

Lecture 4: Molecular evolution, phylogeny and homology

Prof. Anne-Florence Bitbol



Schedule of this class

date	week	lectures	exercises	teacher		
18 feb	1					
25 feb	2	Structural genomics	R exercises	Jacques Rougemont		
4 mar	3					
11 mar	4		R exercises	Anne-Florence Bitbol		
18 mar	5	Population genetics				
25 mar	6		Assignment 1: 25%			
1 apr	7		R exercises	Raphaëlle Luisier		
8 apr	8	Gene expression				
15 apr	9					
22 a	pr	holidays				
29 apr	10	Gene expression	Assignment 2: 25%	Raphaëlle Luisier		
6 may	11		Mini-projects	Jacques Rougemont		
13 may	12	Daniel Cara alternativ				
20 may	13	Regulation, chromatin				
27 may	14					
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Report: 50%

Schedule of this class

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Lecture 1: Feb 18
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Lecture 2: Feb 25

Lecture 3: March 4

Lecture 4: March 11

Lecture 5: March 18 – Assignment 1 available on March 20

Lecture 6: March 25 – Problem class devoted to assignment 1; deadline on March 28

Lecture 7: April 1

Lecture 8: April 8

Lecture 9: April 15 – Assignment 2 available on April 18

Lecture 10: April 29 – Problem class devoted to assignment 2; deadline on May 2

Lecture 11: May 6 – Mini-projects available on April 28; choose yours by May 6

Lecture 12: May 13

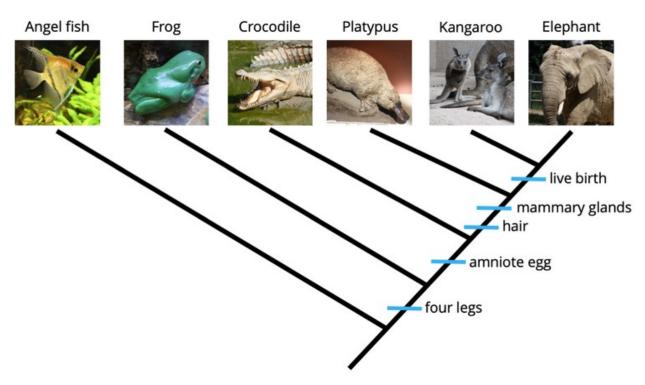
Lecture 13: May 20

Lecture 14: May 27 – Mini-project deadline on May 30

Motivation

Studying the evolution of species and genes

Traditionally: based on traits such as physical or morphological features



Group animals using shared characters

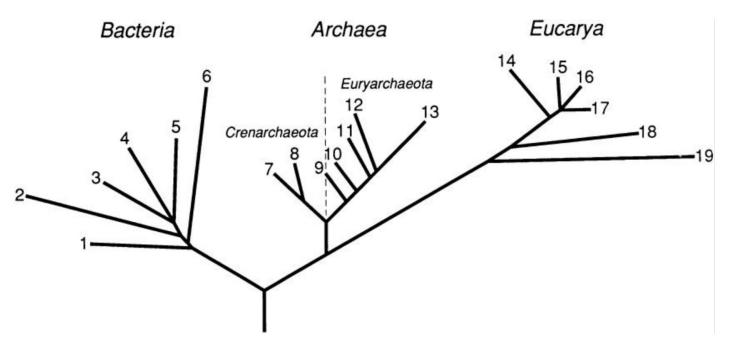
(assume that traits only appeared once)

Digital atlas of ancient life

Motivation

Studying the evolution of species and genes

- Traditionally: based on traits such as physical or morphological features
- More modern: based on molecular sequence data



3 major domains of life

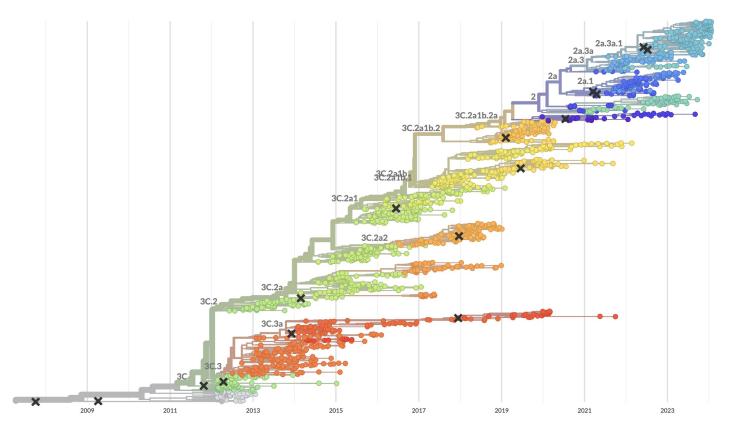
Inferred from analyzing (rRNA) sequences

