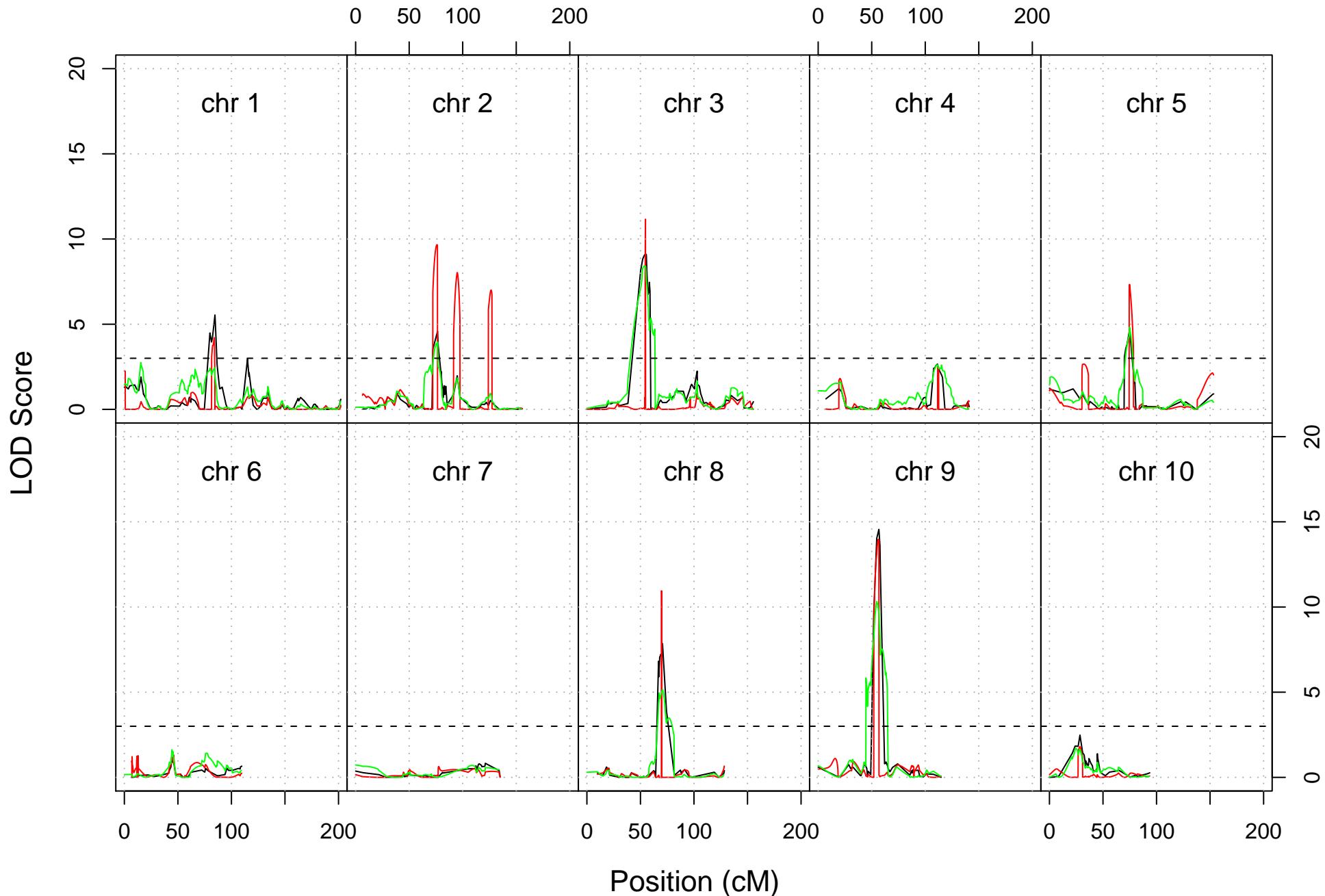
LOD Scores for CML333 population, days to anthesis

ICIM = red, CIM = green, Stepwise = black



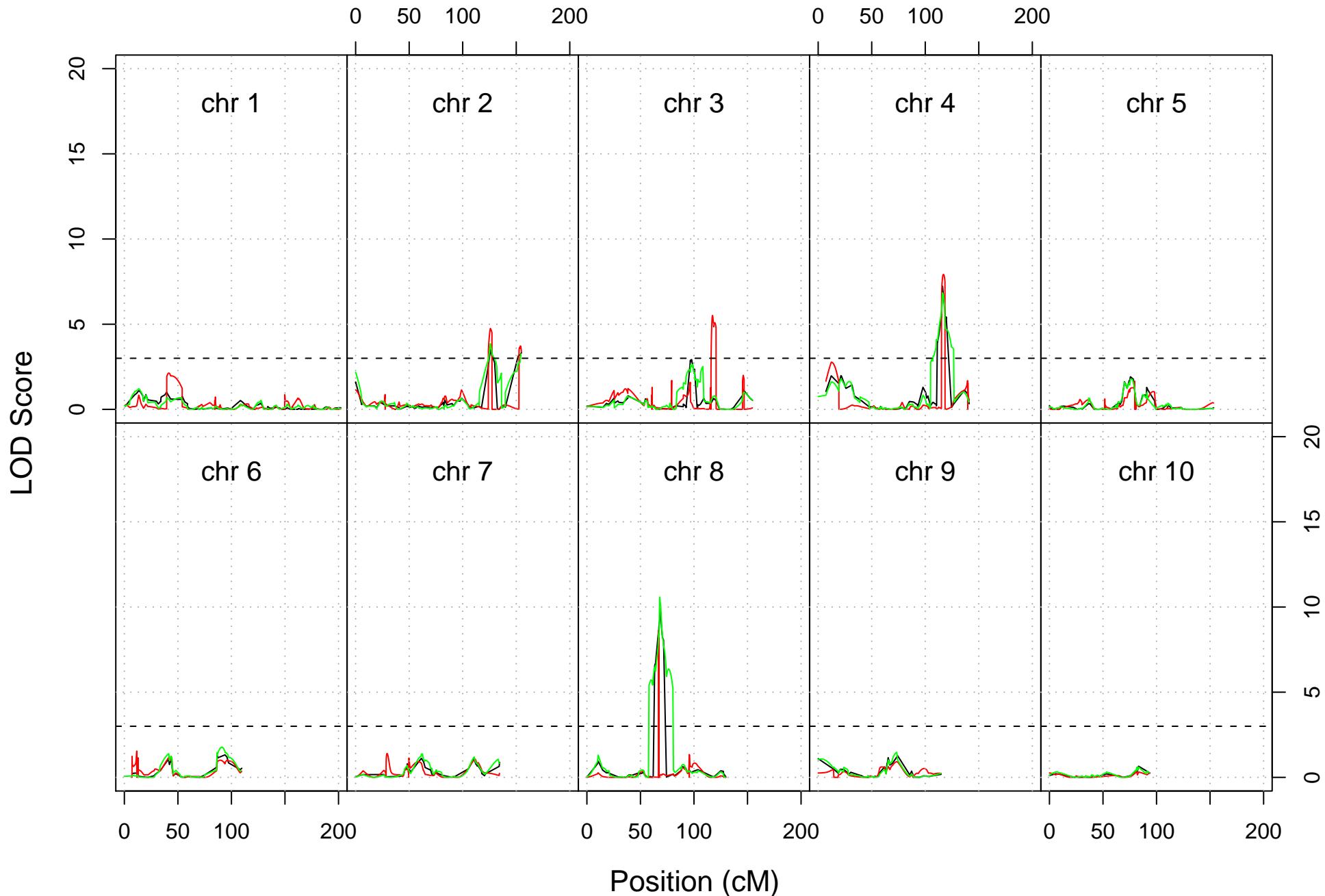
LOD Scores for CML52 population, days to anthesis

ICIM = red, CIM = green, Stepwise = black



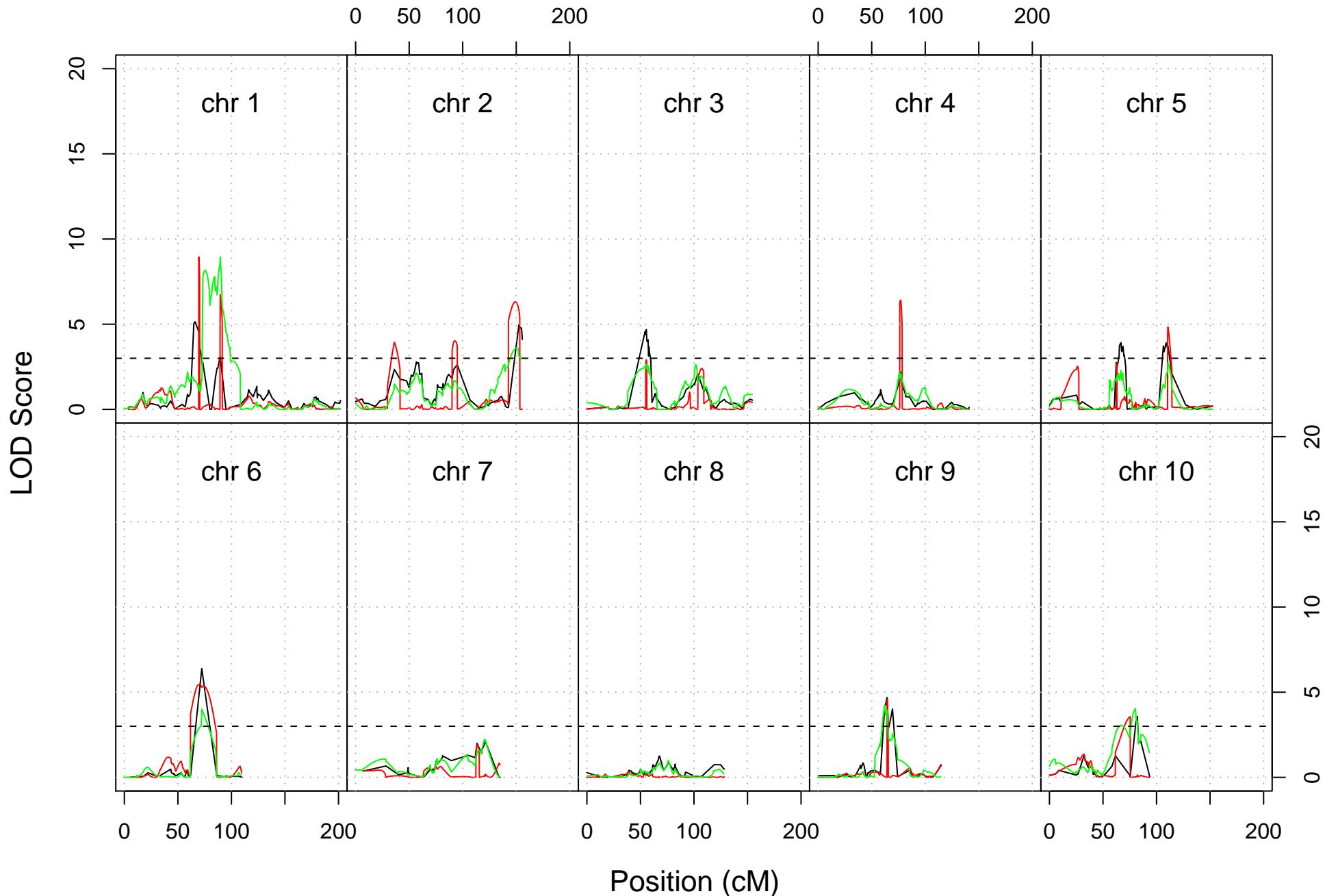
LOD Scores for CML69 population, days to anthesis

ICIM = red, CIM = green, Stepwise = black



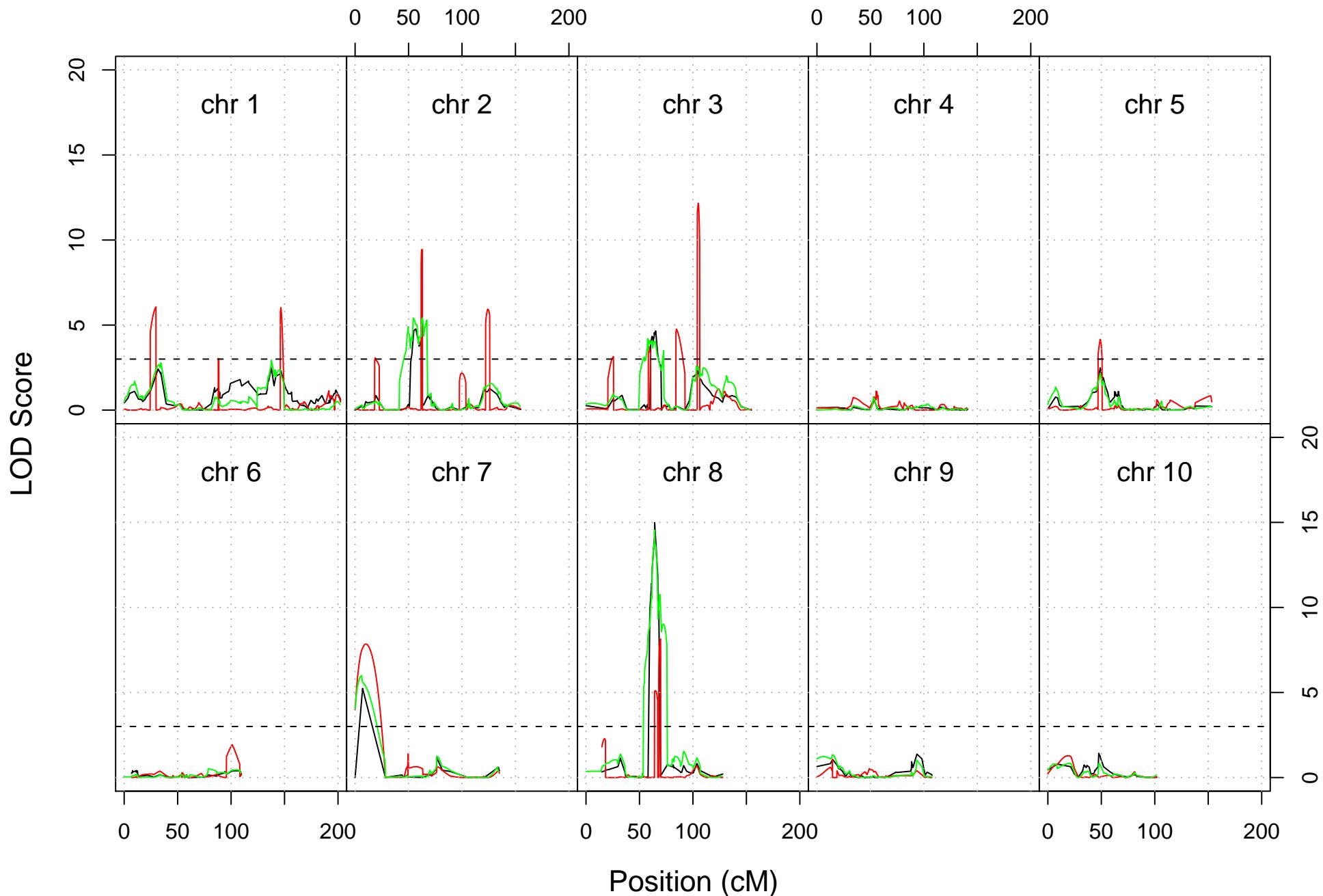
LOD Scores for HP301 population, days to anthesis

ICIM = red, CIM = green, Stepwise = black



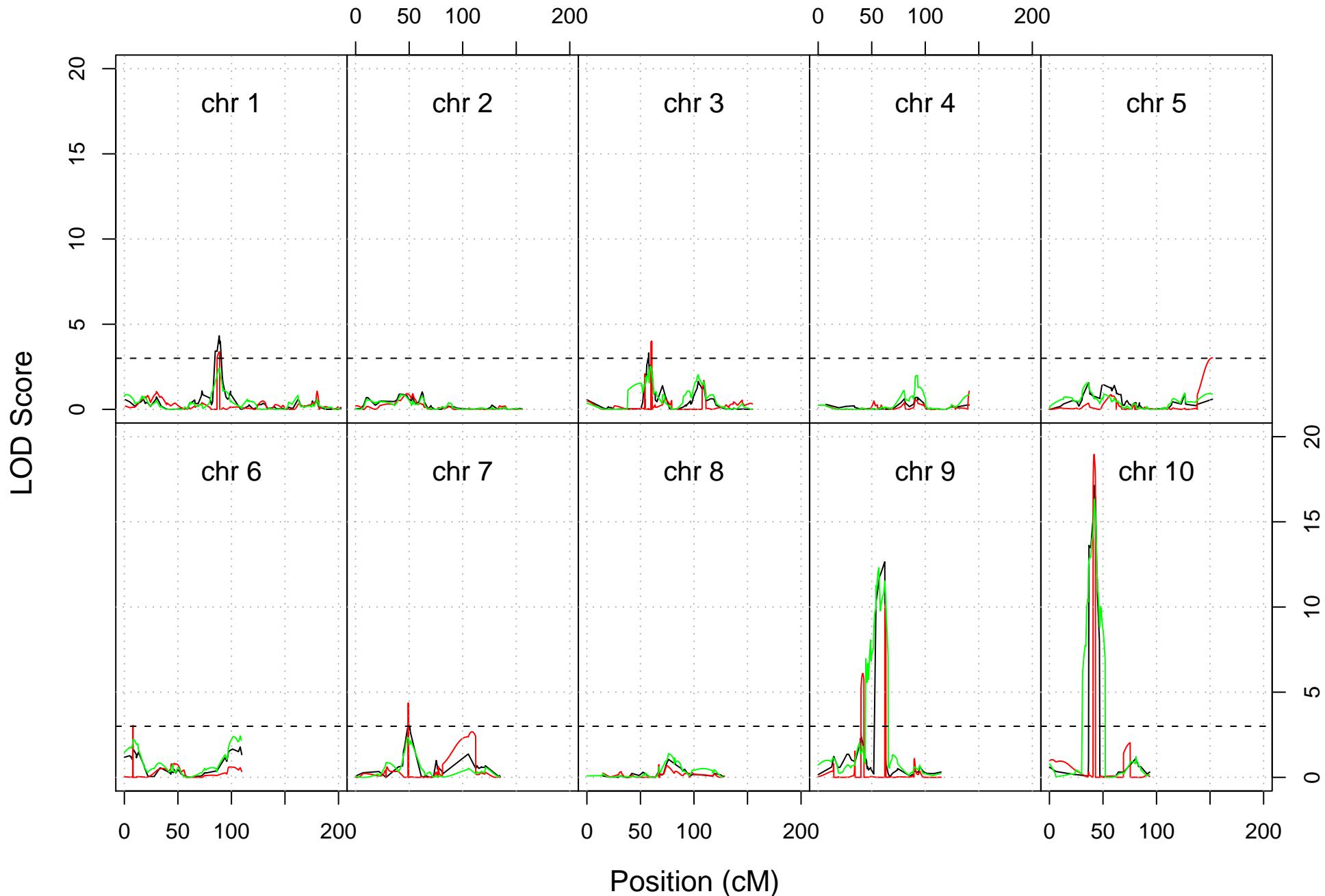
LOD Scores for IL14H population, days to anthesis

