

Class 9: Structural Bioinformatics

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PDB Statistics

The PDB is the main database for structural information on biomolecules. Let's see what it contains

Download a CSV file from the PDB site (accessible from "Analyze" > "PDB Statistics" > "by Experimental Method and Molecular Type". Move this CSV file into your RStudio project and use it to answer the following questions:

```
db<- read.csv("PDB.csv")
#db
```

```
knitr::kable(db)
```

Molecular.Type	X.ray	EM	NMR	Multiple.methods	Neutron	Other	Total
Protein (only)	152,809	9,421	12,117	191	72	32	174,642
Protein/Oligosaccharide	9,008	1,654	32	7	1	0	10,702
Protein/NA	8,061	2,944	281	6	0	0	11,292
Nucleic acid (only)	2,602	77	1,433	12	2	1	4,127
Other	163	9	31	0	0	0	203
Oligosaccharide (only)	11	0	6	1	0	4	22

Q1: What percentage of structures in the PDB are solved by X-Ray and Electron Microscopy.

```
xray.total <- sum(as.numeric(gsub(",","",db$X.ray) ))
em.total <- sum(as.numeric(gsub(",","",db$EM) ))
```

Turn this into a function to get the total of all the columns

```
# x will be the input
sumcomma <- function(x) {
  #substitutue the comma and convert to numeric
  sum(as.numeric(gsub(",", "", x) ) )
}
```

For Xray:

```
sumcomma(db$X.ray)/sumcomma(db$Total)
```

```
[1] 0.8590264
```

For EM:

```
round(sumcomma(db$EM)/sumcomma(db$Total), 2)
```

```
[1] 0.07
```

Q2: What proportion of structures in the PDB are protein?

```
round( sumcomma(db$Total[1]) / sumcomma(db$Total) , 2)
```

```
[1] 0.87
```

Q3: Type HIV in the PDB website search box on the home page and determine how many HIV-1 protease structures are in the current PDB?

SKIPPED

Visualizing the HIV-1 protease structure

Q4: Water molecules normally have 3 atoms. Why do we see just one atom per water molecule in this structure?

The structure is too low a resolution to see H atoms. You need a sub 1 Angstrom resolution to see Hydrogen.

Q5: There is a critical “conserved” water molecule in the binding site. Can you identify this water molecule? What residue number does this water molecule have

HOH 308

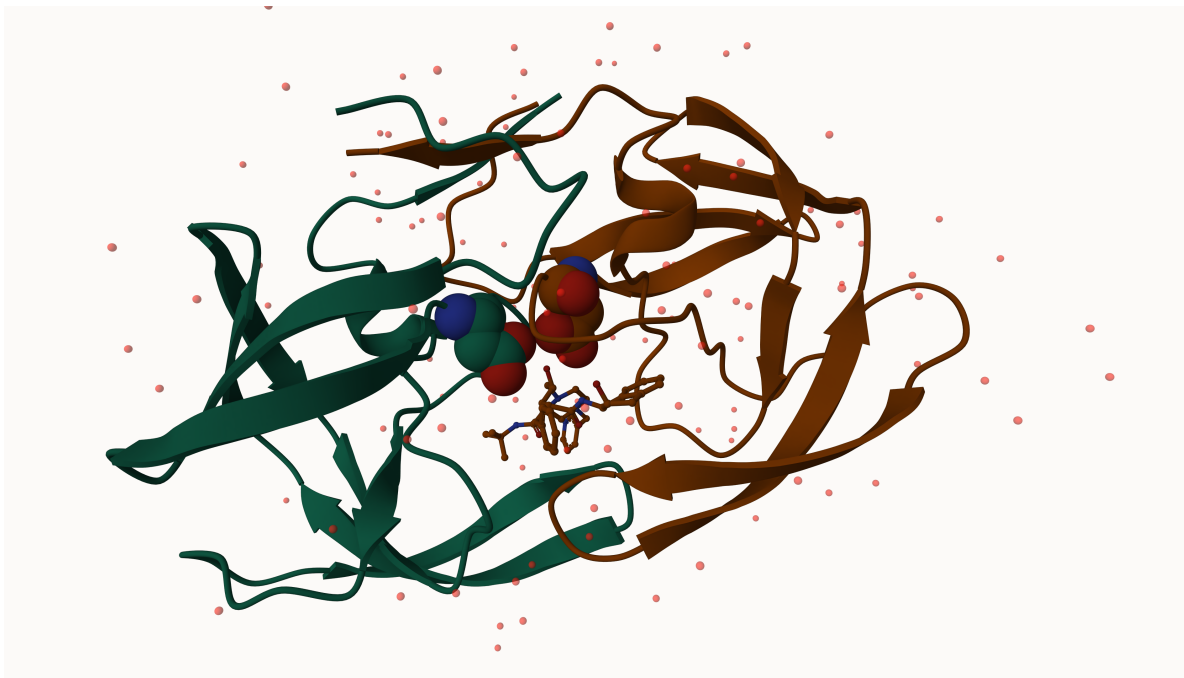


Figure 1: HIV-PR structure from MERK with a bound drug

Working with structures in R

We can use the `bio3d` package to read and perform bioinformatics calculations on PDB structures.

```
library(bio3d)

pdb <- read.pdb("1hsg")
```

Note: Accessing on-line PDB file

```
pdb
```

```
Call: read.pdb(file = "1hsg")
```

```
Total Models#: 1
Total Atoms#: 1686, XYZs#: 5058 Chains#: 2 (values: A B)
```

Protein Atoms#: 1514 (residues/Calpha atoms#: 198)
 Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)