Woese et al 1990 – redrawn in Pace et al 2012

Motivation

Studying the evolution of species and genes

At shorter timescales: understanding the phylogeny of a virus can help design vaccines

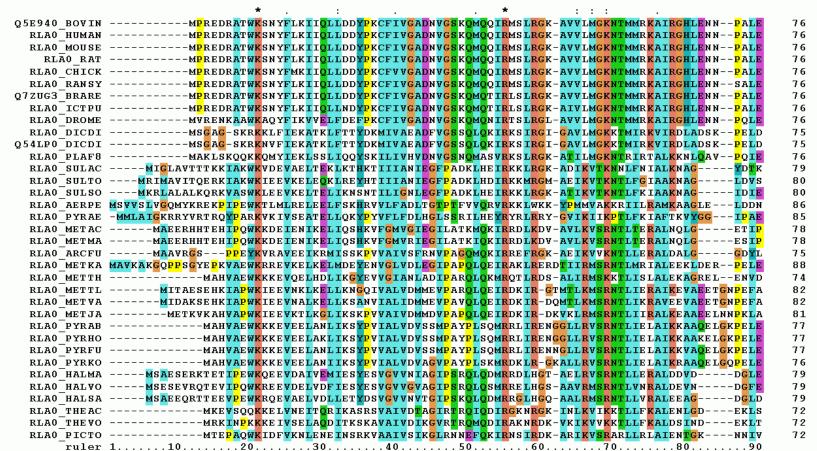


Influenza A/H3N2 evolution (sequence coding for hemagglutinin protein)

https://nextstrain.org/flu/seasonal/h3n2/ha/12y

Starting point: biological sequence data

Multiple sequence alignments, of amino-acid sequences or nucleotide sequences

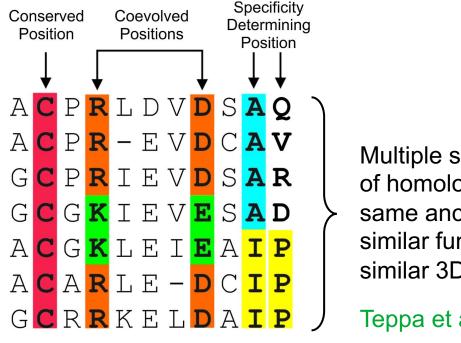


Acidic ribosomal protein P0 (first 90 positions) from several organisms

Row = sequence Column = site (given position in 3D structure)

Colors = level of conservation

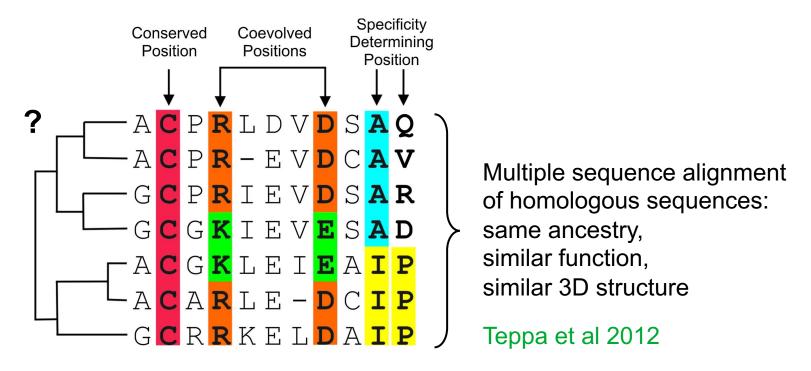
Starting point: biological sequence data



Multiple sequence alignment of homologous sequences: same ancestry, similar function, similar 3D structure

Teppa et al 2012

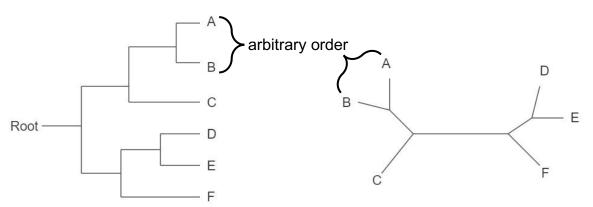
Starting point: biological sequence data



Goal: infer evolutionary tree (phylogenetic tree) from sequence data **Simplification:** ignore coevolution – assume each site (column) evolves independently

Structure of a phylogenetic tree

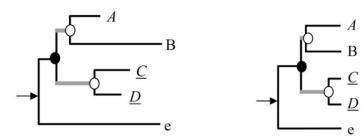
- Assume only bifurcations (2 branches, not more, from 1 node)
- Rooted versus unrooted trees:



Leaf nodes = observed species/sequences Internal nodes = hypothetical ancestors

Root = hypothetical common ancestor of all leaves (difficult to know where it is)

- Important features of a tree:
 - Branching events → tree topology
 - Branch length may represent nothing or evolutionary distances (phylogram) or time (chronogram)

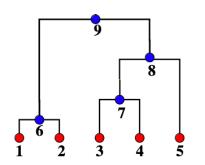


Inferring a phylogenetic tree from sequence data

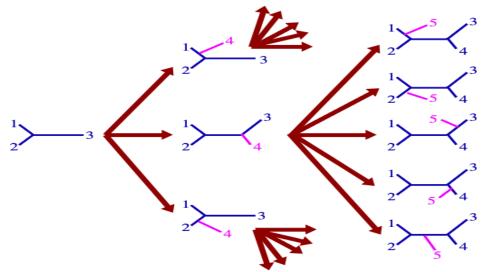
- Distance-based methods: start from evolutionary distances between sequences, and construct a tree based on them, using clustering algorithms
 - Unweighted Paired Group Mean Arithmetic (UPGMA)
 - Neighbor Joining (NJ)
- Character-based methods: use a score that quantifies how well a tree describes the raw data, and find the tree with the best score. These methods directly aim to fit the states (characters, i.e. amino acids or nucleotides) observed at each site in each sequence to a tree
 - Maximum parsimony
 - Maximum likelihood
 - Bayesian (maximum a posteriori)
- Difficulty: many possible trees!
- Phylogenetic tree construction has been shown to be NP-complete (no solution in polynomial time) for many models

Counting possible trees with n sequences (n leaf nodes)

- Move from leaves to root → 2 edges join at each internal node (bifurcating tree)
- → at each internal node, the number of edges decreases by one
 - → there are n-1 internal nodes
- Thus, there are 2n-1 total nodes (and 2n-2 total edges) in the rooted tree
- And there are 2n-2 nodes and 2n-3 edges in the unrooted tree



- Add an extra sequence: extra edge with new (n+1)th leaf can be added at any edge
 - → 2n-3 times more unrooted trees with n+1 leaves than with n leaves
- Thus, 1x3x5x...x(2n-5) unrooted trees with n leaves [1 with 3, 3 with 4...] notation: (2n-5)!!

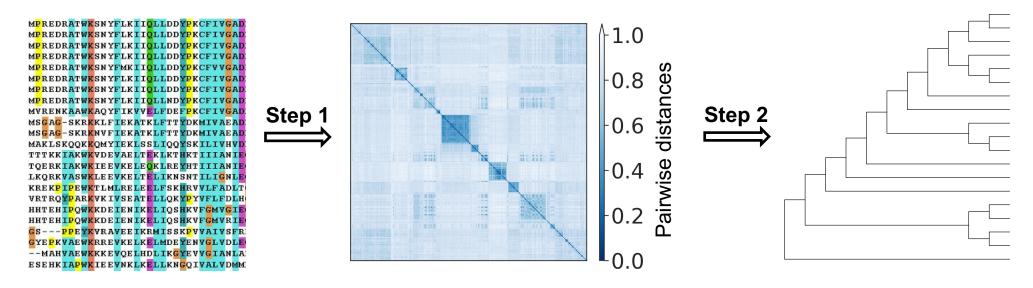


(2n-5)!! = 1x3x5x...x(2n-5) unrooted trees This is already $>10^{20}$ for n=20...

Volker Roth, U. of Basel

General method

- **Step 1**: MSA → pairwise distances between sequences
- Step 2: pairwise distances between sequences → tree matching observed distances



- **Limitation:** restricting to pairwise distances leaves out some information contained in the raw data (sequences). Two different pairs of sequences can have the same distance
- However, a lot of evolutionary information is contained in distances

Step 1: Determining pairwise distances between sequences

- **Ideally:** evolutionary distance number of mutations that actually occurred between two sequences (= sum of the number of mutations that occurred from their last common ancestor to each of them)
- **Hamming distance:** count sites that differ between two sequences (can then divide by number of sites to obtain number between 0 and 1)

AGATC AGGCA

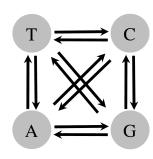
Between these two nucleotide sequences, Hamming distance=3/5

Limitation: evolutionary distance is underestimated due to multiple substitutions at the same site Example: if $A \to T \to A$, we observe no difference but 2 mutations occurred

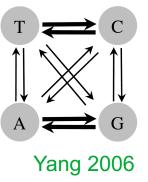
• Jukes-Cantor distance: simplest correction that takes into account multiple substitutions

Evolutionary model where:

- each site evolves independently of others
- all substitutions are equally likely: rate λ
 (Jukes-Cantor 1969)