ICIM = red, CIM = green, Stepwise = black



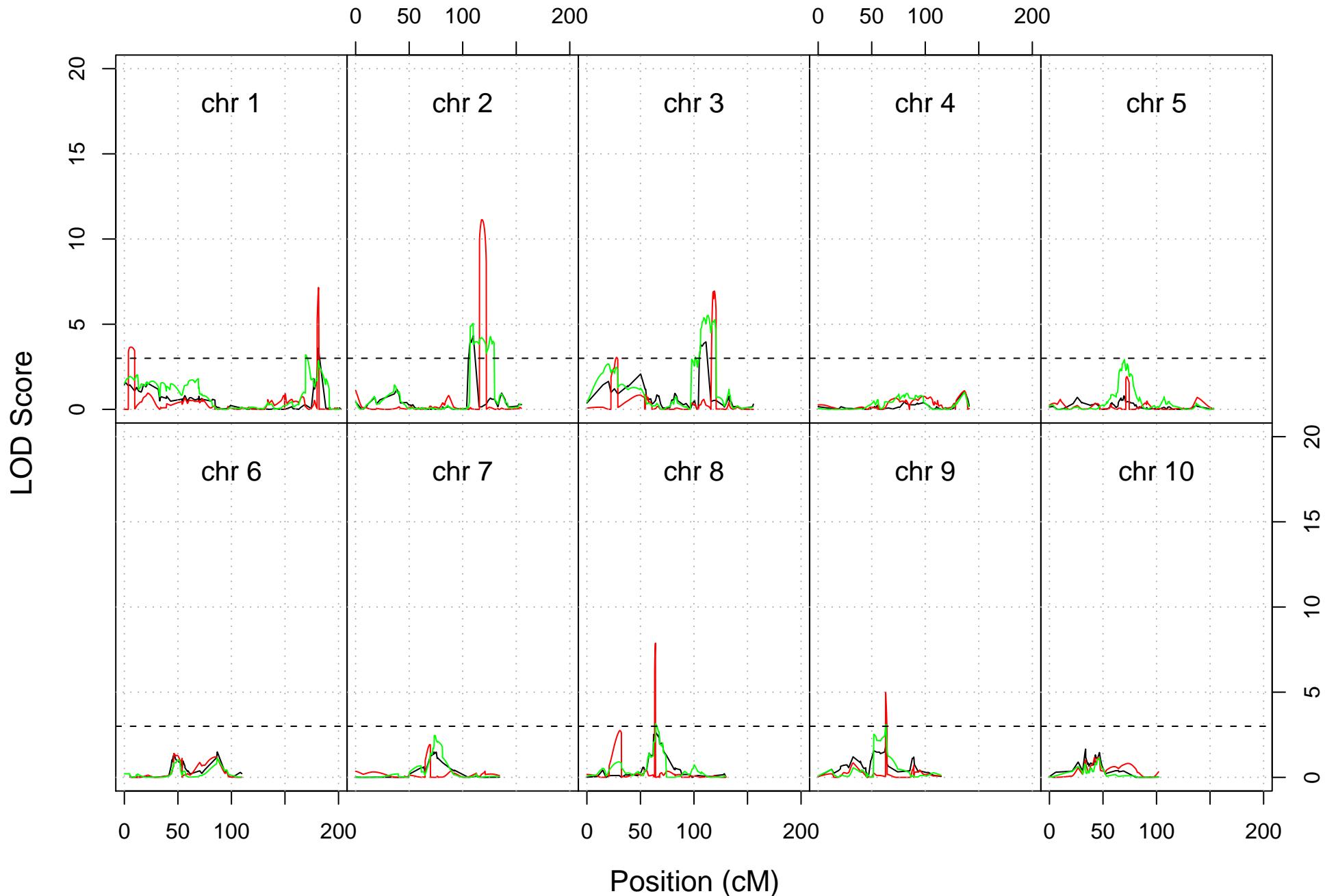
LOD Scores for KI11 population, days to anthesis

ICIM = red, CIM = green, Stepwise = black



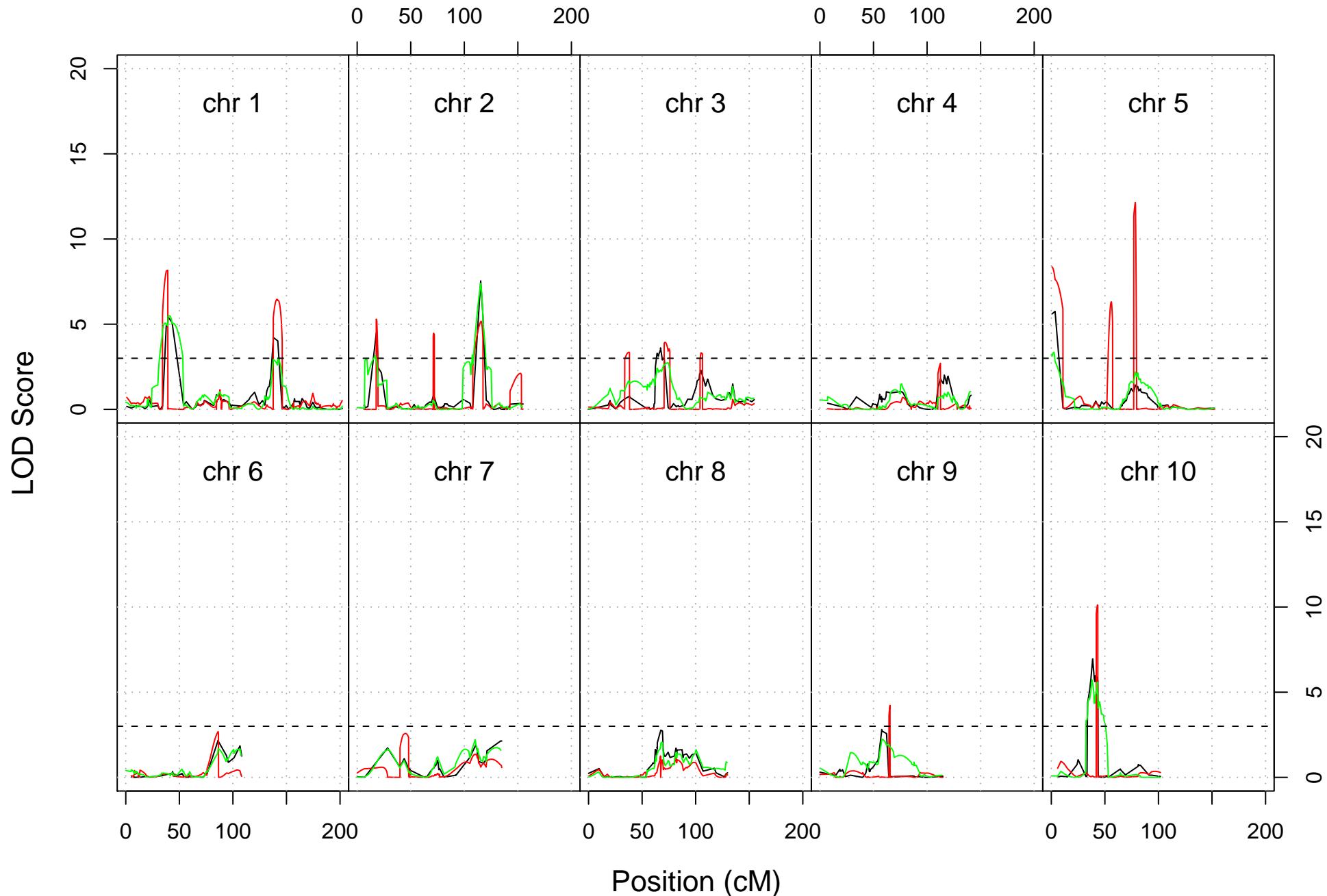
LOD Scores for KI3 population, days to anthesis

ICIM = red, CIM = green, Stepwise = black



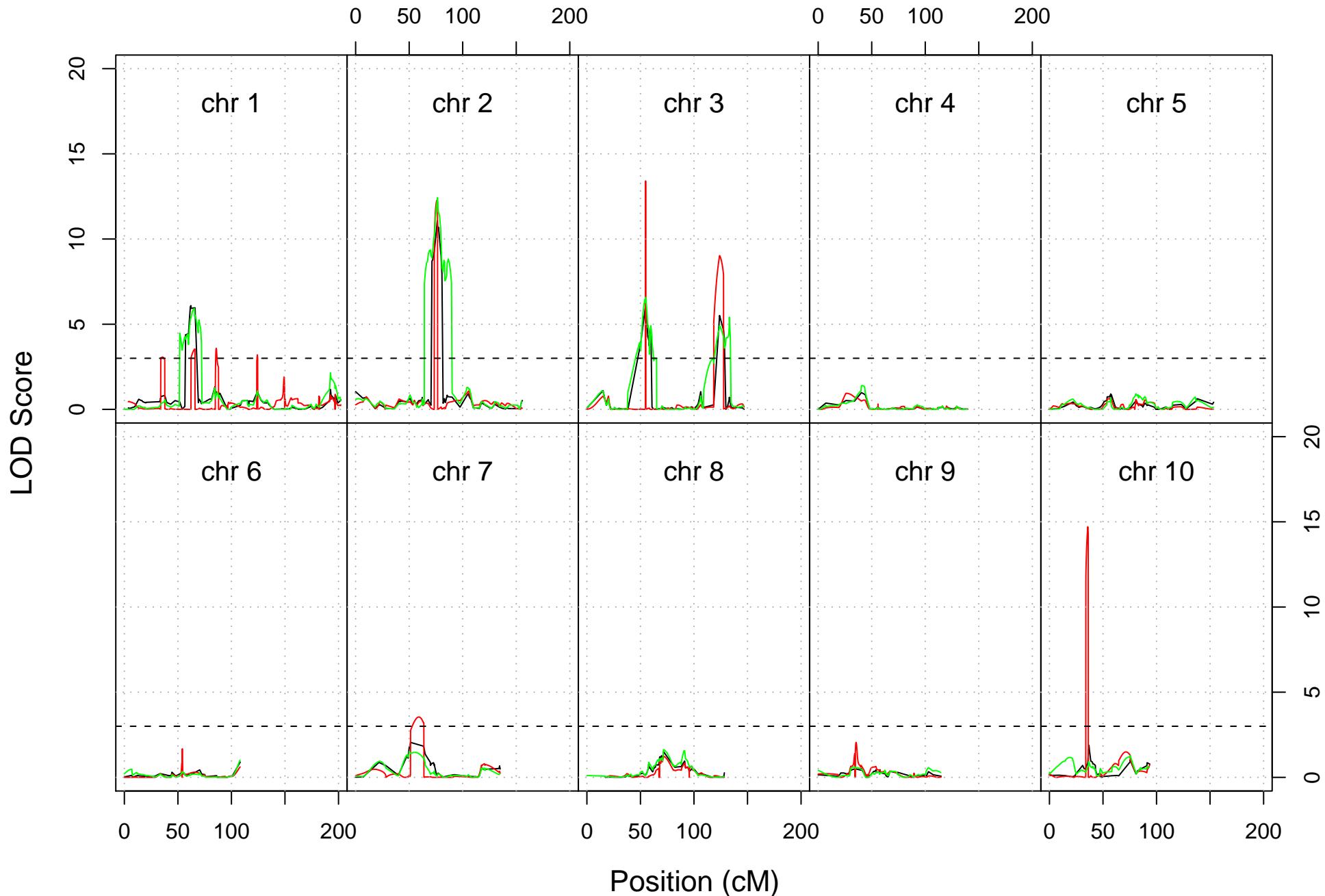
LOD Scores for KY21 population, days to anthesis

ICIM = red, CIM = green, Stepwise = black



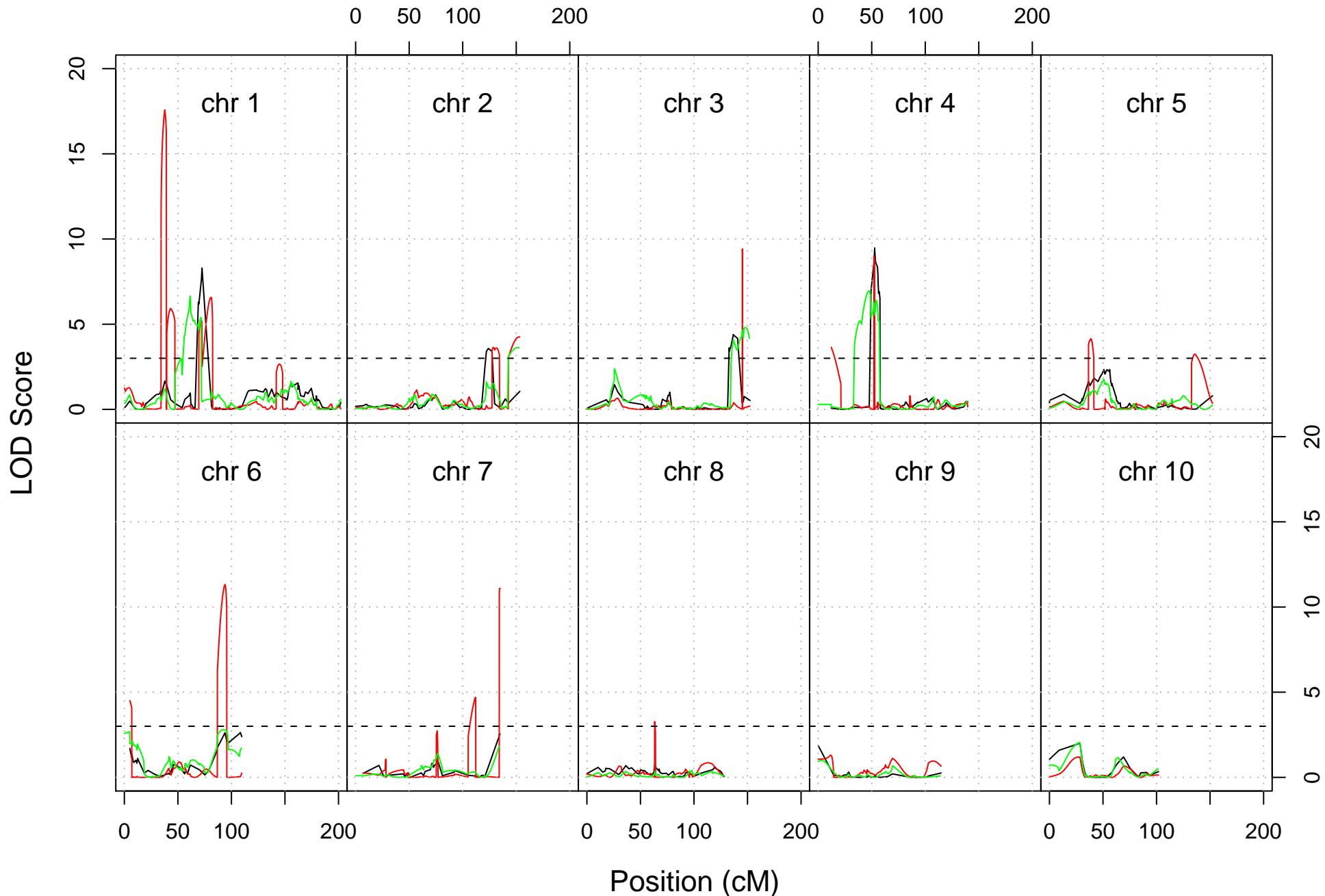
LOD Scores for M162W population, days to anthesis

ICIM = red, CIM = green, Stepwise = black



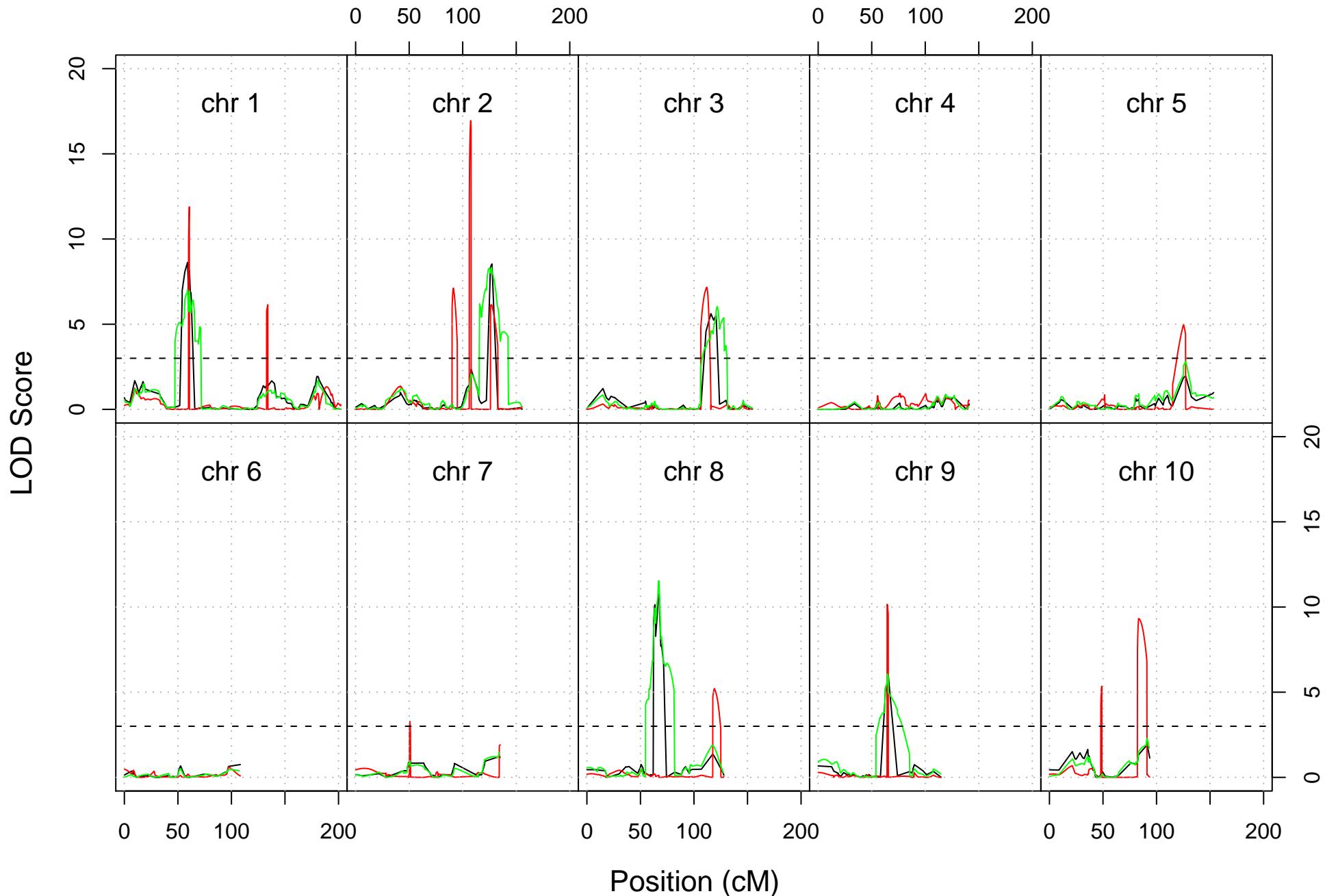
LOD Scores for M37W population, days to anthesis

ICIM = red, CIM = green, Stepwise = black



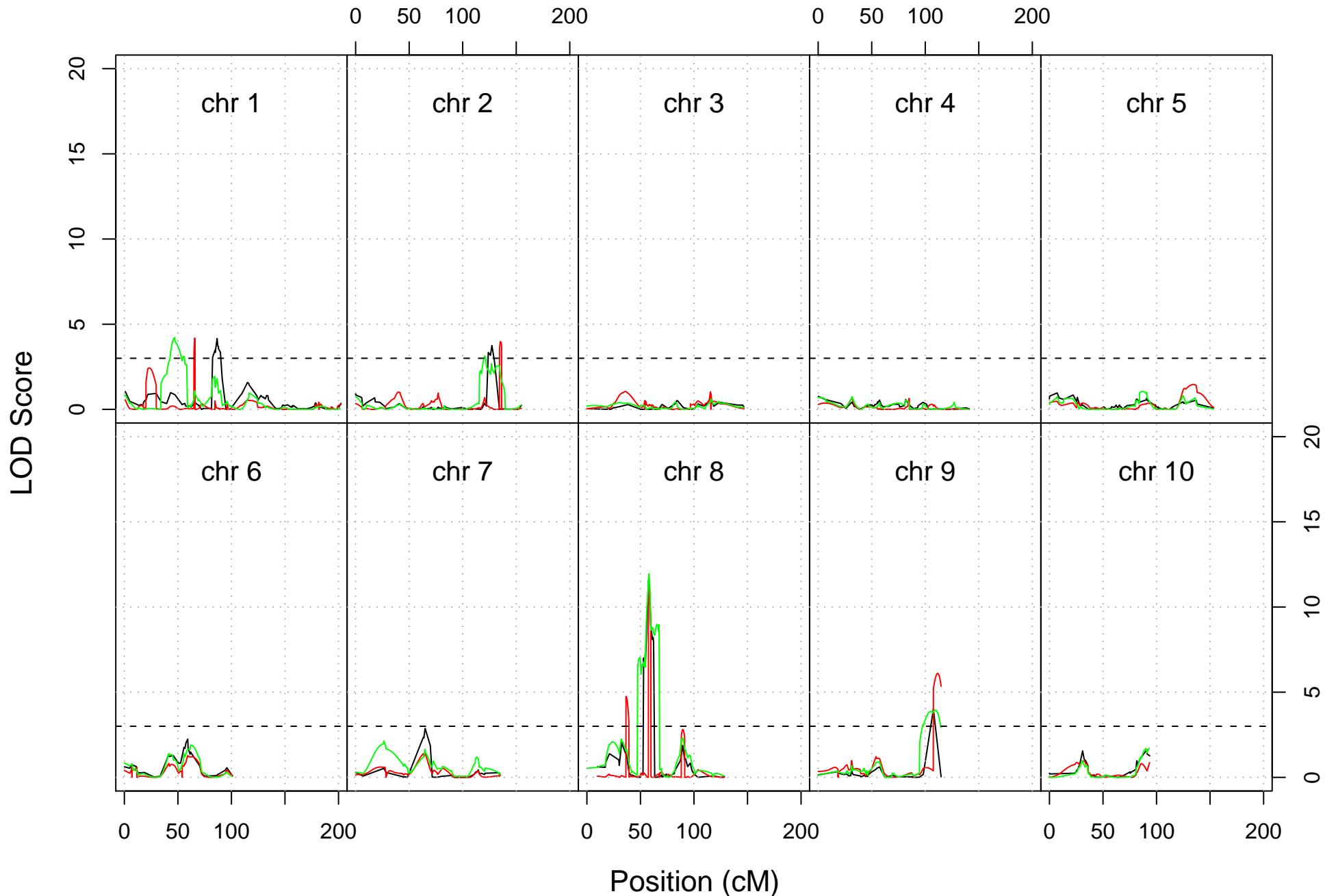
LOD Scores for MO18W population, days to anthesis

