# Modeling the cardiac action potentials in Python

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## 1 Introduction

The heart contraction is triggered by an electrical potential that propagates along all the myocardium, the cardiac muscle. The heart itself is able to produce the electrical impulse that determines this contraction. This is possible owing to the excitability of the heart cells, the cardiomyocytes, which, when suitably stimulated, are able to produce a variation in membrane voltage [1].

Differently from skeletal muscle cells, the cardiomyocyte are able to autonomously activate, independently of a nervous stimulus. The electrical activity of the heart originates at the sinoatrial node (SAN), a group of cardiac pacemaker cells located in the right atrium, see Figure 1(a). In normal conditions, SAN cells generate an electric signal that propagates throughout the right atrium to the left atrium. The activation front reaches the atrioventricular node (AV) located at the base of the atria. The AV conducts the signal activating the specialized fibers of the Purkinje network that spread as a tree-like on the endocardial surface of the ventricles. These Purkinje terminations transmit the electric signal to the ventricular walls and cardiac excitation then propagates throughout the ventricles, see Figure 1(a) [2].

#### 1.1 Cardiac action potentials

The contraction of cardiac cells is initiated by an electrical activation due to an action potential (AP), a depolarizing transitory membrane current that raises the transmembrane potential of an excitable cell from its resting value ranging between -90 and -80 mV to slightly positive values, followed by a repolarizing current that returns the transmembrane potential to its resting value [2].

At rest, the membrane potential is negative (around -90 mV), whereas when stimulated it reaches a positive value (around -20 mV) in a very short period (about 2 ms). After this depolarization, a plateau around 0 mV is observed corresponding to the refractory period, see Figure 1(b). Then, the repolarization

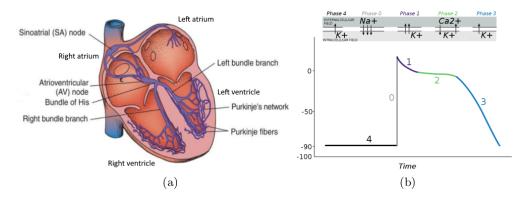


Figure 1: The cardiac conduction system (a) and (b) the characteristic action potential of cardiomyocytes (picture taken from [1]).

phase starts, which brings the potential back to the rest value allowing for a new excitation, see Figure 1(b) [1].

The AP is generated by several ion channels that open and close, and by the resulting currents passing through the membrane. The most important channels are those of calcium, sodium and potassium. In particular, a fast inward sodium current is the main driver of rapid depolarization, a slow inward flux of extra-cellular calcium ions is the main agent behind the characteristic plateau appearing after the depolarization, whereas the outward potassium currents are responsible for the repolarization. The main phases of a typical cardiac AP are displayed in Figure 1(a) [1, 2].

### 2 Cardiac mathematical cell models

At the mathematical level, the AP is described by means of an ionic model, a system of ordinary differential equations (ODE) which models the ionic currents in the cells.

There are three families of ionic models, featuring different levels of complexity and accuracy. The first family, the so-called reduced ionic models, only provide a description of the AP and disregard sub-cellular processes. One of the most celebrated reduced model for cardiac cells is the Aliev-Panfilov (A-P) ionic model [3]. The second family of cardiac cell models is that of the so-called first-generation models. Unlike reduced models, they allow explicit description of the kinetics of different ionic currents. Among them, the Courtemanche-Ramirez-Nattel (CRN) is the most widely used mathematical model for modeling the human atrial AP [4]. Finally, the third family of second-generation cardiac cell models, unlike first-generation models, provides a detailed description of many processes (see [1, 2] for further details).

### 2.1 Aliev-Panfilov ionic model

The A-P is a reduced ionic model that simulates the restitution property of cardiac tissue [3]. It consists of two equations:

$$\begin{cases} \frac{\partial u}{\partial t} &= -ku(u-a)(u-1) - uv \\ \frac{\partial v}{\partial t} &= \epsilon(u,v) \left[ -v - ku(u-a-1) \right] \end{cases},$$

where  $\epsilon(u,v) = \epsilon_0 + \frac{\mu_1 v}{u + \mu_2}$ , with k = 8, a = 0.15,  $\epsilon_0 = 0.002$ ,  $\mu_1 = 0.2$  and  $\mu_2 = 0.3$ . The model involves dimensionless variables u, v and t. The actual AP E, in mV, and the time T, in ms, can be obtained with the formulae:

