## Cardiomyocyte interactome, additional KEGG & Gene Ontology analyses

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# R script for KEGG pathway analysis of cardiomyocyte RNA interactome proteins

Loading libraries and creating custom functions:

```
library("biomaRt")
library("gdata")
library("GO.db")
library("KEGGREST")
library("ggplot2")
library("grid")
library("grid")
library("scales")

# function for conversion of Entrez GeneIDs to KEGG gene IDs
```

```
keggConv.batch <- function(x, max = 100, org = "mmu", id.type = "ncbi-geneid") {</pre>
    if (max > 100) {
         on.exit(print("Maximum number of IDs at a given time is 100"))
        x \leftarrow paste(id.type, x, sep = ":")
         if (length(x > 100)) {
             d1 <- split(x, ceiling(seq_along(x)/max))</pre>
             s1 <- lapply(d1, function(y) {</pre>
                 keggConv(org, y)
             })
             return(unlist(s1))
        } else {
             d1 <- split(x, ceiling(seq_along(x)/10))</pre>
             s1 <- lapply(d1, function(y) {</pre>
                 keggConv(org, y)
             })
             return(unlist(s1))
        }
    }
}
alternative = "greater"
p.adjust.method = "fdr"
```

### Loading data tables.

```
load("data/kegg.brite.rda")
load("data/interactome.rda")
load("data/wcl.rda")
```

### Fetching KEGG identifiers:

```
ids <- unlist(lapply(strsplit(kegg.brite$C, " "), function(x) x[1]))
rownames(kegg.brite) <- ids
total.keggIDs <- keggLink("mmu", "pathway")
save(total.keggIDs, file = "data/total.keggIDs.rda")</pre>
```

We have found a total of 25904 which are used for mapping the WCL and interactome data.

Mapping of WCL protein IDs to KEGG IDs and testing for enrichments against background of all KEGG proteins contained in KEGG pathways.

```
# retrieved Entrez IDs from Biomart
mouse <- useMart("ensembl", dataset = "mmusculus_gene_ensembl")
human <- useMart("ensembl", dataset = "hsapiens_gene_ensembl")
attribs <- listAttributes(mouse)
pages <- attributePages(mouse)</pre>
```

```
hsap.attribs <- listAttributes(human)</pre>
entrez_ids <- getBM(attributes = c("ensembl_gene_id", "entrezgene"), values = wcl[,</pre>
    "ensembl_gene_id"], filters = "ensembl_gene_id", mart = mouse)
wcl.human_homologs <- getBM(attributes = c("ensembl_gene_id", "hsapiens_homolog_ensembl_gene"),</pre>
    values = wcl[, "ensembl_gene_id"], filters = "ensembl_gene_id", mart = mouse)
# remove ensembl gene ids which have duplicated entrez ids
entrez ids <- entrez ids[-which(duplicated(entrez ids$ensembl gene id)), ]</pre>
wcl <- merge(wcl, entrez_ids, by.x = "ensembl_gene_id", by.y = "ensembl_gene_id",
    all.x = T)
wcl.entrezIDs <- unique(wcl[!is.na(wcl$entrezgene), ]$entrezgene)</pre>
# this retrieval is fairly slow, therefore the results were written to
# './data'
wcl.keggIDs <- keggConv.batch(wcl.entrezIDs)</pre>
save(wcl.keggIDs, file = "data/wcl.keggIDs.rda")
wcl.keggQ <- lapply(wcl.keggIDs, function(x) keggGet(x))</pre>
save(wcl.keggQ, file = "data/wcl.keggQ.rda")
wcl.pathways <- unique(unlist(lapply(strsplit(names(unlist(lapply(wcl.keggQ,</pre>
    function(x) x[[1]]$PATHWAY))), "\\."), function(x) x[3]))
save(wcl.pathways, file = "data/wcl.pathways.rda")
wcl.pathways.genes <- lapply(wcl.pathways, function(x) keggLink("genes", x))</pre>
names(wcl.pathways.genes) <- wcl.pathways</pre>
save(wcl.pathways.genes, file = "data/wcl.pathways.genes.rda")
wcl.pathways.genes.entrez_ids <- unique(gsub("mmu:", "", as.character(unlist(wcl.pathways.genes))))</pre>
wcl.df <- kegg.brite[gsub("mmu", "", wcl.pathways), ]</pre>
wcl.df$ID <- rownames(wcl.df)</pre>
wcl.df$total <- rep(0, nrow(wcl.df))</pre>
wcl.df$total <- sapply(rownames(wcl.df), function(x) length(wcl.pathways.genes[[paste("mmu",
    x, sep = "")]]))
wcl.df$count <- rep(0, nrow(wcl.df))</pre>
wcl.df$frac <- rep(0, nrow(wcl.df))</pre>
for (i in rownames(wcl.df)) {
    # print(i)
    kL1 <- keggLink("mmu", paste("mmu", i, sep = ""))</pre>
    wcl.df[i, ]$count <- length(which(wcl.keggIDs %in% kL1))</pre>
    wcl.df[i, ]$frac <- round(length(which(wcl.keggIDs %in% kL1))/length(kL1) *</pre>
        100, 2)
}
# extract list of IDs in pathway
wcl.in_path.IDs <- lapply(rownames(wcl.df), function(x) {</pre>
    kL1 <- keggLink("mmu", paste("mmu", x, sep = ""))</pre>
    in_path <- wcl.keggIDs[which(wcl.keggIDs %in% kL1)]</pre>
})
names(wcl.in_path.IDs) <- rownames(wcl.df)</pre>
```

```
# perform Fisher's Exact Test for each category
bkgd <- length(unique(total.keggIDs))</pre>
smpl <- length(wcl.keggIDs)</pre>
ftl <- apply(wcl.df[1, ], 1, function(x) {</pre>
    ct <- as.integer(x["count"])
    tt <- as.integer(x["total"])</pre>
    m1 <- matrix(c(ct, tt, smpl - ct, bkgd - tt), 2, 2)</pre>
    fisher.test(m1, alternative = alternative)
})
wcl.df$ft_pval <- unlist(lapply(ftl, function(x) {</pre>
    x$p.value
}))
wcl.df$ft_OR <- unlist(lapply(ftl, function(x) {</pre>
    x$estimate
}))
wcl.df$ft_fdr <- p.adjust(wcl.df$ft_pval, method = "fdr")</pre>
save(wcl.df, file = "data/wcl.df.rda")
```