ICIM = red, CIM = green, Stepwise = black



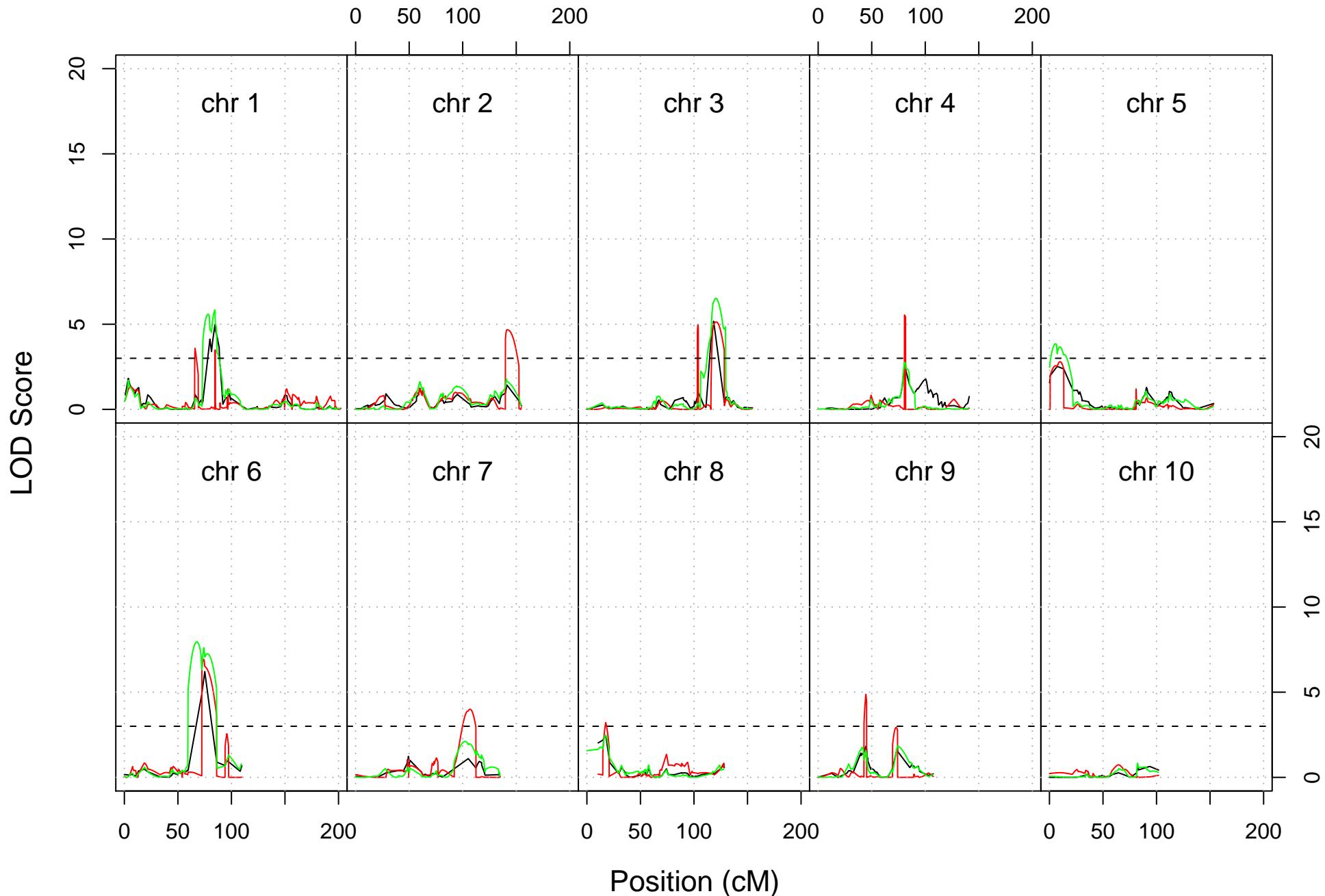
LOD Scores for MS71 population, days to anthesis

ICIM = red, CIM = green, Stepwise = black



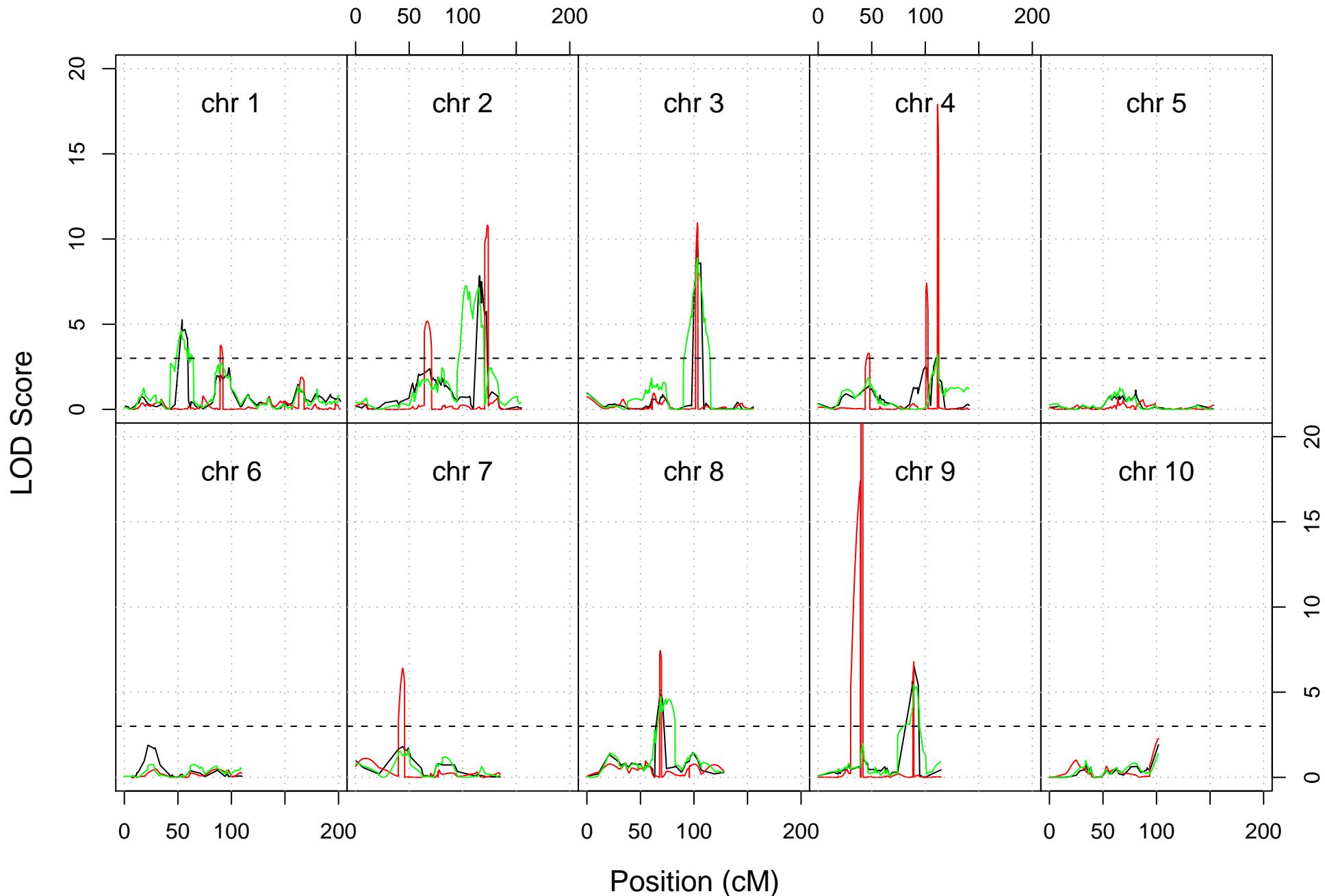
LOD Scores for NC350 population, days to anthesis

ICIM = red, CIM = green, Stepwise = black



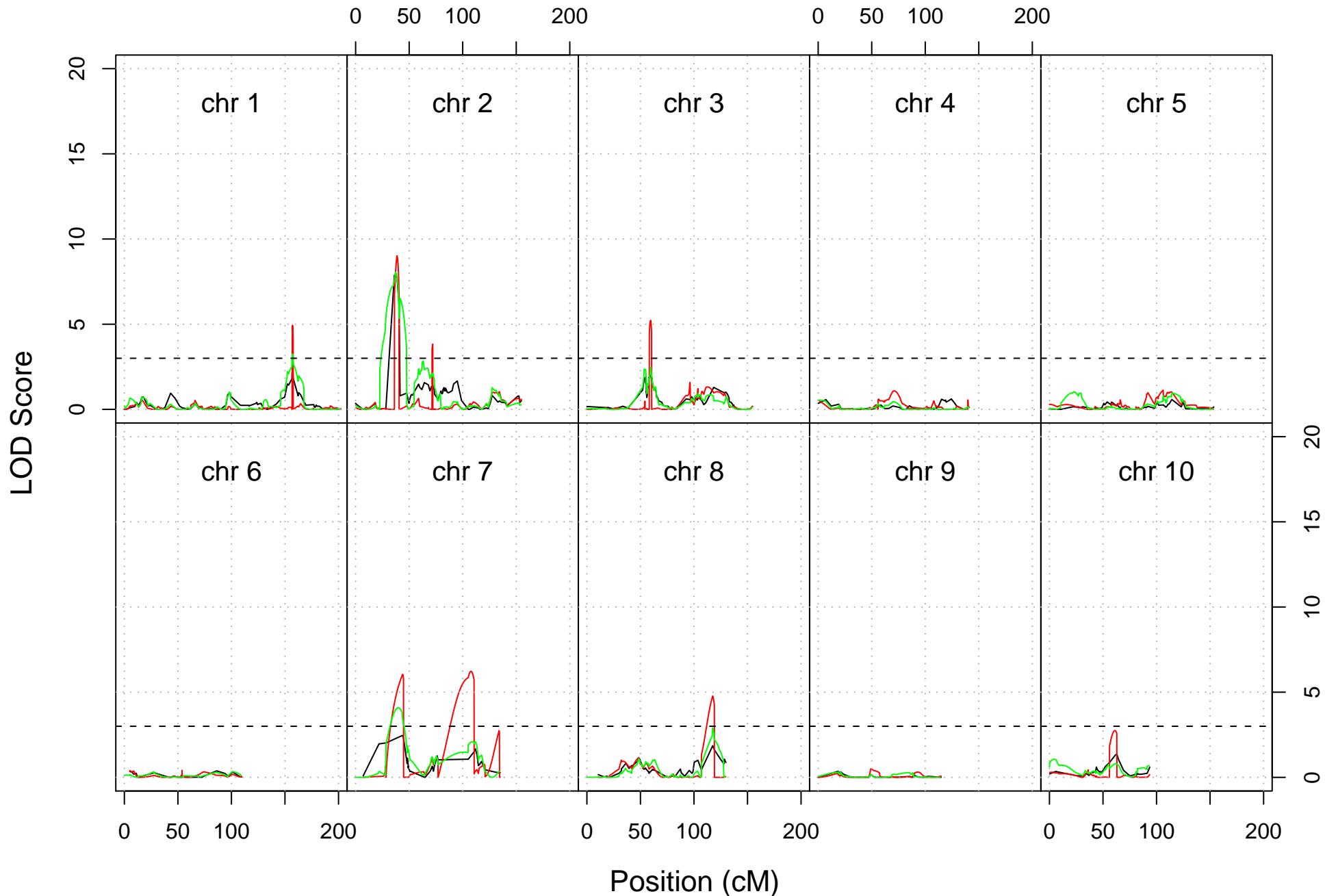
LOD Scores for NC358 population, days to anthesis

ICIM = red, CIM = green, Stepwise = black



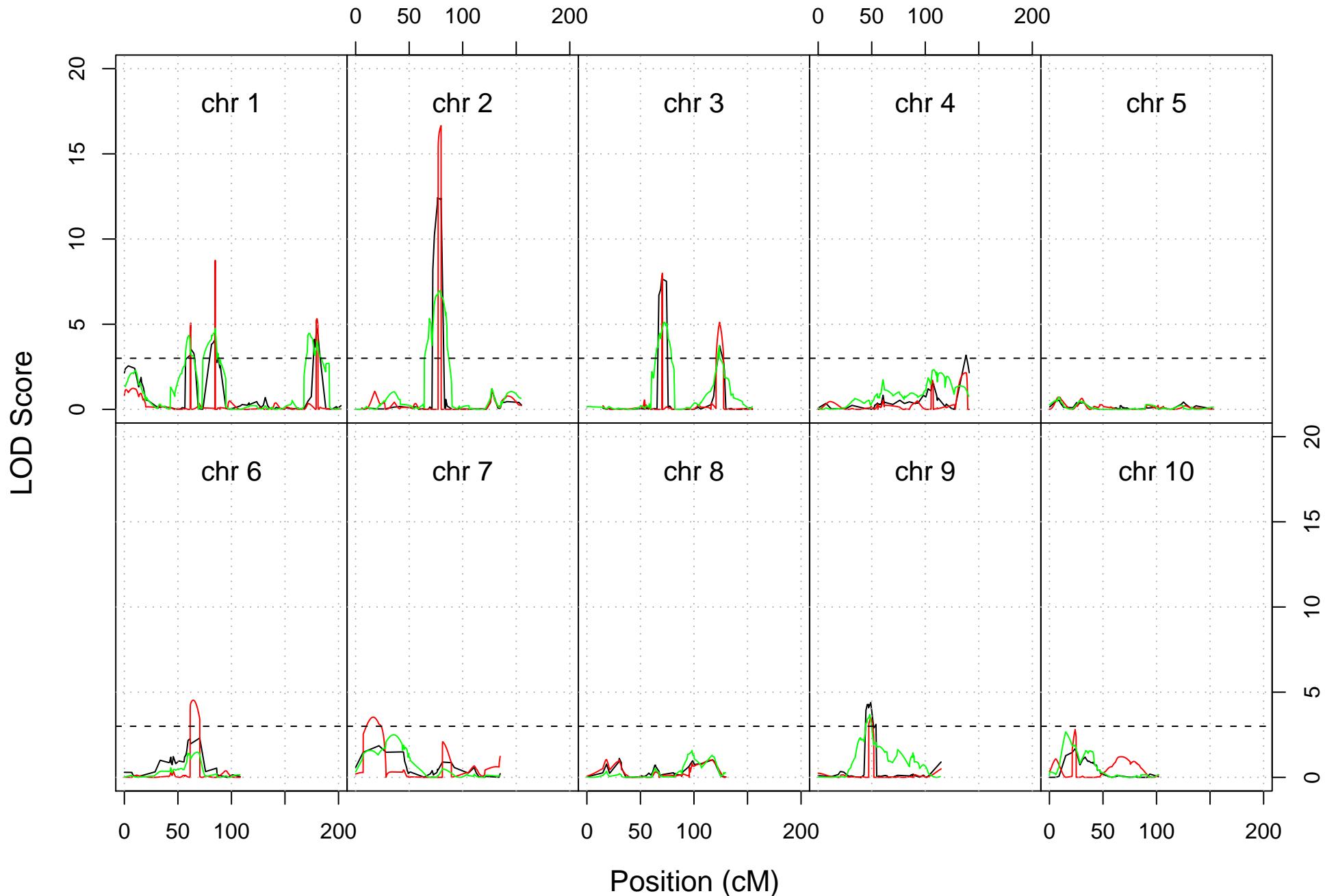
LOD Scores for OH43 population, days to anthesis

ICIM = red, CIM = green, Stepwise = black



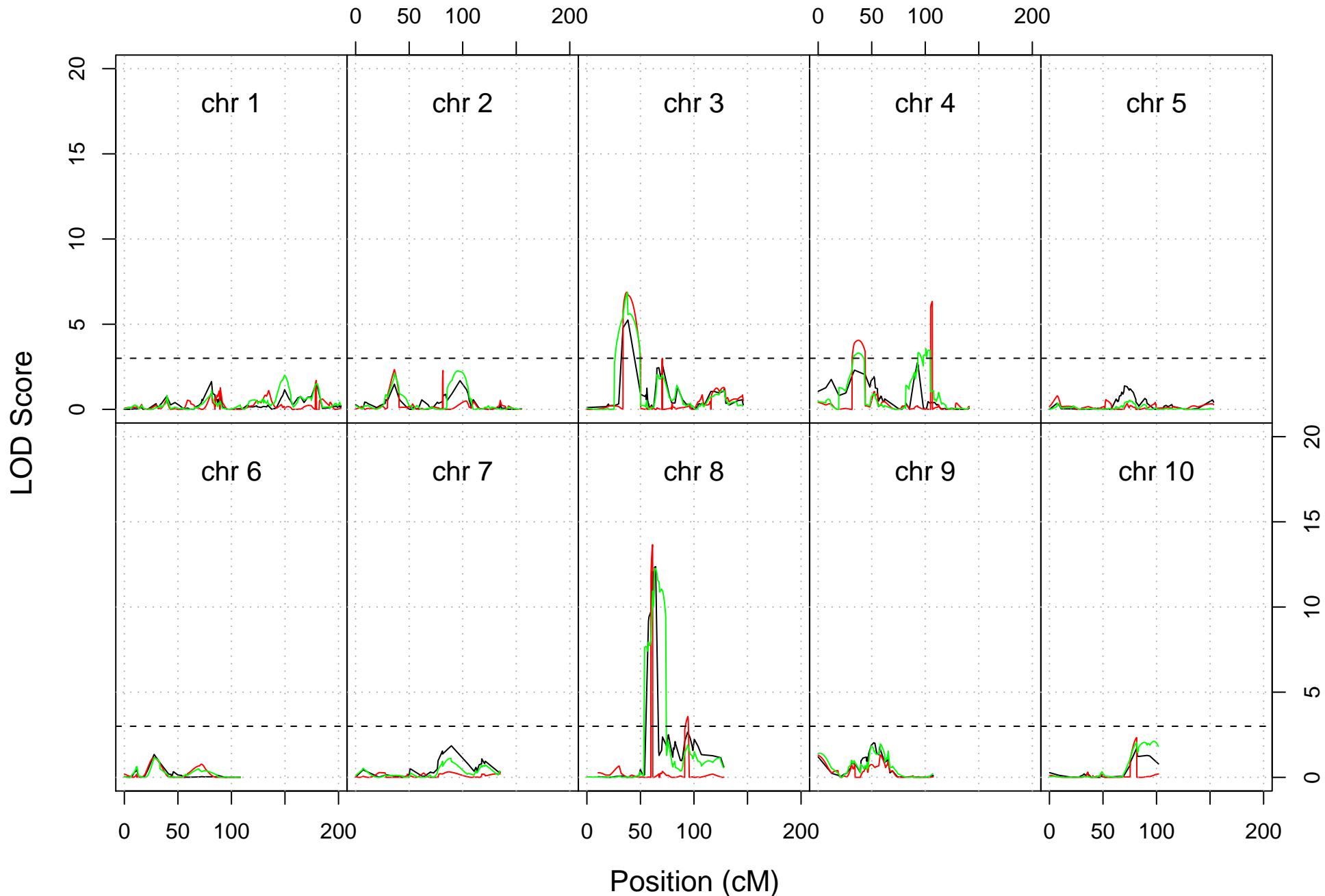
LOD Scores for OH7B population, days to anthesis

