Cardiomyocyte interactome, additional KEGG & Gene Ontology analyses

Sebastian Kurscheid
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R script for KEGG pathway analysis of cardiomyocyte RNA interactome proteins

Loading of the data tables.

```
load("data/kegg.brite.rda")
load("data/interactome.rda")
load("data/wcl.rda")
```

Fetching KEGG identifiers:

```
ids <- unlist(lapply(strsplit(kegg.brite$C, " "), function(x) x[1]))
rownames(kegg.brite) <- ids
total.keggIDs <- keggLink("mmu", "pathway")
save(total.keggIDs, file = "data/total.keggIDs.rda")</pre>
```

We have found a total of length(total.keggIDs) which are used for mapping the WCL and interactome data

Now we proceed to mapping of WCL protein IDs to KEGG IDs and testing for enrichments against background of all KEGG proteins contained in KEGG pathways.

```
# retrieved Entrez IDs from Biomart
mouse <- useMart("ensembl", dataset = "mmusculus_gene_ensembl")</pre>
human <- useMart("ensembl", dataset = "hsapiens_gene_ensembl")</pre>
attribs <- listAttributes(mouse)</pre>
pages <- attributePages(mouse)</pre>
hsap.attribs <- listAttributes(human)
entrez_ids <- getBM(attributes = c("ensembl_gene_id", "entrezgene"), values = wcl[,</pre>
    "ensembl_gene_id"], filters = "ensembl_gene_id", mart = mouse)
wcl.human_homologs <- getBM(attributes = c("ensembl_gene_id", "hsapiens_homolog_ensembl_gene"),
    values = wcl[, "ensembl_gene_id"], filters = "ensembl_gene_id", mart = mouse)
# remove ensembl_gene_ids which have duplicated entrez_ids
entrez_ids <- entrez_ids[-which(duplicated(entrez_ids$ensembl_gene_id)), ]</pre>
wcl <- merge(wcl, entrez_ids, by.x = "ensembl_gene_id", by.y = "ensembl_gene_id",
wcl.entrezIDs <- unique(wcl[!is.na(wcl$entrezgene), ]$entrezgene)</pre>
# this retrieval is fairly slow, therefore the results were written to
wcl.keggIDs <- keggConv.batch(wcl.entrezIDs)</pre>
save(wcl.keggIDs, file = "data/wcl.keggIDs.rda")
```

```
wcl.keggQ <- lapply(wcl.keggIDs, function(x) keggGet(x))</pre>
save(wcl.keggQ, file = "data/wcl.keggQ.rda")
wcl.pathways <- unique(unlist(lapply(strsplit(names(unlist(lapply(wcl.keggQ,</pre>
    function(x) x[[1]]$PATHWAY))), "\\."), function(x) x[3]))
save(wcl.pathways, file = "data/wcl.pathways.rda")
wcl.pathways.genes <- lapply(wcl.pathways, function(x) keggLink("genes", x))</pre>
names(wcl.pathways.genes) <- wcl.pathways</pre>
save(wcl.pathways.genes, file = "data/wcl.pathways.genes.rda")
wcl.pathways.genes.entrez_ids <- unique(gsub("mmu:", "", as.character(unlist(wcl.pathways.genes))))
wcl.df <- kegg.brite[gsub("mmu", "", wcl.pathways), ]</pre>
wcl.df$ID <- rownames(wcl.df)</pre>
wcl.df$total <- rep(0, nrow(wcl.df))</pre>
wcl.df$total <- sapply(rownames(wcl.df), function(x) length(wcl.pathways.genes[[paste("mmu",
    x, sep = "")]]))
wcl.df$count <- rep(0, nrow(wcl.df))</pre>
wcl.df$frac <- rep(0, nrow(wcl.df))</pre>
for (i in rownames(wcl.df)) {
    # print(i)
    kL1 <- keggLink("mmu", paste("mmu", i, sep = ""))</pre>
    wcl.df[i, ]$count <- length(which(wcl.keggIDs %in% kL1))</pre>
    wcl.df[i, ]$frac <- round(length(which(wcl.keggIDs %in% kL1))/length(kL1) *</pre>
        100, 2)
}
# extract list of IDs in pathway
wcl.in_path.IDs <- lapply(rownames(wcl.df), function(x) {</pre>
    kL1 <- keggLink("mmu", paste("mmu", x, sep = ""))</pre>
    in_path <- wcl.keggIDs[which(wcl.keggIDs %in% kL1)]</pre>
})
names(wcl.in_path.IDs) <- rownames(wcl.df)</pre>
# perform Fisher's Exact Test for each category
bkgd <- length(unique(total.keggIDs))</pre>
smpl <- length(wcl.keggIDs)</pre>
ftl <- apply(wcl.df[1, ], 1, function(x) {</pre>
    ct <- as.integer(x["count"])</pre>
    tt <- as.integer(x["total"])</pre>
    m1 <- matrix(c(ct, tt, smpl - ct, bkgd - tt), 2, 2)</pre>
    fisher.test(m1, alternative = alternative)
})
wcl.df$ft_pval <- unlist(lapply(ftl, function(x) {</pre>
    x$p.value
}))
wcl.df$ft_OR <- unlist(lapply(ftl, function(x) {</pre>
    x$estimate
}))
wcl.df$ft_fdr <- p.adjust(wcl.df$ft_pval, method = "fdr")</pre>
```

```
save(wcl.df, file = "data/wcl.df.rda")
```

Mapping of Interactome protein IDs to KEGG IDs and testing for enrichments against background of WCL proteins contained in KEGG pathways.