$$E[mV] = 100u - 80, T[ms] = 12.9t.$$

The model in compact form reads

$$\frac{\partial \mathbf{w}}{\partial t} = F(u, v),\tag{1}$$

where  $\mathbf{w} = [u, v]$  and

$$F(u,v) = [-ku(u-a)(u-1) - uv, \epsilon(u,v) [-v - ku(u-a-1)]].$$

### 2.2 Courtemanche-Ramirez-Nattel ionic model

The CRN is the most widely used mathematical model for modeling the human atrial AP [4]. The CRN was developed using specific formulations of twenty one currents based on data recorded from human atrial myocytes experiments (along with representations of pump, exchange, and background currents).

The CRN consists of twenty one ODE:

$$\begin{cases} \frac{\partial V}{\partial t} &= \frac{-(I_{ion} + I_{st})}{C_m} \\ \frac{\partial \mathbf{c}}{\partial t} &= \mathbf{F}^{(\mathbf{c})} \\ \frac{\partial \mathbf{y}}{\partial t} &= \mathbf{F}^{(\mathbf{y})} \end{cases}$$
(2)

where V is the menbrane AP, the vector  $\mathbf{c} = [Na, K, Ca^{2+}, Ca_{rel}^{2+}, Ca_{up}^{2+}]$  includes the ionic coentrations variable,  $\mathbf{y} = [h, m, j, oa, oi, ua, ui, xr, xs, d, f, fca, u, v, w]$  is the gating varibles vector,  $\mathbf{F^{(c)}} = [F_{Na}, F_K, F_{Ca^{2+}}, F_{Ca^{2+}}, F_{Ca^{2+}}]$  and  $\mathbf{F^{(y)}}$  (with  $F_i^{(y)} = \frac{y_i^{\infty} - y_i}{\tau_i}$  for i = h, m, j, oa, oi, ua, ui, xr, xs, d, f, fca, u, v, w) are the right-hand-side (RHS) vectors of the ionic concentrations and gating varibles ODE, respectively. For the specific definition of RHS vector terms we refer to [4]. Moreover,  $I_{st}$  is the applied current stimulus defined as:

$$I_{st} = -80 \pmod{(t, T_{HB})} > t_{in}^{st}) \pmod{(t, T_{HB})} < t_{end}^{st}),$$

where  $T_{HB}$  is the period of one heartbeat and  $t_{in}^{st}$ ,  $t_{end}^{st}$  is the initial and final applied stimulus time, respectively. Finally, the ionic current  $I_{ion}$  is defined as:

$$I_{ion} = I_{Na} + I_{K1} + I_{to} + I_{Kur} + I_{Kr} + I_{Ks} + I_{Ca,L} + I_{p,Ca} + I_{NaK} + I_{NaCa} + I_{b,Na} + I_{b,Ca},$$

where for the specific definition of each term in  $I_{ion}$  we refer to [4]. The model (2) in compact form reads

$$\frac{\partial \mathbf{Y}}{\partial t} = \mathbf{F},\tag{3}$$

where  $\mathbf{Y} = [V, \mathbf{c}, \mathbf{y}]$  and  $\mathbf{F} = \left[\frac{-(I_{ion} + I_{st})}{C_m}, \mathbf{F^{(c)}}, \mathbf{F^{(y)}}\right]$ .

# 3 Python implementation

In the following section we detail the Python implementation for the A-P and CRN ionic models introduced in Section 2.

#### 3.1 Aliev-Panfilov code

To numerically solve the AP model in Python we use the scipy.integrate package using the function odeint.

```
y = odeint(model, y0, t)
```

The odeint requires three inputs:

- model: function name that returns derivative values at requested y and t values as dydt = model(y,t);
- y0: initial conditions of the differential states;
- t: time points at which the solution should be reported.

An example of using odeint is the following differential equation:

$$\frac{dy(t)}{dt} = -ky(t),\tag{4}$$

with parameter k = 0.3, the initial condition y0 = 5. The Python code (1) first imports the needed Numpy, Scipy, and Matplotlib packages. The model, initial conditions, and time points are defined as inputs to odeint to numerically calculate y(t).