Remark: more sophisticated models exist, e.g. transitions more likely than transversions (Kimura 1980)



Step 1: Determining pairwise distances between sequences

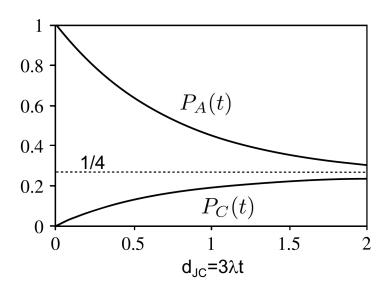
• Jukes-Cantor distance (typed notes): simplest correction that takes into account multiple substitutions

Evolutionary model where:

- each site evolves independently of others
- all substitutions are equally likely: rate λ

$$\frac{dP_A}{dt}(t) = \lambda \left[1 - 4P_A(t)\right]$$

If the initial state at t=0 is A, then: $P_A(t)=\frac{3}{4}e^{-4\lambda t}+\frac{1}{4}$ and $P_C(t)=P_G(t)=P_T(t)=-\frac{1}{4}e^{-4\lambda t}+\frac{1}{4}$



Under the Jukes-Cantor model, the evolutionary distance d_{JC} between two proteins can be estimated from their Hamming distance d_{H} as:

$$d_{JC} = 3\lambda t = -\frac{3}{4}\log\left|1 - \frac{4}{3}d_H\right|$$
 (natural logarithm)

Examples: d_H : 0.1, 0.3, 0.5, 0.7 d_{JC} : 0.11, 0.38, 0.82, 2.03

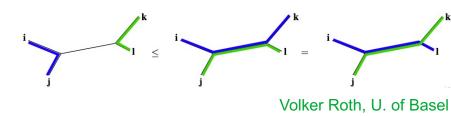
Remarks: For small d_H , $d_{JC} \approx d_H$. If $d_H \rightarrow 3/4$, $d_{JC} \rightarrow \infty$

Step 2: Building a tree from a matrix of pairwise distances between sequences

- **Goal:** Find a phylogenetic tree that agrees with the empirical pairwise distances

 Distance d_{ij}^T between leaves i and j along the tree T should match the empirical d_{ij} , as well as possible
- Least-square approach: find the tree T that minimizes $\sum_{i=1}^{T} \sum_{i \neq i} (d_{ij} d_{ij}^T)^2$ (n: number of leaves)
- Difficult because there are many trees; NP-complete. But efficient (polynomial) approximate algorithms:
 - Unweighted Paired Group Mean Arithmetic (UPGMA)
 - Neighbor Joining (NJ)
- If all leaves have the same distance from the root (all species evolve at the same rate constant molecular clock – "ultrametric tree"), then UPGMA will find the correct topology
- If distances are additive (less strong than ultrametric, rates can differ across species), there exists a
 tree T such that d_{ii}^T=d_{ii}, and NJ works well

Four point condition: For every set of four leaves i, j, k and l, two of the distances $d_{ij}+d_{kl}, d_{ik}+d_{jl}$ and $d_{il}+d_{jk}$ must be equal and larger than the third. For instance $d_{ij}+d_{kl} \leq d_{ik}+d_{jl}=d_{il}+d_{jk}$

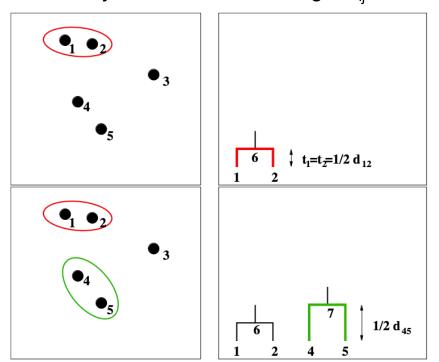


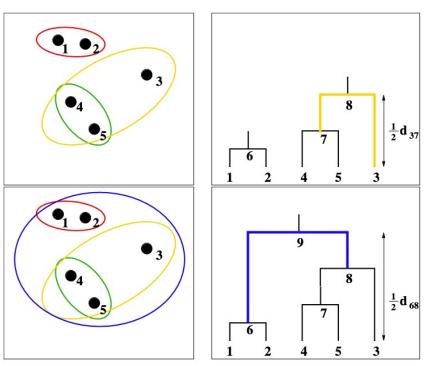
Generally, data is neither ultrametric nor additive, but NJ can often give reasonable approximations

Step 2: Building a tree from a matrix of pairwise distances between sequences

• **UPGMA:** it is a form of hierarchical clustering – iteratively joins two nearest clusters. Initially, each leaf is a cluster.

