

REVEALING SMRT™ BIOLOGY

On-site System Training Workflow

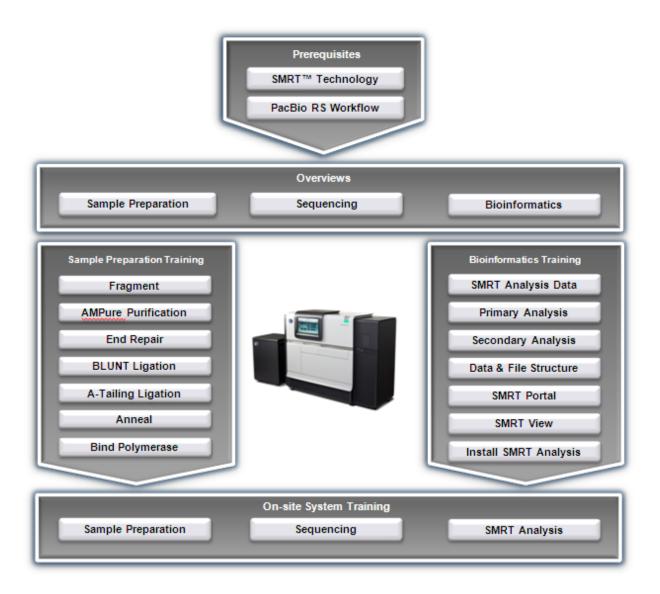
Pacbio RSII系统标准客户培训流程

FU Wei Senior Application Specialist



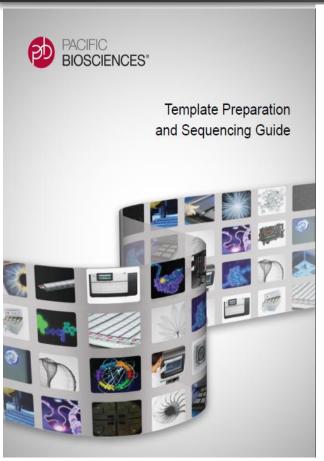
Login to the **PacBio Portal** and perform following tasks

http://www.pacificbiosciences.com/support/pubmap/training.html





Download documents





Procedure & Checklist -2 kb Template Preparation and Sequencing



Procedure & Checklist -10 kb Template Preparation and Sequencing

Before You Begin

To perform this procedure, you must have the Pacific Biosciences®:

- . DNA Template Prep Kit (verify you have the correct kit for your insert size)
- · DNA/Polymerase Binding Kit
- DNA Sequencing Kit
- DNA Control Complex
- SMRT® Cells for standard sequencing

This procedure can be used to prepare 1500 bp to 3 kb libraries from 500 ng up to 750 ng of sheared and concentrated DNA. If preparing larger amounts of DNA, scale all the reaction volumes proportionally (e.g., if the input amount of DNA is double the amount set forth in this procedure, then double all the reaction volumes listed in the tables).

Insert Size Target	Insert Size Range
2 kb	~ 1500 bp to 3

For DNA sequencing, it is recommended

Fragment and Concentrate DN

Use a Covaris® System (with Adaptive Fi concentrate your DNA sample. Note that ple loss is to be expected as a result of th ficient amounts of starting DNA in order to (25 ng/µL) for the End-Repair reaction.

Target Shear	Tube	Shearing	Temp
Size		Volume	Chile
2.0 kb	Mini-tube Clear	200.0	4°C



Sheared and

Concentrated DNA

Amount

Procedure & Checklist -Low-Input 10 kb Library Preparation and Sequencing (MagBead Station)

Before You Begin

To perform this procedure, you must have the PacBio®

DNA Template Prep Kit (verify you have the correct kit for your insert size)
 DNA/Polymerase Binding Kit

Ligation

- MagBead Kit
- DNA Sequencing Kit
- DNA Control Complex
- SMRT® Cells for standard sequencing.

This procedure can be used to prepare 10 kb libraries from 1µg up to 5 µg of sheared and concentrated DNA. If preparing libraries with DNA input amounts great than 5 µg, scale all the reaction volumes proportionally (e.g., if the input amount of DNA is 10 µg, which is double the maximum input amount set forth in this procedure, then double all the reaction volumes listed in the tables).

DNA Damage

Insert Size Target	Insert Size Range	Sheared and Concentrated DNA Amount	Ligation	DNA Damage Repair
10 kb	8 kb to 12 kb	1 to 5 µg	Blunt	Required

For DNA sequencing, it is recommended to perform titrations to ensure optimal loading.