Listing 1: example of using odeint for the example equation (4)

```
import numpy as np
from scipy.integrate import odeint
# function that returns dy/dt
```

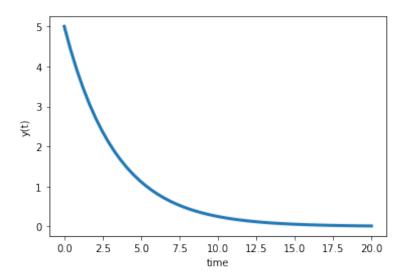


Figure 2: Result of code (1) for solving the equation (4) with odeint.

```
def model(y,t):
    k = 0.3
    dydt = -k * y
    return dydt

# initial condition
y0 = 5

# time points
t = np.linspace(0,20)

# solve ODE
y = odeint(model,y0,t)
```

Figure 2, produced by means of code (2), shows the result of code (1).

Listing 2: Plot the result of code (1)

```
import matplotlib.pyplot as plt

plt.plot(t,y,linewidth=3)
plt.xlabel('time')
plt.ylabel('y(t)')
plt.show()
```

The Python code (3) implements the A-P ionic model using the odeint package. The model, initial conditions, and time points are defined as inputs to odeint to numerically calculate the solution of equation (1).

Listing 3: A-P ionic model Python code

```
import numpy as np
from scipy.integrate import odeint
def AP_{model}(k=8, a=0.15, eps_0=0.002, mu_1=0.2, mu_2=0.3):
  def eps_fun(u, v):
    return eps_0 + mu_1*v/(u+mu_2)
  def fun(y, t):
    u\,,\ v\,=\,y
    eps = eps_fun(u, v)
    dydt = [-k*u*(u-a)*(u-1)-u*v, eps*(-v-k*u*(u-a-1))]
    return dydt
  return fun
def u_to_mV(u):
  return 100 * u - 80
def t_to_ms(t):
  return 12.9 * t
def run():
  t_{-}0 = 0
  t_end = 40
  discretization = (t_end - t_0) * 1000
  t = np.linspace(t_0, t_end, discretization)
  y0 = (0.17, 0)
  solution = odeint(AP_model(), y0, t)
```

### 3.2 Courtemanche-Ramirez-Nattel code

The Python code for solving the CRN ionic model begins with the model constants definition after importing the needed Numpy package:

Listing 4: CRN constants definition

```
import numpy as np
import matplotlib.pyplot as plot
log = np.log;
exp = np.exp;
sqrt = np.sqrt;

# constants
R = 8.3143; # Gas constant
T = 310; # Temperature
Fconst = 96.4867; # Faraday constant
Vi = 13668; # Intracellular volume
Vup = 1109.52; # Sarcoplasmic Reticulum uptake volume
```

```
# Sarcoplasmic Reticulum release volume
Vrel = 96.48;
K0 = 5.4;
                   # Extracellular K+ concentration
Na0 = 140;
                   # Extracellular Na+ concentration
Ca0 = 1.8;
                   # Extracellular Ca2+ concentration
                   # Maximal I_Na conductance
gna = 7.8;
                   # Maximal I_K1 conductance
gk1 = 0.09;
                   # Maximal I_to conductance
gto = 0.1652;
                   # Maximal I_Kr conductance
gkr = 0.0294;
                   # Maximal I_Ks conductance
gks = 0.129;
                   # Maximal I_Cal conductance
gcal = 0.1238;
                   # Maximal I_bCa conductance
gbca = 0.00113;
gbna = 0.000674;
                   # Maximal I_bNa conductance
                   # Maximal I_NaK
Inakmax = 0.60;
Inacamax = 1600;
                   # Maximal I_NaCa
Ipcamax = 0.275;
                   # Maximal I_pCa
Iupmax = 0.005;
                   # Maximal I_up
KQ10 = 3;
                   # Temperature scaling for I_Kur and I_to
                   # Voltage dependence param for I_NaCa
gamma = 0.35;
Kmnai = 10;
                   # [Na+]i Half saturation constant for I_NaK
Kmko = 1.5;
                   \# [K+]o Half saturation constant for I-NaK
Kmna = 87.5;
                   # [Na+]o Half saturation constant for I_NaCa
Kmca = 1.38;
                     [Ca2+]o Half saturation constant for I_NaCa
ksat = 0.1;
                     Saturation factor for L-NaCa
krel = 30;
                   # Maximal release rate for I_rel
kup = 0.00092;
                     [Ca2+]o half-saturation constant for I_up
                   # Maximal Ca2+ conc in uptake compartement
Caupmax = 15;
Cmdnmax = 0.05;
                   # Total calmodulin concentration in myoplasm
Trpnmax = 0.07;
                   # Total troponin concentration in myplasm
                   # Total calsequestrin concentration in myoplasm
Csgnmax = 10;
Kmcmdn = 0.00238;
                   # [Ca2+]i half saturation constant for calmodulin
Kmtrpn = 0.0005;
                   # [Ca2+]i half saturation constant for troponin
Kmcsqn = 0.8;
                   # [Ca2+]rel half saturation constant for I_up
                   # Membrane capacitance
CCconv = 1e6*2e-4; \# Conversion factor
```