ICIM = red, CIM = green, Stepwise = black



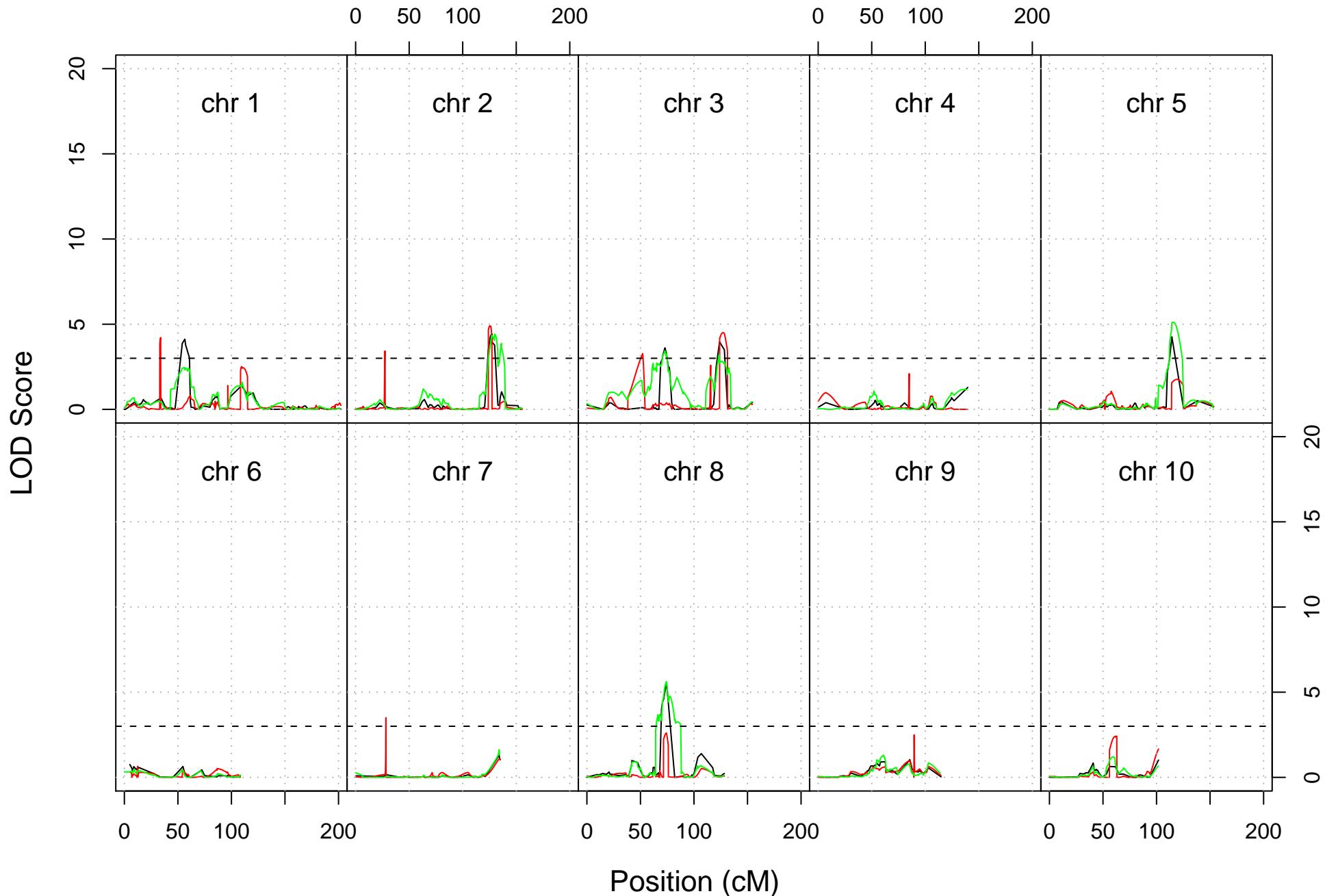
LOD Scores for P39 population, days to anthesis

ICIM = red, CIM = green, Stepwise = black



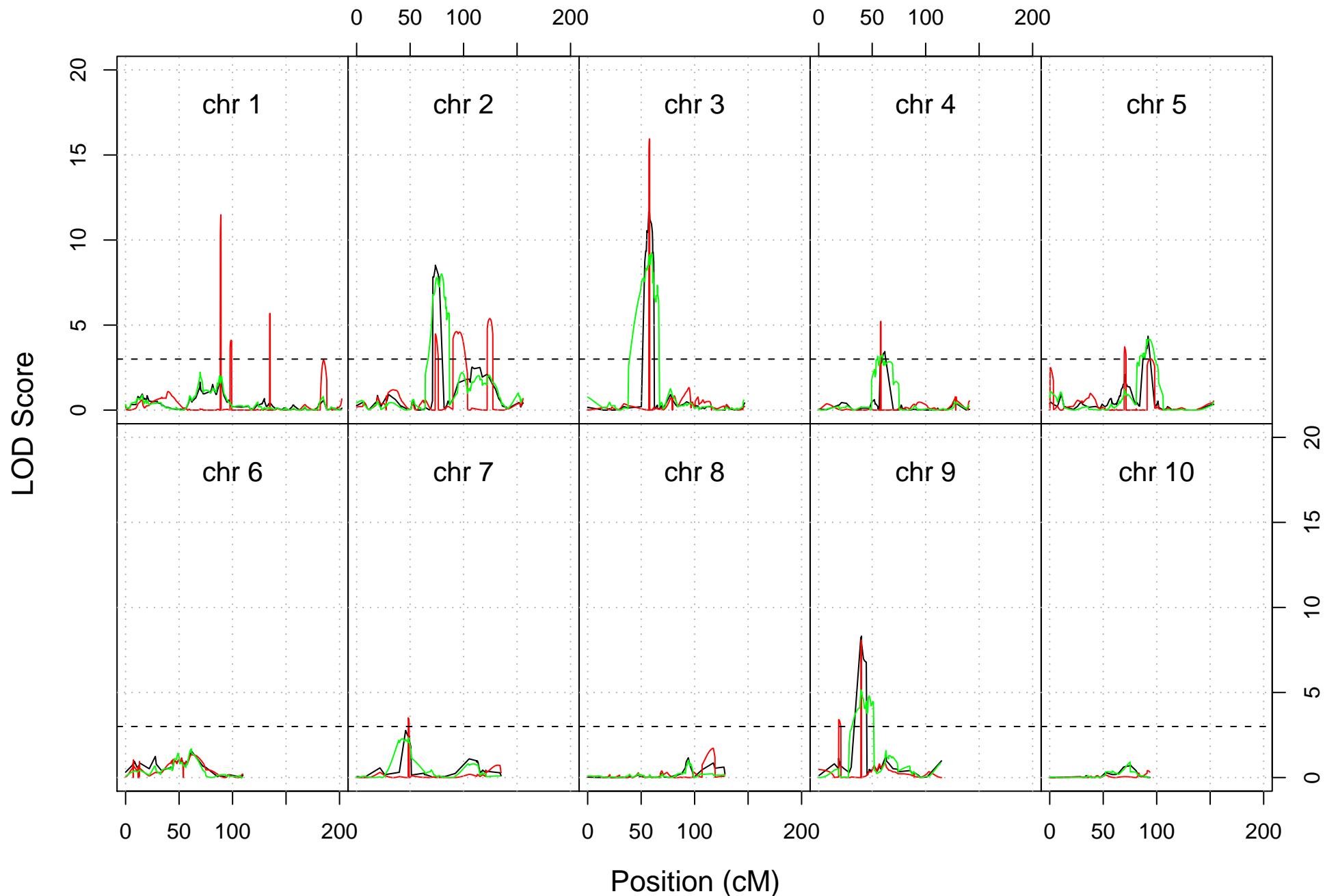
LOD Scores for TX303 population, days to anthesis

ICIM = red, CIM = green, Stepwise = black



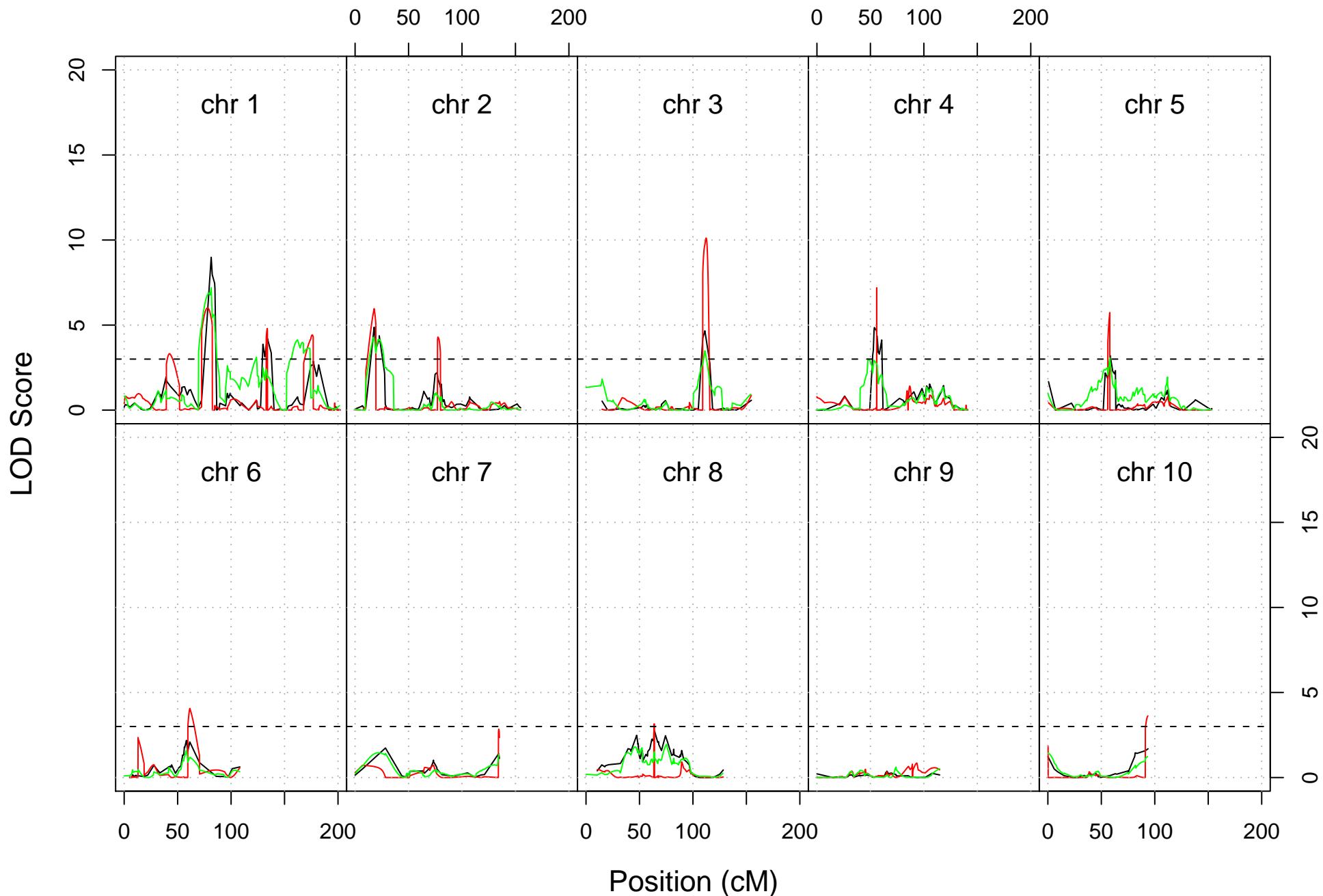
LOD Scores for TZI8 population, days to anthesis

ICIM = red, CIM = green, Stepwise = black



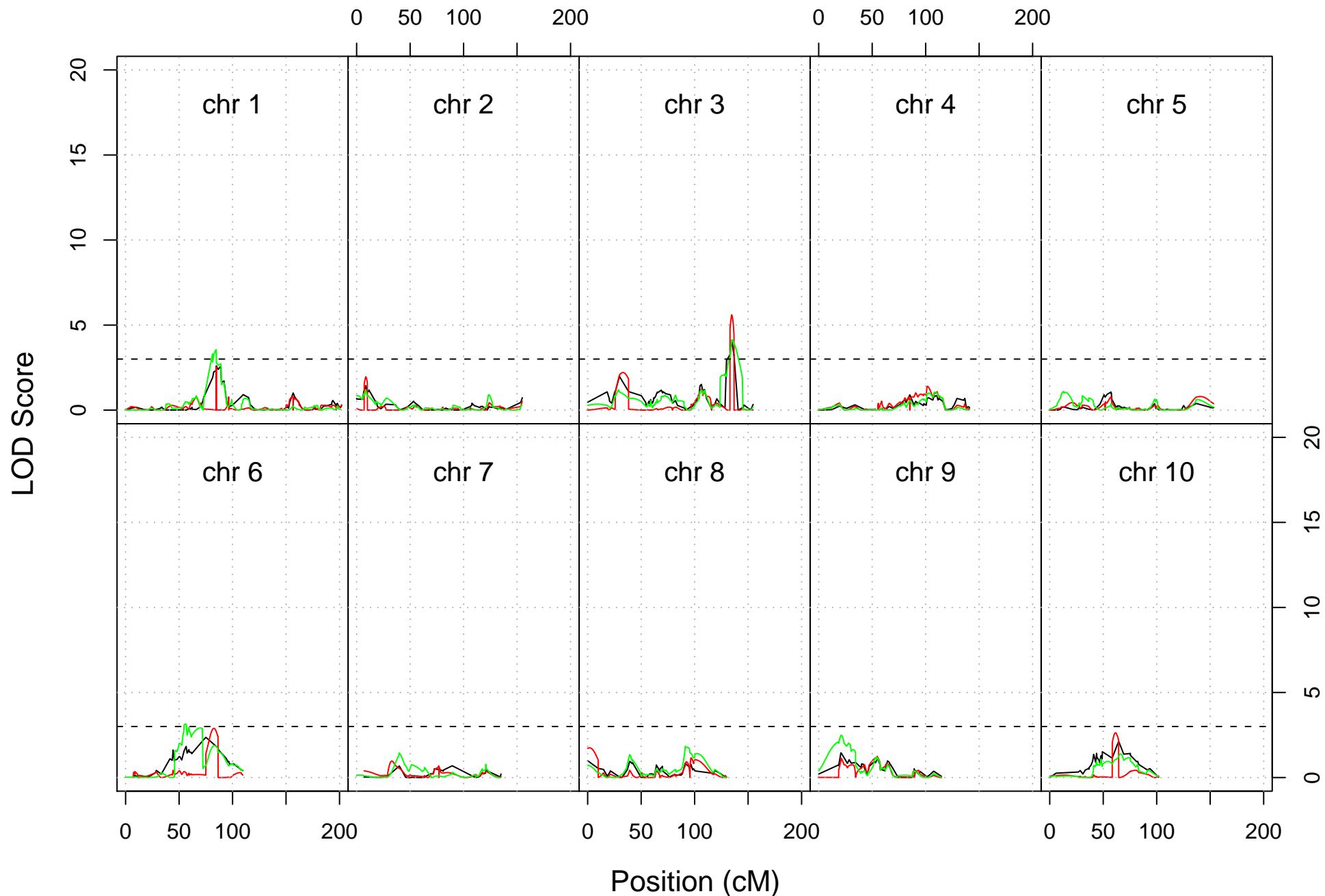
LOD Scores for B97 population, days to silk

ICIM = red, CIM = green, Stepwise = black



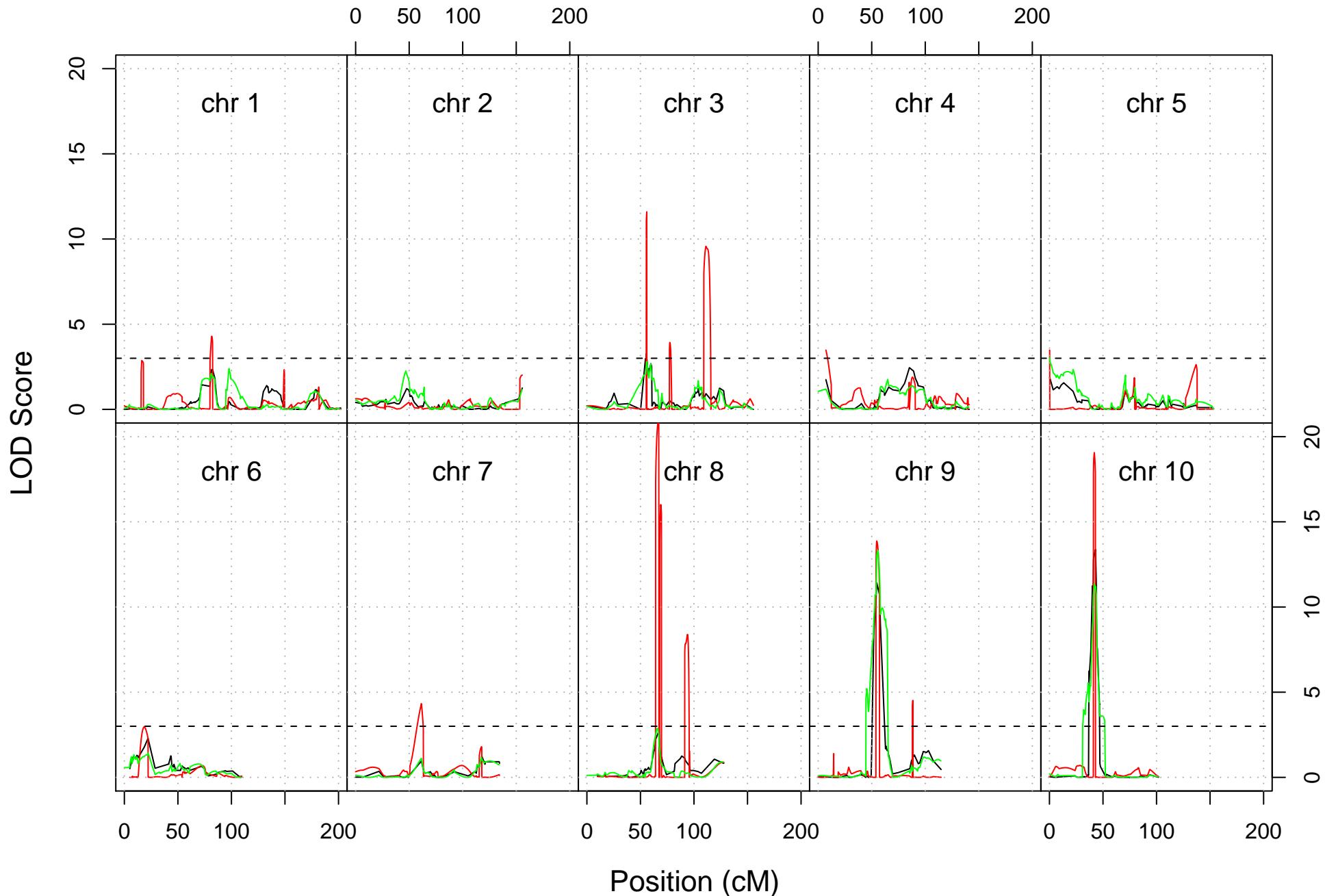
LOD Scores for CML103 population, days to silk