Mapping of Interactome protein IDs to KEGG IDs and testing for enrichments against background of WCL proteins contained in KEGG pathways.

```
interactome.entrez_ids <- getBM(attributes = c("ensembl_gene_id", "entrezgene"),</pre>
    values = interactome[, "ensembl_gene_id"], filters = "ensembl_gene_id",
interactome.human_homologs <- getBM(attributes = c("ensembl_gene_id", "hsapiens_homolog_ensembl_gene"),
    values = interactome[, "ensembl_gene_id"], filters = "ensembl_gene_id",
# remove ensembl_gene_ids which have duplicated entrez_ids
interactome.entrez ids <- interactome.entrez ids[-which(duplicated(interactome.entrez ids$ensembl gene
interactome <- merge(interactome, interactome.entrez_ids, by.x = "ensembl_gene_id",</pre>
    by.y = "ensembl_gene_id", all.x = T)
interactome.entrezIDs <- unique(interactome[!is.na(interactome$entrezgene),</pre>
    ]$entrezgene)
save(interactome.entrezIDs, file = "data/interactome.entrezIDs")
interactome.keggIDs <- keggConv.batch(interactome.entrezIDs)</pre>
save(interactome.keggIDs, file = "data/interactome.keggIDs.rda")
interactome.keggQ <- lapply(interactome.keggIDs, function(x) keggGet(x))</pre>
save(interactome.keggQ, file = "data/interactome.keggQ.rda")
interactome.pathways <- unique(unlist(lapply(strsplit(names(unlist(lapply(interactome.keggQ,
    function(x) x[[1]]$PATHWAY))), "\\."), function(x) x[3]))
save(interactome.pathways, file = "data/interactome.pathways.rda")
interactome.pathways.genes <- lapply(interactome.pathways, function(x) keggLink("genes",</pre>
    x))
```

```
names(interactome.pathways.genes) <- interactome.pathways</pre>
save(interactome.pathways.genes, file = "data/interactome.pathways.genes.rda")
interactome.pathways.genes.entrez_ids <- unique(gsub("mmu:", "", as.character(unlist(interactome.pathwa
# create dataframe for counting hits in pathways
interactome.df <- kegg.brite[gsub("mmu", "", interactome.pathways), ]</pre>
interactome.df$source <- rep("Interactome", nrow(interactome.df))</pre>
interactome.df$ID <- rownames(interactome.df)</pre>
\# we are now using WCL as background to test for enrichment
i1 <- intersect(rownames(interactome.df), rownames(wcl.df))</pre>
interactome.df$total <- rep(0, nrow(interactome.df))</pre>
interactome.df[i1, ]$total <- wcl.df[i1, ]$count</pre>
interactome.df$count <- rep(0, nrow(interactome.df))</pre>
interactome.df$frac <- rep(0, nrow(interactome.df))</pre>
for (i in rownames(interactome.df)) {
    kL1 <- keggLink("mmu", paste("mmu", i, sep = ""))</pre>
    interactome.df[i, ]$count <- length(which(interactome.keggIDs %in% kL1))</pre>
    interactome.df[i, ]$frac <- round(length(which(interactome.keggIDs %in%))</pre>
        kL1))/length(kL1) * 100, 2)
}
# extract list of IDs in pathway
interactome.in_path.IDs <- lapply(rownames(interactome.df), function(x) {</pre>
    kL1 <- keggLink("mmu", paste("mmu", x, sep = ""))</pre>
    in_path <- interactome.keggIDs[which(interactome.keggIDs %in% kL1)]</pre>
})
# perform Fisher's Exact Test for each category
bkgd <- length(unique(wcl.keggIDs))</pre>
smpl <- length(interactome.keggIDs)</pre>
ftl <- apply(interactome.df, 1, function(x) {</pre>
    ct <- as.integer(x["count"])</pre>
    tt <- as.integer(x["total"])</pre>
    m1 <- matrix(c(ct, tt, smpl - ct, bkgd - tt), 2, 2)</pre>
    fisher.test(m1, alternative = alternative)
})
interactome.df$ft_pval <- unlist(lapply(ftl, function(x) {</pre>
    x$p.value
}))
interactome.df$ft_OR <- unlist(lapply(ftl, function(x) {</pre>
    x$estimate
}))
interactome.df$ft_fdr <- p.adjust(interactome.df$ft_pval, method = p.adjust.method,
    n = nrow(wcl.df)
save(interactome.df, file = "data/interactome.df.rda")
```