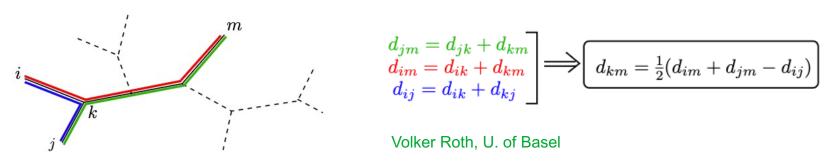
Find the 2 clusters i,j with the smallest distance d_{ij} . Group them into new cluster, and compute distance from it to all other ones as a weighted average: $d_{kl} = (\frac{n_i}{n_i + n_j})d_{il} + (\frac{n_j}{n_i + n_j})d_{jl}$ Connect i,j to new node k, at height $d_{ij}/2$. Iterate.





Step 2: Building a tree from a matrix of pairwise distances between sequences

• **NJ:** The idea is to find direct ancestor of 2 species, join them, iterate. Initially, each leaf is a cluster. For each node i, compute $u_i = \sum_{k \neq i} \frac{d_{ik}}{(n-2)}$: average distance to all other leaves k Choose i and j with smallest value of d_{ij} - u_i - u_j (thus i and j are close together and far from the rest) Join i and j with ancestor k. The distances between k and other leaves m is defined as follows:



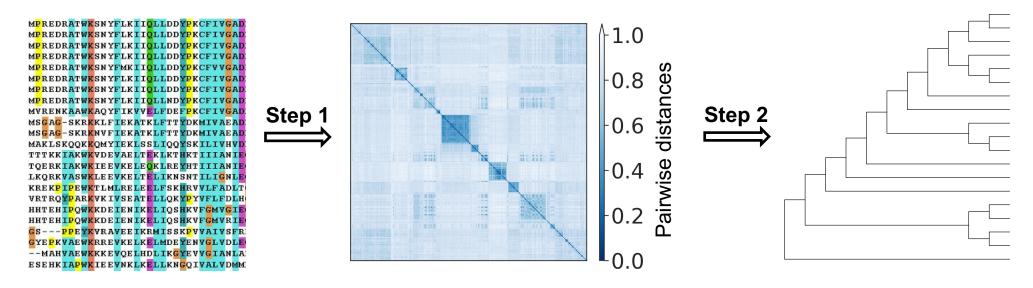
Branch lengths from i and j to the new node k are calculated as: $d_{ik} = \frac{1}{2}(d_{ij} + u_i - u_j)$, $d_{jk} = \frac{1}{2}(d_{ij} + u_j - u_i)$ Iterate.

Reminder: limitations:

- Starting from distances → we lose information from data (+ distance calculation is approximate)
- Generally, data is not ultrametric or additive, but NJ can often give reasonable approximations

General method

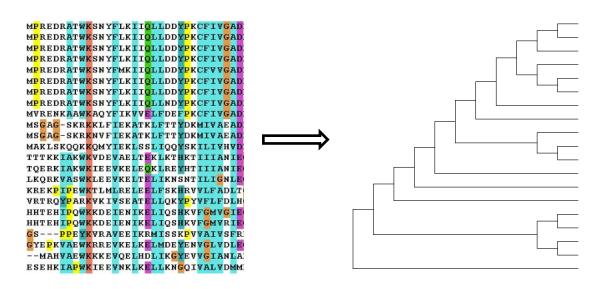
- **Step 1**: MSA → pairwise distances between sequences
- Step 2: pairwise distances between sequences → tree matching observed distances



- **Limitation:** restricting to pairwise distances leaves out some information contained in the raw data (sequences). Two different pairs of sequences can have the same distance
- However, a lot of evolutionary information is contained in distances

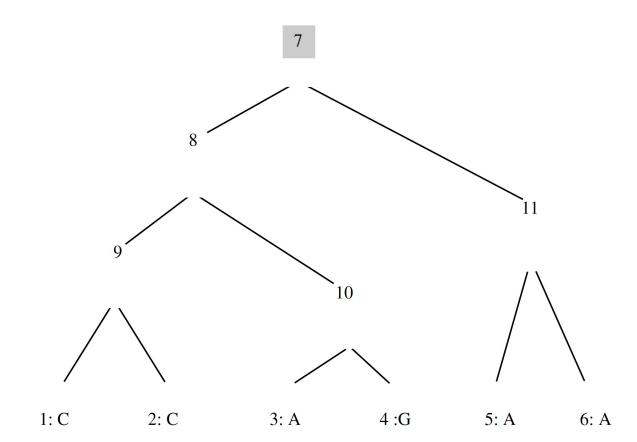
General method

- MSA → tree explaining the MSA
- Use a score that quantifies how well a tree describes the raw data, and find the tree with the best score.
 These methods directly aim to fit the states (characters, i.e. amino acids or nucleotides) observed at each site in each sequence to a tree
- Simplifying assumption: each site evolves independently from all others



