GS scripts

#Research #fbox #scripts

Genome assembly

```
# trim
zcat fastq_pass/*.gz > raw.fq
porechop-runner.py -i raw.fq -o raw.t.fq &> trim.log &
# filter reads length > 5000
cat raw.t.fq|seqkit fx2tab|awk -F '\t' 'length($2)>5000'|
segkit tab2fx > raw.t.5k.fg
# correct with canu
# results ${i}.5k.correctedReads.fasta.gz
canu -correct genomeSize=130m -d cor_5k -p canu_cor.5k
-nanopore raw.t.5k.fq
# flye assemble
python flye --nano-raw ${i}.5k.correctedReads.fasta.gz --
out-dir flye_out --genome-size 130m --threads 96 &>flye.log
# ragtag correct
ragtag.py correct ${i}.fa assembly.fasta --aligner nucmer
-o cor_out -f 5000 --nucmer-params '--mum --mincluster 100
--maxgap 300 -t 96'
# ragtag scaffold
ragtag.py scaffold ${i}.fa cor_out/ragtag.correct.fasta --
aligner nucmer -o scaf_out -f 5000 --nucmer-params '--mum --
mincluster 100 -- maxgap 300 -t 96'
Remove bacteria
cat ${i}.fa|seqkit fx2tab|grep -v Chr|seqkit tab2fx > $
{i}.unplaced.fa
blastn -db nt -num_threads 48 -query ${i}.unplaced.fa
-outfmt "6 gsegid ssegid pident length mismatch gapopen
qstart gend sstart send evalue bitscore stitle" > $
{i}.unplaced.blast.txt
cat ${i}.unplaced.blast.txt|grep 'Escherichia coli'|cut -f1|
sort|uniq|seqkit grep -v -f - ${i}.fa > clean/${i}.fa
Polishing
bwa-mem2 index ${i}.fa
bwa-mem2 mem -t 48 -M ${i}.genome.fa *fq.gz|samtools sort -@
48 - o ${i}_R1.bam
```

```
java -Xmx24G -jar pilon-1.24.jar --genome ${i}.fa --frags $
\{i\}_R1.bam
RepeatMasking
# RepeatModeler
BuildDatabase -name ${i}DB ${i}.fa
RepeatModeler -database ${i}DB -pa 16 &> $
{i}.RepeatModeler.log.txt &
getorf -minsize 90 ${i}DB-families.fa -outseq ${i}-
families.orf.fa
#cel_pro = path to celegans protein
blastp -query ${i}-families.orf.fa -db $cel_pro -evalue
1e-09 -outfmt 6 -seg yes -num_threads 12 -out $
{i}.orf.blastp.txt
# run interpro to filter out those with protein domain
interpro/interproscan:5.65-97.0 --input ${i}-families.orf.fa
--disable-precalc
# combine blastp and interpro results
cat ${i}.orf.blastp.txt |cut -d "#" -f1|sort|uniq|sed 's/$/
#/g' > ${i}.orf.blastp.families.txt
cat ${i}-families.orf.fa.tsv|cut -d "#" -f1|sort|uniq|sed
's/$/#/g' > ${i}.orf.interpro.families.txt
cat ${i}.*.families.txt|sort|uniq > ${i}.filter.txt
# filter and make the final repeat library
seqkit fx2tab ${i}DB-families.fa|grep -vf ${i}.filter.txt|
seqkit tab2fx > ${i}-families.filtered.fa
# RepeatMasker
RepeatMasker -lib ${i}-families.filtered.fa -xsmall -pa 6
-gff ${i}.fa -e ncbi -gccalc &>${i}.RepeatMasker.log &
Prediction - mapping
trim_galore --cores 4 --paired *.fastq
STAR --runThreadN 24 --runMode genomeGenerate --genomeDir $
{i} index --genomeFastaFiles ${i}.fa
# mapping
STAR --runThreadN 24 --genomeDir ${i}_index --
outFileNamePrefix ${i} --readFilesIn *.fq
# samtools sort results
samtools sort ${i}Aligned.out.sam -@ 24 -o ${i}.bam
samtools index ${i}.bam
```

we can use samtools flagstat to check the mapping quality
samtools flagstat \${i}.bam

Prediction - AUGUSTUS

bam2wig \${i}.bam > \${i}.wig
cat \${i}.wig |wig2hints.pl --width=10 --margin=10 -minthresh=2 --minscore=4 --src=W --type=ep --radius=4.5 > \$
{i}.hints.exon.gff

bam2hints --in AF.bam --out=\${i}.hints.intron.gff

cat *.gff > \${i}.hints.gff

augustus --extrinsicCfgFile=extrinsic.M.RM.E.W.cfg -softmasking=True --hintsfile=\${i}.hints.gff -uniqueGeneId=true --protein=on --introns=on --start=on -stop=on --cds=on --codingseq=on --gff3=on --progress=true -species=caenorhabditis \${i}.fa.masked > \${i}.gff 2>\$
{i}.gff.log

OrthoFinder

OrthoFinder
orthofinder -f . -M msa -A muscle -T iqtree -X &>
orthofinder.log &