KEGG contains three levels/hierarchies (A>B>C), here we summarize the enrichment at B level:

```
# -----interactome-summarizing data at 'B' level before doing Fisher's
# Exact test-----
interactome.B.df <- data.frame(matrix(ncol = 5, nrow = length(unique(interactome.df$B))))</pre>
colnames(interactome.B.df) <- c("B", "A", "total", "count", "source")</pre>
interactome.B.df$B <- unique(interactome.df$B)</pre>
interactome.B.df$A <- sapply(unique(interactome.df$B), function(x) {
    A <- unique(interactome.df[which(interactome.df$B %in% x), "A"])
})
interactome.B.df$source <- rep("Interactome", nrow(interactome.B.df))</pre>
interactome.B.df$total <- sapply(unique(interactome.df$B), function(x) {</pre>
    tot <- sum(interactome.df[which(interactome.df$B %in% x), "total"])
})
interactome.B.df$count <- sapply(unique(interactome.df$B), function(x) {</pre>
    count <- sum(interactome.df[which(interactome.df$B %in% x), "count"])</pre>
})
bkgd <- length(unique(wcl.keggIDs))</pre>
smpl <- length(interactome.keggIDs)</pre>
ftl <- apply(interactome.B.df, 1, function(x) {</pre>
    ct <- as.integer(x["count"])
    tt <- as.integer(x["total"])</pre>
    m1 <- matrix(c(ct, tt, smpl - ct, bkgd - tt), 2, 2)
    fisher.test(m1, alternative = alternative)
})
interactome.B.df$ft_pval <- unlist(lapply(ftl, function(x) {</pre>
    x$p.value
}))
interactome.B.df$ft_OR <- unlist(lapply(ftl, function(x) {</pre>
    x$estimate
}))
interactome.B.df$ft_fdr <- p.adjust(interactome.B.df$ft_pval, method = p.adjust.method,
    n = nrow(wcl.df)
save(interactome.B.df, file = "data/interactome.B.df.rda")
```