Non-protein/nucleic Atoms#: 172 (residues: 128)
 Non-protein/nucleic resid values: [HOH (127), MK1 (1)]

Protein sequence:

PQITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWKPKMIGGIGGFIKVRQYD
 QILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNFPQITLWQRPLVTIKIGGQLKE
 ALLDTGADDTVLEEMSLPGRWKPKMIGGIGGFIKVRQYDQILIEICGHKAIGTVLVGPTP
 VNIIGRNLLTQIGCTLNF

+ attr: atom, xyz, seqres, helix, sheet,
 calpha, remark, call

`attributes(pdb)`

\$names

[1] "atom" "xyz" "seqres" "helix" "sheet" "calpha" "remark" "call"

\$class

[1] "pdb" "sse"

`head(pdb$atom)`

	type	eleno	elety	alt	resid	chain	resno	insert	x	y	z	o	b
1	ATOM	1	N	<NA>	PRO	A	1	<NA>	29.361	39.686	5.862	1	38.10
2	ATOM	2	CA	<NA>	PRO	A	1	<NA>	30.307	38.663	5.319	1	40.62
3	ATOM	3	C	<NA>	PRO	A	1	<NA>	29.760	38.071	4.022	1	42.64
4	ATOM	4	O	<NA>	PRO	A	1	<NA>	28.600	38.302	3.676	1	43.40
5	ATOM	5	CB	<NA>	PRO	A	1	<NA>	30.508	37.541	6.342	1	37.87
6	ATOM	6	CG	<NA>	PRO	A	1	<NA>	29.296	37.591	7.162	1	38.40
	segid elesy charge												
1	<NA>	N	<NA>										
2	<NA>	C	<NA>										
3	<NA>	C	<NA>										
4	<NA>	O	<NA>										
5	<NA>	C	<NA>										
6	<NA>	C	<NA>										

Q7: How many amino acid residues are there in this pdb object?

From the output above I can see: 198

Q8: Name one of the two non-protein residues?

HOH (Water)

Q9: How many protein chains are in this structure?

2 chains (A and B)

Predicting functional motions of a single structure

Read an ADK structure

```
adk <- read.pdb("6s36")
```

Note: Accessing on-line PDB file

PDB has ALT records, taking A only, rm.alt=TRUE

```
adk
```

Call: read.pdb(file = "6s36")

Total Models#: 1

Total Atoms#: 1898, XYZs#: 5694 Chains#: 1 (values: A)

Protein Atoms#: 1654 (residues/Calpha atoms#: 214)

Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)

Non-protein/nucleic Atoms#: 244 (residues: 244)

Non-protein/nucleic resid values: [CL (3), HOH (238), MG (2), NA (1)]

Protein sequence:

```
MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGDMRLRAAVKSGSELGKQAKDIMDAGKLV  
DELVIALVKERIAQEDCRNGFLLDGFPRTPQADAMKEAGINVDYVLEFDVPDELIVDKI  
VGRRVHAPSGRVYHVKNPPKVEGKDDVTGEELTTRKDDQEETVRKRLVEYHQMTAPLIG  
YYSKEAEAGNTKYAKVDGTPVAEVRADLEKILG
```

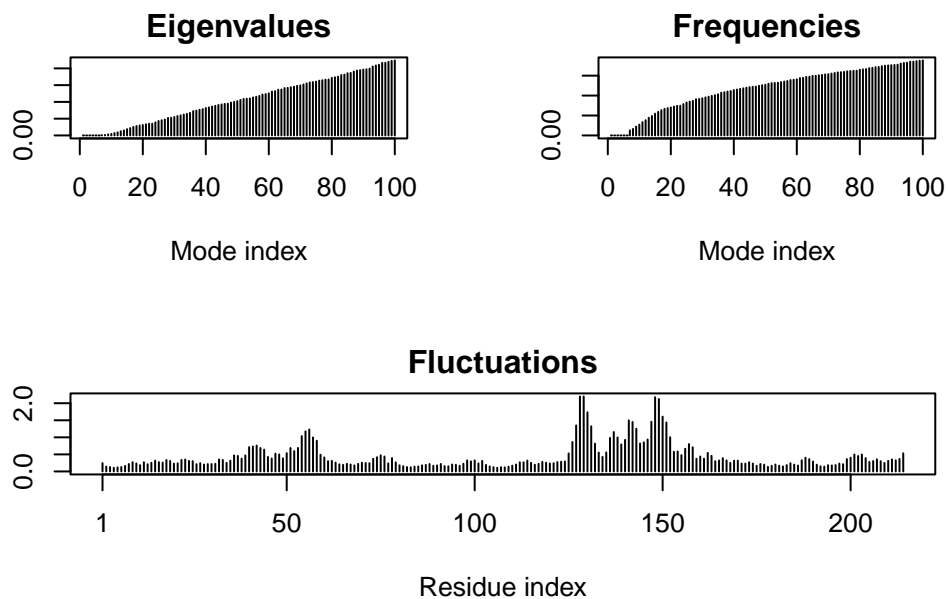
```
+ attr: atom, xyz, seqres, helix, sheet,  
      calpha, remark, call
```

Perform a prediction of flexibility with a technique called NMA (normal mode analysis).

```
# Perform flexibility prediction  
m <- nma(adk)
```

```
Building Hessian...      Done in 0.03 seconds.  
Diagonalizing Hessian... Done in 0.37 seconds.
```

```
plot(m)
```



Write out a “movie” (a.k.a trajectory) of the motion for viewing in M01star.

```
mktrj(m, file="adk_m7.pdb")
```