babette: BEAUti 2, BEAST2 and Tracer for R

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Summary

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cumbersome.

- 1. In the field of phylogenetics, BEAST2 is one of the most widely used software tools. It comes with the graphical user interfaces BEAUti 2, DensiTree and Tracer, to create BEAST2 configuration files and to interpret BEAST2's output files. However, when many different alignments or model setups are required, a workflow of graphical user interfaces is
- 2. Here, we present a free, libre and open-source package, babette:

 'BEAUti 2, BEAST2 and Tracer for R', for the R programming language.

 babette creates BEAST2 input files, runs BEAST2 and parses its results,

 all from an R function call.
- 3. We describe babette's usage and the novel functionality it provides compared to the original tools and we give some examples.
 - 4. As babette is designed to be of high quality and extendable, we conclude by describing the further development of the package.

Keywords: computational biology, evolution, phylogenetics, BEAST2, R

23 1 Introduction

Phylogenies are commonly used to explore evolutionary hypotheses. Not only can phylogenies show us how species (or other evolutionary units) rela other. but we al stimate relevant parameters such as extinction and speciation rates. There are many phylogenetics tools available to obtain an estimate of 27 the phylogenetic tree of a given set of species. BEAST2 (Bouckaert et al. 2014) is one of the most widely used ones. It creates a posterior of jointly-estimated phylogenies and model parameters, from one or more DNA, RNA or amino acid alignments (see figure 1 for an overview of the workflow). It has a graphical 31 and a command-line interface, that both need a configuration file containing 32 alignments and model parameters. BEAST2 is bundled with BEAUti 2 (Drum-33 mond et al. 2012) ('BEAUti' from now on), a desktop application to create a BEAST2 configuration file. BEAUti has a user-friendly graphical user interface, with helpful default settings. As such, BEAUti is an attractive alternative to 36 manual and error-prone editing of BEAST2 configuration files. However, BEAUti cannot be called from a command-line script. This implies that when the user wants to explore the consequences of various settings, this must be done manually. This is the common warflow when using a few alignments and doing a superficial analysis of sensitivity of the reconstructed tree to model settings. For exploring many trees (for instance from simulations) and for more thorough sensitivity analysis, one would like to loop through multiple (simulated) alignments, nucleotide substitution models, clock models and tree priors. One such tool to replace BEAUti is BEASTmasteR (Matzke 2015), which 45 focuses on morphological traits and tip-dating, but also supports DNA data. andreds of lines of R code to setup the BEAST2 BEASTmasteR, however, take model configuration and a Microsoft Excel file to specify alignment files. BEAST2 is also associated with Tracer (Rambaut & Drummond 2007) and 49

- $_{50}\,$ Densi Tree (Bouckaert & Heled 2014). Both are desktop applications to an a-
- by lyze the output of BEAST2, each with a user-friendly graphical user interface.
- 52 Tracer's purpose is to analyze the parameter estimates generated from a BEAST
- ⁵³ 2 run. It shows, among others, the effective sample size (ESS) and time series
- ('the trace', hence the name) of each variable in the MCMC run. Both ESS and
- trace are needed to assess the strength of the inference. DensiTree visualizes the
- 56 phylogenies of a BEAST2 posterior, with many options to improve the
- of many phylogenies.
- However, for exploring the output of many BEAST2 runs, one would like a
- 59 script to collect all parameters' ESSes, parameter traces and posterior phyloge-
- nies. There is no single package that offers a complete solution, but examples
- of R packages that offer a partial solution are BEASTmasteR, rBEAST (Ratmann
- 62 2015) and RBeast (Faria & Suchard 2015). BEASTmasteR and RBeast are in-
- complete packages, where BEAST does not aim to be a package to be used by
- 64 most BEAST2 users.
- Here, we present babette: BEAUti 2, BEAST2 and Tracer for R, which
- creates BEAST2 configuration files, runs BEAST2, and analyzes its results, all
- from an R function call. This will save time, tedious mouse clicking and reduces
- the chances of errors in such repetitive actions. The interface of babette mimics
- 69 the tools it is based on. This familiarity helps both beginner and experienced
- ₇₀ BEAST2 users to make the step from those tools to babette. babette enables
- 71 the creation of a single-script pipeline from sequence alignments to posterior
- 72 analysis in R.