Interactome proteins are annotated as RNA-related and un-related proteins, based on annotation analysis (upstream of these steps):

```
interactome.go_rna_unrelated.df$source <- rep("GO_RNA_unrelated", nrow(interactome.go_rna_unrelated.df)
interactome.go_rna_unrelated.df$ID <- rownames(interactome.go_rna_unrelated.df)
interactome.go_rna_unrelated.df$total <- rep(0, nrow(interactome.go_rna_unrelated.df))</pre>
# we are now using WCL as background to test for enrichment
i1 <- intersect(rownames(interactome.go_rna_unrelated.df), rownames(wcl.df))</pre>
interactome.go_rna_unrelated.df[i1,]$total <- wcl.df[i1,]$count</pre>
interactome.go rna unrelated.df$count <- rep(0, nrow(interactome.go rna unrelated.df))
for (i in rownames(interactome.go_rna_unrelated.df)) {
  kL1 <- keggLink("mmu", paste("mmu", i, sep = ""))</pre>
  interactome.go_rna_unrelated.df[i, ]$count <- length(which(interactome.go_rna_unrelated.keggIDs %in%)
# extract list of IDs in pathway
interactome.go_rna_unrelated.in_path.IDs <- lapply(rownames(interactome.go_rna_unrelated.df), function(
  kL1 <- keggLink("mmu", paste("mmu", x, sep = ""))</pre>
  in_path <- interactome.go_rna_unrelated.keggIDs[which(interactome.go_rna_unrelated.keggIDs %in% kL1)]</pre>
names(interactome.go_rna_unrelated.in_path.IDs) <- rownames(interactome.go_rna_unrelated.df)</pre>
# perform Fisher's Exact Test for each category
# Using WCL as background
bkgd <- length(unique(wcl.keggIDs))</pre>
smpl <- length(interactome.go rna unrelated.keggIDs)</pre>
ftl <- apply(interactome.go_rna_unrelated.df, 1, function (x) {
  ct <- as.integer(x["count"])</pre>
  tt <- as.integer(x["total"])</pre>
 m1 <- matrix(c(ct, tt, smpl - ct, bkgd - tt), 2, 2)
  fisher.test(m1, alternative = alternative)
})
interactome.go_rna_unrelated.df$ft_pval <- unlist(lapply(ftl, function(x) {x$p.value}))</pre>
interactome.go_rna_unrelated.df$ft_OR <- unlist(lapply(ftl, function(x) {x$estimate}))</pre>
interactome.go_rna_unrelated.df$ft_fdr <- p.adjust(interactome.go_rna_unrelated.df$ft_pval, method = p.</pre>
save(interactome.go_rna_unrelated.df, file = "data/interactome.go_rna_unrelated.df.rda")
# summarizing data at "B" level before doing Fisher's Exact test
interactome.go_rna_unrelated.B.df <- data.frame(matrix(ncol = 5, nrow = length(unique(interactome.go_rn
colnames(interactome.go_rna_unrelated.B.df) <- c("B", "A", "total", "count", "source")</pre>
interactome.go_rna_unrelated.B.df$B <- unique(interactome.go_rna_unrelated.df$B)
interactome.go_rna_unrelated.B.df$A <- sapply(unique(interactome.go_rna_unrelated.df$B), function(x) {A
interactome.go_rna_unrelated.B.df$source <- rep("GO_RNA_unrelated", nrow(interactome.go_rna_unrelated.B
interactome.go_rna_unrelated.B.df$total <- sapply(unique(interactome.go_rna_unrelated.df$B), function(x
interactome.go_rna_unrelated.B.df$count <- sapply(unique(interactome.go_rna_unrelated.df$B), function(x</pre>
# using WCL as background
ftl <- apply(interactome.go_rna_unrelated.B.df, 1, function (x) {
  ct <- as.integer(x["count"])</pre>
  tt <- as.integer(x["total"])</pre>
  m1 <- matrix(c(ct, tt, smpl - ct, bkgd - tt), 2, 2)</pre>
  fisher.test(m1, alternative = alternative)
```

```
interactome.go_rna_unrelated.B.df$ft_pval <- unlist(lapply(ftl, function(x) {x$p.value}))
interactome.go_rna_unrelated.B.df$ft_OR <- unlist(lapply(ftl, function(x) {x$estimate}))
interactome.go_rna_unrelated.B.df$ft_fdr <- p.adjust(interactome.go_rna_unrelated.B.df$ft_pval, method
save(interactome.go_rna_unrelated.B.df, file = "data/interactome.go_rna_unrelated.B.df")</pre>
```