```
interactome.entrez_ids <- getBM(attributes = c("ensembl_gene_id", "entrezgene"),</pre>
    values = interactome[, "ensembl_gene_id"], filters = "ensembl_gene_id",
    mart = mouse)
interactome.human_homologs <- getBM(attributes = c("ensembl_gene_id", "hsapiens_homolog_ensembl_gene"),
    values = interactome[, "ensembl_gene_id"], filters = "ensembl_gene_id",
    mart = mouse)
# remove ensembl gene ids which have duplicated entrez ids
interactome.entrez_ids <- interactome.entrez_ids[-which(duplicated(interactome.entrez_ids$ensembl_gene_
interactome <- merge(interactome, interactome.entrez_ids, by.x = "ensembl_gene_id",</pre>
    by.y = "ensembl_gene_id", all.x = T)
interactome.entrezIDs <- unique(interactome[!is.na(interactome$entrezgene),</pre>
    ]$entrezgene)
save(interactome.entrezIDs, file = "data/interactome.entrezIDs")
interactome.keggIDs <- keggConv.batch(interactome.entrezIDs)</pre>
save(interactome.keggIDs, file = "data/interactome.keggIDs.rda")
interactome.keggQ <- lapply(interactome.keggIDs, function(x) keggGet(x))</pre>
save(interactome.keggQ, file = "data/interactome.keggQ.rda")
interactome.pathways <- unique(unlist(lapply(strsplit(names(unlist(lapply(interactome.keggQ,
    function(x) x[[1]]$PATHWAY))), "\\."), function(x) x[3]))
save(interactome.pathways, file = "data/interactome.pathways.rda")
interactome.pathways.genes <- lapply(interactome.pathways, function(x) keggLink("genes",</pre>
names(interactome.pathways.genes) <- interactome.pathways</pre>
save(interactome.pathways.genes, file = "data/interactome.pathways.genes.rda")
interactome.pathways.genes.entrez_ids <- unique(gsub("mmu:", "", as.character(unlist(interactome.pathwa
# create dataframe for counting hits in pathways
interactome.df <- kegg.brite[gsub("mmu", "", interactome.pathways), ]</pre>
interactome.df$source <- rep("Interactome", nrow(interactome.df))</pre>
interactome.df$ID <- rownames(interactome.df)</pre>
# we are now using WCL as background to test for enrichment
i1 <- intersect(rownames(interactome.df), rownames(wcl.df))</pre>
interactome.df$total <- rep(0, nrow(interactome.df))</pre>
interactome.df[i1, ]$total <- wcl.df[i1, ]$count</pre>
interactome.df$count <- rep(0, nrow(interactome.df))</pre>
interactome.df$frac <- rep(0, nrow(interactome.df))</pre>
for (i in rownames(interactome.df)) {
```

```
kL1 <- keggLink("mmu", paste("mmu", i, sep = ""))</pre>
    interactome.df[i, ]$count <- length(which(interactome.keggIDs %in% kL1))</pre>
    interactome.df[i, ] $frac <- round(length(which(interactome.keggIDs %in%
        kL1))/length(kL1) * 100, 2)
}
# extract list of IDs in pathway
interactome.in path.IDs <- lapply(rownames(interactome.df), function(x) {</pre>
    kL1 <- keggLink("mmu", paste("mmu", x, sep = ""))</pre>
    in_path <- interactome.keggIDs[which(interactome.keggIDs %in% kL1)]</pre>
})
# perform Fisher's Exact Test for each category
bkgd <- length(unique(wcl.keggIDs))</pre>
smpl <- length(interactome.keggIDs)</pre>
ftl <- apply(interactome.df, 1, function(x) {</pre>
    ct <- as.integer(x["count"])</pre>
    tt <- as.integer(x["total"])</pre>
    m1 <- matrix(c(ct, tt, smpl - ct, bkgd - tt), 2, 2)</pre>
    fisher.test(m1, alternative = alternative)
})