$_{\scriptscriptstyle 3}$ 2 Description

- babette is written in the R programming language (R Core Team 2013) and en-
- ables the full BEAST2 workflow from an R function call, in a similar way to what

BEAUti, DensiTree and Tracer do. babette's main function is run_beast2,
which configures BEAST2, runs it and parses its output. run_beast2 needs
at least the name of a FASTA file containing a DNA alignment. The default
settings for the other arguments of run_beast2 are identical to BEAUti's and
BEAST2's default settings. Per alignment, a site model, clock model and tree
prior can be chosen. Multiple alignments can be used, each with its own (unlinked) site model, clock model and tree prior.

babette currently has 61 exported functions to set up a BEAST2 configuration file. babette is an alternative for majority of BEAUti use cases. Because
of BEAUti's high number of plugins, babette uses a software architecture that
is designed to be extended. Furthermore, babette has 7 exported function

is designed to be extended. Furthermore, babette has 7 exported function
run and help run BEAST2. One function is used to run BEAST2, others allow the user to check if a BEAST2 configuration file is indeed valid. Finally,
babette has 20 exported function to parse the BEAST2 output files and analyze
the created posterior. babette gives the same ESSes and summary statistics
as Tracer. The data is formatted as such, that it can easily be visualized using
ggplot2 (for a trace, similar to Tracer) or phangorn (Schliep 2011) (for the
phylogenies in a posterior, similar to DensiTree).

Currently, babette does not replace all functionality in BEAUti, as it does not provide 3 out of 7 tree priors, nor does it support RNA alignments or use of morphological data. The many plug-ins of BEAUti are not yet supported by babette. babette does not support all command-line arguments of BEAST2, does not provide the more specialized Tracer analysis options, nor is it as feature-rich in plotting options as DensiTree. Up until now, the babette features implemented are those requested by users. Further extension of babette will be based on future user requests.

$_{\scriptscriptstyle 02}$ 3 Usage

```
In R, the functions of a package need to be loaded in the global namespace first:
103
    library(babette)
   BEAUti, and likewise babette, needs at least a FASTA filename to produce a
105
   BEAST2 configuration file. In BEAUti, this is achieved by loading a FASTA file,
106
    then saving an output file using a common save file dialog. After this, BEAST2
   needs to be applied to the created configuration file. It creates multiple files
108
   storing the posterior. These output files must be parsed by either Tracer of
   DensiTree. In babette, all this is achieved by:
110
    out <- run_beast2("anthus_aco.fas")</pre>
111
    This code will create a (temporary) BEAST2 configuration file, from the FASTA
112
   file with name anthus_aco.fas (which is supplied with the package, from
    (Van Els & Norambuena 2018)), using the same default settings as BEAUti,
114
    which are, among others, a Jukes-Cantor site model, a strict clock, and a Yule
   birth tree prior. babette will then execute BEAST2 using that file, and parses
116
    the output. The returned data structure, named out, is a list of parameter
117
   estimates (called estimates), posterior phylogenies (called anthus_aco_trees,
118
   named after the alignment's name) and MCMC operator performance (operators).
119
    An example of using a different site model, clock model and tree prior is:
    out <- run_beast2(
      "anthus_aco.fas",
122
      site_models = create_hky_site_model(),
123
      clock_models = create_rln_clock_model(),
124
      tree_priors = create_bd_tree_prior()
125
   )
126
```

This code uses an HKY site model, a relaxed log-normal clock model and a birth-

death tree prior, each with their default settings in BEAUti. Table 1 shows an overview of all functions to create site models, clock models and tree priors. Note that the arguments' names site_models, clock_models and tree_priors are plural, as each of these can be (a list of) one or more elements. Each of these arguments must have the same number of elements, so that each alignment has its own site model, clock model and tree prior. An example of two alignments, each with its own site model, is:

```
out <- run_beast2(

c("anthus_aco.fas", "anthus_nd2.fas"),

site_models = list(

create_tn93_site_model(),

create_gtr_site_model()

140 )
```

babette also uses the same default prior distributions as BEAUti for each of
the site models, clock models and tree priors. For example, by default, a Yule
tree prior assumes that the birth rate follows a uniform distribution, from minus
infinity to plus infinity. This assumption implies that negative and positive birth
rates are just as likely, where a negative birth rate is biologically impossible (note
that in practice, this usually works out just fine). One may prefer an exponential
distribution instead, as this would assume only positive birth rates, and makes
high birth rates unlikely. To do this in babette:

```
out <- run_beast2(
    "anthus_aco.fas",
    tree_priors = create_yule_tree_prior(
    birth_rate_distr = create_exp_distr()
    )
```