#### RNA-unrelated

```
#-----GO RNA related-----
interactome.go_rna_related <- interactome[-which(interactome$G0 == "unrelated"),]</pre>
interactome.go_rna_related.entrezIDs <- unique(interactome.go_rna_related[!is.na(interactome.go_rna_rel
interactome.go_rna_related.keggIDs <- keggConv.batch(interactome.go_rna_related.entrezIDs)</pre>
# we are testing this subset of "interactome", therefore we include all the pathways from "interactome"
interactome.go_rna_related.df <- interactome.df</pre>
i1 <- intersect(rownames(interactome.go_rna_related.df), rownames(wcl.df))</pre>
interactome.go_rna_related.df$total <- rep(0, nrow(interactome.go_rna_related.df))</pre>
interactome.go_rna_related.df[i1,]$total <- wcl.df[i1,]$count</pre>
interactome.go_rna_related.df$source <- rep("GO_RNA_related", nrow(interactome.go_rna_related.df))</pre>
interactome.go_rna_related.df$ID <- rownames(interactome.go_rna_related.df)</pre>
interactome.go_rna_related.df$count <- rep(0, nrow(interactome.go_rna_related.df))</pre>
interactome.go_rna_related.df$frac <- rep(0, nrow(interactome.go_rna_related.df))</pre>
for (i in rownames(interactome.go_rna_related.df)) {
 kL1 <- keggLink("mmu", paste("mmu", i, sep = ""))</pre>
  interactome.go_rna_related.df[i, ]$count <- length(which(interactome.go_rna_related.keggIDs %in% kL1)
}
# extract list of IDs in pathway
interactome.go_rna_related.in_path.IDs <- lapply(rownames(interactome.go_rna_related.df), function(x) {
  kL1 <- keggLink("mmu", paste("mmu", x, sep = ""))</pre>
  in_path <- interactome.go_rna_related.keggIDs[which(interactome.go_rna_related.keggIDs %in% kL1)]
})
names(interactome.go_rna_related.in_path.IDs) <- rownames(interactome.go_rna_related.df)</pre>
# perform Fisher's Exact Test for each category
# Using WCL as background
bkgd <- length(unique(wcl.keggIDs))</pre>
smpl <- length(interactome.go_rna_related.keggIDs)</pre>
ftl <- apply(interactome.go_rna_related.df, 1, function (x) {</pre>
  ct <- as.integer(x["count"])</pre>
  tt <- as.integer(x["total"])</pre>
 m1 <- matrix(c(ct, tt, smpl - ct, bkgd - tt), 2, 2)
 fisher.test(m1, alternative = alternative)
})
interactome.go_rna_related.df$ft_pval <- unlist(lapply(ftl, function(x) {x$p.value}))</pre>
```

```
interactome.go_rna_related.df$ft_OR <- unlist(lapply(ftl, function(x) {x$estimate}))</pre>
interactome.go_rna_related.df$ft_fdr <- p.adjust(interactome.go_rna_related.df$ft_pval, method = p.adju
save(interactome.go_rna_related.df, file = "data/interactome.go_rna_related.df")
# summarizing data at "B" level before doing Fisher's Exact test
interactome.go_rna_related.B.df <- data.frame(matrix(ncol = 5, nrow = length(unique(interactome.go_rna_
colnames(interactome.go_rna_related.B.df) <- c("B", "A", "total", "count", "source")</pre>
interactome.go_rna_related.B.df$B <- unique(interactome.go_rna_related.df$B)
interactome.go_rna_related.B.df$A <- sapply(unique(interactome.go_rna_related.df$B), function(x) {A <-
interactome.go_rna_related.B.df$source <- rep("GO_RNA_related", nrow(interactome.go_rna_related.B.df))</pre>
interactome.go_rna_related.B.df$total <- sapply(unique(interactome.go_rna_related.df$B), function(x) {t
interactome.go_rna_related.B.df$count <- sapply(unique(interactome.go_rna_related.df$B), function(x) {c</pre>
ftl <- apply(interactome.go_rna_related.B.df, 1, function (x) {</pre>
  ct <- as.integer(x["count"])</pre>
 tt <- as.integer(x["total"])</pre>
 m1 \leftarrow matrix(c(ct, tt, smpl - ct, bkgd - tt), 2, 2)
 fisher.test(m1, alternative = alternative)
})
interactome.go_rna_related.B.df$ft_pval <- unlist(lapply(ftl, function(x) {x$p.value}))</pre>
interactome.go_rna_related.B.df$ft_OR <- unlist(lapply(ftl, function(x) {x$estimate}))</pre>
interactome.go_rna_related.B.df$ft_fdr <- p.adjust(interactome.go_rna_related.B.df$ft_pval, method = p.
save(interactome.go_rna_related.B.df, file = "data/interactome.go_rna_related.B.df.rda")
```