interactome.df$ft_pval <- unlist(lapply(ftl, function(x) {</pre>
    x$p.value
}))
interactome.df$ft_OR <- unlist(lapply(ftl, function(x) {</pre>
    x$estimate
}))
interactome.df$ft_fdr <- p.adjust(interactome.df$ft_pval, method = p.adjust.method,</pre>
    n = nrow(wcl.df)
save(interactome.df, file = "data/interactome.df.rda")
```

KEGG contains three levels/hierarchies (A>B>C), here we summarize the enrichment at B level:

```
# -----interactome-summarizing data at 'B' level before doing Fisher's
# Exact test-----
interactome.B.df <- data.frame(matrix(ncol = 5, nrow = length(unique(interactome.df$B))))</pre>
colnames(interactome.B.df) <- c("B", "A", "total", "count", "source")</pre>
interactome.B.df$B <- unique(interactome.df$B)</pre>
interactome.B.df$A <- sapply(unique(interactome.df$B), function(x) {</pre>
    A <- unique(interactome.df[which(interactome.df$B %in% x), "A"])
})
interactome.B.df$source <- rep("Interactome", nrow(interactome.B.df))</pre>
interactome.B.df$total <- sapply(unique(interactome.df$B), function(x) {</pre>
    tot <- sum(interactome.df[which(interactome.df$B %in% x), "total"])</pre>
})
interactome.B.df$count <- sapply(unique(interactome.df$B), function(x) {</pre>
    count <- sum(interactome.df[which(interactome.df$B %in% x), "count"])</pre>
})
bkgd <- length(unique(wcl.keggIDs))</pre>
smpl <- length(interactome.keggIDs)</pre>
```

```
ftl <- apply(interactome.B.df, 1, function(x) {
   ct <- as.integer(x["count"])
   tt <- as.integer(x["total"])
   m1 <- matrix(c(ct, tt, smpl - ct, bkgd - tt), 2, 2)
   fisher.test(m1, alternative = alternative)
})

interactome.B.df$ft_pval <- unlist(lapply(ftl, function(x) {
        x$p.value
}))

interactome.B.df$ft_OR <- unlist(lapply(ftl, function(x) {
        x$estimate
}))

interactome.B.df$ft_fdr <- p.adjust(interactome.B.df$ft_pval, method = p.adjust.method,
        n = nrow(wcl.df))

save(interactome.B.df, file = "data/interactome.B.df.rda")</pre>
```

We are splitting the Interactome proteins into RNA-related and un-related proteins, based on annotation analysis (upstream of these steps): First, RNA-unrelated

```
#-----GO RNA unrelated-----
# subset the interactome table
interactome.go rna unrelated <- interactome[which(interactome$GO == "unrelated"),]
interactome.go_rna_unrelated.entrezIDs <- unique(interactome.go_rna_unrelated[!is.na(interactome.go_rna
interactome.go_rna_unrelated.keggIDs <- keggConv.batch(interactome.go_rna_unrelated.entrezIDs)
# dataframe for count data
interactome.go_rna_unrelated.df <- interactome.df</pre>
interactome.go_rna_unrelated.df$source <- rep("GO_RNA_unrelated", nrow(interactome.go_rna_unrelated.df)
interactome.go_rna_unrelated.df$ID <- rownames(interactome.go_rna_unrelated.df)</pre>
interactome.go_rna_unrelated.df$total <- rep(0, nrow(interactome.go_rna_unrelated.df))</pre>
# we are now using WCL as background to test for enrichment
i1 <- intersect(rownames(interactome.go_rna_unrelated.df), rownames(wcl.df))</pre>
interactome.go_rna_unrelated.df[i1,]$total <- wcl.df[i1,]$count</pre>
interactome.go_rna_unrelated.df$count <- rep(0, nrow(interactome.go_rna_unrelated.df))</pre>
for (i in rownames(interactome.go_rna_unrelated.df)) {
 kL1 <- keggLink("mmu", paste("mmu", i, sep = ""))</pre>