Then there is the defininion of the CRN initial conditions:

Listing 5: CRN initial conditions

```
y0 = np.zeros(21);
y0[0] = -81.2;
                         # v
y0[1] = 11.2;
                         # Na
y0[2] = 139;
                         # K
y0[3] = 1.02*1e-4;
                         # Ca2+
y0[4] = 1.49;
                         # Ca2+rel
y0[5] = 1.49;
                         # Ca2+up
                         # h
y0[6] = 0.965;
y0[7] = 2.91 * 1e-03;
                         # m
y0[8] = 0.978;
                         # j
y0[9] = 3.04 * 1e-02;
                         # oa
v0[10] = 0.999;
                         # oi
y0[11] = 4.96 * 1e - 3;
                         # ua
y0[12] = 0.999;
                         # ui
y0[13] = 3.29 * 1e - 5;
                         # xr
```

```
y0[14] = 0.0187; # xs

y0[15] = 1.37*1e-4; # d

y0[16] = 0.999; # f

y0[17] = 0.775; # fca

y0[18] = 0.0; # u

y0[19] = 1.0; # v

y0[20] = 0.999; # w
```

The ionic currents definition follows:

Listing 6: CRN ionic currents

```
def ENa(y):
 return R * T/Fconst * log(Na0/y[1])
def EK(y):
 return R * T/Fconst * log(K0/y[2])
def ECa(y):
 return R * T/(2*Fconst) * log(Ca0/y[3])
def INa(y):
 return gna * y[7] * * 3 * y[6] * y[8] * (y[0] - ENa(y))
def IK1(y):
 return gk1 * (y[0] - EK(y)) / (1 + exp(0.07 * (y[0] + 80)))
def Ito(y):
 return gto * y[9]**3 * y[10] * (y[0] - EK(y))
def gkur(y):
 return 0.005 + 0.05 / (1 + \exp(-(y[0]-15)/13))
def Ikur(y):
 return gkur(y) * y[11]**3 * y[12] * (y[0] - EK(y))
def IKr(y):
 return gkr * y[13] * (y[0] - EK(y)) / (1 + exp((y[0] + 15)/22.4))
def IKs(y):
 return gks * y[14]**2 * (y[0] - EK(y))
def ICaL(y):
 return gcal * y[15] * y[16] * y[17] * (y[0] - 65)
def fnak(y):
  return 1 / (1 + 0.1245 * \exp(-0.1*Fconst*y[0]/R/T) + 0.0365 / 7 *
   (\exp(Na0/67.3) - 1) * \exp(-Fconst*y[0]/R/T))
def INaK(y):
  return Inakmax * fnak(y) * K0 / (1 + (Kmnai/y[1]) **1.5) / (K0 +
   Kmko)
def INaCanum(y):
  return Inacamax * (exp(gamma * Fconst * y[0]/R/T)*y[1]**3*Ca0 -
   \exp((gamma-1)*Fconst*y[0]/R/T)*Na0**3*y[3])
def INaCaden(y):
  return (Kmna**3 + Na0**3)*(Kmca + Ca0)*(1 + ksat*(exp((gamma-1)*
   Fconst*y[0]/R/T))
def INaCa(y):
 return INaCanum(y) / INaCaden(y)
def Ibca(y):
 return gbca * (y[0] - ECa(y))
def Ibna(y):
 return gbna * (y[0] - ENa(y))
def Ipca(y):
return Ipcamax * y[3]/(0.0005 + y[3])
```

Then there is the RHS definition of the AP equation:

Listing 7: CRN RHS of the AP equation

```
def Vfunc(t,y):
  return -(Iion(y) + Ist(t))/Cm
```

The gating variables definition follows:

