babette: BEAUti 2, BEAST2 and Tracer for R

- Richèl J.C. Bilderbeek¹ and Rampal S. Etienne¹
- ¹Groningen Institute for Evolutionary Life Sciences, University of
- Groningen, Groningen, The Netherlands

March 6, 2018

Summary

10

11

12

19

20

- 1. In the field of phylogenetics, BEAST2 is one of the most widely used software tools. It comes with the graphical user interfaces BEAUti 2, DensiTree and Tracer, to create BEAST2 configuration files and to interpret BEAST2's output files. However, when many different alignments or model setups are required, a workflow of graphical user interfaces is cumbersome.
- 2. Here, we present a free, libre and open-source package, babette:
 'BEAUti 2, BEAST2 and Tracer for R', for the R programming language.
 babette creates BEAST2 input files, runs BEAST2 and parses its results, all from an R function call.
- 3. We describe babette's usage and the novel functionality it provides compared to the original tools and we give some examples.
 - 4. As babette is designed to be of high quality and extendable, we conclude by describing the further development of the package.

Keywords: computational biology, evolution, phylogenetics, BEAST2, R

23 1 Introduction

Phylogenies are commonly used to explore evolutionary hypotheses. Not only 24 can phylogenies show us how species (or other evolutionary units) are related to each other, but we also estimate relevant parameters such as extinction and speciation rates. There are many phylogenetics tools available to obtain an es-27 timate of the phylogenetic tree of a given set of species. BEAST2 (Bouckaert et al. 2014) is one of the most widely used ones. It creates a posterior of jointly estimated phylogenies and model parameters, from one or more DNA, RNA or amino acid alignments (see figure 1 for an overview of the workflow). It has a 31 graphical and a command-line interface, that both need a configuration file con-32 taining alignments and model parameters. BEAST2 is bundled with BEAUti 33 2 (Drummond et al. 2012) ('BEAUti' from now on), a desktop application to create a BEAST2 configuration file. BEAUti has a user-friendly graphical user interface, with helpful default settings. As such, BEAUti is an attractive alternative to manual and error-prone editing of BEAST2 configuration files. However, BEAUti cannot be called from a command-line script. This implies that when the user wants to explore the consequences of various settings, this must be done manually. This is the managable workflow when using a few 40 alignments and doing a superficial analysis of sensitivity of the reconstructed tree to model settings. For exploring many trees (for instance from simulations) and for more thorough sensitivity analysis, one would like to loop through multiple (simulated) alignments, nucleotide substitution models, clock models and tree priors. One such tool to replace BEAUti is BEASTmasteR (Matzke 2015), 45 which focuses on morphological traits and tip-dating, but also supports DNA data. BEASTmasteR, however, requires hundreds of lines of R code to setup the BEAST2 model configuration and a Microsoft Excel file to specify alignment files.

BEAST2 is also associated with Tracer (Rambaut & Drummond 2007) and 50 DensiTree (Bouckaert & Heled 2014). Both are desktop applications to analyze the output of BEAST2, each with a user-friendly graphical user inter-52 face. Tracer's purpose is to analyze the parameter estimates generated from a BEAST2 run. It shows, among others, the effective sample size (ESS) and time series ('the trace', hence the name) of each variable in the MCMC run. Both ESS and trace are needed to assess the strength of the inference. DensiTree visualizes the phylogenies of a BEAST2 posterior, with many options to improve the simultaneous display of many phylogenies. However, for exploring the output of many BEAST2 runs, one would like a 59 script to collect all parameters' ESSes, parameter traces and posterior phylogenies. There is no single package that offers a complete solution, but examples 61 of R packages that offer a partial solution are rBEAST (Ratmann 2015) and RBeast (Faria & Suchard 2015). RBeast provides some plotting options and

parsing of BEAST2 output files, but the plotting functions are too specific for general use, where the functions for parsing call those of babette. rBEAST was

developed to test a particular biological hypothesis (Ratmann et al. 2016), and

hence was not designed for general use.