  interactome.go_rna_unrelated.df[i, ]$count <- length(which(interactome.go_rna_unrelated.keggIDs %in%
}
# extract list of IDs in pathway
interactome.go_rna_unrelated.in_path.IDs <- lapply(rownames(interactome.go_rna_unrelated.df), function(
  kL1 <- keggLink("mmu", paste("mmu", x, sep = ""))</pre>
  in_path <- interactome.go_rna_unrelated.keggIDs[which(interactome.go_rna_unrelated.keggIDs %in% kL1)]
})
names(interactome.go_rna_unrelated.in_path.IDs) <- rownames(interactome.go_rna_unrelated.df)</pre>
# perform Fisher's Exact Test for each category
# Using WCL as background
bkgd <- length(unique(wcl.keggIDs))</pre>
smpl <- length(interactome.go_rna_unrelated.keggIDs)</pre>
```

```
ftl <- apply(interactome.go_rna_unrelated.df, 1, function (x) {
  ct <- as.integer(x["count"])</pre>
  tt <- as.integer(x["total"])</pre>
  m1 \leftarrow matrix(c(ct, tt, smpl - ct, bkgd - tt), 2, 2)
  fisher.test(m1, alternative = alternative)
})
interactome.go rna unrelated.df$ft pval <- unlist(lapply(ftl, function(x) {x$p.value}))
interactome.go_rna_unrelated.df$ft_OR <- unlist(lapply(ftl, function(x) {x$estimate}))</pre>
interactome.go_rna_unrelated.df$ft_fdr <- p.adjust(interactome.go_rna_unrelated.df$ft_pval, method = p.
save(interactome.go_rna_unrelated.df, file = "data/interactome.go_rna_unrelated.df.rda")
# summarizing data at "B" level before doing Fisher's Exact test
interactome.go_rna_unrelated.B.df <- data.frame(matrix(ncol = 5, nrow = length(unique(interactome.go_rn
colnames(interactome.go_rna_unrelated.B.df) <- c("B", "A", "total", "count", "source")</pre>
interactome.go_rna_unrelated.B.df$B <- unique(interactome.go_rna_unrelated.df$B)</pre>
interactome.go_rna_unrelated.B.df$A <- sapply(unique(interactome.go_rna_unrelated.df$B), function(x) {A
interactome.go_rna_unrelated.B.df$source <- rep("GO_RNA_unrelated", nrow(interactome.go_rna_unrelated.B
interactome.go_rna_unrelated.B.df$total <- sapply(unique(interactome.go_rna_unrelated.df$B), function(x
interactome.go_rna_unrelated.B.df$count <- sapply(unique(interactome.go_rna_unrelated.df$B), function(x
# using WCL as background
ftl <- apply(interactome.go_rna_unrelated.B.df, 1, function (x) {
  ct <- as.integer(x["count"])</pre>
  tt <- as.integer(x["total"])</pre>
  m1 <- matrix(c(ct, tt, smpl - ct, bkgd - tt), 2, 2)</pre>
  fisher.test(m1, alternative = alternative)
})
interactome.go_rna_unrelated.B.df$ft_pval <- unlist(lapply(ftl, function(x) {x$p.value}))
interactome.go_rna_unrelated.B.df$ft_OR <- unlist(lapply(ftl, function(x) {x$estimate}))</pre>
interactome.go_rna_unrelated.B.df$ft_fdr <- p.adjust(interactome.go_rna_unrelated.B.df$ft_pval, method
save(interactome.go_rna_unrelated.B.df, file = "data/interactome.go_rna_unrelated.B.df")
Now, RNA-related:
#-----GO RNA related-----
interactome.go_rna_related <- interactome[-which(interactome$GO == "unrelated"),]</pre>
interactome.go_rna_related.entrezIDs <- unique(interactome.go_rna_related[!is.na(interactome.go_rna_rel
interactome.go_rna_related.keggIDs <- keggConv.batch(interactome.go_rna_related.entrezIDs)</pre>
# we are testing this subset of "interactome", therefore we include all the pathways from "interactome"
interactome.go_rna_related.df <- interactome.df</pre>
i1 <- intersect(rownames(interactome.go_rna_related.df), rownames(wcl.df))</pre>