Listing 8: CRN gating variables

```
#hfunc
def alphah(y):
  return 0.135*\exp(-(y[0] + 80)/6.8)*(y[0] < -40)
def betah(y):
  return 3.56 * \exp(0.079*y[0]) + 3.1*1e5 * \exp(0.35 * y[0]) * (y[0])
    < -40) + (y[0] >= -40) * 1/0.13 / (1 + exp(-(y[0]+10.66) /
    11.1))
def tauh(y):
  return 1 / (alphah(y) + betah(y))
def hinf(y):
  return alphah(y) * tauh(y)
def hfunc(y):
 return (hinf(y) - y[6]) / (tauh(y))
#mfunc
def alpham(y):
  return 0.32 * (y[0] + 47.13) / (1 - \exp(-0.1*(y[0] + 47.13))) * (y[0] + 47.13))
    [0] := -47.13) + (y[0] == -47.13) * 3.2
def betam(y):
  return 0.08 * \exp(-y[0]/11)
def taum(y):
 return 1 / (alpham(y) + betam(y))
def minf(y):
 return alpham(y) * taum(y)
def mfunc(y):
 return (minf(y) - y[7]) / (taum(y))
#jfunc
def alphaj(y):
  return (-126140 * \exp(0.2444*y[0]) - 3.474 * 1e-5 * \exp(-0.04391*y
    [0]) * (y[0] + 37.78)/(1 + \exp(0.311*(y[0] + 79.23))) * (y[0] <
     -40)
```

```
def betaj(y):
  return 0.1212 * \exp(-0.01052*y[0]) / (1 + \exp(-0.1378*(y[0]) +
   (40.14)) * (y[0] < -40) + 0.3 * exp(-2.535 * 1e-7)/(1 + exp(-0.1))
    * (y[0] + 32))) * (y[0] >= -40)
def tauj(y):
  def jinf (y):
 return alphaj(y) * tauj(y)
def jfunc(y):
 return (jinf(y) - y[8]) / (tauj(y))
#oafunc
def alphaoa(y):
 return 0.65 / (\exp(-(y[0] + 10)/8.5) + \exp(-(y[0] - 30)/59.0))
def betaoa(y):
 return 0.65 / (2.5 + \exp((y[1] + 82)/17))
def tauoa(y):
 \frac{\text{return}}{1} / \left( \text{alphaoa}(y) + \text{betaoa}(y) \right) / KQ10
def oainf(y):
 return 1 / (1 + \exp(-(y[0] + 20.47)/17.54))
def oafunc(y):
 return (oainf(y) - y[9]) / (tauoa(y))
#oifunc
def alphaoi(y):
  return 1 / (18.53 + \exp((y[0] + 113.7)/10.95))
def betaoi(y):
  return 1 / (35.56 + \exp(-(y[0] + 1.26)/7.44))
def tauoi(y):
  return 1 / (alphaoi(y) + betaoi(y)) / KQ10
def oiinf(y):
 return 1 / (1 + \exp((y[0] + 43.1)/5.3))
def oifunc(y):
  return (oiinf(y) - y[10]) / (tauoi(y)) 
#uafunc
def alphaua(y):
 return 0.65 / (\exp(-(y[0] + 10)/8.5) + \exp(-(y[0] - 30)/59))
def betaua(y):
 return 0.65 / (2.5 + \exp((y[0] + 82)/17))
def tauua(y):
 return 1 / (alphaua(y) + betaua(y)) / KQ10
def uainf(y):
 return 1 / (1 + \exp(-(y[1] + 30.3)/9.6))
def uafunc(y):
 return (uainf(y) - y[11]) / (tauua(y))
#uifunc
def alphaui(y):
  return 1 / (21 + \exp(-(y[0]-185)/28))