63

Here, we present babette: BEAUti 2, BEAST2 and Tracer for R, which creates BEAST2 (v.2.4.7) configuration files, runs BEAST2, and analyzes its results, all from an R function call. This will save time, tedious mouse clicking and reduces the chances of errors in such repetitive actions. The interface of babette mimics the tools it is based on. This familiarity helps both beginner and experienced BEAST2 users to make the step from those tools to babette. babette enables the creation of a single-script pipeline from sequence alignments to posterior analysis in R.

6 2 Description

babette is written in the R programming language (R Core Team 2013) and enables the full BEAST2 workflow from an R function call, in a similar way to what BEAUti, DensiTree and Tracer do. babette's main function is run_beast2, which configures BEAST2, runs it and parses its output. run_beast2 needs 80 at least the name of a FASTA file containing a DNA alignment. The default settings for the other arguments of run_beast2 are identical to BEAUti's and BEAST2's default settings. Per alignment, a site model, clock model and tree prior can be chosen. Multiple alignments can be used, each with its own (un-84 linked) site model, clock model and tree prior. babette currently has 100 exported functions to set up a BEAST2 configuration file. babette can currently handle a majority of BEAUti use cases. Because of BEAUti's high number of plugins, babette uses a software architecture that is designed to be extended. Furthermore, babette has 12 exported functions to run and help run BEAST2. One function is used to run BEAST2, others allow the user to check if a BEAST2 configuration file is indeed valid. Finally, babette has 21 exported function to parse the BEAST2 output files and analyze the created posterior. babette gives the same ESSes and summary 93 statistics as Tracer. The data is formatted such that it can easily be visualized using ggplot2 (for a trace, similar to Tracer) or phangorn (Schliep 2011) (for the phylogenies in a posterior, similar to DensiTree). Currently, babette does not replace all functionality in BEAUti, as it does 97 not provide 3 out of 7 tree priors, nor does it support RNA alignments or 98 use of morphological data. The many plug-ins of BEAUti are not yet supported by babette. babette does not support all command-line arguments of 100 BEAST2, does not provide the more specialized Tracer analysis options, nor is it 101 as feature-rich in plotting options as DensiTree. Up until now, the babette fea-102

tures implemented are those requested by users. Further extension of babette will be based on future user requests.

3 Usage

127

```
In R, the functions of a package need to be loaded in the global namespace first:
106
    library(babette)
107
   BEAUti, and likewise babette, needs at least a FASTA filename to produce a
108
   BEAST2 configuration file. In BEAUti, this is achieved by loading a FASTA file,
    then saving an output file using a common save file dialog. After this, BEAST2
110
   needs to be applied to the created configuration file. It creates multiple files
   storing the posterior. These output files must be parsed by either Tracer of
112
   DensiTree. In babette, all this is achieved by:
    out <- run_beast2("anthus_aco.fas")</pre>
114
    This code will create a (temporary) BEAST2 configuration file, from the FASTA
   file with name anthus_aco.fas (which is supplied with the package, from
116
    (Van Els & Norambuena 2018)), using the same default settings as BEAUti,
117
    which are, among others, a Jukes-Cantor site model, a strict clock, and a Yule
118
   birth tree prior. babette will then execute BEAST2 using that file, and parses
119
    the output. The returned data structure, named out, is a list of parameter
120
    estimates (called estimates), posterior phylogenies (called anthus_aco_trees,
121
   named after the alignment's name) and MCMC operator performance (operators).
122
    An example of using a different site model, clock model and tree prior is:
123
    out <- run_beast2(
124
      "anthus_aco.fas",
125
      site_models = create_hky_site_model(),
126
      clock_models = create_rln_clock_model(),
```

```
tree_priors = create_bd_tree_prior()

129

This code uses an HKY site model, a relaxed log-normal clock model and a birth-
```

