

Aligning reads: tools and theory



Genome

chrX: 52139280 152139290 152139300 152139310 152139320 152139330  
--->CGCCGTCCCTCAGAAATGGAAACCTCGCTTCTCTCTGCCCCACAATGCGCAAGTCAG

Sequence read

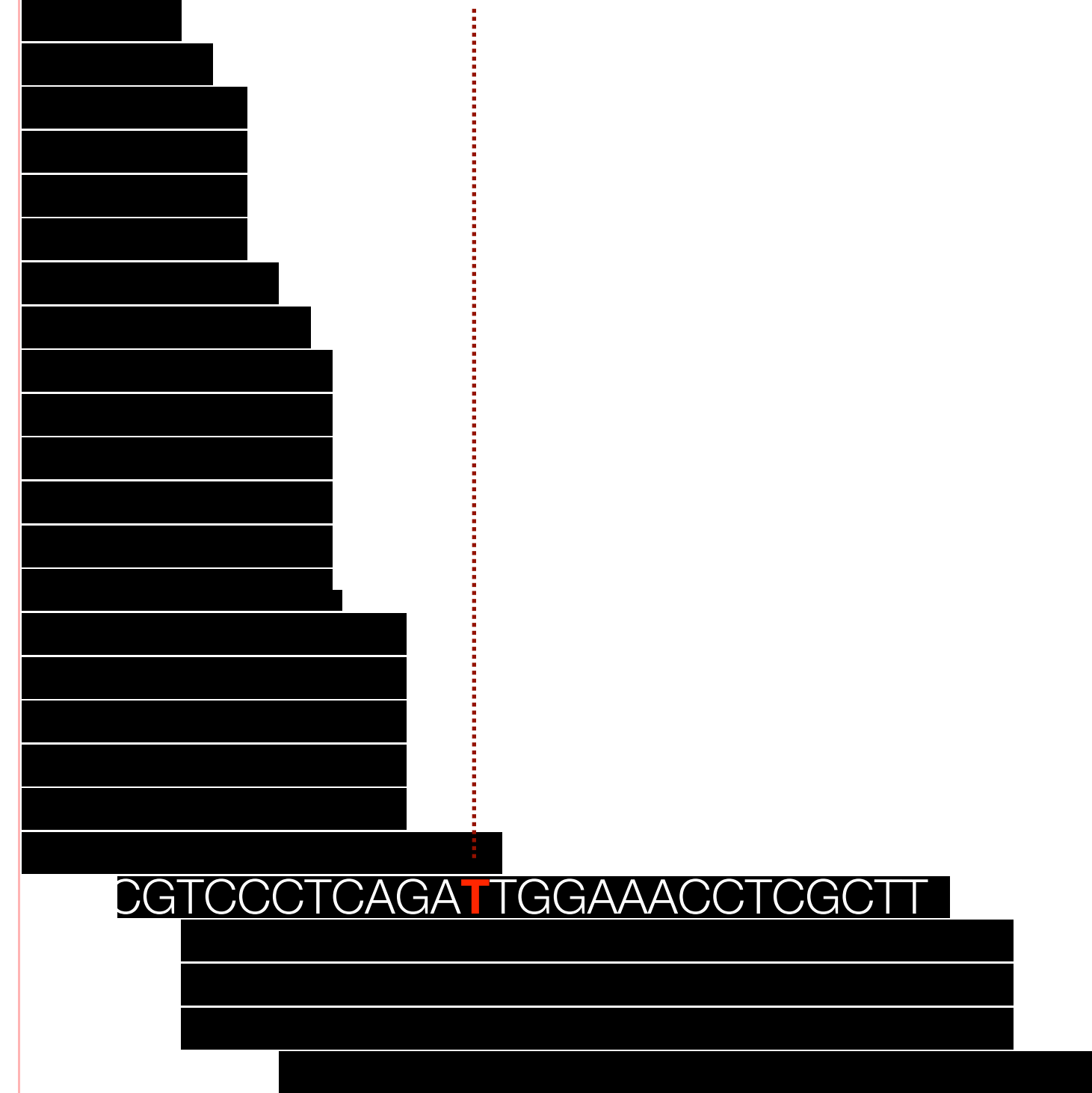
CGTCCCTCAGAAATGGAAACCTCGCTT

A simple case of string matching

Genome

chrX: 52139280 152139290 152139300 152139310 152139320 152139330  
--->CGCCGTCCCTCAGAAATGGAAACCTCGCTTCTCTCTGCCCCACAATGCGCAAGTCAG