Within this same example, one may specify the initial shape parameters of the exponential distribution. In BEAST2's implementation, an exponential distribution bution has one shape parameter: its mean, which can be set to any value with BEAUti. Within babette, to set the mean value of the exponential distribution to a fixed (non-estimated) value, do:

```
out <- run_beast2(
161
      "anthus_aco.fas",
162
      tree_priors = create_yule_tree_prior(
163
        birth_rate_distr = create_exp_distr(
164
          mean = create_mean_param(
             value = 1.0,
             estimate = FALSE
167
          )
168
        )
169
      )
170
   )
171
```

Our initial motivation to create babette was that we wanted to fix the crown age of a phylogeny. BEAUti assumes that a phylogeny has a crown age that needs to be jointly commated with the phylogeny and other parameters. It does not allow for fixing the crown age. Without babette, one needs to manually edit the BEAST2 configuration file (Bouckaert & Vaughan 2017), which is tedious and prone to errors. Fixing the crown ages is especially useful for theoretical experiments, as this allows for one less source of variation. This is how to specify a fixed crown age with babette:

```
180  out <- run_beast2(
181     "anthus_aco.fas",
182     posterior_crown_age = 15
183  )</pre>
```

```
babette allows for the same functionality as Tracer. Tracer works on the val-
    ues of the parameter estimates sampled in the BEAST2 run. This is called
185
    the "trace" (hence the name). The start of the trace is usually discarded, as
186
    an MCMC algorithm (such as used by BEAST2) first has to converge to its
187
    equilibrium. The start of the trace, called the "burn-in", will be removed,
188
    its parameter estimates are not representative. By default, Tracer discards the
189
    first 10% of all the parameter estimates. To remove a 20% burn-in from all
190
    parameter estimates in babette, the following code can be used:
191
    traces <- remove_burn_ins(
192
      out $ estimates,
193
      burn_in_fraction = 0.2
194
    )
195
    Tracer shows the ESSes of each posterior's variables. These ESSes are important
    to determine the strength of the inference. As a rule of thumb, an ESS of 200 is
197
    acceptable for any parameter estimate. To calculate the effective sample sizes
198
    (of all estimated variables) in babette:
199
    esses <- calc_esses(
200
      traces.
201
      sample_interval = 1000
202
    )
203
    Tracer displays multiple summary statistics for each estimated variable: the
204
    mean and its standard error, standard deviation, variance, median, mode, geo-
    metric mean, 95% highest posterior density interval, auto-correlation time and
206
    effective sample size. It displays these statistics per variable. In babette, these
    summary statistics are collected for all estimated parameters at once
208
    sum_stats <- calc_summary_stats(</pre>
209
      traces,
210
```

```
sample_interval = 1000
211
    )
212
    babette allows for the same functionality as DensiTree. DensiTree displays the
213
    phylogenies in a posterior at the same time scale, drawn one over one another,
214
    allowing to see the uncertainty in topology and branch lengths. Within the
215
    object out, the posterior phylogenies are stored as anthus_aco_trees, and can
216
    be plotted as such:
217
    plot_densitree(out$anthus_aco_trees)
218
```

babette resources 4 219

235

```
babette is free, libre and open source software available from the official R pack-
220
    age archive at http://cran.r-project.org/src/contrib/PACKAGES.html#babette
221
    and is licensed under the GNU General Public License v3.0. babette uses the
222
    Travis CI (https://travis-ci.org) continuous integration service, which is
    known to significantly increase the number of bugs exposed (Vasilescu et al.
224
    2015) and increases the speed at which new features are added (Vasilescu et al.
    2015). babette has a 100% code coverage, which correlates with code quality
226
    (Horgan et al. 1994; Del Frate et al. 1995). babette follows Hadley Wickham's
    style guide (Wickham 2015), which improves software quality (Fang 2001).
228
    babette dependends on multiple packages, which are ape (Paradis et al. 2004),
229
    beautier (Bilderbeek 2018b), beastier (Bilderbeek 2018a), devtools (Wick-
230
    ham & Chang 2016), geiger (Harmon et al. 2008), ggplot2 (Wickham 2009),
231
    knitr (Xie 2017), phangorn (Schliep 2011), rmarkdown (Allaire et al. 2017),
232
    seqinr (Charif & Lobry 2007), stringr (Wickham 2017), testit (Xie 2014)
233
    and tracerer (Bilderbeek 2018c).
234
       babette's development takes place on GitHub, https://github.com/richelbilderbeek/
```

babette, which accommodates collaboration (Perez-Riverol et al. 2016) and im proves transparency (Gorgolewski & Poldrack 2016). babette's GitHub facili tates feature requests and has guidelines how to do so.

babette's documentation is extensive. All functions are documented in the
package's internal documentation. For quick use, each exported function shows
a minimal example. For easy exploration, each exported function's documentation links to related functions. Additionally, babette has a vignette that
demonstrates extensively how to use it. The GitHub documentation helps to get
started, with a dozen examples of BEAUti screenshots with equivalent babette
code.