Fragment and Concentrate DNA

Use a Covaris® g-TUBE™ to shear your DNA sample. The most up-to-date guidance on how to use the g-TUBE, along with recommended centrifuges and centrifugation speeds, can be found in the g-TUBE user manual available for download from the Covaris website. Depending upon the quality of your sample, approximately 20% to 50% sample loss is to be expected as a result of the shearing and concentration process. Therefore, be sure to have sufficient amounts of starting DNA in order to have at least 1 µg of sheared and concentrated DNA (30 ng/µL) for the Damage Repair reaction.

Note that a Hydroshear[®] Shearing Device can also be used to shear your DNA sample. However, because Hydrodynamic shearing performance can vary with each shearing assembly, we recommend optimizing the shearing whenever a new shearing assembly is used.

Before You Begin

To perform this procedure, you must have the Pacific Biosciences®:

- . DNA Template Prep Kit (verify you have the correct kit for your insert size)
- · DNA/Polymerase Binding Kit
- DNA Sequencing Kit
- DNA Control Complex
- SMRT® Cells for standard sequencing.

This procedure can be used to prepare 10 kb libraries from 5 µg of sheared and concentrated DNA. For sheared DNA libraries and PCR products greater than 4 kb, any DNA damage (generated during DNA extraction and PCR amplifications) must be repaired using the DNA Damage Repair reagents provided by Pacific Biosciences and according to the instructions as set forth in this Procedure. Common types of damage may include abasic sites, cytosine deamination, and oxidation.

> all the reaction volumes proportionally (e.g., if the input amount of procedure, then double all the reaction volumes listed in the tables).

ge	Sheared and Concentrated DNA Amount	Ligation	DNA Damage Repair
	5 µg	Blunt	Required

to perform titrations to ensure optimal loading.

А

near your DNA sample using the following conditions. Note that a, approximately 20% to 50% sample loss is to be expected as a result in Therefore, be sure to have sufficient amounts of starting DNA in I concentrated DNA (140 ng/µL) for the Damage Repair reaction.

ance can vary with each shearing assembly, we recommend optimiza assembly is used.

Speed Code	Cycles	Shearing Volume
SC8 - SC10	20	200 µL

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3

Lab equipment and supplies

Covaris or Hydroshear



or



Agilent 2100 Bioanalyzer



- QubitTM Quantitation Platform
- Fluorometer



• Agencourt ® AMPureXP® Beads



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- Low bind tubes
- PCR System
- Centrifuge (Plate)
- Vortex Genie



P6/C4 reagents&Consumables

厂家	类型	货号	描述
		000-448-888	Sample Plate (50 pack)
		000-882-901	Sample Plate Septa (50 pack)
		000-947-365	Mixing Plate (160 pack)
	耗材	100-243-500	Mixing Plate (40 pack)
	4673	100-237-900	Sequencing plate septa
		001-292-541	Tube septa
		100-154-200	Pipet Tips (10 racks)
		001-252-068	Pipet Tips (50 racks)
	试剂	100-171-800	SMRT Cell 8Pac V3 (8 Cells)
		100-209-300	SMRT Cell Oil
PCB		100-133-600	MagBead Kit
		100-265-900	AMPure PB (5ml)
		100-259-100	SMRTbell Template Prep Kit 1.0
	FF-6 11.7	100-286-300	SMRTbell HT Template Prep Kit 1.0
		100-356-200	DNA Sequencing Reagent 4.0
		100-356-400	DNA Sequencing Reagent 4.0 (10 package)
		100-372-700	DNA/Polymerase Binding Kit 6.0 V2
		100-356-500	DNA Internal Control Complex (P6)
	其他	100-514-900	SMRTbell TM Barcoded Adapter Complete Prep Kit - 96
		100-465-900	SMRTbell™ Damage Repair Kit
		100-466-100	Barcoded Universal F/R Primers Plate - 96

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On-site System Training

- Duration: 5 days
- Covers the end-to-end workflow:



- Primary Users
 - Scientists
 - Research Associates
 - Bioinformaticians

- Standard Training
 - Lambda only
 - 10 kb
 - 5 days total
 - Standard Training Kit
 - Help customers with their samples once training is complete

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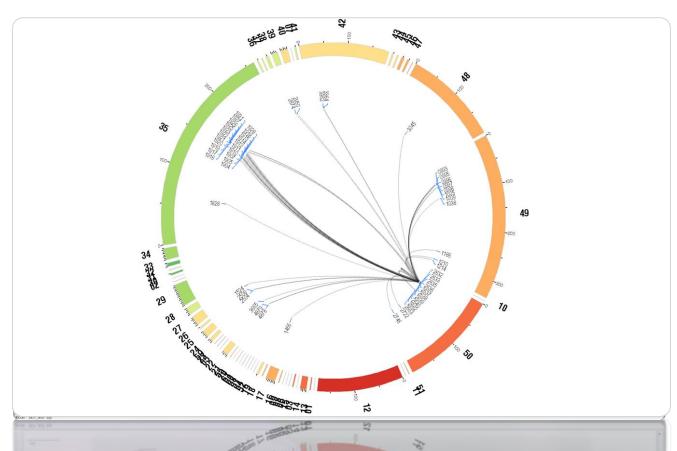
Training Agenda:

	Time	Day 1 (Monday)	Day 2 (Tuesday)	Day 3 (Wednesday)	Day 4 (Thursday)
	l l	Training Overview	10 kb Library Construction (Contd.)	Annealing/Bindinfg	Data Analysis
	9:00 AM	Technology Overview	Heat Kill Ligase	Data Analysis Overview	
	9:30 AM	Workflow Overview	Exo Treatment	•SMRT Porta	•SMRT Portal
	10:00 AM	vvorkilow Overview	Double Ampure Purification	Calculator Overview	•SMRT View
	10:30 AM		Quantitation, Bioanalyzer, Break		
	11:00 AM	10 kb Lambda Library Shearing	Quantitation, Bioanalyzer, Break	Hands On Pactice	
	11:30 AM	Purification of sheared DNA	MDC Diadia	Calculator	Metrics Review
	12:00 PM	Lunch Break	MBS Binding	Lunch Break	Lunch Break
	1:00 PM	Quantitation (Qubit, nanodrop). BioAnalyzer		Troubleshooting Overview	Instrument RS Overview
	1:30 PM	nansarspy. Elia sially25	Training Run RS Remote Set Up	****	
	2:00 PM	10Kb Library prep		RS remote Overview	Laboratory Setup overview
	2:30 PM	PreCR	Instrument Run	No remote overwew	Educationy Cottap ever new
	3:00 PM	End Repair	mstrument Kun		
	3:30 PM	Purify			
	4:00 PM	Quantitation/Quality	Additional Training	Hands On Pactice RS remote	
P	4:30 PM	Blunt Ligation (safe stopping point), Store at 4 C overnight			
α	5:00 PM	Review: Day 1	Review: Day 2	Review: Day 3	Review: Day 4
	ZHIIMA'	A Gene Group Company			DIOCOILING

Sequencing Workflow

Template Preparation

Run Design Polymerase Binding Instrument Run Primary Analysis Secondary Analysis Tertiary Analysis



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SMRT POR SARROUMER View



After Training

- Sign-off training completion form
- Complete the quiz
- Follow up on bioinformatics by PacBio Tech Support
 - Technical bioinformatics issues
 - Q&A
- Contact PacBio Tech support
 - Toll Free Telephone +1 (877) 920-7222
 - Email <u>techsupport@pacificbiosciences.com</u>
 - Logging a case on <u>PacBio Portal</u>

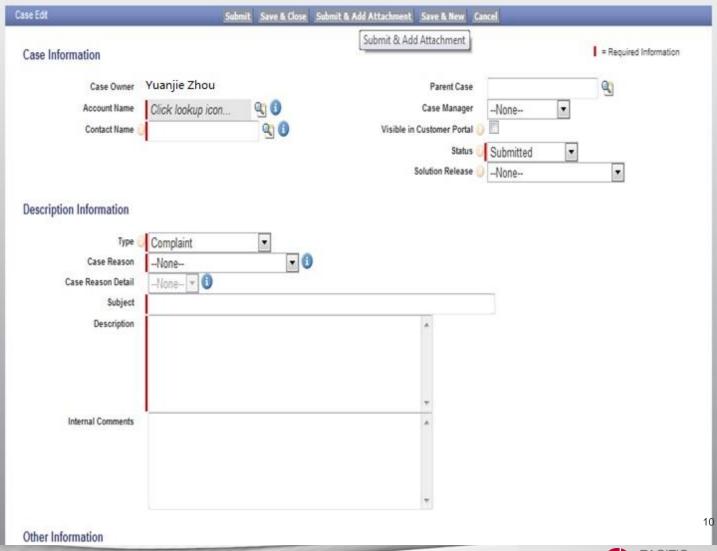






Case Edit

	Search Search All search Advanced Search
	Create New
	Recent Items
	2011-05-23 Office Hour -Low Yield Troubleshooting, Error Messages, PM
	2011-05-02 Office Hour - Web Based Calculator
	2011-04-18 Office Hour - Release v1.2.1 Field Webinar
	2011-03-28 Office Hour - SMRT Cell Loading
	Repair stage pi
	2011-03-16 Office Hour - Annealing and Binding Calculator
	Cholera Webinar by Eric Schadt
	Recorded Webinar - Release v1.2 End-User Training
https://na5.sale	sforce.com/500/e





SMRT® Technology



SMRT® Cell

Thank you for attention