death tree prior, each with their default settings in BEAUti. Table 1 shows an overview of all functions to create site models, clock models and tree priors. Note that the arguments' names site_models, clock_models and tree_priors are plural, as each of these can be (a list of) one or more elements. Each of these arguments must have the same number of elements, so that each alignment has its own site model, clock model and tree prior. An example of two alignments, each with its own site model, is:

```
out <- run_beast2(
c("anthus_aco.fas", "anthus_nd2.fas"),
site_models = list(
create_tn93_site_model(),
create_gtr_site_model()

143 )</pre>
```

babette also uses the same default prior distributions as BEAUti for each of
the site models, clock models and tree priors. For example, by default, a Yule
tree prior assumes that the birth rate follows a uniform distribution, from minus
infinity to plus infinity. This assumption implies that negative and positive birth
rates are just as likely, where a negative birth rate is biologically impossible (note
that in practice, this usually works out just fine). One may prefer an exponential
distribution instead, as this would assume only positive birth rates, and makes
high birth rates unlikely. To do this in babette:

```
153  out <- run_beast2(
154  "anthus_aco.fas",</pre>
```

```
birth_rate_distr = create_exp_distr()
156
      )
157
   )
158
   In this same example, one may specify the initial shape parameters of the expo-
   nential distribution. In BEAST2's implementation, an exponential distribution
160
   has one shape parameter: its mean, which can be set to any value with BEAUti.
    Within babette, to set the mean value of the exponential distribution to a fixed
162
    (non-estimated) value, do:
163
    out <- run_beast2(
164
      "anthus_aco.fas",
165
      tree_priors = create_yule_tree_prior(
166
         birth_rate_distr = create_exp_distr(
167
           mean = create_mean_param(
              value = 1.0,
169
              estimate = FALSE
           )
171
         )
172
      )
173
   )
174
    Our initial motivation to create babette was that we wanted to fix the crown
175
   age of a phylogeny. BEAUti assumes that a phylogeny has a crown age that
176
   needs to be jointly estimated with the phylogeny and other parameters. It does
177
    not allow for fixing the crown age. Without babette, one needs to manually edit
178
    the BEAST2 configuration file (Bouckaert & Vaughan 2017), which is tedious
179
```

tree_priors = create_yule_tree_prior(

155

180

181

a fixed crown age with babette:

and prone to errors. Fixing the crown ages is especially useful for theoretical

experiments, as this allows for one less source of variation. This is how to specify

```
out <- run_beast2(
183
       "anthus_aco.fas",
184
      posterior_crown_age = 15
185
    )
186
    babette allows for the same functionality as Tracer. Tracer works on the values
    of the parameter estimates sampled in the BEAST2 run. This is called the
188
    "trace" (hence the name). The start of the trace is usually discarded, as an
    MCMC algorithm (such as used by BEAST2) first has to converge to its equi-
190
    librium. The start of the trace, called the "burn-in", will be removed, because
191
    its parameter estimates are not representative. By default, Tracer discards the
    first 10% of all the parameter estimates. To remove a 20% burn-in from all
193
    parameter estimates in babette, the following code can be used:
    traces <- remove_burn_ins(</pre>
195
      out $ estimates,
      burn_in_fraction = 0.2
197
    )
198
    Tracer shows the ESSes of each posterior's variables. These ESSes are important
199
    to determine the strength of the inference. As a rule of thumb, an ESS of 200 is
200
    acceptable for any parameter estimate. To calculate the effective sample sizes
201
    (of all estimated variables) in babette:
202
    esses <- calc_esses(
203
      traces,
204
      sample_interval = 1000
205
    )
206
    Tracer displays multiple summary statistics for each estimated variable: the
    mean and its standard error, standard deviation, variance, median, mode, geo-
208
    metric mean, 95% highest posterior density interval, auto-correlation time and
209
```

```
effective sample size. It displays these statistics per variable. In babette, these
    summary statistics are collected for all estimated parameters at once:
211
    sum_stats <- calc_summary_stats(</pre>
212
       traces,
213
       sample_interval = 1000
214
    )
215
    babette allows for the same functionality as DensiTree. DensiTree displays the
216
    phylogenies in a posterior at the same time scale, drawn one over one another,
217
    allowing to see the uncertainty in topology and branch lengths. Within the
218
    object out, the posterior phylogenies are stored as anthus_aco_trees, and can
219
    be plotted as such:
    plot_densitree(out$anthus_aco_trees)
```

