DNA extraction from *N. furzeri* embryos

Materials

- DNeasy Blood & Tissue (Qiagen)

1. Remove chorion and yolk from embryos. Also, remove YSL/membrane as much as possible. Collect processed embryos in Eppendorf with PBS, on ice.
2. After all embryos are processed, wash x 2 with PBS
3. Remove PBS supernatant. Add 180 ul Buffer ATL. Vortex briefly.
   (Optional) If RNA-free genomic DNA is required, add 4ul RNase A (100 mg/ml) here, vortex, and incubate for 2 min at RT.
4. Add 20 ul proteinase K. Mix thoroughly by vortexing, and incubate at 56'C overnight (or until embryos are completely lysed).
5. Vortex for 15 s. Add 200 ul Buffer AL and vortex. Then add 200 ul 100% EtOH, and vortex again.
6. Transfer the mixture from Step 5 (including any precipitate) into the DNeasy Mini spin column placed in a 2 ml collection tube. Centrifuge at >6000 g for 1 min. Discard flow-through and collection tube.
7. Place the DNase Mini spin column in a new 2 ml collection tube, add 500 ul Buffer AW1, and centrifuge for 1 min at 6,000 g to dry the DNase membrane. Discard flow-through and collection tube.
8. Place the DNase Mini spin column in a new 2 ml collection tube, add 500 ul Buffer AW2, and centrifuge for 3 min at 20,000 g to dry the DNase membrane. Discard flow-through and collection tube.
9. Place the DNase Mini spin column in a clean 1.5 ml Eppendorf tube, and pipet 150 ul Buffer AE directly onto the DNase membrane. Incubate at RT for 1 min, and then centrifuge for 1 min at 6000 g to elute.
10. (Recommended & Optional) For maximum DNA yield, repeat elution once as described in Step 8 with the same elute and tube.
11. Determine DNA concentration by Nanodrop.

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