Sequence reads



Difficult in practice

- Volume of data: ~3 Gbp
- ~50% of genome is repeat regions that cannot be covered by reads
  - Simple repeats, tandem, interspersed
  - Transposons
  - Segmental duplications where mapping is unclear
- Gap or unfinished regions
  - peri-centromere, sub-telomere
  - ~5Mb unique to ethnic groups (e.g., African, Asian)
- Finishing errors(1/10,000bp), miscalled base incorporated

Challenges:  
Human genome is large and complex

- ▶ Genome Reference Consortium: "...working to create assemblies that better represent diversity and provide more robust substrates for genome analysis."
- ▶ novel assembly algorithm
- ▶ correcting assembly errors (fix patches)
- ▶ addition of new alternate loci (patches)
- ▶ filling in gaps



Challenges:  
Genome is continuously changing

- Ensembl, UCSC and NCBI all use the same genome assemblies or builds provided by the GRC (i.e GrCh37 = hg19)
- Patches are provided by the GRC, but incorporated as updates by each database at different intervals
- At any point in time, the sequence can vary between databases but coordinates are unchanged
- **Always use the same biological database for all reference data!**

Challenges:  
Sources of genome reference sequence

- Closing the gaps; more complete genome information
- 8000 nucleotides altered
- Several misassembled regions corrected
- 261 alternate loci across 178 regions (improved diversity)
- Sequence information for centromeres

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### Human species advised to move to GRCh37

Posted on April 15, 2015 by [jovialscientist](#)

BOSTON. The entire human species has been advised to convert their genome to GRCh37 by the [GATK Best Practices team](#) at the Broad Institute, *The ScienceWeb* has learned.

GRCh37 is the *previous* version of the human genome reference. Last year, a rogue team of militant terrorist bioinformaticians within the Genome Reference Consortium released GRCh38, a hellish combination of core chromosomes, patches, unplaced contigs and alternate loci. In one fell swoop they broke every single bioinformatics pipeline ever written.

"Enough is enough" said Geraldine Van Damme, former martial arts expert and now head of the GATK team. "We took one look at GRCh38 and though 'that's it, we're sticking to GRCh37 and never moving'. We're therefore recommending that every human on the planet converts their genome to GRCh37. They should use CRISPR or something. It's going to make our lives a lot easier" she finished.

However, not everyone agrees. Deanna Cathedral, formerly Head of Anything Useful at the National Church of Biology Idiots (NCBI) said: "This reminds of the early days of the human genome project, when Frankie Collins suggested we try and genetically modify everyone to be haploid. It's just not realistic" she concluded.

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# GRCh37 vs. GRCh38



# LiftOver at UCSC

You can obtain corresponding coordinates of a different genome build, if you have a set of coordinates from a known build using the **LiftOver tool (UCSC)**

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**Lift Genome Annotations**

This tool converts genome coordinates and genome annotation files between assemblies. The input data can be pasted into the text box, or uploaded from a file. If a pair of assemblies cannot be selected from the pull-down menus, a direct lift between them is unavailable. However, a sequential lift may be possible. Example: lift from Mouse, May 2004, to Mouse, Feb. 2006, and then from Mouse, Feb. 2006 to Mouse, July 2007 to achieve a lift from mm5 to mm9.

Original Genome:  

Human

Original Assembly:  

Mar. 2006 (NCBI36/hg18)

New Genome:  

Human

New Assembly:  

Feb. 2009 (GRCh37/hg19)



- Short reads: 50-150 bp (versus a very long reference)
  - Non-unique alignment
  - Sensitive to sequencing errors
- Massive amount of short reads: one lane produces  $\geq 150$  million 100 nucleotide reads
- Small insert size: 200-500 bp libraries

Challenges: short read NGS data

<b>Reference</b>	ATCTCCATAGGACTAGAAAGTAG
Substitution	ATCTCCATAG <b>C</b> ACTAGAAAGTAG
Deletion	ATCTCCATAGGAC <b>-</b> AGAAAGTAG
Insertion	ATCTCCATAGGACTAGAAAGT <b>T</b> AG
3bp deletion	ATCTC <b>- - -</b> AGGACTAGAAAGTAG