222 4 babette resources

```
babette is free, libre and open source software available from the official R pack-
223
    age archive at http://cran.r-project.org/src/contrib/PACKAGES.html#babette
224
    and is licensed under the GNU General Public License v3.0. babette uses the
225
    Travis CI (https://travis-ci.org) continuous integration service, which is
226
   known to significantly increase the number of bugs exposed (Vasilescu et al.
227
    2015) and increases the speed at which new features are added (Vasilescu et al.
228
   2015). babette has a 100% code coverage, which correlates with code quality
    (Horgan et al. 1994; Del Frate et al. 1995). babette follows Hadley Wickham's
230
   style guide (Wickham 2015), which improves software quality (Fang 2001).
   babette depends on multiple packages, which are ape (Paradis et al. 2004),
232
   beautier (Bilderbeek 2018b), beastier (Bilderbeek 2018a), devtools (Wick-
   ham & Chang 2016), geiger (Harmon et al. 2008), ggplot2 (Wickham 2009),
```

```
knitr (Xie 2017), phangorn (Schliep 2011), rmarkdown (Allaire et al. 2017),
    seqinr (Charif & Lobry 2007), stringr (Wickham 2017), testit (Xie 2014)
236
    and tracerer (Bilderbeek 2018c). babette is tested to give a clean error mes-
237
    sage for incorrect input, by calling babette one million times with random or
238
    random sensible inputs, using the Peregrine high performance computer cluster.
239
    The scripts to do so, are supplied with babette.
240
       babette's development takes place on GitHub, https://github.com/richelbilderbeek/
241
    babette, which accommodates collaboration (Perez-Riverol et al. 2016) and im-
242
    proves transparency (Gorgolewski & Poldrack 2016). babette's GitHub facili-
    tates feature requests and has guidelines how to do so.
244
       babette's documentation is extensive. All functions are documented in the
    package's internal documentation. For quick use, each exported function shows
246
    a minimal example. For easy exploration, each exported function's documen-
    tation links to related functions. Additionally, babette has a vignette that
248
    demonstrates extensively how to use it. There is documentation on the GitHub
    to get started, with a dozen examples of BEAUti screenshots with equivalent
250
    babette code. Finally, babette has some tutorial video's that can be down-
251
    loaded or viewed on YouTube, https://goo.gl/weKaaU.
252
```

5 Citation of babette

```
Scientists using babette in a published paper can cite this article, and/or cite
the babette package directly. To obtain this citation from within an R script,
use:

> citation("babette")
```

58 6 Acknowledgements

- 259 Thanks to Yacine Ben Chehida and Paul van Els for supplying their BEAST2
- use cases. Thanks again to Paul van Els for the sharing his FASTA files for use by
- this package. Thanks to Leonel Herrera-Alsina, Raphael Scherrer and Giovanni
- 262 Laudanno for their comments on this package and article. We would like to
- thank the Center for Information Technology of the University of Groningen
- for their support and for providing access to the Peregrine high performance
- 265 computing cluster.

266 References

- Allaire, J., Xie, Y., McPherson, J., Luraschi, J., Ushey, K., Atkins, A., Wick-
- ham, H., Cheng, J. & Chang, W. (2017) rmarkdown: Dynamic Documents
- for R. R package version 1.8.
- Bilderbeek, R.J. (2018a) beastier: BEAST2 from R. https://github.com/
- richelbilderbeek/beastier [Accessed: 2018-02-28].
- Bilderbeek, R.J. (2018b) beautier: BEAUti 2 from R. https://github.com/
- 273 richelbilderbeek/beautier [Accessed: 2018-02-28].
- Bilderbeek, R.J. (2018c) tracerer: Tracer from R. https://github.com/
- richelbilderbeek/tracerer [Accessed: 2018-02-28].
- Bouckaert, R. & Heled, J. (2014) Densitree 2: Seeing trees through the forest.
- bioRxiv, p. 012401.
- Bouckaert, R., Heled, J., Kühnert, D., Vaughan, T., Wu, C.H., Xie, D., Suchard,
- M.A., Rambaut, A. & Drummond, A.J. (2014) Beast 2: a software platform
- for bayesian evolutionary analysis. *PLoS Comput Biol*, **10**, e1003537.