Parsimony

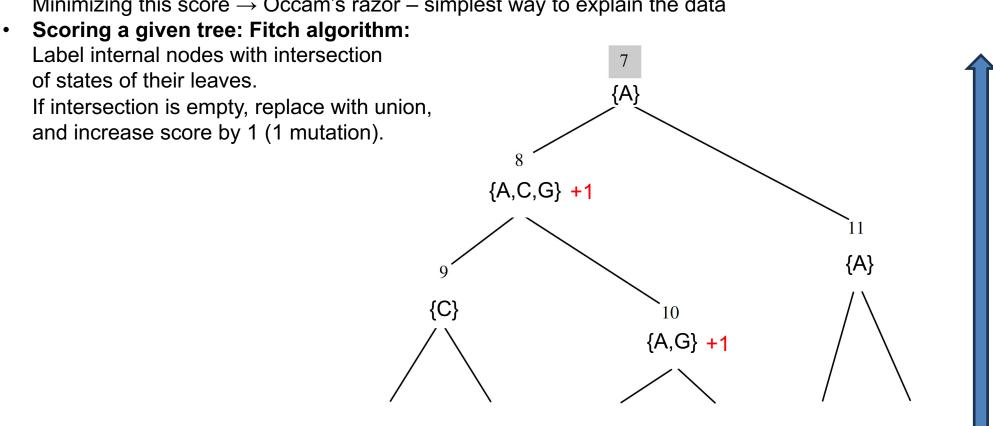
- Score: total number of substitutions (mutations) along all edges of the tree
 Minimizing this score → Occam's razor simplest way to explain the data
- Scoring a given tree:



Parsimony

Score: total number of substitutions (mutations) along all edges of the tree
 Minimizing this score → Occam's razor – simplest way to explain the data

1: C



2: C

3: A

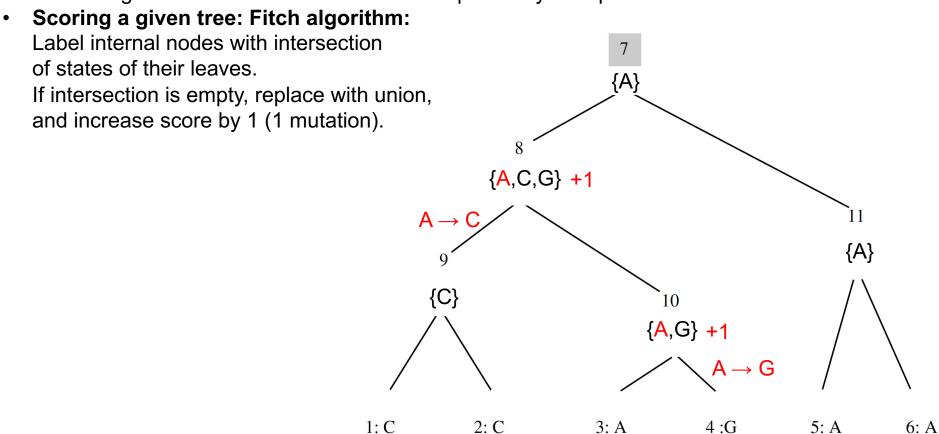
5: A

6: A

4 :G

Parsimony

Score: total number of substitutions (mutations) along all edges of the tree
 Minimizing this score → Occam's razor – simplest way to explain the data



Parsimony

Score: total number of substitutions (mutations) along all edges of the tree
 Minimizing this score → Occam's razor – simplest way to explain the data

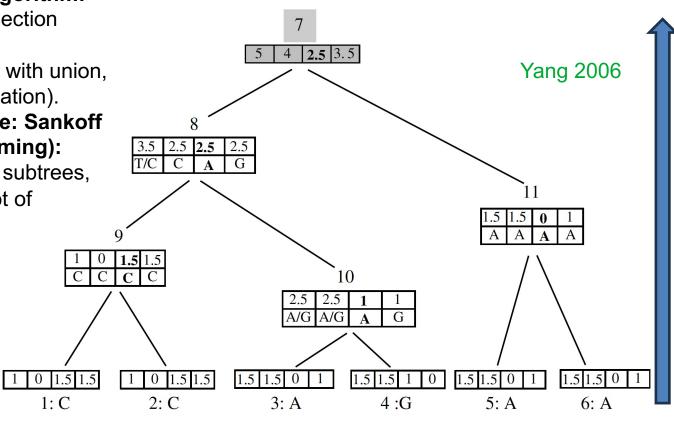
Scoring a given tree: Fitch algorithm:
 Label internal nodes with intersection of states of their leaves.