def betaui(y):
 return \exp((y[0]-158)/16)
def tauui(y):
 return 1 / (alphaui(y) + betaui(y)) / KQ10
def uiinf(y):
 return 1 / (1 + \exp((y[0] - 99.45)/27.48))
def uifunc(y):
return (uiinf(y) - y[12]) / (tauui(y))
```

```
#xrfunc
def alphaxr(y):
 return 0.0003 * (y[0] + 14.1) / (1 - \exp(-(y[0] + 14.1)/5))
def betaxr(y):
  return 7.3898 * 1e-5 * (y[0] - 3.3328) / (exp((y[0] - 3.3328))
    /5.1237) - 1)
def tauxr(y):
 \frac{\text{return } 1 / (alphaxr(y) + betaxr(y))}{}
def xrinf(y):
 return 1 / (1 + \exp(-(y[0] + 14.1)/6.5))
def xrfunc(y):
 return (xrinf(y) - y[13]) / (tauxr(y))
#xs func
def alphaxs(y):
 return 4 * 1e-5 * (y[0] - 19.9) / (1 - exp(-(y[0]-19.9)/17))
def betaxs(y):
 return 3.5 * 1e-5 * (y[0]-19.9) / (exp((y[0]-19.9)/9) - 1)
def tauxs(y):
 return 0.5 / (alphaxs(y) + betaxs(y))
def xsinf(y):
 return 1 / sqrt(1 + exp(-(y[0] - 19.9)/12.7))
def xsfunc(y):
 return (xsinf(y) - y[14]) / (tauxs(y))
#dfunc
def taud(y):
  return (1 - \exp(-(y[0]+10)/6.24)) / (0.035*(y[0] + 10)*(1 + \exp(-(y[0]+10)/6.24)))
   y[0] + 10)/6.24))
def dinf(y):
 return 1 / (1 + \exp(-(y[0] + 10)/8))
def dfunc(y):
 return (dinf(y) - y[15]) / (taud(y))
#ffunc
def tauf(y):
 return 9 / (0.0197 * \exp(-0.0337 * 0.0337 * (y[0]+10)**2) + 0.02)
def finf(y):
 return 1 / (1 + \exp((y[0]+28)/6.9))
def ffunc(y):
 return (finf(y) - y[16]) / (tauf(y))
#fca func
def fcainf(y):
 return 1 / (1 + y[3]/0.00035)
def fcafunc(y):
 return (fcainf(y) - y[17]) / 2
#ufunc
def Fn(y):
  return 1e-12 * Vrel * Irel(y) - 5*1e-13/Fconst*(0.5 * ICaL(y)-
   INaCa(y)/5
def uinf(y):
  return 1 / (1 + \exp(-(\operatorname{Fn}(y) - 3.4175 * 1e - 13) / (13.67 * 1e - 16)))
def ufunc(y):
 return (uinf(y) - y[18]) / 8
#vfunc
def tauv(y):
return 1.91 + 2.09 / (1 + \exp(-(Fn(y) - 3.4175 * 1e-13)/(13.67 * 1e-13))
```

```
\begin{array}{c} e-16)))\\ \text{def } vinf(y):\\ \text{return } 1-1 \ / \ (1+\exp(-(Fn(y)-6.835*1e-14)/(13.67*1e-16)))\\ \text{def } vfunc(y):\\ \text{return } (vinf(y)-y[19]) \ / \ tauv(y)\\ \#wfunc\\ \text{def } tauw(y):\\ \text{return } 6.0*(1-\exp(-(y[0]-7.9)/5)) \ / \ (1+0.3*\exp(-(y[0]-7.9)/5)) \ / \ (y[0]-7.9)\\ \text{def } winf(y):\\ \text{return } 1-1 \ / \ (1+\exp(-(y[0]-40)/17))\\ \text{def } wfunc(y):\\ \text{return } (winf(y)-y[20]) \ / \ tauw(y) \end{array}
```