interactome.go_rna_related.df$total <- rep(0, nrow(interactome.go_rna_related.df))</pre>
interactome.go_rna_related.df[i1,]$total <- wcl.df[i1,]$count</pre>
interactome.go_rna_related.df$source <- rep("GO_RNA_related", nrow(interactome.go_rna_related.df))
interactome.go_rna_related.df$ID <- rownames(interactome.go_rna_related.df)</pre>
interactome.go_rna_related.df$count <- rep(0, nrow(interactome.go_rna_related.df))</pre>
interactome.go_rna_related.df$frac <- rep(0, nrow(interactome.go_rna_related.df))</pre>
for (i in rownames(interactome.go_rna_related.df)) {
```

```
kL1 <- keggLink("mmu", paste("mmu", i, sep = ""))</pre>
  interactome.go_rna_related.df[i, ]$count <- length(which(interactome.go_rna_related.keggIDs %in% kL1)
}
# extract list of IDs in pathway
interactome.go_rna_related.in_path.IDs <- lapply(rownames(interactome.go_rna_related.df), function(x) {</pre>
 kL1 <- keggLink("mmu", paste("mmu", x, sep = ""))</pre>
  in path <- interactome.go rna related.keggIDs[which(interactome.go rna related.keggIDs %in% kL1)]
})
names(interactome.go_rna_related.in_path.IDs) <- rownames(interactome.go_rna_related.df)</pre>
# perform Fisher's Exact Test for each category
# Using WCL as background
bkgd <- length(unique(wcl.keggIDs))</pre>
smpl <- length(interactome.go_rna_related.keggIDs)</pre>
ftl <- apply(interactome.go_rna_related.df, 1, function (x) {
  ct <- as.integer(x["count"])</pre>
 tt <- as.integer(x["total"])</pre>
 m1 <- matrix(c(ct, tt, smpl - ct, bkgd - tt), 2, 2)
 fisher.test(m1, alternative = alternative)
})
interactome.go_rna_related.df$ft_pval <- unlist(lapply(ftl, function(x) {x$p.value}))</pre>
interactome.go_rna_related.df$ft_OR <- unlist(lapply(ftl, function(x) {x$estimate}))</pre>
interactome.go_rna_related.df$ft_fdr <- p.adjust(interactome.go_rna_related.df$ft_pval, method = p.adju
save(interactome.go_rna_related.df, file = "data/interactome.go_rna_related.df")
# summarizing data at "B" level before doing Fisher's Exact test
interactome.go_rna_related.B.df <- data.frame(matrix(ncol = 5, nrow = length(unique(interactome.go_rna_
colnames(interactome.go_rna_related.B.df) <- c("B", "A", "total", "count", "source")</pre>
interactome.go_rna_related.B.df$B <- unique(interactome.go_rna_related.df$B)</pre>
interactome.go_rna_related.B.df$A <- sapply(unique(interactome.go_rna_related.df$B), function(x) {A <-
interactome.go_rna_related.B.df$source <- rep("GO_RNA_related", nrow(interactome.go_rna_related.B.df))</pre>
interactome.go_rna_related.B.df$total <- sapply(unique(interactome.go_rna_related.df$B), function(x) {t
interactome.go_rna_related.B.df$count <- sapply(unique(interactome.go_rna_related.df$B), function(x) {c
ftl <- apply(interactome.go_rna_related.B.df, 1, function (x) {
  ct <- as.integer(x["count"])</pre>
  tt <- as.integer(x["total"])</pre>
 m1 <- matrix(c(ct, tt, smpl - ct, bkgd - tt), 2, 2)
 fisher.test(m1, alternative = alternative)
})
interactome.go_rna_related.B.df$ft_pval <- unlist(lapply(ftl, function(x) {x$p.value}))</pre>
interactome.go_rna_related.B.df$ft_OR <- unlist(lapply(ftl, function(x) {x$estimate}))</pre>
interactome.go_rna_related.B.df$ft_fdr <- p.adjust(interactome.go_rna_related.B.df$ft_pval, method = p.