ICIM = red, CIM = green, Stepwise = black



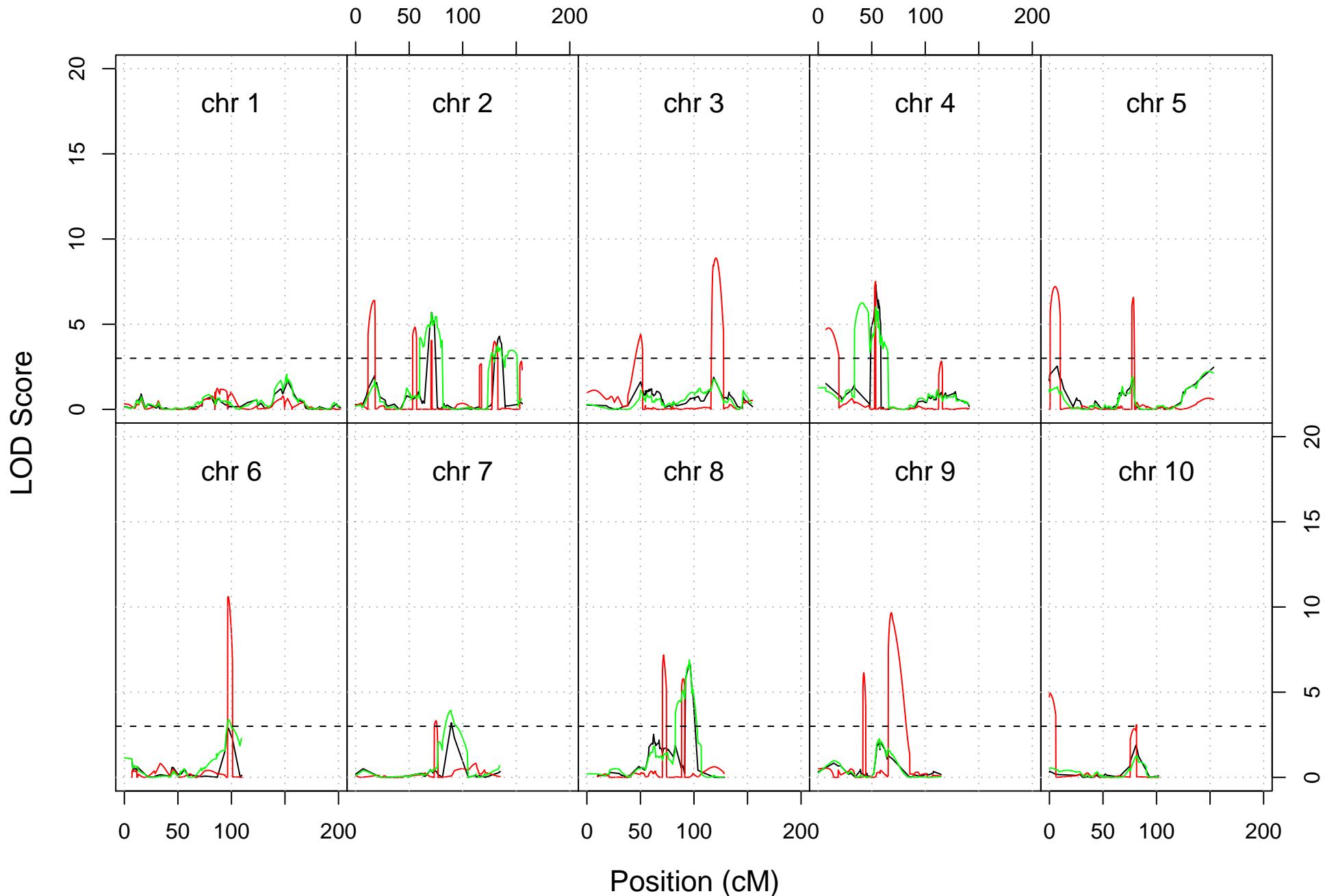
LOD Scores for CML228 population, days to silk

ICIM = red, CIM = green, Stepwise = black



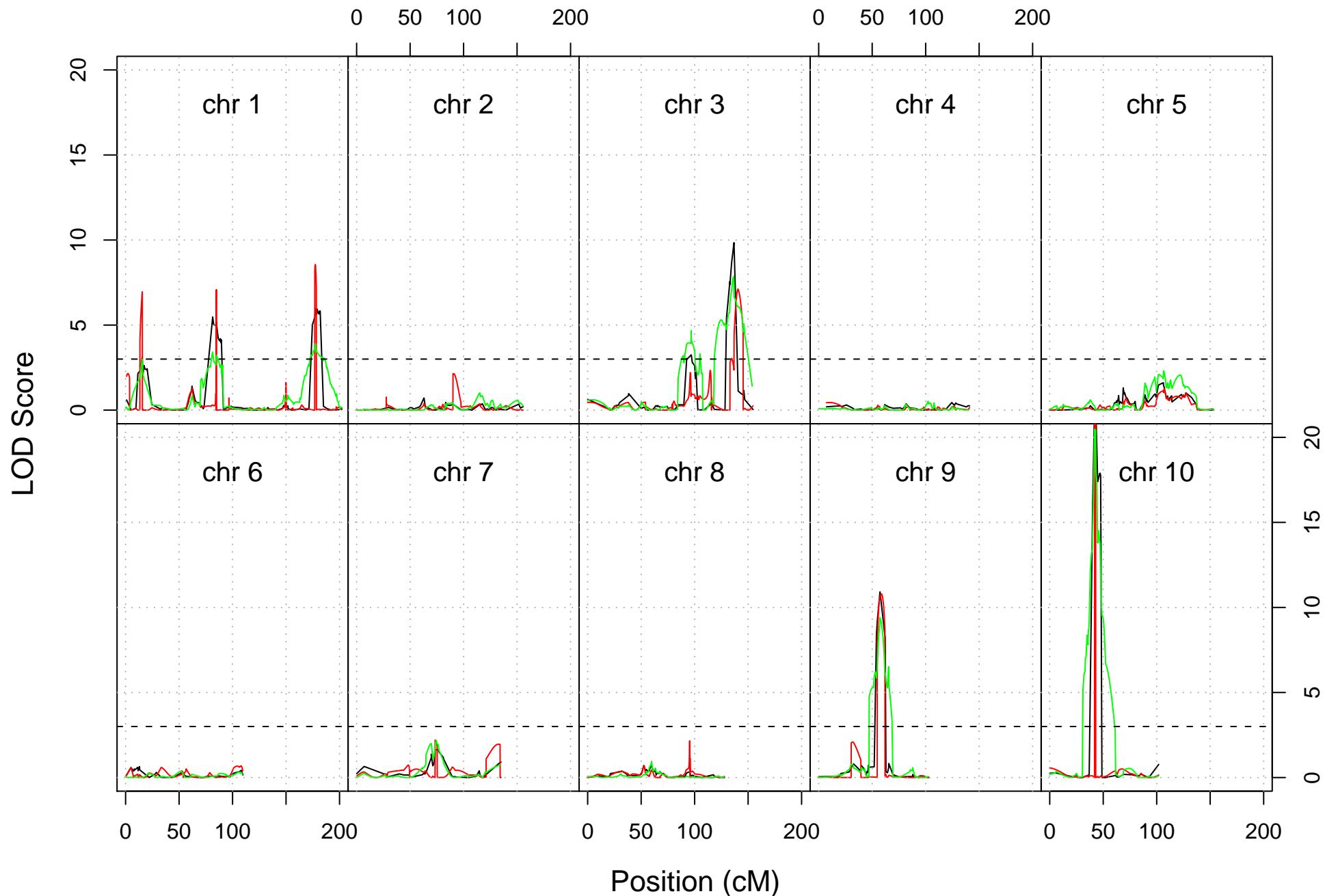
LOD Scores for CML247 population, days to silk

ICIM = red, CIM = green, Stepwise = black



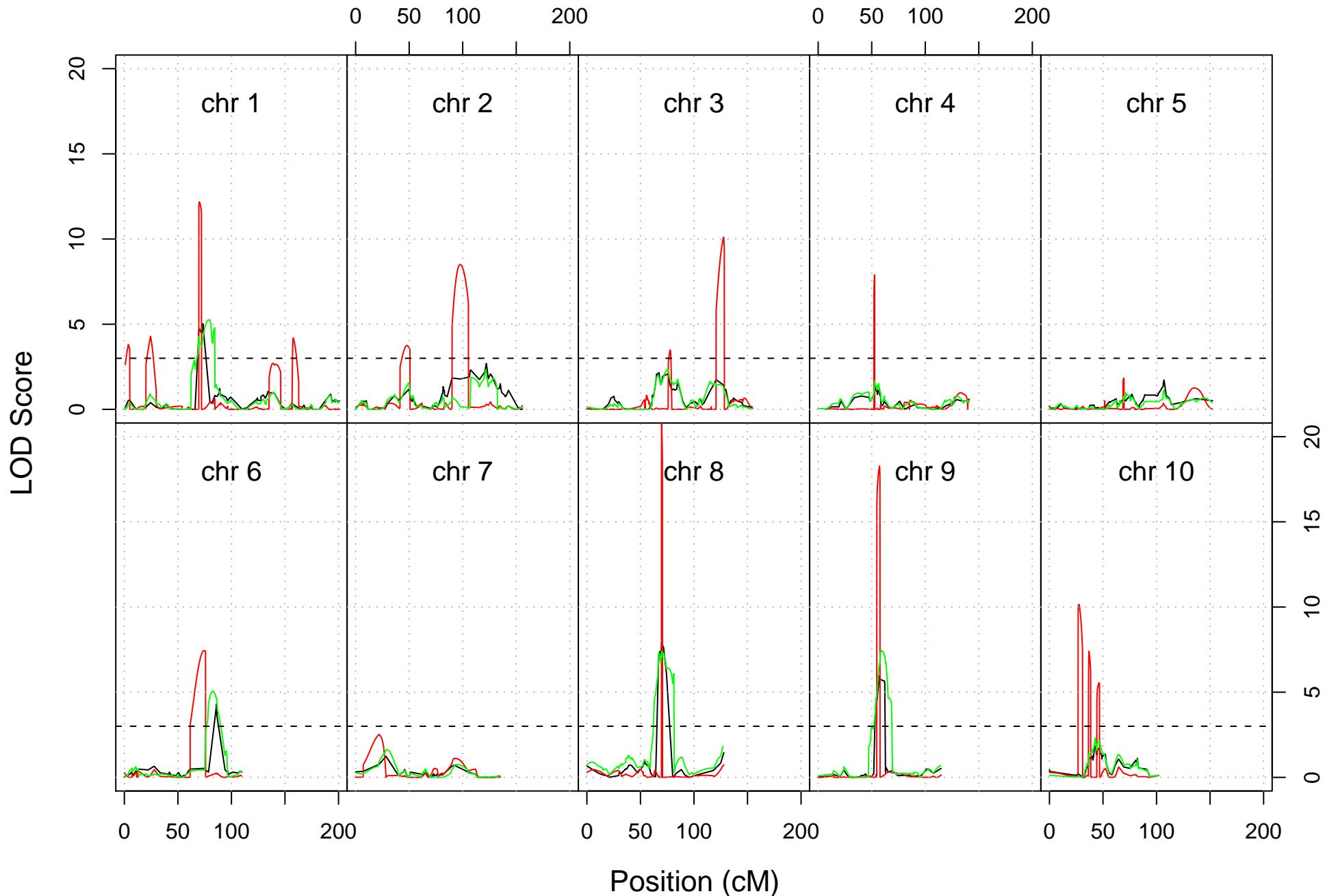
LOD Scores for CML277 population, days to silk

ICIM = red, CIM = green, Stepwise = black



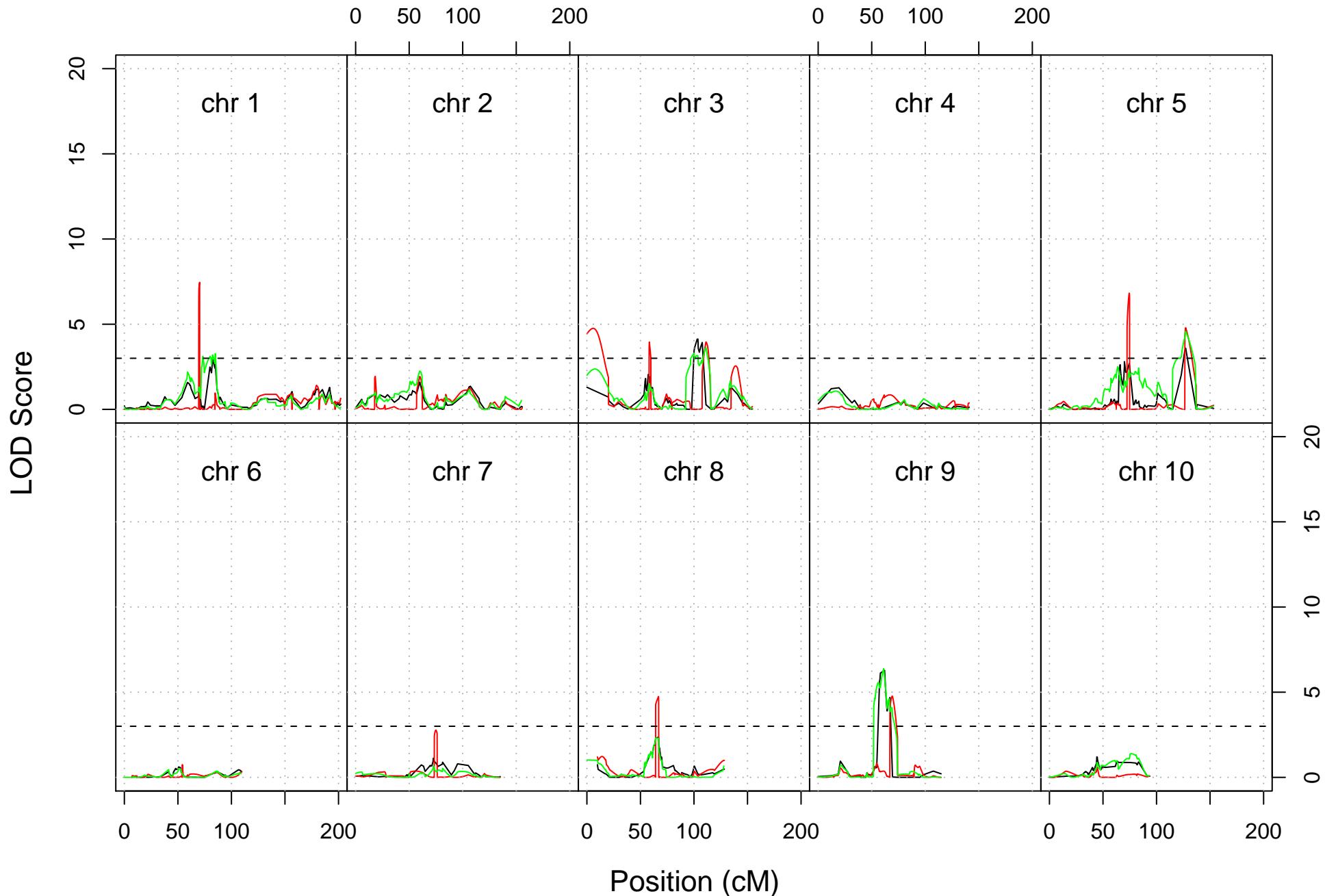
LOD Scores for CML322 population, days to silk

ICIM = red, CIM = green, Stepwise = black



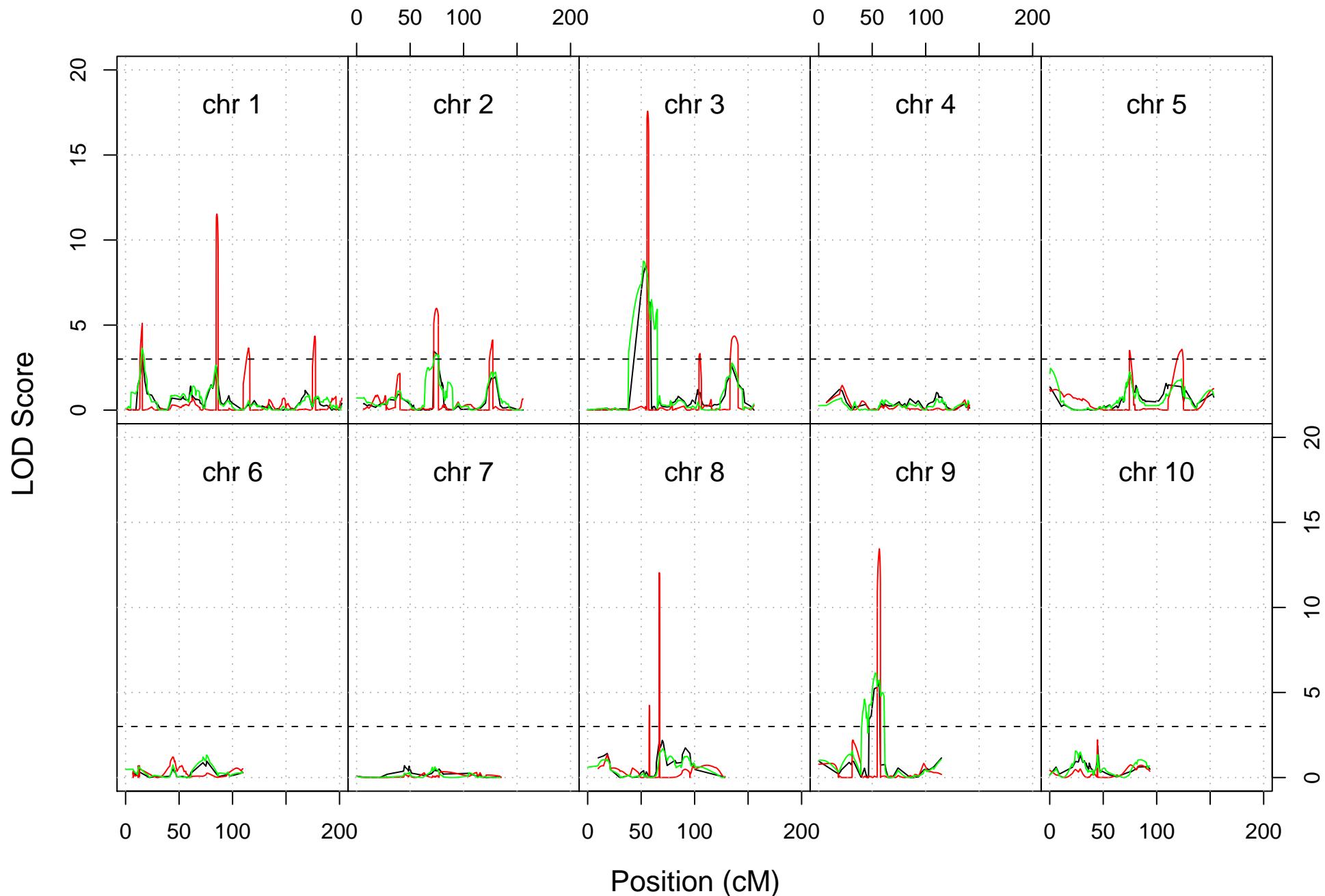
LOD Scores for CML333 population, days to silk