²⁴⁶ 5 Citation of babette

Scientists using babette in a published paper can cite this article, and/or cite
the babette package directly. To obtain this citation from within an R script,
use:

250 > citation("babette")

$_{^{251}}$ 6 Acknowledgements

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References

- Allaire, J., Xie, Y., McPherson, J., Luraschi, J., Ushey, K., Atkins, A., Wick-
- ham, H., Cheng, J. & Chang, W. (2017) rmarkdown: Dynamic Documents
- for R. R package version 1.8.
- Bilderbeek, R.J. (2018a) beastier: BEAST2 from R. https://github.com/
- richelbilderbeek/beastier [Accessed: 2018-02-28].
- Bilderbeek, R.J. (2018b) beautier: BEAUti 2 from R. https://github.com/
- richelbilderbeek/beautier [Accessed: 2018-02-28].
- Bilderbeek, R.J. (2018c) tracerer: Tracer from R. https://github.com/
- richelbilderbeek/tracerer [Accessed: 2018-02-28].
- Bouckaert, R. & Heled, J. (2014) Densitree 2: Seeing trees through the forest.
- bioRxiv, p. 012401.
- Bouckaert, R., Heled, J., Kühnert, D., Vaughan, T., Wu, C.H., Xie, D., Suchard,
- M.A., Rambaut, A. & Drummond, A.J. (2014) Beast 2: a software platform
- for bayesian evolutionary analysis. *PLoS Comput Biol*, **10**, e1003537.
- Bouckaert, R. & Vaughan, T. (2017) Fix starting tree. http://www.beast2.
- org/fix-starting-tree [Accessed: 2018-02-28].
- ²⁷⁶ Charif, D. & Lobry, J. (2007) SeqinR 1.0-2: a contributed package to the R
- 277 project for statistical computing devoted to biological sequences retrieval and
- analysis. U. Bastolla, M. Porto, H. Roman & M. Vendruscolo, eds., Struc-
- tural approaches to sequence evolution: Molecules, networks, populations, Bi-
- ological and Medical Physics, Biomedical Engineering, pp. 207–232. Springer
- ²⁸¹ Verlag, New York. ISBN: 978-3-540-35305-8.

- Del Frate, F., Garg, P., Mathur, A.P. & Pasquini, A. (1995) On the correlation
- between code coverage and software reliability. Software Reliability Engi-
- neering, 1995. Proceedings., Sixth International Symposium on, pp. 124–132.
- 285 IEEE.
- Drummond, A.J., Suchard, M.A., Xie, D. & Rambaut, A. (2012) Bayesian phy-
- logenetics with beauti and the beast 1.7. Molecular biology and evolution, 29,
- 288 1969-1973.
- Fang, X. (2001) Using a coding standard to improve program quality. Quality
- 290 Software, 2001. Proceedings. Second Asia-Pacific Conference on, pp. 73–78.
- 291 IEEE.
- Faria, N. & Suchard, M.A. (2015) RBeast. https://github.com/beast-dev/
- 293 RBeast [Accessed: 2018-03-02].
- 294 Gorgolewski, K.J. & Poldrack, R. (2016) A practical guide for improving trans-
- parency and reproducibility in neuroimaging research. bioRxiv, p. 039354.
- ²⁹⁶ Harmon, L., Weir, J., Brock, C., Glor, R. & Challenger, W. (2008) Geiger:
- investigating evolutionary radiations. *Bioinformatics*, **24**, 129–131.
- ²⁹⁸ Horgan, J.R., London, S. & Lyu, M.R. (1994) Achieving software quality with
- testing coverage measures. Computer, 27, 60–69.
- 300 Matzke, N.J. (2015) BEASTmasteR: R tools for automated conversion of
- NEXUS data to BEAST2 XML format, for fossil tip-dating and other uses.
- https://github.com/nmatzke/BEASTmasteR [Accessed: 2018-02-28].
- Paradis, E., Claude, J. & Strimmer, K. (2004) APE: analyses of phylogenetics
- and evolution in R language. *Bioinformatics*, **20**, 289–290.