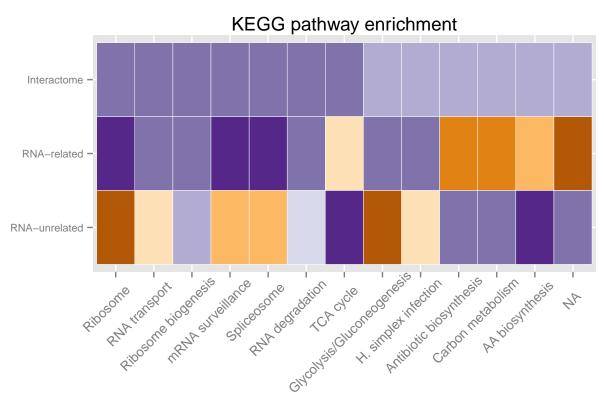
#### **RNA-related:**

### Plotting the results of the enrichment analysis:

```
library(ggplot2)
library(grid)
library(scales)
load("data/interactome.df.rda")
load("data/interactome.go_rna_unrelated.df.rda")
load("data/interactome.go_rna_related.df")
dfC <- rbind(interactome.df[, c("A", "B", "C", "ft_OR", "ft_fdr", "source", "count")],</pre>
             interactome.go_rna_related.df[, c("A", "B", "C", "ft_OR", "ft_fdr", "source", "count")],
             interactome.go_rna_unrelated.df[, c("A", "B", "C", "ft_OR", "ft_fdr", "source", "count")]
)
dfC$source <- as.factor(dfC$source)</pre>
dfC$source <- factor(dfC$source, levels = levels(dfC$source)[c(3,1,2)])</pre>
select1 <- unique(as.character(dfC[which(dfC$ft_fdr <= 0.1 & dfC$ft_OR > 1),]$C))
select1.pathIDs <- paste("mmu", unlist(lapply(strsplit(select1, "\\ "), function(x) x[1])), sep = "")</pre>
dfC <- dfC[which(dfC$C %in% select1),]</pre>
dfC$C <- as.factor(as.character(dfC$C))</pre>
dfC$ft_OR.cut <- cut(log2(dfC$ft_OR), breaks = c(-Inf,-4:4), right = F)</pre>
```

```
dfC$C <- factor(dfC$C, levels = levels(dfC$C)[dfC[dfC$source == "Interactome", "C"][order(dfC[which(dfC
# formatting labels etc for plotting
11 <- levels(dfC$ft OR.cut)</pre>
11 <- gsub("\\[", "", 11)</pre>
11 <- gsub("\\)", "", 11)
levels(dfC$ft_OR.cut) <- 11</pre>
11 <- as.character(levels(dfC$C))</pre>
11 <- unlist(lapply(strsplit(l1, " "), function(x) {</pre>
  for (i in 2:length(x)){
    if (i == 2){
      v \leftarrow x[i]
    } else {
      v <- paste(v, x[i])</pre>
  }
  return(v)
}))
levels(dfC$C) <- 11</pre>
levels(dfC$source)[2:3] <- c("RNA-related", "RNA-unrelated")</pre>
levels(dfC$C)[3] <- "Ribosome biogenesis"</pre>
levels(dfC$C)[6] <- "TCA cycle"</pre>
levels(dfC$C)[7] <- "mRNA surveillance"</pre>
levels(dfC$C)[11] <- "AA biosynthesis"</pre>
levels(dfC$C)[8] <- "H. simplex infection"</pre>
levels(dfC$C)[9] <- "Antibiotic biosynthesis"</pre>
levels(dfC$C)[12] <- "Glycolysis/Gluconeogenesis"</pre>
flevels <- levels(dfC$source)</pre>
11 \leftarrow factor(dfC\$C, levels = levels(dfC\$C)[c(1,2,3,7,4,5,6,12,8,9,10,11)])
dfC$C <- 11
p1 \leftarrow ggplot(data = dfC, aes(y = source, x = C)) +
      geom_tile(aes(fill = ft_OR.cut), colour = "white") +
      scale_fill_manual(values = brewer_pal(pal = "PuOr")(8), labels = levels(dfC$ft_OR.cut)) + #
      theme(axis.text.y = element_text(angle = 0, size = 8), axis.title = element_blank()) +
      guides(fill = guide_legend(label.position = "bottom", direction = "horizontal")) +
      theme(axis.text.x = element_text(angle = 45, vjust = 0.9, hjust = 0.8, size = 10)) +
      labs(fill = "Log2 OR") +
      scale_y_discrete(limits = rev(flevels)) +
      theme(legend.position = c(0.4,-1.92),
             legend.text = element_text(size = 4),
             legend.text.align = 0.5,
             legend.title = element_text(size = 4, vjust = 5),
             legend.key.size = unit(3.5, "mm"),
             legend.key.width = unit(3.5, "mm"),
             legend.margin = unit(0, "mm"),
             panel.margin = unit(1, "mm")) +
```





# R script for cardiovascular-associated GO term analysis of cardiomyocyte RNA interactome proteins

### Load libraries

```
library(gdata)
library(biomaRt)
library(GO.db)
library(ggplot2)

# make lists from GO.db
go.term <- as.list(GOTERM)
go.bp.offspring <- as.list(GOBPOFFSPRING)
go.cc.offspring <- as.list(GOCCOFFSPRING)</pre>
```

### Connect to Biomart

```
human <- useMart("ensembl", dataset = "hsapiens_gene_ensembl")
mouse <- useMart("ensembl", dataset = "mmusculus_gene_ensembl")</pre>
```

```
attribs <- listAttributes(mouse)
filters <- listFilters(mouse)
attribs.hsap <- listAttributes(human)</pre>
```