ICIM = red, CIM = green, Stepwise = black



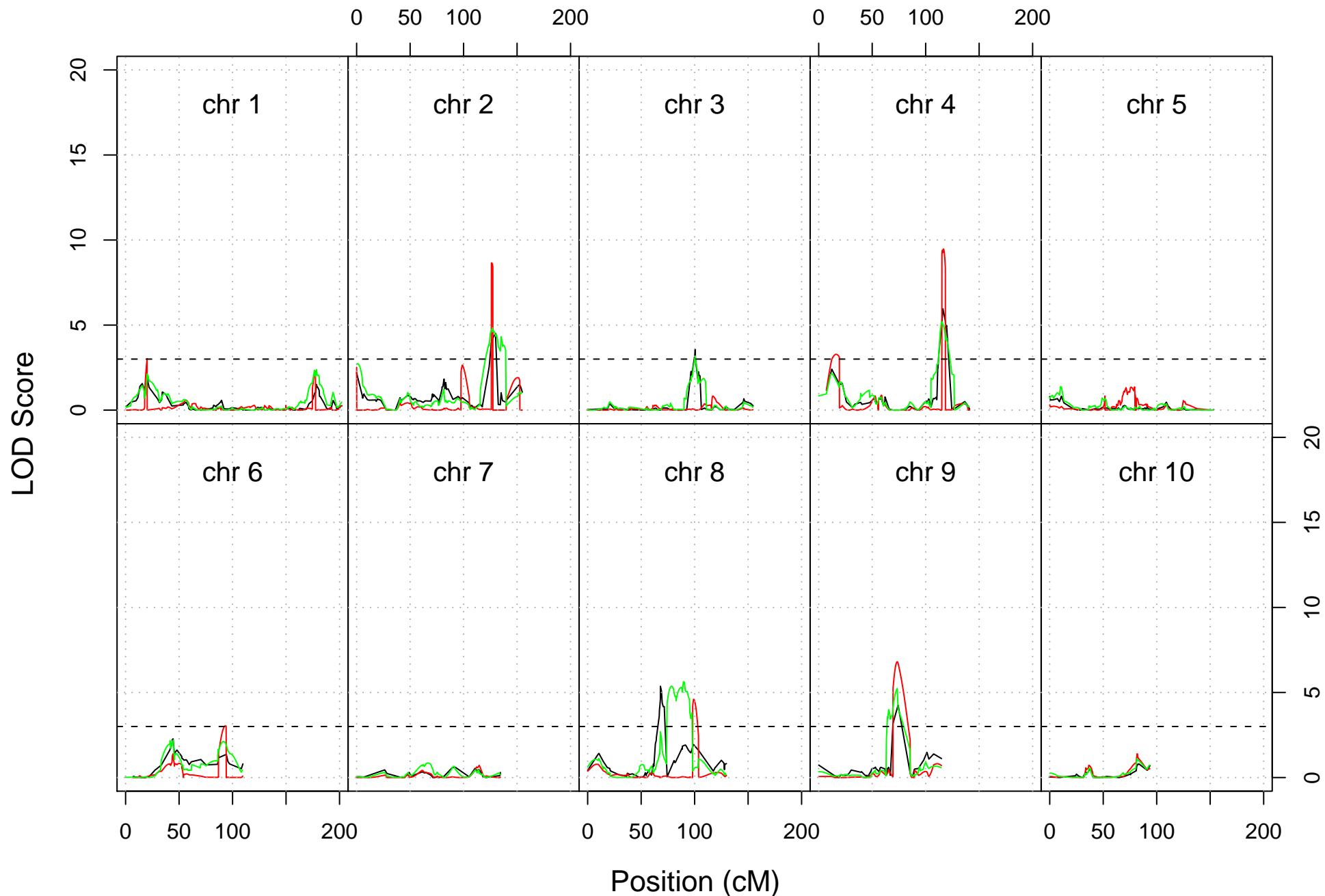
LOD Scores for CML52 population, days to silk

ICIM = red, CIM = green, Stepwise = black



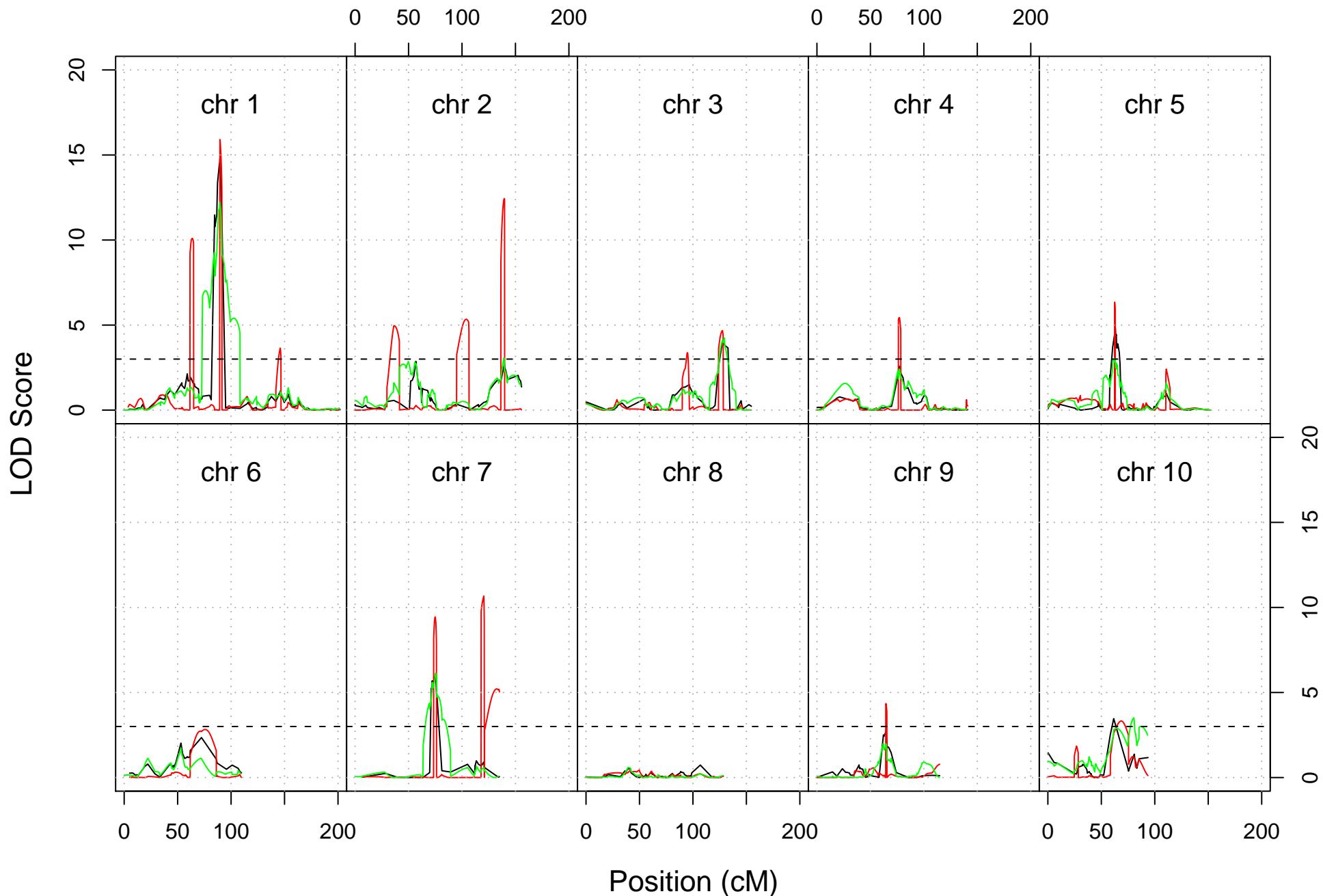
LOD Scores for CML69 population, days to silk

ICIM = red, CIM = green, Stepwise = black



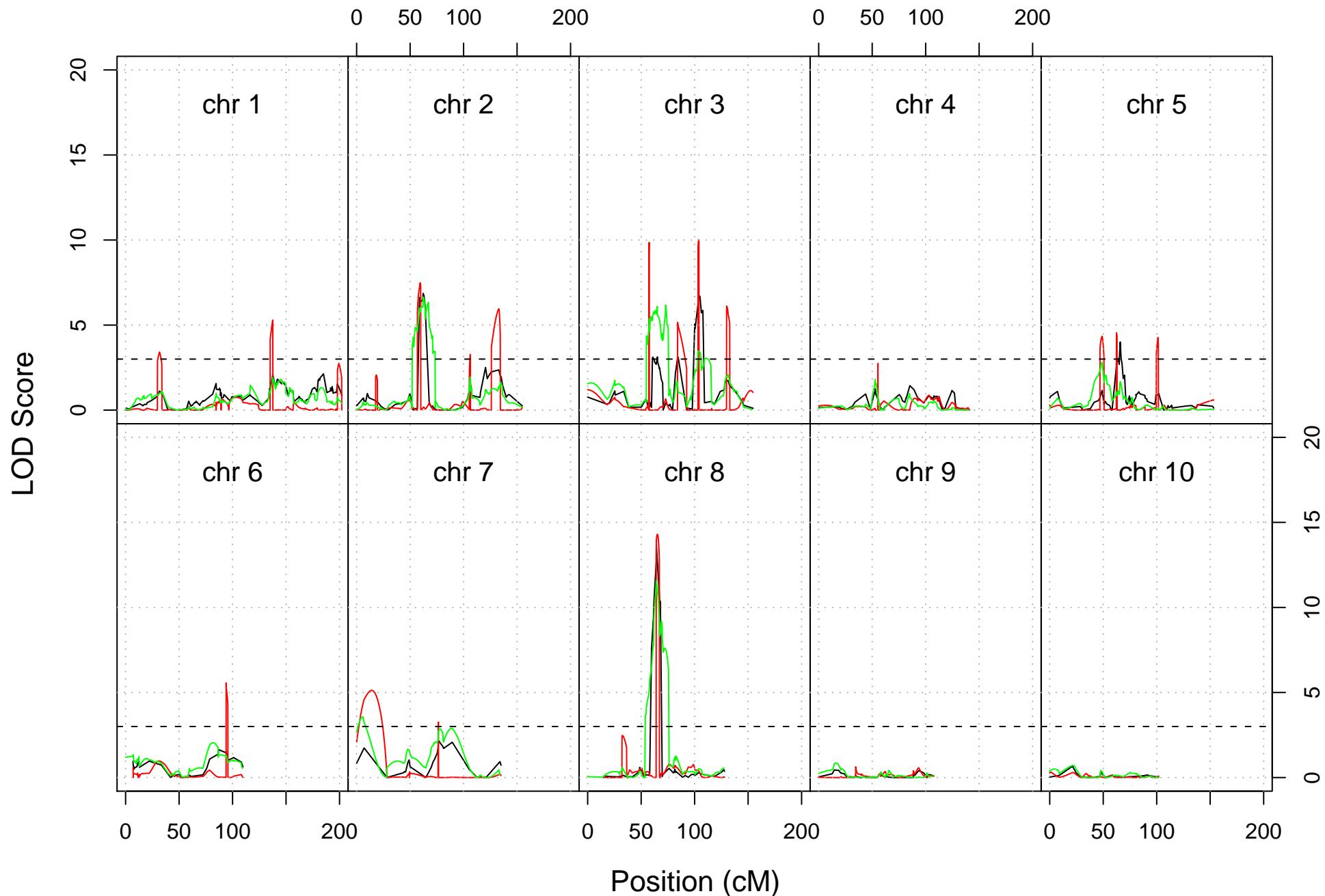
LOD Scores for HP301 population, days to silk

ICIM = red, CIM = green, Stepwise = black



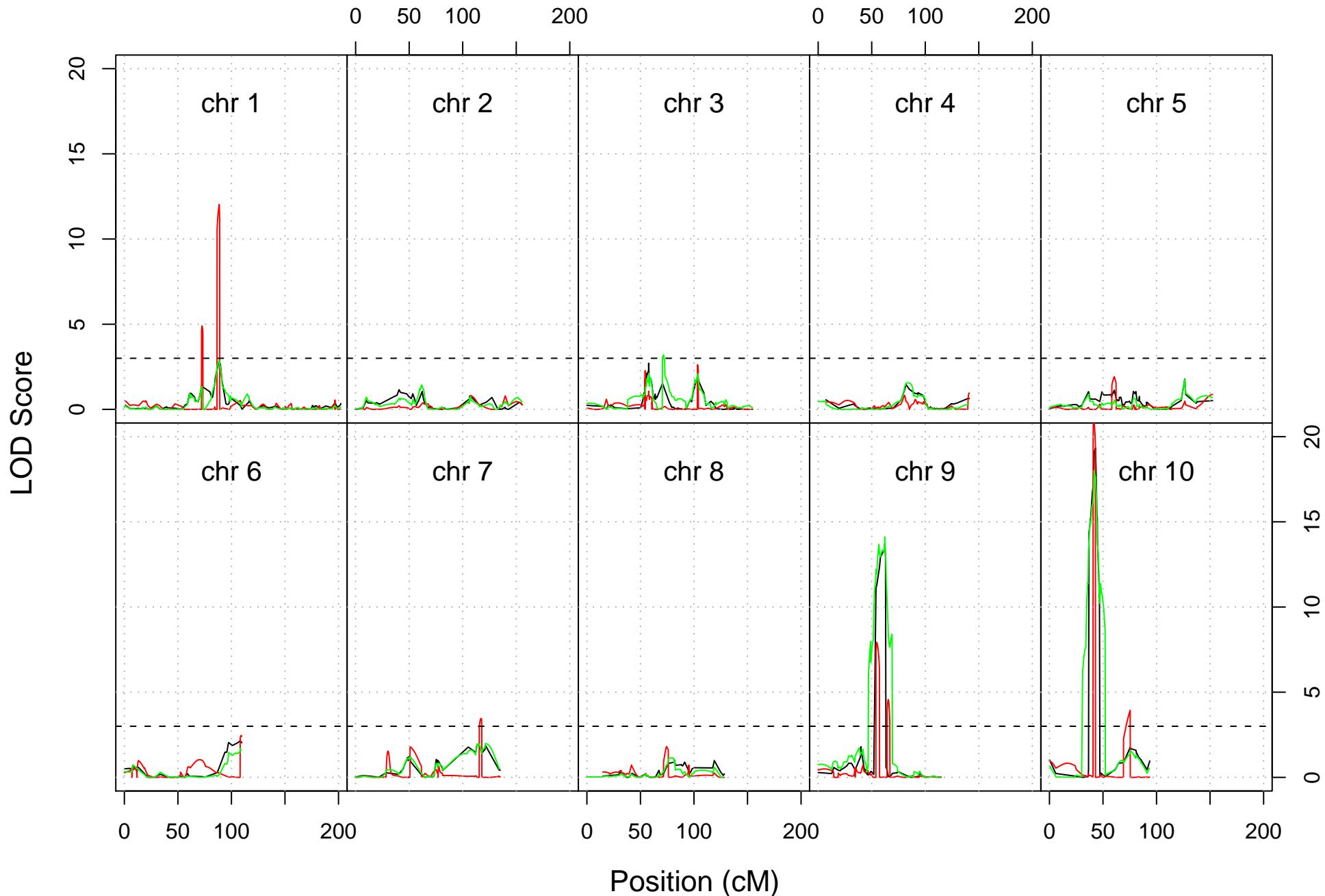
LOD Scores for IL14H population, days to silk

ICIM = red, CIM = green, Stepwise = black



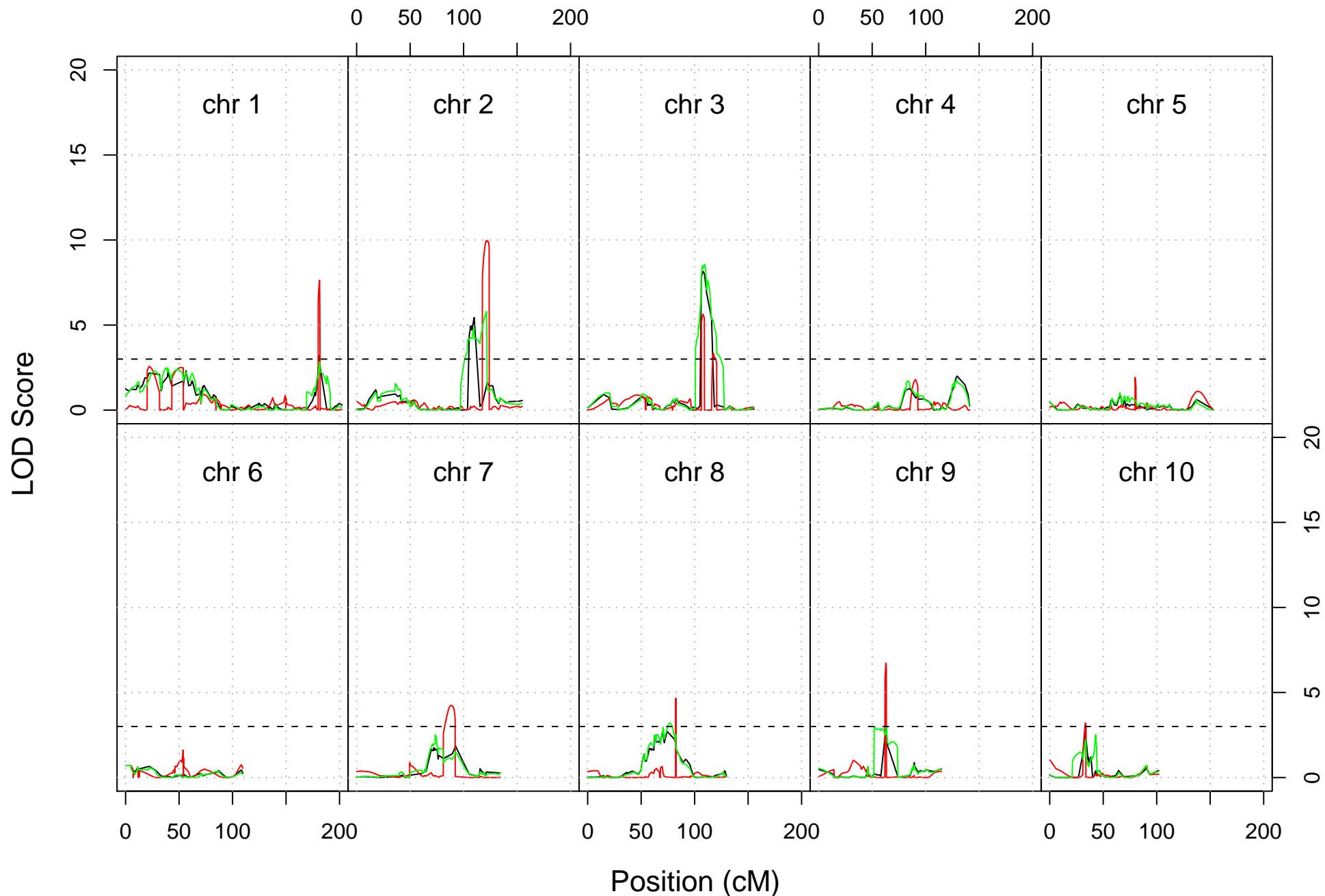
LOD Scores for KI11 population, days to silk

ICIM = red, CIM = green, Stepwise = black



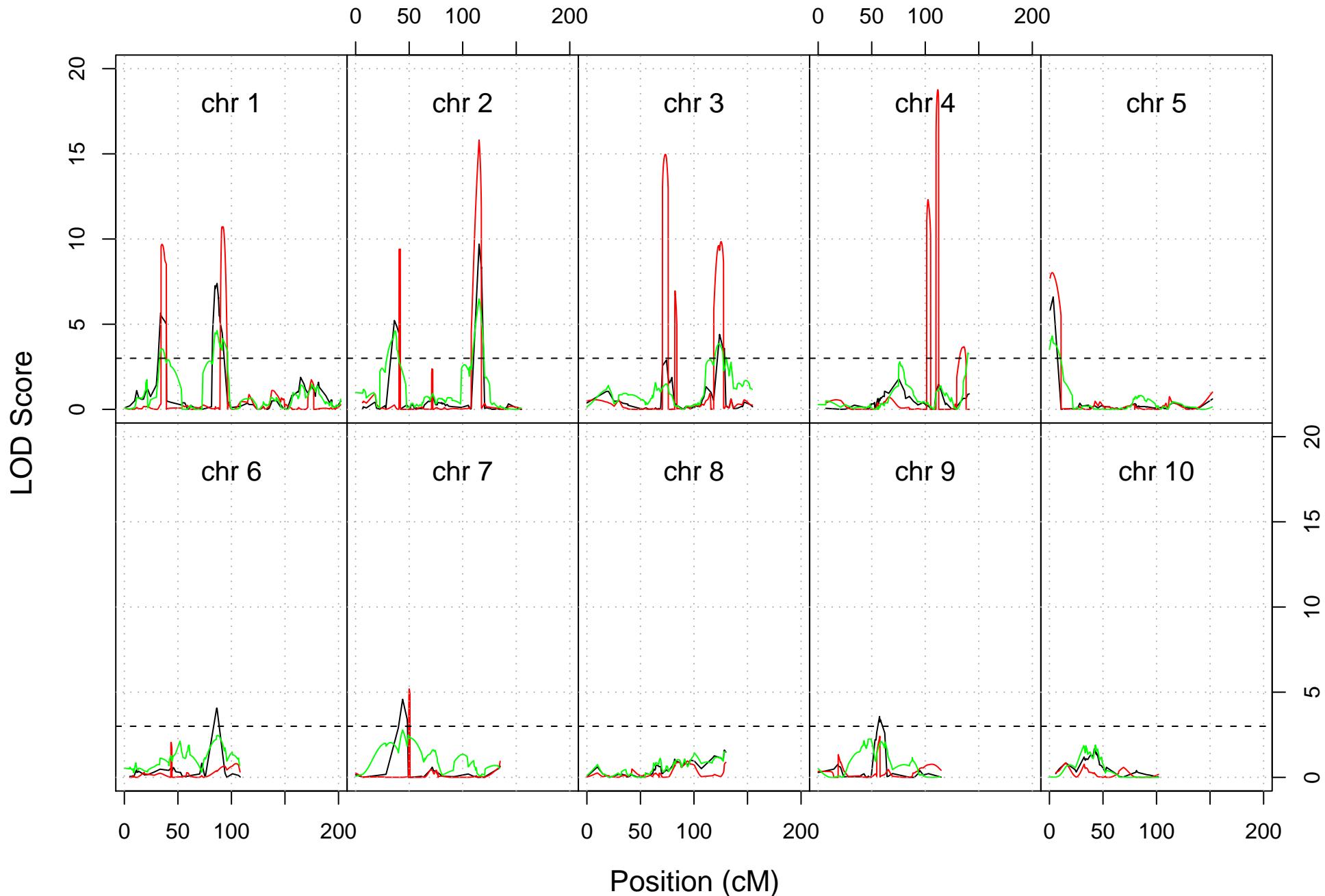
LOD Scores for KI3 population, days to silk