- Perez-Riverol, Y., Gatto, L., Wang, R., Sachsenberg, T., Uszkoreit, J., Lepre-
- vost, F., Fufezan, C., Ternent, T., Eglen, S.J., Katz, D.S. et al. (2016) Ten
- simple rules for taking advantage of git and github. bioRxiv, p. 048744.
- R Core Team (2013) R: A Language and Environment for Statistical Computing.
- R Foundation for Statistical Computing, Vienna, Austria.
- Rambaut, A. & Drummond, A.J. (2007) Tracer v1.4. Available from
- http://beast.bio.ed.ac.uk/Tracer.
- Ratmann, O. (2015) rBEAST. https://github.com/olli0601/rBEAST [Ac-
- cessed: 2018-03-02].
- Schliep, K. (2011) phangorn: phylogenetic analysis in Bioinformatics, 27,
- ³¹⁵ 592–593.
- Van Els, P. & Norambuena, H.V. (2018) A revision of species limits in neotrop-
- ical pipits anthus based on multilocus genetic and vocal data. *Ibis*.
- Vasilescu, B., Yu, Y., Wang, H., Devanbu, P. & Filkov, V. (2015) Quality and
- productivity outcomes relating to continuous integration in github. Proceed-
- ings of the 2015 10th Joint Meeting on Foundations of Software Engineering,
- pp. 805–816. ACM.
- Wickham, H. (2009) ggplot2: elegant graphics for data analysis. Springer New
- York.
- Wickham, H. (2015) R packages: organize, test, document, and share your code.
- O'Reilly Media, Inc.
- Wickham, H. (2017) stringr: Simple, Consistent Wrappers for Common String
- Operations. R package version 1.2.0.

- Wickham, H. & Chang, W. (2016) devtools: Tools to Make Developing R Pack-
- ages Easier. R package version 1.12.0.9000.
- 330 Xie, Y. (2014) testit: A Simple Package for Testing R Packages. R package
- version 0.4, http://CRAN.R-project.org/package=testit.
- 332 Xie, Y. (2017) knitr: A General-Purpose Package for Dynamic Report Genera-
- tion in R. R package version 1.17.

Name	Description
run_beast2	Run BEAST2
create_gtr_site_model	Create a GTR site model
create_hky_site_model	Create an HKY site model
create_jc69_site_model	Create a Jukes-Cantor site model
create_tn93_site_model	Create a TN93 site model
create_rln_clock_model	Create a relaxed log-normal clock model
create_strict_clock_model	Create a strict clock model
create_bd_tree_prior	Create a birth-death tree prior
create_cbs_tree_prior	Create a coalescent Bayesian skyline tree prior
create_ccp_tree_prior	Create a coalescent constant-population tree prior
create_cep_tree_prior	Create a coalescent exponential-population tree prior
create_yule_tree_prior	Create a Yule tree prior
create_beta_distr	Create a beta distribution
create_exp_distr	Create an exponential distribution
create_gamma_distr	Create a gamma distribution
create_inv_gamma_distr	Create an inverse gamma distribution
create_laplace_distr	Create a Laplace distribution
create_log_normal_distr	Create a log-normal distribution
create_normal_distr	Create a normal distribution
create_one_div_x_distr	Create a 1/X distribution
create_poisson_distr	Create a Poisson distribution
create_uniform_distr	Create a uniform distribution

Table 1: babette's main functions

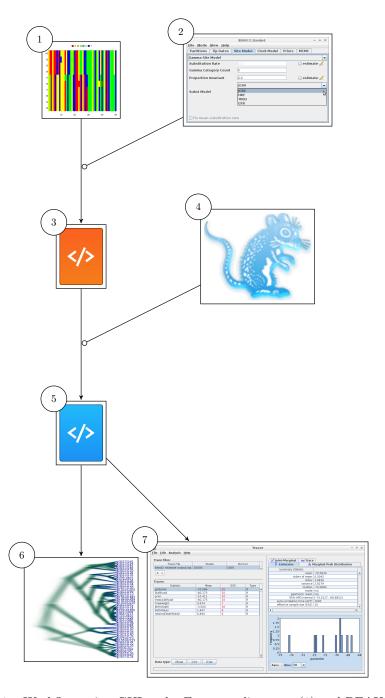


Figure 1: Workflow using GUI tools. From an alignment (1) and BEAUti (2), a BEAST2 configuration file (3) is created. BEAST2 (4) uses that file to infer a posterior, storing it in multiple files (5). These results are visualized using DensiTree (6) and Tracer (7). babette allows for the same workflow, all from an R function call.