Challenges: non-exact matching

# Local alignment vs Global alignment

- ▶ **Local alignment** matches the query with a *substring* (k-mer) of the reference
  - ▶ Tailored towards finding *regions of highly similar sequence* and aligning around those by working outwards to align the rest

## Local Alignment

```
5' ACTACTAGATTACTTACGGATCAGGTACTTTAGAGGCTTGCAACCA 3'
      |||| ||||| ||||| ||||| |||||
5' TACTCACGGATGAGGTACTTTAGAGGC 3'
```

## Global Alignment

```
5' ACTACTAGATTACTTACGGATCAGGTACTTTAGAGGCTTGCAACCA 3'
||||| ||||| ||||| ||||| |||||
5' ACTACTAGATT----ACGGATC--GTACTTTAGAGGCTAGCAACCA 3'
```

- ▶ A **global alignment** performs end-to-end alignment between the query and the reference



**Reference**    ATCTCCATAGGACTAGGAAGTAG

Substitution    ATCTCCATAG**C**ACTAGGAAGTAG

Deletion        ATCTCCATAGGAC**-**AGGAAGTAG

Insertion        ATCTCCATAGGACTAGGAAGT**T**AG

3bp deletion    ATCTC**---**AGGACTAGGAAGTAG

General concepts: edit distance

Reference CGTCCCTCAGATTGGAA—CCTCGCTT

Read TCCCTCAGAATGGAAACCTCGCT

Edit distance =3

General concepts: edit distance

# Building an index

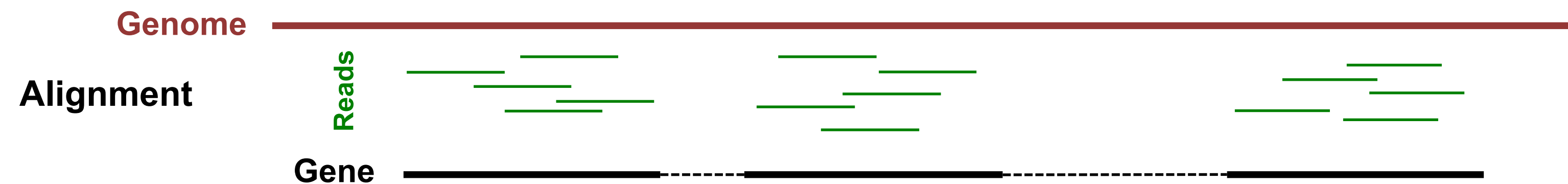
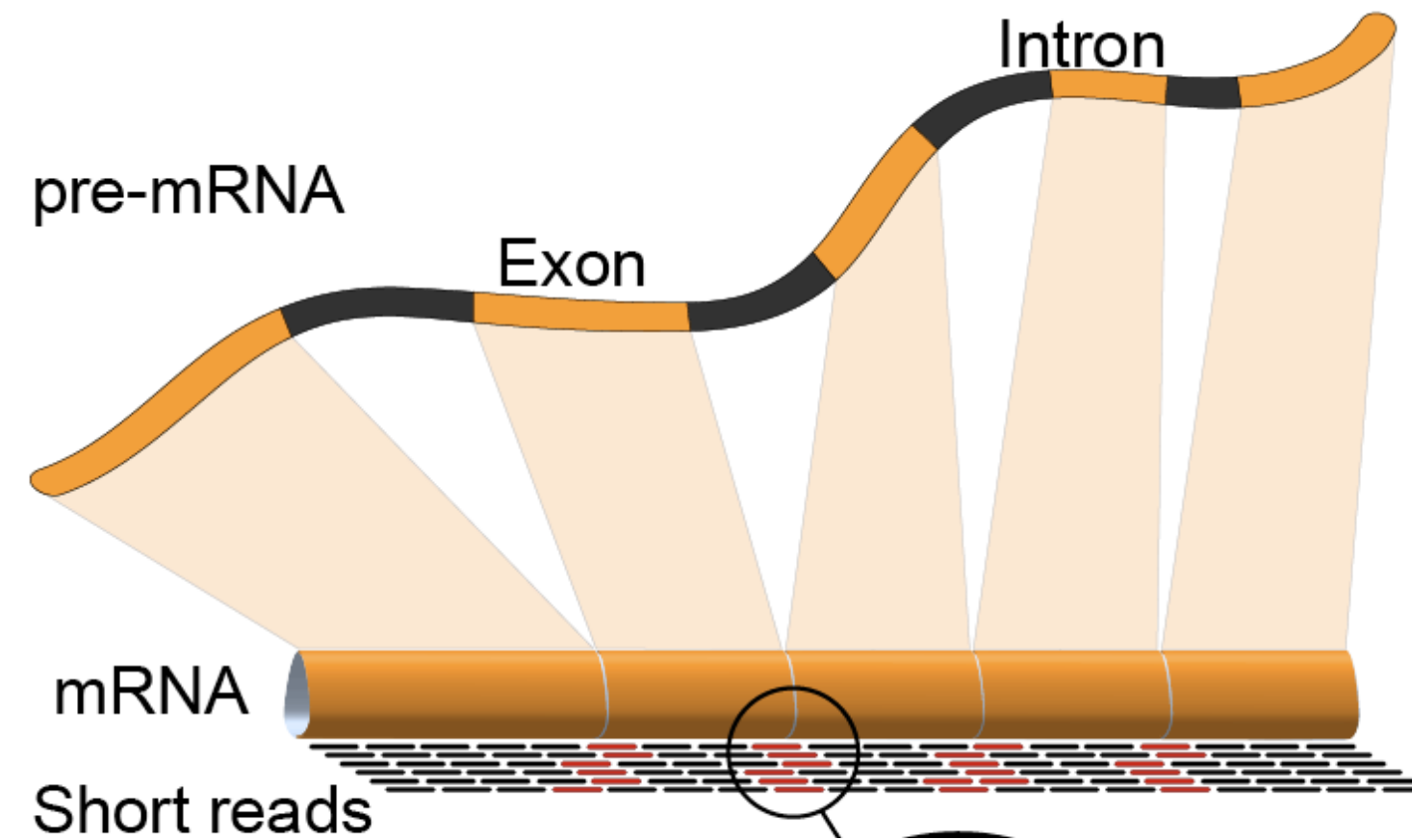
- ▶ For each read we need to scan the entire corpus as fast as possible
- ▶ Having an index of the reference genome provides an efficient way to search
- ▶ Once index is built, it can be queried any number of times
- ▶ Indexes are genome and tool-specific