 If intersection is empty, replace with union, and increase score by 1 (1 mutation).

 Remark – scoring a given tree: Sankoff algorithm (dynamic programming):
 Mutation score matrix → score subtrees, for all possible states of the root of

the subtree: T,C,A,G.

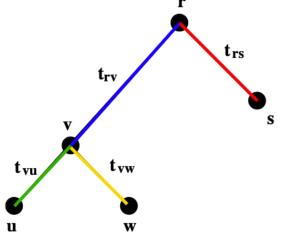
	То					
From	T	С	A	G		
T	0	1	1.5	1.5		
C	1	0	1.5	1.5		
Α	1.5	1.5	0	1		
G	1.5	1.5	1	0		



Maximum likelihood

- Score: likelihood of the data (MSA) given the model (the tree, with topology and branch lengths, under a certain nucleotide evolution model)
- Scoring a given tree:
 - (1) Assume that each site (each MSA column) evolves independently of others
 - (2) Assume that the probability of a node having a certain state only depends on the state of its parent node and on the branch length (genetic distance) t between them
 - (3) Assume that nucleotide frequencies P(x) are fixed through the phylogeny

Using (1), the likelihood is
$$\mathcal{L} = \prod_{i=1}^{L} P\left(x_1^{(i)}, \dots, x_D^{(i)} \middle| T\right)$$
 with L: number of sites; D: number of sequences



Observed data at one given site i, for D=3: u, w, s (= x_1 , x_2 , x_3)

$$P(u,w,s|T) = \sum_{r,v} P(u,w,s,v,r|T)$$
 Sum over ancestral states r, v

$$= \sum_{r,v} P(r) P(s|r,t_{rs}) P(v|r,t_{rv}) P(w|v,t_{vw}) P(u|v,t_{vu})$$
(2) & (3), using Bayes' theorem $P(y|x) = P(x|y) \frac{P(y)}{P(x)}$

For each branch, use a nucleotide evolution model, e.g. Jukes-Cantor

Maximum likelihood

Reminder: Jukes-Cantor distance (see typed notes):

Evolutionary model where:

- each site evolves independently of others
- all substitutions are equally likely: rate λ

$$\frac{dP_A}{dt}(t) = \lambda \left[1 - 4P_A(t)\right]$$

If the initial state at t=0 is A, then: $P_A(t) = \frac{3}{4}e^{-4\lambda t} + \frac{1}{4}$ and $P_C(t) = P_G(t) = P_T(t) = -\frac{1}{4}e^{-4\lambda t} + \frac{1}{4}$

• Thus, for one given site i, the score of a branch is $P(s|r,d_{rs})=-\frac{1}{4}e^{-4d_{rs}/3}+\frac{1}{4}$ if $r\neq s$ $P(s|r,d_{rs})=\frac{3}{4}e^{-4d_{rs}/3}+\frac{1}{4}$ if r=s

where d_{rs} is the evolutionary distance between the two nodes (recall that d=3 λ t)

$$\rightarrow$$
 express $\mathcal{L} = \prod_{i=1}^{L} P\left(x_1^{(i)}, \dots, x_D^{(i)} \middle| T\right)$

- For a given tree topology, branch lengths can be chosen to maximize this likelihood
- These results can be compared across trees topologies, to find the tree with the highest likelihood

Bayesian approach

• **Score:** posterior probability of the model (the tree, with topology and branch lengths, under a certain nucleotide evolution model) given the data (the MSA)

Scoring a given tree:

Reminder: Bayes' theorem: $P(\text{model}|\text{data}) = P(\text{data}|\text{model}) \times \frac{P(\text{model})}{P(\text{data})}$ It is difficult to access the posterior in analytic form

Strategy: draw samples (generate tree) from the posterior, using the Metropolis method

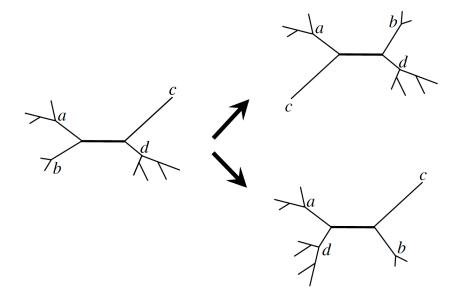
Comparison of approaches

- Currently, maximum likelihood and Bayesian methods are considered the most accurate ones
- But they are computationally intensive
- Parsimony is intuitive; distance-based methods are efficient and can be used as starting points
- Distance-based methods can be good enough, e.g. for relatively close sequences