Defining the RHS of the concentration equations:

Listing 9: RHS of the concentration equations

```
def Nafunc(y):
  return CCconv * (-3*INaK(y) - 3*INaCa(y) - Ibna(y) - INa(y)) / (
   Fconst * Vi)
def Kfunc(v):
  return CCconv * (2*INaK(y) - IK1(y) - Ito(y) - Ikur(y) - IKr(y) -
   IKs(y)) / (Fconst*Vi)
def B1(y):
  return CCconv * (2*INaCa(y) - Ipca(y) - ICaL(y) - Ibca(y))/(2*
   Fconst*Vi) + (Vup*(Iupleak(y) - Iup(y)) + Irel(y)*Vrel)/Vi
def B2(y):
  return 1 + Trpnmax*Kmtrpn/(y[3] + Kmtrpn)**2 + Cmdnmax * Kmcmdn/(y
   [3] + \text{Kmcmdn})**2
def Cafunc(y):
 return B1(y)/B2(y)
def Caupfunc(y):
 return Iup(y) - Iupleak(y) - Itr(y) * Vrel / Vup
def Carelfunc(y):
  return (Itr(y) - Irel(y))/(1 + Csqnmax * Kmcsqn/(y[4] + Kmcsqn)
   **2)
```

Defining the RHS of of all the ODE:

Listing 10: Vector of RHS of all the ODE

```
 \begin{array}{l} \textbf{def } F(t\,,y) \colon \\ \textbf{vF} = \textbf{np.array} \left( \left( \textbf{Vfunc}(t\,,y) \,,\,\, \textbf{Nafunc}(y) \,,\,\, \textbf{Kfunc}(y) \,,\,\, \textbf{Cafunc}(y) \,,\,\, \\ \textbf{Carelfunc}(y) \,,\,\, \textbf{Caupfunc}(y) \,,\,\, \textbf{hfunc}(y) \,,\,\, \textbf{mfunc}(y) \,,\,\, \textbf{jfunc}(y) \,,\,\, \textbf{oafunc}(y) \,,\,\, \textbf{oifunc}(y) \,,\,\, \textbf{uifunc}(y) \,,\,\, \textbf{xrfunc}(y) \,,\,\, \textbf{xsfunc}(y) \,,\,\, \textbf{dfunc}(y) \,,\,\, \textbf{ffunc}(y) \,,\,\, \textbf{fcafunc}(y) \,,\,\, \textbf{ufunc}(y) \,,\,\, \textbf{vfunc}(y) \,,\,\, \textbf{wfunc}(y))) \,;\,\, \\ \textbf{return} \quad \textbf{vF} \end{array}
```

Finally, the CRN model (3) is numerically solved by means of the forward Euler method. Introducing the discrete times  $t^n = ndt$  (where  $dt = \frac{t_{end} - t_0}{n}$  with  $t_0$ ,  $t_{end}$  the initial and final times, respectively and  $n \in \mathbb{N}_0^+$ ) and denoting  $\mathbf{A}^n$  the time discretization of the generic vector variable  $\mathbf{A}$ , one step of the Euler method

from  $t_n = t_0 + ndt$  to  $t_{n+1} = t_0 + (n+1)dt$  is:

$$\mathbf{Y}^{n+1} = \mathbf{Y}^n + dt \mathbf{F}^n. \tag{5}$$

The time discretization (5) is performed by means of code (11).

Listing 11: CRN ionic model time integration

```
THB = 800;
                    # heartbeat length
nHB = 1;
                    # number of heartbeats
t_i = 0;
                                       # initial time
dt = 0.01;
                                      # time step
t_end = nHB * THB;
                                       # final time
tt = np.arange(t_in, t_end+dt, dt); # time vector
\dim = y0. size;
Y = np.zeros((dim, tt.size)); # Inizialize solution
Y[:,0] = y0;
                               # Vector of initial conditions
index = 1;
t = t_i n;
for i in tt[1:]:
  y0 = y0 + dt * F(i, y0);
  Y[:,index] = y0;
  t \ = \ t \ + \ dt \ ;
  index = index + 1;
```

### 4 Numerical results

In this section we show the results obtained with the A-P and CRN codes fully detailed in Section 3.

### 4.1 Aliev-Panfilov

Figure 3, obtained by means of the plotter code (12), display the time trace of the AP (both in dimensional and adimensional forms), the gating variable and the phase diagram for the A-P ionic model.

Listing 12: A-P plotter

```
import matplotlib.pyplot as plot

def pictures(t, solution):
    fig = plot.figure(1, figsize=(10, 12))

ax1 = fig.add_subplot(221)
    ax1.plot(t, solution[:, 0],'k',linewidth=2)
    ax1.set_title('Adimensional action potential u')
    ax1.set_xlabel('t [-]')
    ax1.set_ylabel('u [-]')
```