ICIM = red, CIM = green, Stepwise = black



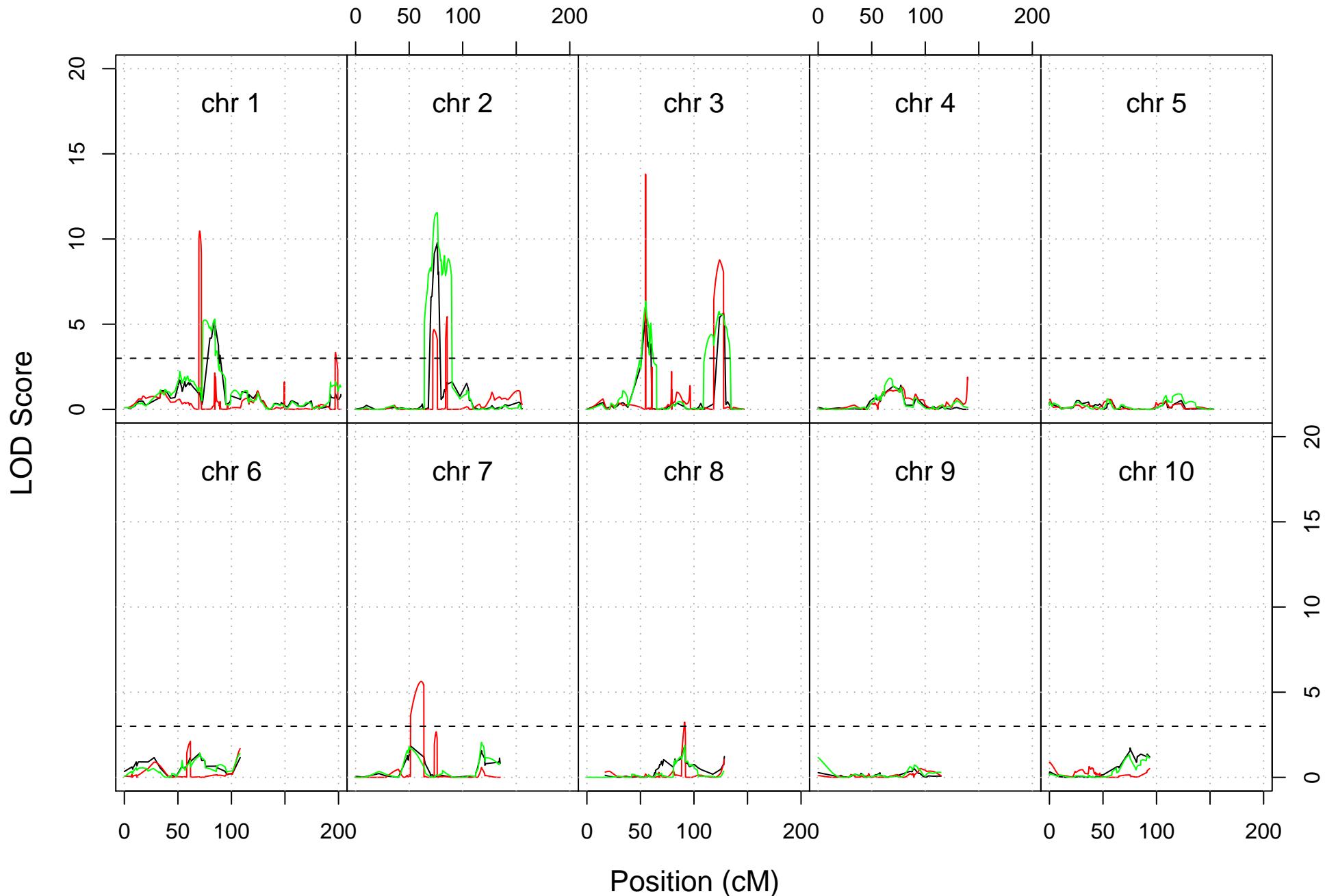
LOD Scores for KY21 population, days to silk

ICIM = red, CIM = green, Stepwise = black



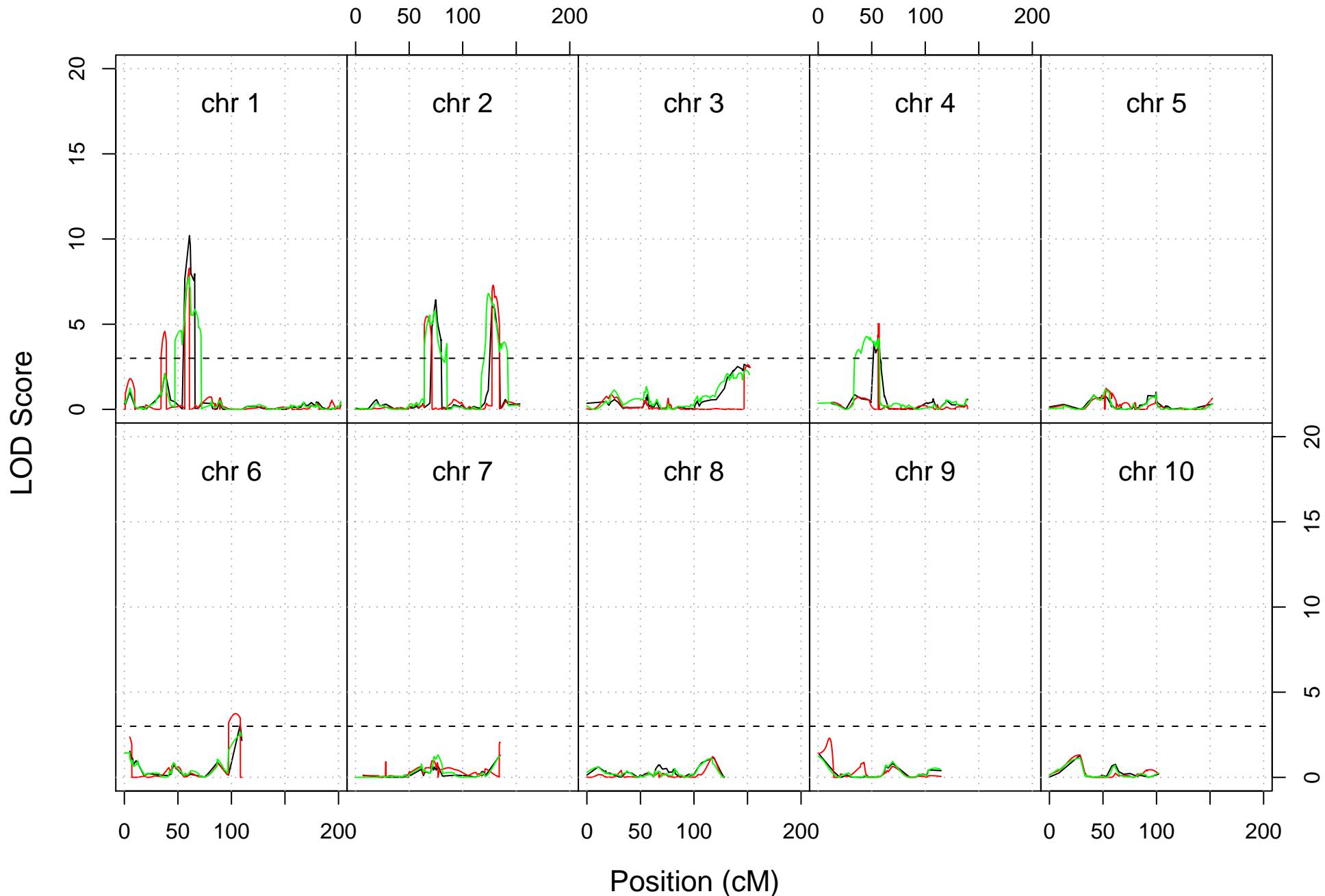
LOD Scores for M162W population, days to silk

ICIM = red, CIM = green, Stepwise = black



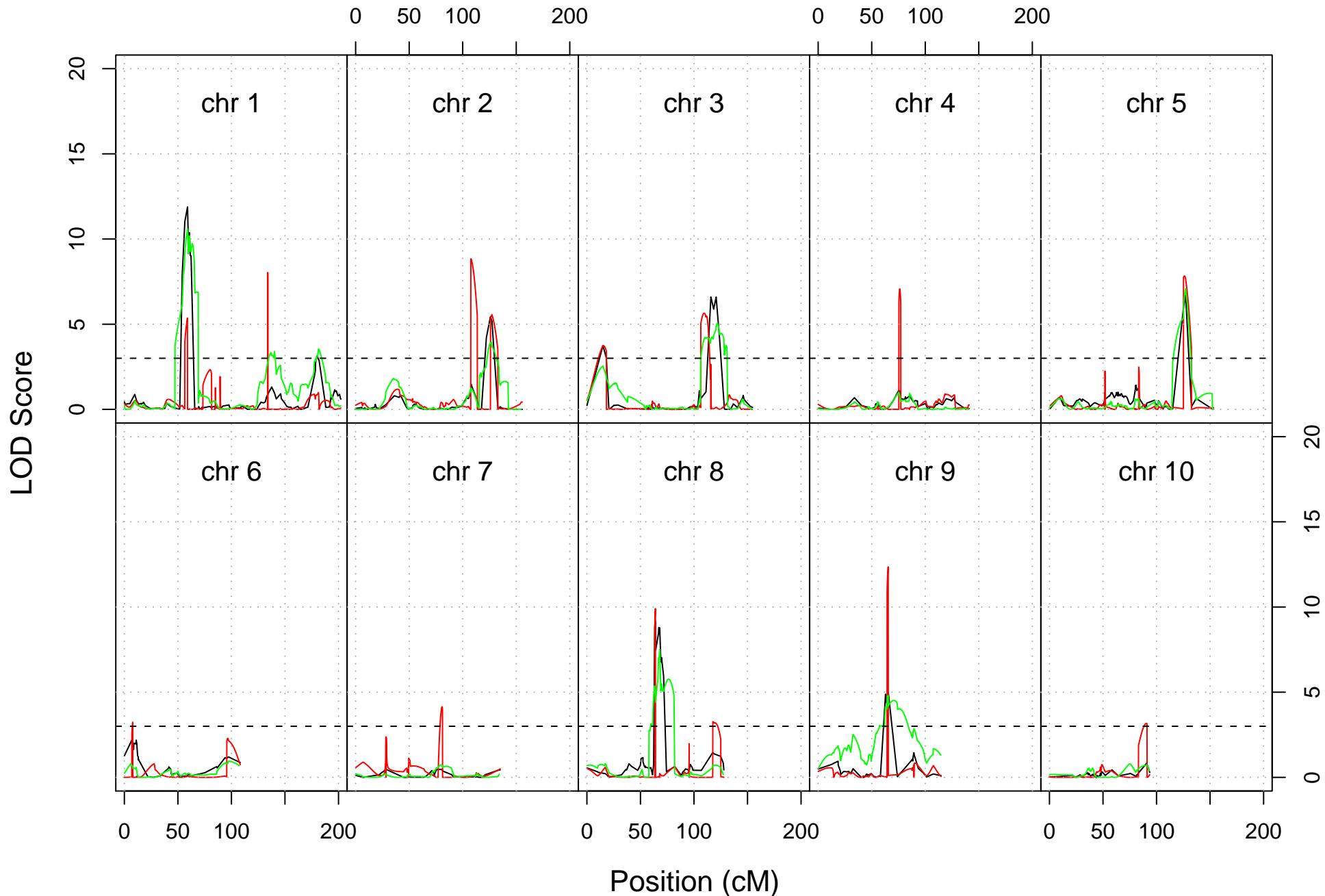
LOD Scores for M37W population, days to silk

ICIM = red, CIM = green, Stepwise = black



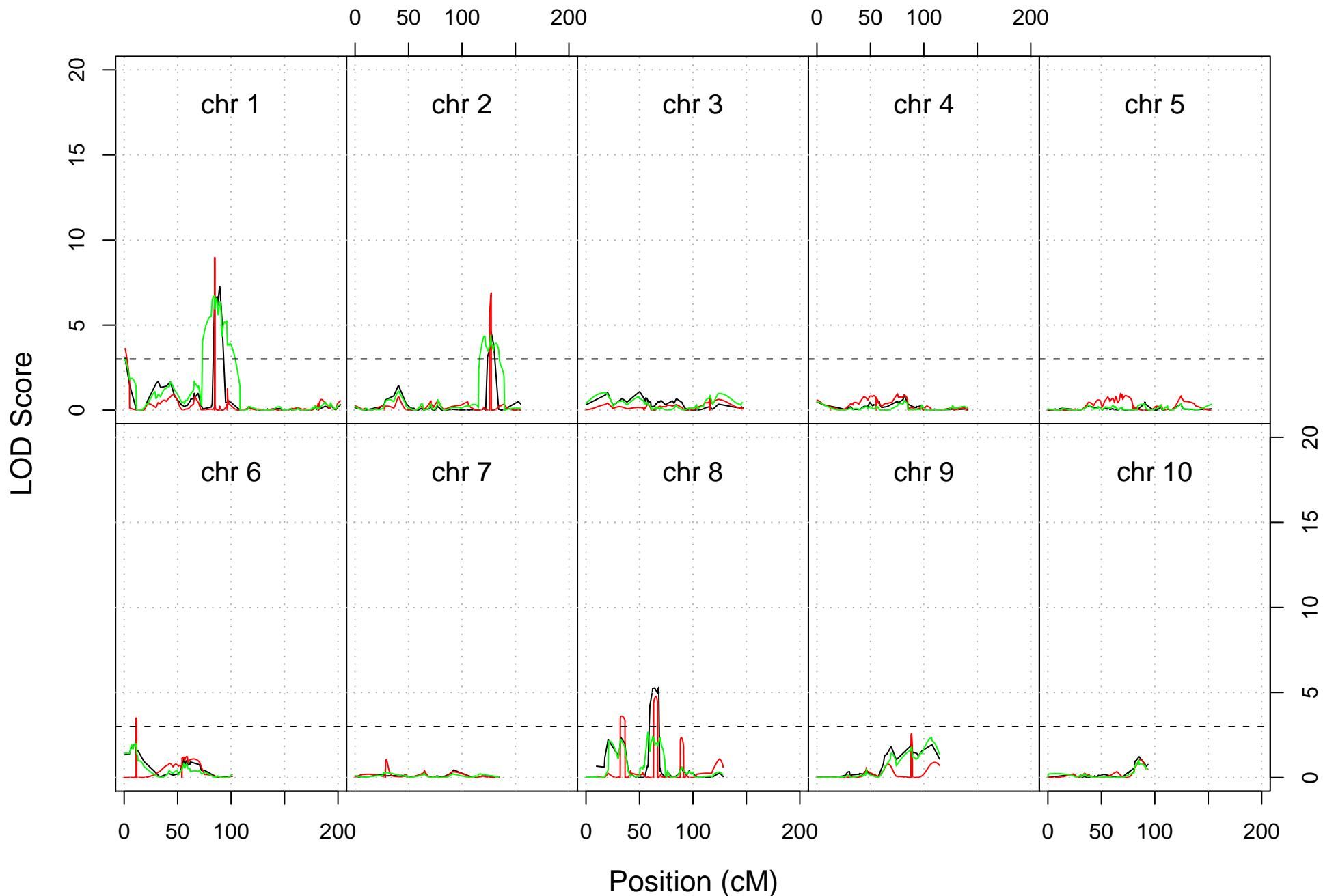
LOD Scores for MO18W population, days to silk

ICIM = red, CIM = green, Stepwise = black



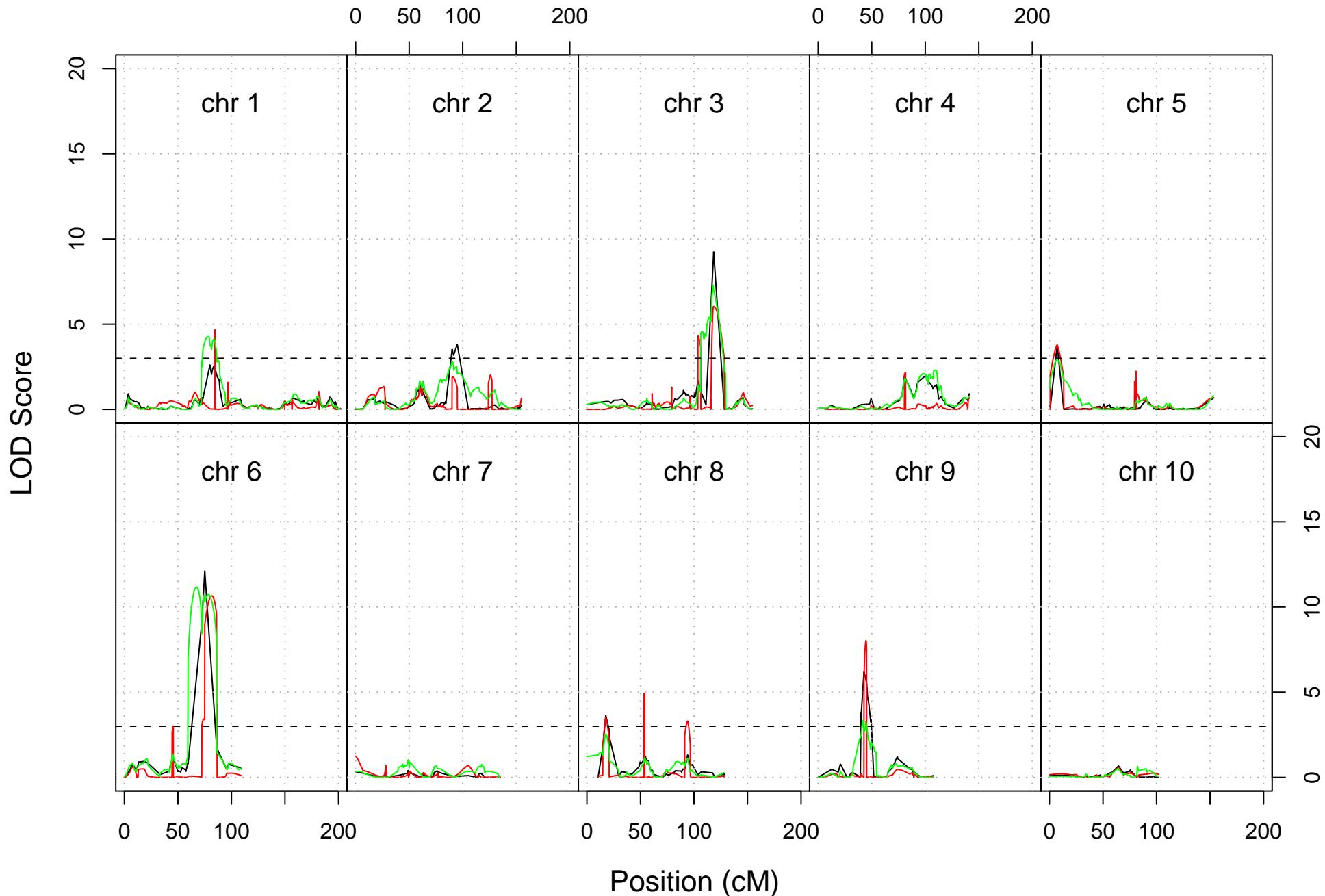
LOD Scores for MS71 population, days to silk