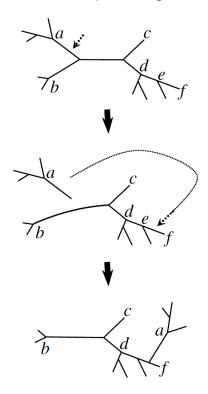
Remark: search over possible trees

- So far, we mainly looked at how to score a given tree
- Next, need (in principle) to score all possible trees. But there are many trees!
- Heuristic search strategies exist

- Search over possible trees: heuristic strategies
 - **Idea:** starting from a tree, construct neighboring trees by elementary operations
 - Nearest-neighbor interchange (NNI)



Subtree pruning and regrafting (SPR)

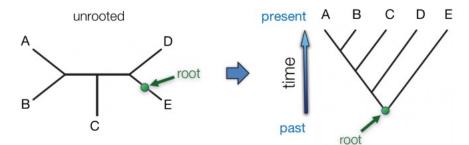


Yang 2006

Phylogeny inference: rooting a tree

Rooting a tree

- So far, we mainly saw how to select an unrooted tree
- Root = position of the most common recent ancestor; tells us the direction of evolution
- Position of the root affects interpretation about relatedness of sequences



EMBL-EBI online training

Where to place the root? (Remark: there are even more rooted trees than unrooted trees)

A common strategy is **outgroup rooting**:

- Include a sequence that is known to be more distant from the sequences considered than they are among them (e.g. from species information)
- Place the root where this outgroup joins the rest of the tree
- In the illustration above, E would be the outgroup

Phylogeny inference: estimating confidence

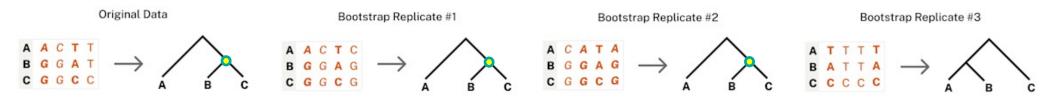
Bootstrapping

- Reminder: usually, in phylogeny inference methods, each site (each column of the MSA) is assumed to evolve independently
- → This can be exploited to estimate confidence

Method:

- Resample sites (columns) from the MSA: sample sites uniformly at random with replacement, to form a new MSA with the same number of columns. This new MSA is called a bootstrap replicate.
- Infer trees for multiple bootstrap replicates
- Bootstrap support value = percentage of bootstrapped trees that contain a particular node
- Higher value (close to 100) means better confidence

Example:



Highlighted node present in 67% of bootstrap replicate trees → score 67

Phylogeny inference: limitations

Assumptions

- Independently evolving sites
- Nucleotide evolution model
- Same mutation rate at each site, or limited number of classes of mutation rates

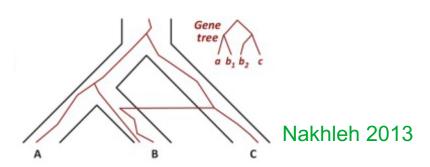
Conserved Positions Position Coevolved Positions Specificity Determining Position A C P R L D V D S A Q A C P R I E V D C A V G C P R I E V D S A R G C G K I E V E S A D A C G K L E I E A I P A C A R L E - D C I P G C R R K E L D A I P

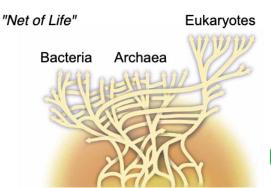
Approximations

- Starting from distances in distance-based methods
- Search over all trees can't be done exhaustively → heuristic search strategies

More fundamentally

- Horizontal gene transfer between species → trees of different genes are inconsistent with each other
- No fundamental species notion in prokaryotes





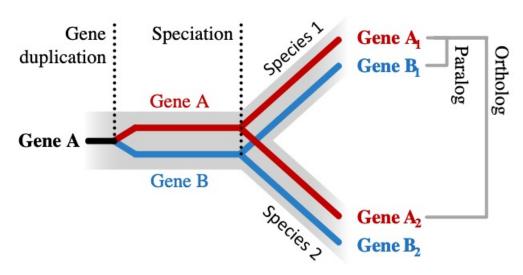
Doolittle 2000

Homologs, orthologs and paralogs

Definitions

Reminder: Gene A_1 in species 1 and gene A_2 in species 2 are homologous if they share a common ancestor Two homologous genes are:

- Orthologous if they diverged at a speciation event
- Paralogous if they diverged at a duplication event



Generally, orthologs preserve the same function, while paralogs do not and become more different

Practical approximate way to find orthologs

Reciprocal best hits: pairs (**g**,**h**) of genes from genomes (**G**,**H**) such that **g** is the gene in **G** most similar to **h**, and **h** is the gene in **H** most similar to **g**