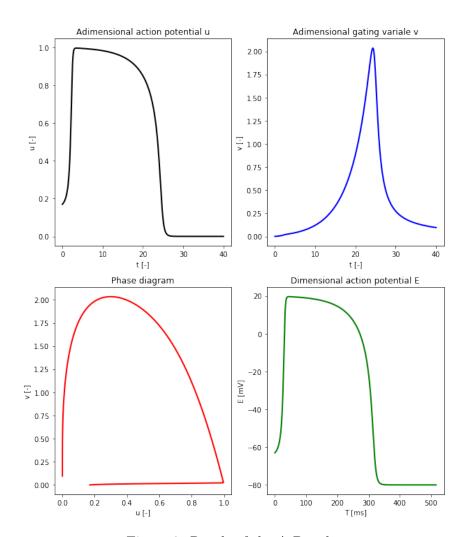


Figure 3: Result of the A-P code.

```
ax2 = fig.add_subplot(222)
ax2.plot(t, solution[:, 1],'b',linewidth=2)
ax2.set_title('Adimensional gating variale v')
ax2.set_xlabel('t [-]')
ax2.set_ylabel('v [-]')

ax3 = fig.add_subplot(223)
ax3.plot(solution[:, 0], solution[:, 1],'r',linewidth=2)
ax3.set_title('Phase diagram')
ax3.set_xlabel('u [-]')
ax3.set_ylabel('v [-]')
E = u_to_mV(solution[:, 0])
T = t_to_ms(t)
```

```
ax4 = fig.add_subplot(224)
ax4.set_title('Dimensional action potential E')
ax4.plot(T, E, 'g', linewidth=2)
ax4.set_xlabel('T [ms]')
ax4.set_ylabel('E [mV]')
plot.show()
```

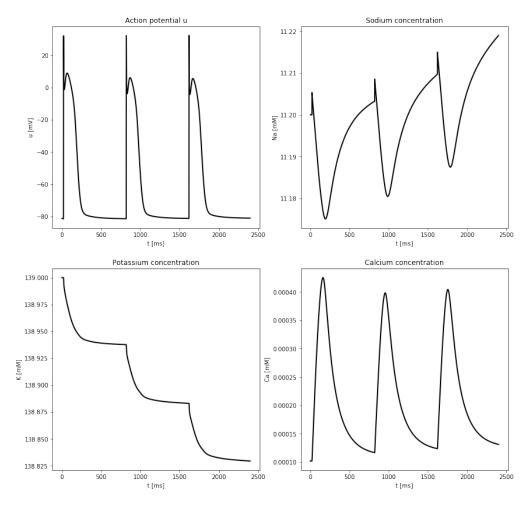


Figure 4: Result of the CRN code.

# 4.2 Courtemanche-Ramirez-Nattel

Figure 4, obtained by means of the plotter code (13), display the time trace of the AP and three concentrations (Sodium, Potassium and Calcium) of the CRN ionic model.

```
import matplotlib.pyplot as plot
def pictures (time, solution):
fig = plot.figure(1, figsize = (14, 14))
ax1 = fig.add_subplot(221)
ax1.plot(time, solution[0,:], 'k', linewidth=2)
ax1. set_title ('Action potential u')
ax1.set_xlabel('t [ms]')
ax1.set_ylabel('u [mV]')
ax2 = fig.add_subplot(222)
ax2.plot(time, solution[1,:],'k',linewidth=2)
ax2.set_title('Sodium concentration')
ax2.set_xlabel('t [ms]')
ax2.set_ylabel('Na [mM]')
ax3 = fig.add_subplot(223)
ax3.plot(time, solution[2,:], 'k', linewidth=2)
ax3.set_title('Potassium concentration')
ax3.set_xlabel('t [ms]')
ax3.set_ylabel('K [mM]')
ax4 = fig.add_subplot(224)
ax4.set_title('Calcium concentration')
ax4.plot(time, solution[3,:], 'k', linewidth=2)
ax4.set_xlabel('t [ms]')
ax4.set_ylabel('Ca [mM]')
plot.show()
```

## References

- [1] A. Quarteroni, L. Dedè, A. Manzoni, and C. Vergara. *Mathematical Modelling of the Human Cardiovascular System: Data, Numerical Approximation, Clinical Applications.* Cambridge University Press, 2019.
- [2] P.C. Franzone, L.F. Pavarino, and S. Scacchi. *Mathematical Cardiac Electrophysiology*, volume 13. Springer, 2014.
- [3] R.R. Aliev and A.V. Panfilov. A simple two-variable model of cardiac excitation. *Chaos, Solitons & Fractals*, 7:293–301, 1996.
- [4] M. Courtemanche, R.J. Ramirez, and S. Nattel. Ionic mechanisms underlying human atrial action potential properties: insights from a mathematical model. *American Journal of Physiology-Heart and Circulatory Physiology*, 275:H301–H321, 1998.