save(interactome.go_rna_related.B.df, file = "data/interactome.go_rna_related.B.df.rda")
```

Plotting the results of the enrichment analysis:

```
library(ggplot2)
library(grid)
```

```
library(scales)
load("data/interactome.df.rda")
load("data/interactome.go_rna_unrelated.df.rda")
load("data/interactome.go_rna_related.df")
dfC <- rbind(interactome.df[, c("A", "B", "C", "ft_OR", "ft_fdr", "source", "count")],</pre>
              interactome.go_rna_related.df[, c("A", "B", "C", "ft_OR", "ft_fdr", "source", "count")],
              interactome.go_rna_unrelated.df[, c("A", "B", "C", "ft_OR", "ft_fdr", "source", "count")]
dfC$source <- as.factor(dfC$source)</pre>
dfC$source <- factor(dfC$source, levels = levels(dfC$source)[c(3,1,2)])</pre>
select1 <- unique(as.character(dfC[which(dfC$ft_fdr <= 0.1 & dfC$ft_OR > 1),]$C))
select1.pathIDs <- paste("mmu", unlist(lapply(strsplit(select1, "\\ "), function(x) x[1])), sep = "")</pre>
dfC <- dfC[which(dfC$C %in% select1),]</pre>
dfC$C <- as.factor(as.character(dfC$C))</pre>
dfCft_OR.cut \leftarrow cut(log2(dfCft_OR), breaks = c(-Inf,-4:4), right = F)
dfC$C <- factor(dfC$C, levels = levels(dfC$C)[dfC[dfC$source == "Interactome", "C"][order(dfC[which(dfC
# formatting labels etc for plotting
11 <- levels(dfC$ft_OR.cut)</pre>
11 <- gsub("\\[", "", 11)
11 <- gsub("\\)", "", 11)
levels(dfC$ft_OR.cut) <- 11</pre>
11 <- as.character(levels(dfC$C))</pre>
11 <- unlist(lapply(strsplit(l1, " "), function(x) {</pre>
  for (i in 2:length(x)){
    if (i == 2){
      v <- x[i]
    } else {
      v <- paste(v, x[i])</pre>
  }
 return(v)
}))
levels(dfC$C) <- 11</pre>
levels(dfC$source)[2:3] <- c("RNA-related", "RNA-unrelated")</pre>
levels(dfC$C)[3] <- "Ribosome biogenesis"</pre>
levels(dfC$C)[6] <- "TCA cycle"</pre>
levels(dfC$C)[7] <- "mRNA surveillance"</pre>
levels(dfC$C)[11] <- "AA biosynthesis"</pre>
levels(dfC$C)[8] <- "H. simplex infection"</pre>
levels(dfC$C)[9] <- "Antibiotic biosynthesis"</pre>
levels(dfC$C)[12] <- "Glycolysis/Gluconeogenesis"</pre>
flevels <- levels(dfC$source)</pre>
```

```
11 \leftarrow factor(dfC\$C, levels = levels(dfC\$C)[c(1,2,3,7,4,5,6,12,8,9,10,11)])
dfC$C <- 11
p1 \leftarrow ggplot(data = dfC, aes(y = source, x = C)) +
      geom_tile(aes(fill = ft_OR.cut), colour = "white") +
      scale_fill_manual(values = brewer_pal(pal = "PuOr")(8), labels = levels(dfC$ft_OR.cut)) + #
      theme(axis.text.y = element_text(angle = 0, size = 8), axis.title = element_blank()) +
      guides(fill = guide legend(label.position = "bottom", direction = "horizontal")) +
      theme(axis.text.x = element_text(angle = 45, vjust = 0.9, hjust = 0.8, size = 10)) +
      labs(fill = "Log2 OR") +
      scale_y_discrete(limits = rev(flevels)) +
      theme(legend.position = c(0.4,-1.92),
            legend.text = element_text(size = 4),
            legend.text.align = 0.5,
            legend.title = element_text(size = 4, vjust = 5),
            legend.key.size = unit(3.5, "mm"),
            legend.key.width = unit(3.5, "mm"),
            legend.margin = unit(0, "mm"),
            panel.margin = unit(1, "mm")) +
      ggtitle("KEGG pathway enrichment")
plot(p1)
```