- Bouckaert, R. & Vaughan, T. (2017) Fix starting tree. http://www.beast2.
- org/fix-starting-tree [Accessed: 2018-02-28].
- ²⁸³ Charif, D. & Lobry, J. (2007) SeqinR 1.0-2: a contributed package to the R
- project for statistical computing devoted to biological sequences retrieval and
- analysis. U. Bastolla, M. Porto, H. Roman & M. Vendruscolo, eds., Struc-
- tural approaches to sequence evolution: Molecules, networks, populations, Bi-
- ological and Medical Physics, Biomedical Engineering, pp. 207–232. Springer
- ²⁸⁸ Verlag, New York. ISBN: 978-3-540-35305-8.
- Del Frate, F., Garg, P., Mathur, A.P. & Pasquini, A. (1995) On the correlation
- between code coverage and software reliability. Software Reliability Engi-
- neering, 1995. Proceedings., Sixth International Symposium on, pp. 124–132.
- 292 IEEE.
- Drummond, A.J., Suchard, M.A., Xie, D. & Rambaut, A. (2012) Bayesian phy-
- logenetics with beauti and the beast 1.7. Molecular biology and evolution, 29,
- ²⁹⁵ 1969–1973.
- Fang, X. (2001) Using a coding standard to improve program quality. Quality
- 297 Software, 2001. Proceedings. Second Asia-Pacific Conference on, pp. 73–78.
- 298 IEEE.
- Faria, N. & Suchard, M.A. (2015) RBeast. https://github.com/beast-dev/
- RBeast [Accessed: 2018-03-02].
- 301 Gorgolewski, K.J. & Poldrack, R. (2016) A practical guide for improving trans-
- parency and reproducibility in neuroimaging research. bioRxiv, p. 039354.
- Harmon, L., Weir, J., Brock, C., Glor, R. & Challenger, W. (2008) Geiger:
- investigating evolutionary radiations. *Bioinformatics*, **24**, 129–131.

- Horgan, J.R., London, S. & Lyu, M.R. (1994) Achieving software quality with
- testing coverage measures. Computer, 27, 60–69.
- 307 Matzke, N.J. (2015) BEASTmasteR: R tools for automated conversion of
- NEXUS data to BEAST2 XML format, for fossil tip-dating and other uses.
- https://github.com/nmatzke/BEASTmasteR [Accessed: 2018-02-28].
- Paradis, E., Claude, J. & Strimmer, K. (2004) APE: analyses of phylogenetics
- and evolution in R language. *Bioinformatics*, **20**, 289–290.
- Perez-Riverol, Y., Gatto, L., Wang, R., Sachsenberg, T., Uszkoreit, J., Lepre-
- vost, F., Fufezan, C., Ternent, T., Eglen, S.J., Katz, D.S. et al. (2016) Ten
- simple rules for taking advantage of git and github. bioRxiv, p. 048744.
- R Core Team (2013) R: A Language and Environment for Statistical Computing.
- R Foundation for Statistical Computing, Vienna, Austria.
- Rambaut, A. & Drummond, A.J. (2007) Tracer v1.4. Available from
- http://beast.bio.ed.ac.uk/Tracer.
- Ratmann, O. (2015) rBEAST. https://github.com/olli0601/rBEAST [Ac-
- cessed: 2018-03-02].
- Ratmann, O., Van Sighem, A., Bezemer, D., Gavryushkina, A., Jurriaans, S.,
- Wensing, A., De Wolf, F., Reiss, P., Fraser, C. et al. (2016) Sources of hiv
- infection among men having sex with men and implications for prevention.
- Science translational medicine, 8, 320ra2-320ra2.
- Schliep, K. (2011) phangorn: phylogenetic analysis in R. Bioinformatics, 27,
- ₃₂₆ 592–593.
- Van Els, P. & Norambuena, H.V. (2018) A revision of species limits in neotrop-
- ical pipits anthus based on multilocus genetic and vocal data. *Ibis*.

