The Goodman–Kruskal Coefficient and Its Applications in Genetic Diagnosis of Cancer

Szymon Jaroszewicz, Dan A. Simovici*, Member, IEEE, Winston P. Kuo, and Lucila Ohno-Machado

Abstract—Increasing interest in new pattern recognition methods has been motivated by bioinformatics research. The analysis of gene expression data originated from microarrays constitutes an important application area for classification algorithms and illustrates the need for identifying important predictors. We show that the Goodman–Kruskal coefficient can be used for constructing minimal classifiers for tabular data, and we give an algorithm that can construct such classifiers.

Index Terms—Bonferroni correction, classification, gene expression, predictors.

I. Introduction

LOBAL gene expression profiling using DNA microarrays has emerged as a rapid means to explore, classify, and predict the biological processes underlying human diseases. In the field of cancer, this revolutionary technology permits the simultaneous measurement of the transcription of tens of thousands of genes and of their relative expression between normal, dysplastic, and malignant cells. Since the first report of microarrays [25], the number of cancer and microarray related publications has increased exponentially, from approximately 30 publications in the first four years (1995–1999) to almost 600 publications in the past 24 months. Microarrays have evolved from depositing DNA onto a solid support to measure gene expression profiles to DNA copy number analysis [comparative genomic hybridization arrays (CGH)] [23] and more recently protein and antibody spotted microarrays [9].

The construction of gene expression databases requires technologies that can accurately and reproducibly measure changes in global mRNA expression levels. Ideally, these technologies should be able to screen all gene transcripts, be applicable across a wide range of cell and tissue types, require minimal amounts of biological material, and be capable of processing a large number of samples. There are currently two commonly used types of DNA microarrays: spotted complementary DNA (cDNA) microarrays [3] and short oligonucleotide arrays

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- S. Jaroszewicz is with the Department of Computer Science, University of Massachusetts, Boston, MA 02125 USA (e-mail: sj@cs.umb.edu).
- *D. A. Simovici is with the Department of Computer Science, University of Massachusetts, Boston, MA 02125 USA (e-mail: dsim@cs.umb.edu).
- W. P. Kuo and L. Ohno-Machado are with the Decision System Group, Brigham and Womens' Hospital, Boston, MA 02115 USA and also with the Division of Health Sciences and Technology, Harvard–MIT, Cambridge, MA 02139 USA (e-mail: wkuo@dsg.bwh.harvard.edu; machado@dsg.bwh.harvard.edu).

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(Affymetrix GeneChips) [15]. cDNA arrays represent a popular platform in which double-stranded polymerase chain reaction (PCR) products amplified from expressed sequence tag (EST) clones are robotically spotted onto glass slides. The average product size ranges from 100 to 1000 nucleotides in length. cDNA arrays offer versatility where project specific arrays can be custom-designed. For example, it is possible to build cancer-specific and chromosome-specific arrays, and they can be employed in the simultaneous analysis of two different biological samples (e.g., normal versus malignant tissues, tumors of different stages, or cancer cell lines). More importantly is the opportunity to discover novel genes since, in addition to well-characterized genes, ESTs of unknown function can be spotted on the DNA arrays. This approach has been used by several groups for cancer research [2], [4], [22]. GeneChips, on the other hand, are manufactured by synthesizing one nucleotide at a time onto a glass slide and consist of short oligonucleotides. These oligonucleotide arrays have also been used widely in cancer research [1], [8], [18]. In general, there are advantages and disadvantages to both microarray technology platforms; however, the crucial difference between the way cDNA and oligonucleotide microarrays are commonly used is that cDNA experiments return the amount of each transcript relative to another sample, whereas oligonucleotide experiments return an absolute amount of each transcript. This implies a major difference in the ability to group and universally compare across different microarray platforms [13]. Regardless of the technology platform chosen, microarray experiments yield far more information than we were used to processing in biological experiments.