### Load data files

```
load("data/wcl.rda")
load("data/interactome.rda")
load("data/cv.assoc.proteins.rda")
```

### ID mapping

```
# biomaRt attribute uniprot_swissprot
mmus.cv.assoc <- getBM(attributes = c("ensembl_gene_id", "uniprot_swissprot"),</pre>
    filters = "uniprot_swissprot", values = cv.assoc.proteins[which(cv.assoc.proteins$Taxon ==
        "10090"), "ID"], mart = mouse)
hsap.cv.assoc <- getBM(attributes = c("ensembl_gene_id", "uniprot_swissprot"),
    filters = "uniprot_swissprot", values = cv.assoc.proteins[which(cv.assoc.proteins$Taxon ==
        "9606"), "ID"], mart = human)
hsap.cv.assoc.mmus.homologs <- getBM(attributes = c("ensembl_gene_id", "mmusculus_homolog_ensembl_gene"
    filters = "uniprot_swissprot", values = cv.assoc.proteins[which(cv.assoc.proteins$Taxon ==
        "9606"), "ID"], mart = human)
i1 <- intersect(unique(mmus.cv.assoc$ensembl_gene_id), interactome$ensembl_gene_id)</pre>
i2 <- intersect(unique(hsap.cv.assoc.mmus.homologs$mmusculus_homolog_ensembl_gene),</pre>
    interactome$ensembl_gene_id)
i3 <- c(i1[which(!i1 %in% intersect(i1, i2))], i2[which(!i2 %in% intersect(i1,
    i2))])
interactome.go_ids <- getBM(attributes = c("ensembl_gene_id", "go_id"), filters = "ensembl_gene_id",
    values = interactome$ensembl_gene_id, mart = mouse)
\# subtract genes which are in common between interactome and WCL
wcl <- wcl[-which(wcl$ensembl_gene_id %in% interactome$ensembl_gene_id), ]</pre>
wcl.go_ids <- getBM(attributes = c("ensembl_gene_id", "go_id"), filters = "ensembl_gene_id",
   values = wcl$ensembl_gene_id, mart = mouse)
cv.go_terms.bp <- c("GO:0007507", "GO:0048738", "GO:0008015", "GO:0050878",
    "GD:0001944", "GD:0042060", "GD:0006979", "GD:0016055", "GD:0006520", "GD:0050817",
    "GD:0006629", "GD:0006936", "GD:0048771", "GD:0051145", "GD:0007517", "GD:0042692",
    "GO:0048659")
cv.go_terms.cc <- c("GD:0005739", "GD:0005578")</pre>
```