- Vasilescu, B., Yu, Y., Wang, H., Devanbu, P. & Filkov, V. (2015) Quality and
- productivity outcomes relating to continuous integration in github. Proceed-
- ings of the 2015 10th Joint Meeting on Foundations of Software Engineering,
- pp. 805–816. ACM.
- Wickham, H. (2009) ggplot2: elegant graphics for data analysis. Springer New
- York.
- Wickham, H. (2015) R packages: organize, test, document, and share your code.
- O'Reilly Media, Inc.
- Wickham, H. (2017) stringr: Simple, Consistent Wrappers for Common String
- Operations. R package version 1.2.0.
- Wickham, H. & Chang, W. (2016) devtools: Tools to Make Developing R Pack-
- ages Easier. R package version 1.12.0.9000.
- 341 Xie, Y. (2014) testit: A Simple Package for Testing R Packages. R package
- version 0.4, http://CRAN.R-project.org/package=testit.
- 343 Xie, Y. (2017) knitr: A General-Purpose Package for Dynamic Report Genera-
- tion in R. R package version 1.17.

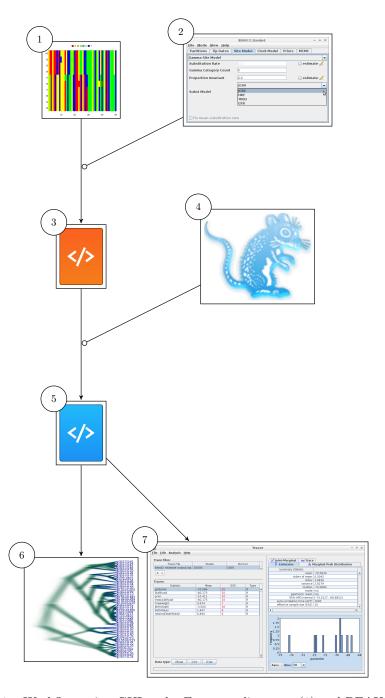


Figure 1: Workflow using GUI tools. From an alignment (1) and BEAUti (2), a BEAST2 configuration file (3) is created. BEAST2 (4) uses that file to infer a posterior, storing it in multiple files (5). These results are visualized using DensiTree (6) and Tracer (7). babette allows for the same workflow, all from an R function call.

Name	Description
run_beast2	Run BEAST2
create_gtr_site_model	Create a GTR site model
create_hky_site_model	Create an HKY site model
create_jc69_site_model	Create a Jukes-Cantor site model
create_tn93_site_model	Create a TN93 site model
create_rln_clock_model	Create a relaxed log-normal clock model
create_strict_clock_model	Create a strict clock model
create_bd_tree_prior	Create a birth-death tree prior
create_cbs_tree_prior	Create a coalescent Bayesian skyline tree prior
create_ccp_tree_prior	Create a coalescent constant-population tree prior
create_cep_tree_prior	Create a coalescent exponential-population tree prior
create_yule_tree_prior	Create a Yule tree prior
create_beta_distr	Create a beta distribution
create_exp_distr	Create an exponential distribution
create_gamma_distr	Create a gamma distribution
create_inv_gamma_distr	Create an inverse gamma distribution
create_laplace_distr	Create a Laplace distribution
create_log_normal_distr	Create a log-normal distribution
create_normal_distr	Create a normal distribution
create_one_div_x_distr	Create a 1/X distribution
create_poisson_distr	Create a Poisson distribution
create_uniform_distr	Create a uniform distribution

Table 1: babette's main functions