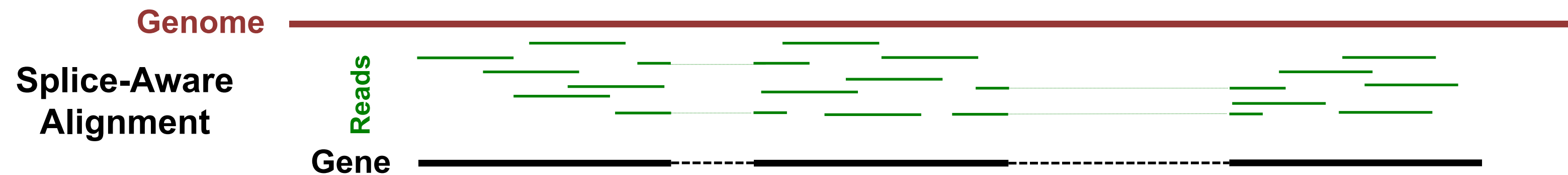


# Alignment tools can be grouped based on indexing method

- ▶ Some examples include:
  - ▶ Hash-based
  - ▶ Suffix arrays
  - ▶ Burrows-Wheeler Transform



Versus



# Splice-aware alignment

Splice-aware alignment tools:

HISAT2, STAR, MapSplice, SOAPSsplice, Passion, SpliceMap,  
RUM, ABMapper, CRAC, GMAP-GSNAP, HMMSplicer, Olego,  
BLAT

There are excellent aligners available that are not splice-aware. These are useful for aligning directly to genes. However, you will lose isoform information.

Bowtie2, BWA, Novoalign (not free), SOAPaligner

# Splice-aware alignment



- ▶ Use the genome and GTF from the same source (i.e. Ensembl, NCBI, UCSC)
- ▶ Choose an aligner that can allow for a read to be “split” across distant regions to account for splice events
- ▶ Evaluate your computational resources and use an aligner that would work best within the confines of the available memory and CPU

## Alignment for RNA-seq