ICIM = red, CIM = green, Stepwise = black



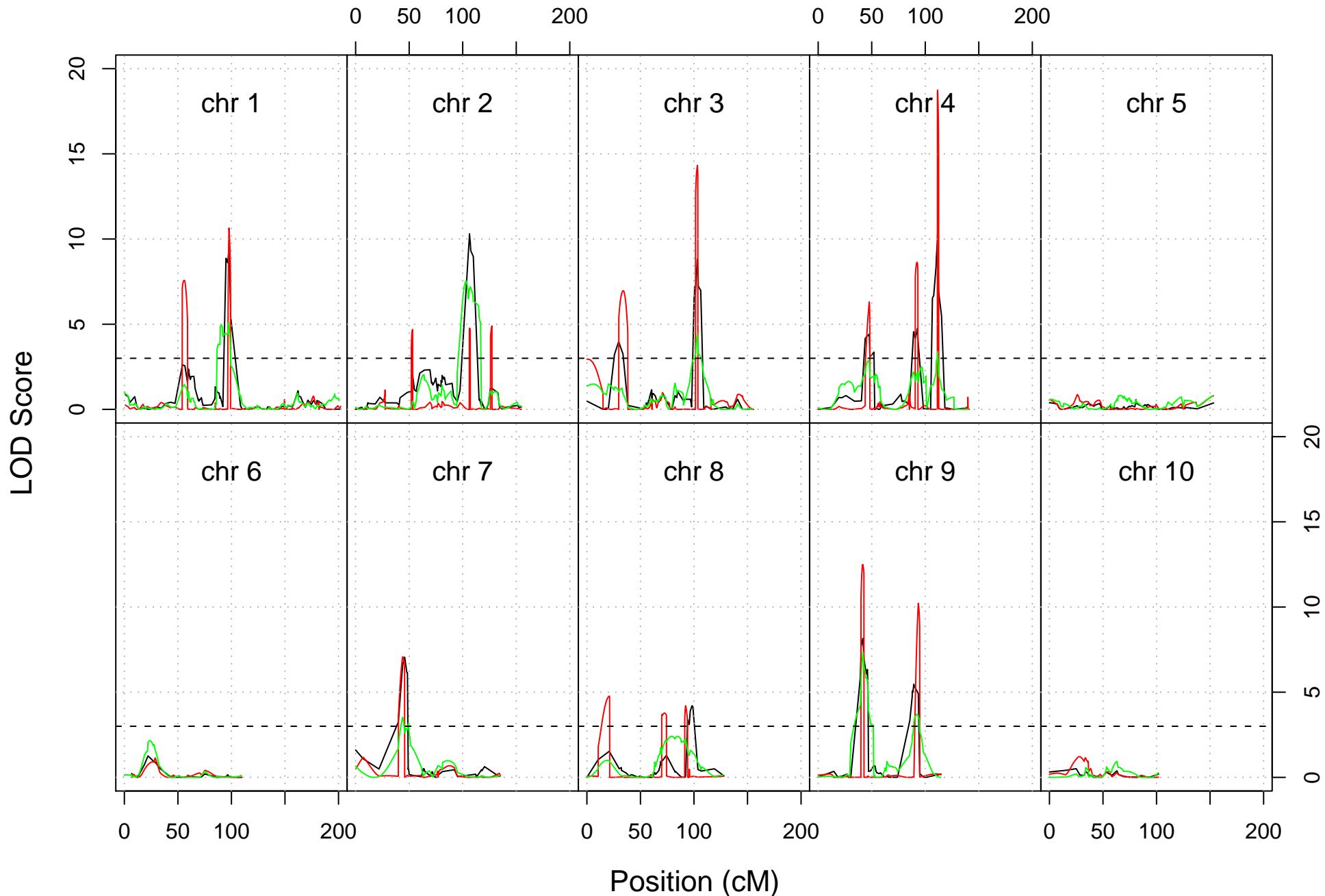
LOD Scores for NC350 population, days to silk

ICIM = red, CIM = green, Stepwise = black



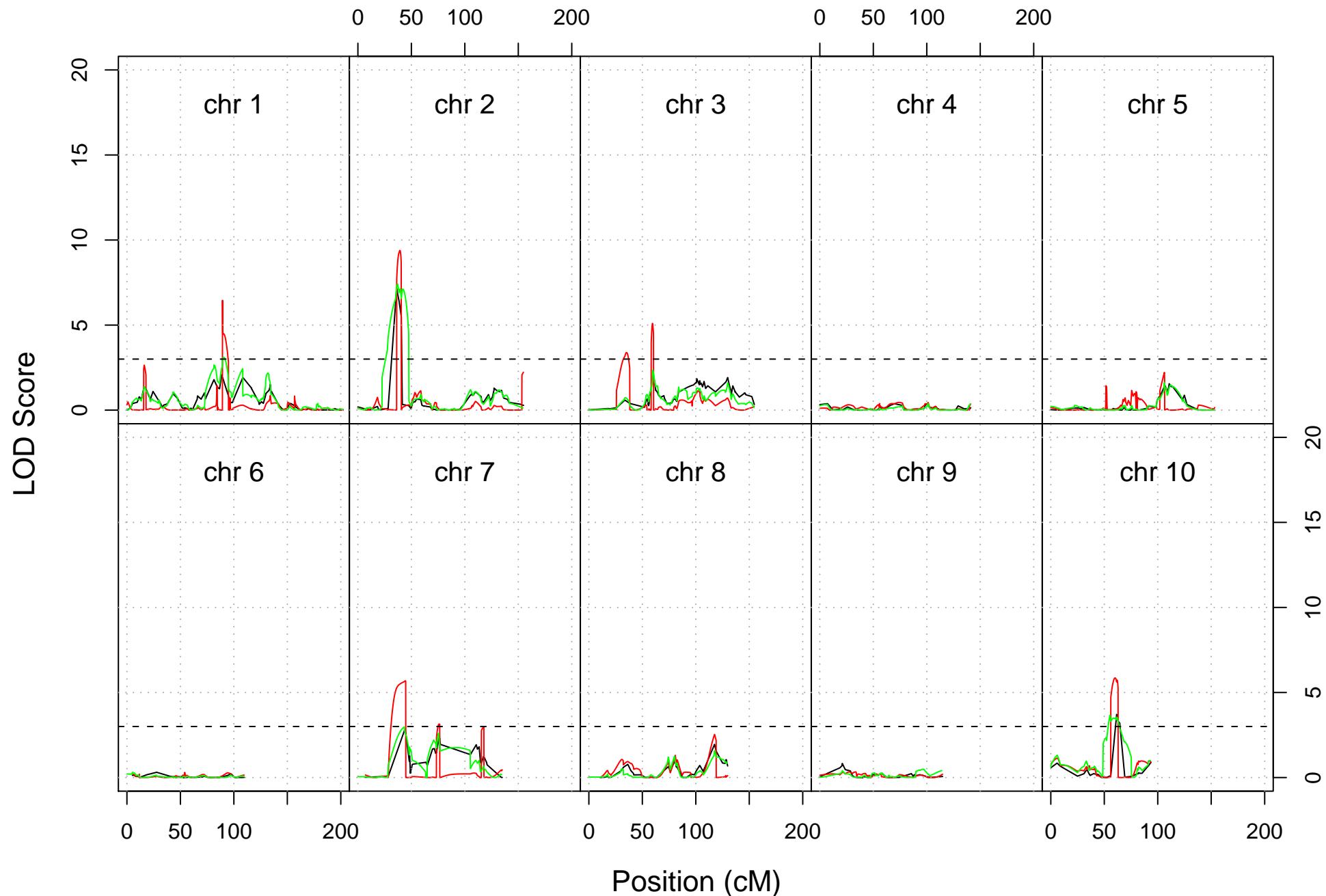
LOD Scores for NC358 population, days to silk

ICIM = red, CIM = green, Stepwise = black



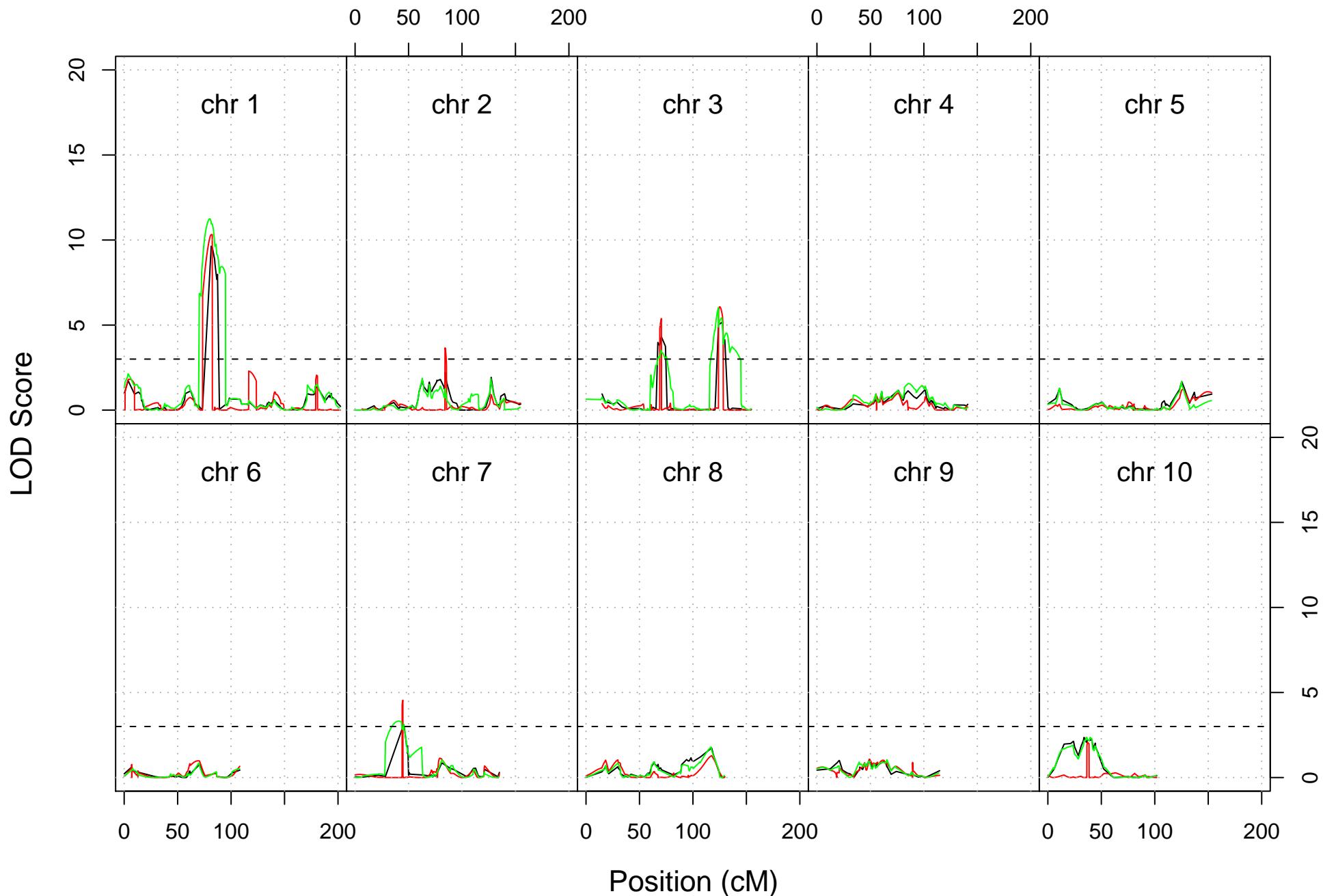
LOD Scores for OH43 population, days to silk

ICIM = red, CIM = green, Stepwise = black



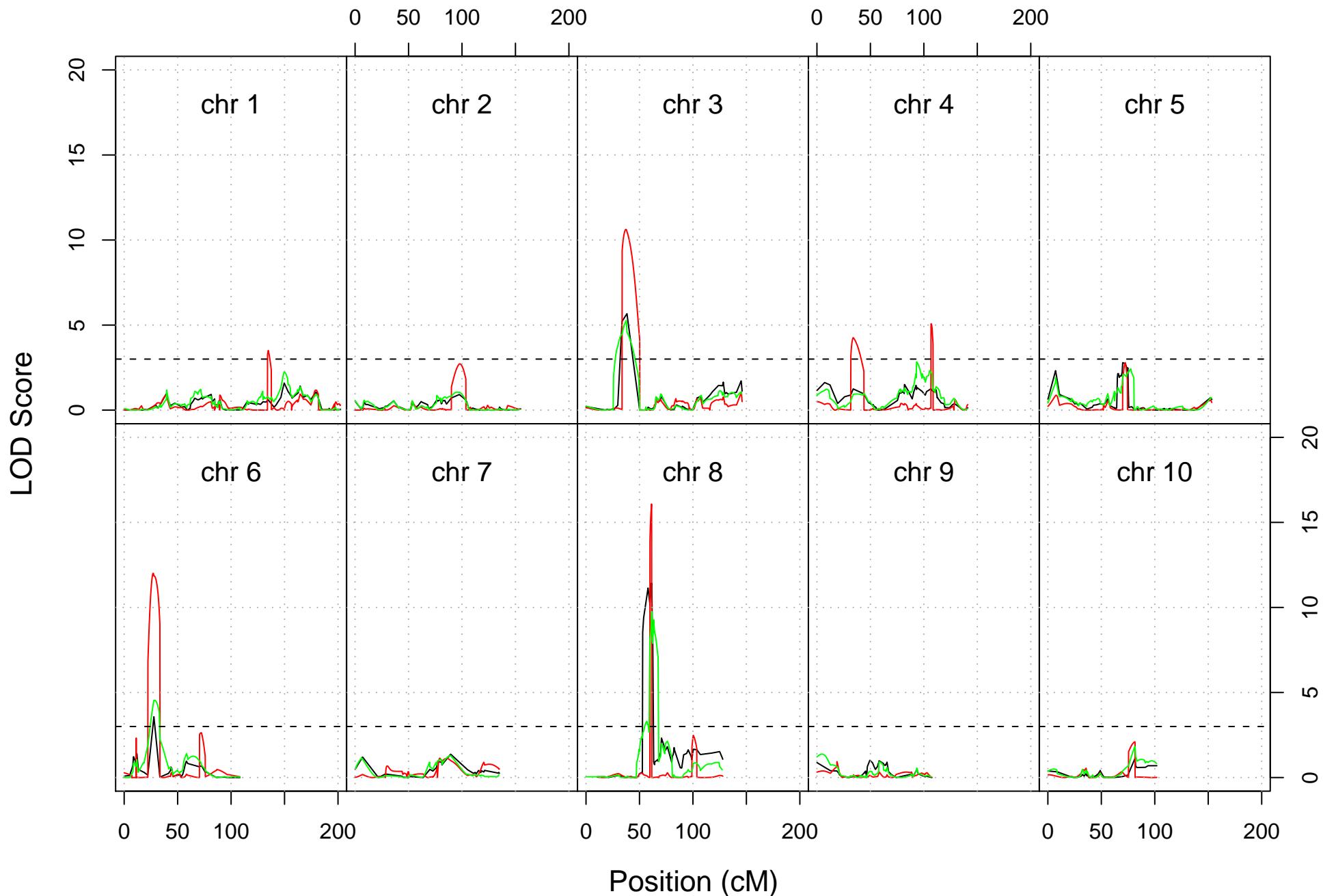
LOD Scores for OH7B population, days to silk

ICIM = red, CIM = green, Stepwise = black



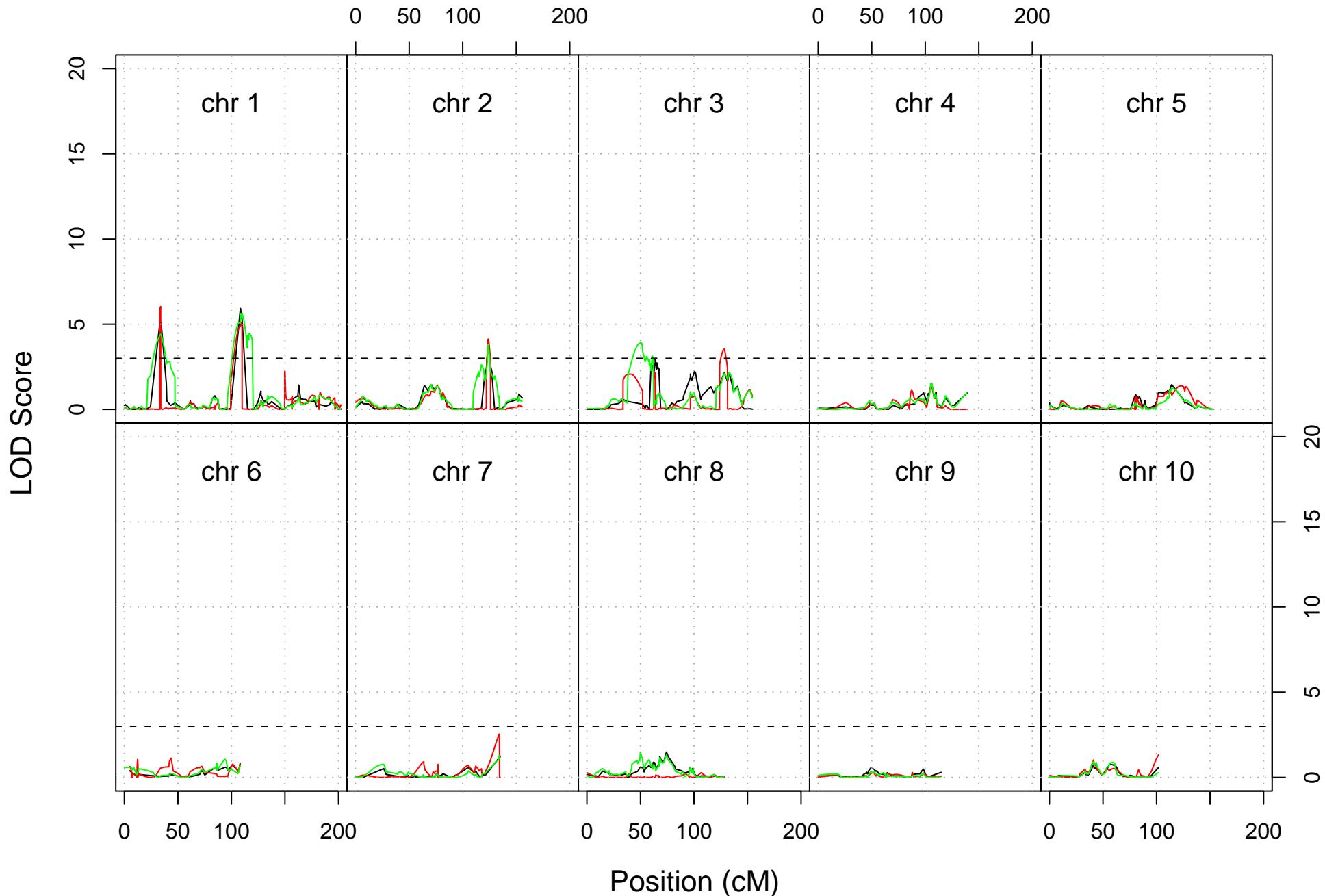
LOD Scores for P39 population, days to silk

ICIM = red, CIM = green, Stepwise = black



LOD Scores for TX303 population, days to silk

ICIM = red, CIM = green, Stepwise = black



LOD Scores for TZI8 population, days to silk

ICIM = red, CIM = green, Stepwise = black