### Count term frequencies in Interactome and WCL data and produce bar plots:

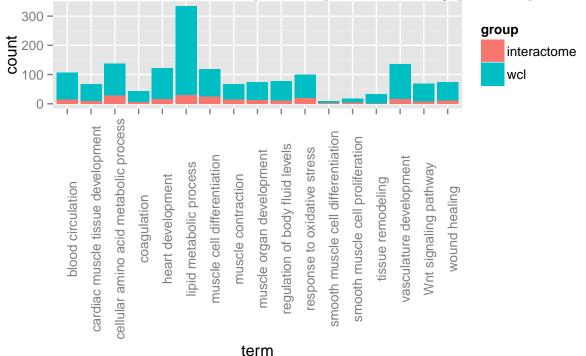
```
interactome.cv.go_bp.offsp <- sapply(cv.go_terms.bp, function(x) length(unique(interactome.go_ids[which
        unlist(go.bp.offspring[x])), "ensembl_gene_id"])))
interactome.cv.go_cc.offsp <- sapply(cv.go_terms.cc, function(x) length(unique(interactome.go_ids[which</pre>
```

```
unlist(go.cc.offspring[x])), "ensembl_gene_id"])))
interactome.cv.go_bp.offsp.IDs <- sapply(cv.go_terms.bp, function(x) unique(interactome.go_ids[which(in</pre>
    unlist(go.bp.offspring[x])), "ensembl_gene_id"]))
interactome.cv.go_cc.offsp.IDs <- sapply(cv.go_terms.cc, function(x) unique(interactome.go_ids[which(in
    unlist(go.cc.offspring[x])), "ensembl_gene_id"]))
wcl.cv.go_bp.offsp <- sapply(cv.go_terms.bp, function(x) length(unique(wcl.go_ids[which(wcl.go_ids$go_ids$go_ids)])</pre>
    unlist(go.bp.offspring[x])), "ensembl_gene_id"])))
wcl.cv.go_cc.offsp <- sapply(cv.go_terms.cc, function(x) length(unique(wcl.go_ids[which(wcl.go_ids$go_i
    unlist(go.cc.offspring[x])), "ensembl_gene_id"])))
wcl.cv.go_bp.offsp.IDs <- sapply(cv.go_terms.bp, function(x) unique(wcl.go_ids[which(wcl.go_ids$go_id %
    unlist(go.bp.offspring[x])), "ensembl_gene_id"]))
wcl.cv.go cc.offsp.IDs <- sapply(cv.go terms.cc, function(x) unique(wcl.go ids[which(wcl.go ids$go id %
    unlist(go.cc.offspring[x])), "ensembl_gene_id"]))
df.go_bp.interactome <- as.data.frame(interactome.cv.go_bp.offsp)</pre>
colnames(df.go_bp.interactome) <- "count"</pre>
df.go_bp.interactome$id <- rownames(df.go_bp.interactome)</pre>
df.go_bp.interactome$group <- "interactome"</pre>
df.go_bp.wcl <- as.data.frame(wcl.cv.go_bp.offsp)</pre>
colnames(df.go_bp.wcl) <- "count"</pre>
df.go_bp.wcl$group <- "wcl"</pre>
df.go_bp.wcl$id <- rownames(df.go_bp.wcl)</pre>
df.go_bp <- rbind(df.go_bp.interactome, df.go_bp.wcl)</pre>
df.go_bp$term <- sapply(df.go_bp$id, function(x) go.term[x][[1]]@Term)
n1 <- length(unique(interactome.go_ids[which(interactome.go_ids$go_id %in% unlist(go.bp.offspring[cv.go
    "ensembl_gene_id"]))
n2 <- length(unique(wcl.go_ids[which(wcl.go_ids$go_id %in% unlist(go.bp.offspring[cv.go_terms.bp])),
    "ensembl_gene_id"]))
```

Plotting the frequency of GO BP term descendants of major cardiovascular-associated GO terms

```
hist.go_bp <- ggplot(df.go_bp, aes(term, count, group = group, fill = group)) +
    geom_bar(postion = "dodge", stat = "identity")
hist.go_bp <- hist.go_bp + theme(axis.text.x = element_text(angle = 90))
hist.go_bp <- hist.go_bp + labs(title = paste("CV-associated gene counts\n in GO BP terms for Interactor
    n1, "] and WCL only [N = ", n2, "]", sep = ""))
print(hist.go_bp)</pre>
```

### CV-associated gene counts n GO BP terms for Interactome [N = 148] and WCL only [N = 809]



```
df.go_cc.interactome <- as.data.frame(interactome.cv.go_cc.offsp)</pre>
colnames(df.go_cc.interactome)[1] <- "count"</pre>
df.go_cc.interactome$group <- "interactome"</pre>
df.go_cc.interactome$id <- rownames(df.go_cc.interactome)</pre>
df.go_cc.interactome$term <- sapply(rownames(df.go_cc.interactome), function(x) go.term[x][[1]]@Term)
df.go_cc.wcl <- as.data.frame(wcl.cv.go_cc.offsp)</pre>
colnames(df.go_cc.wcl) <- "count"</pre>
df.go_cc.wcl$group <- "wcl"</pre>
df.go_cc.wcl$id <- rownames(df.go_cc.wcl)</pre>
df.go_cc.wcl$term <- sapply(rownames(df.go_cc.wcl), function(x) go.term[x][[1]]@Term)
df.go_cc <- rbind(df.go_cc.interactome, df.go_cc.wcl)</pre>
df.go_cc$group <- as.factor(df.go_cc$group)</pre>
n1 <- length(unique(interactome.go_ids[which(interactome.go_ids$go_id %in% unlist(go.cc.offspring[cv.go
    "ensembl_gene_id"]))
n2 <- length(unique(wcl.go_ids[which(wcl.go_ids$go_id %in% unlist(go.cc.offspring[cv.go_terms.cc])),
    "ensembl_gene_id"]))
```

Plotting the frequency of GO CC term descendants of major cardiovascular-associated GO terms

```
hist.go_cc <- ggplot(df.go_cc, aes(term, count, group = group, fill = group)) +
    geom_bar(postion = "dodge", stat = "identity")
hist.go_cc <- hist.go_cc + theme(axis.text.x = element_text(angle = 0))</pre>
```

```
hist.go_cc <- hist.go_cc + labs(title = paste("CV-associated gene counts\n in GO CC terms for Interactor
    n1, "] and WCL only [N = ", n2, "]", sep = ""))
print(hist.go_cc)</pre>
```