Classical empirical approaches to anticancer therapy strategies are still being used, although global approaches to identify new targets for anticancer drugs represent a faster strategy. However, several bottlenecks still exist and translating microarrays findings into clinical applications remains a work in progress. Recent microarray studies involving cancer research have focused on tumor classification, outcome prediction. and progression of disease. Combining expression analysis with emerging technologies, such as CGH microarrays, laser capture microdissection (LCM) [27], and single-nucleotide polymorphisms (SNPs) and proteomics analyses may provide new possibilities to understand the genetic changes associated with cancer etiology and development. Ultimately, this may allow the discovery of new biomarkers for disease diagnosis and prognosis prediction and development of therapeutic strategies for cancer treatment.

Tumor classification using microarray data can be implemented using algorithms that identify minimal classification

criteria for tabular data. We present such an algorithm starting with the Goodman–Kruskal association index (see [7] and [16]), which is one of two measures of association introduced in [7] that can be naturally interpreted as a probability of misclassification.

Let X, Y be two discrete random variables. We assume that we deal with a finite probability space where the elementary events are pairs of values (a_i, b_j) , where a_i is a value of X and b_j is a value of Y. The Goodman–Kruskal coefficient of X and Y is defined by

$$\begin{aligned} \mathsf{GK}(X,Y) &= \sum_{i=1}^{l} P(X = a_i) (1 - \max_{1 \le j \le k} P(Y = b_j \, | \, X = a_i)) \\ &= 1 - \sum_{i=1}^{l} P(X = a_i) \max_{1 \le j \le k} P(Y = b_j \, | \, X = a_i). \end{aligned}$$

Goodman and Kruskal adopt the classification rule that prescribes that an elementary event is to be classified in the class that has the maximal probability. In the absence of any knowledge about X, an elementary event will be classified in the Y class b_j if b_j corresponds to the highest value among the probabilities $P(Y=b_j)$ for $1 \le j \le k$. If $P(Y=b_j \mid X=a_i)$ is the probability of predicting the value b_j for Y when $X=a_i$, then an event that has the component $X=a_i$ will be classified in the Y class b_j if j is the number for which $P(Y=b_j \mid X=a_i)$ has the largest value. The probability of misclassification committed by applying this rule is $1-\max_{1\le j\le k} P(Y=b_j \mid X=a_i)$. Thus, $\mathsf{GK}(X,Y)$ is the expected probability that in a randomly chosen case the value of Y will be incorrectly predicted from X.

The Goodman–Kruskal association index $\lambda_{Y|X}$, commonly used in literature, is the relative reduction in the probability of prediction error and is given as follows:

$$\lambda_{Y \mid X} = 1 - \frac{\mathsf{GK}(X, Y)}{1 - \max_{1 \le j \le k} P(Y = b_j)}.$$

In other words, $\lambda_{Y|X}$ is the proportion of the relative error in predicting the value of Y that can be eliminated by knowledge of the X value.

In the remainder of the paper, we use exclusively the Goodman-Kruskal coefficient.

In the next section we formulate a definition of the Goodman–Kruskal coefficient **GK** within a purely algebraic setting, using partitions of finite sets. The main advantage of this formulation is that we can use the properties of the partially ordered set of partitions of a set in our considerations.

Starting from the properties of GK discussed in Section II, we formulate an algorithm that identifies minimal classification criteria for tabular data. We applied this algorithm to two well-known sets of data discussed in [14] and in [8]. The data set of [14] is used to differentiate between four types of child-hood tumors known collectively as round blue cell tumors. The second set, presented in [8], is used to distinguish between acute myeloid leukemia (AML) and acute lymphoblastic leukemia (ALL). Both data sets were intensively explored in recent publications [20], [21], [29] using fuzzy sets, neural networks, and support vector machines.

II. THE GOODMAN-KRUSKAL COEFFICIENT FOR PARTITIONS

Let S be a finite set. A partition of S is a family of its subsets $\{B_1, B_2, \ldots, B_k\}$ such that $\bigcup_{i=1}^k B_i = S$ and $B_i \cap B_j = \emptyset$ for all $1 \leq i < j \leq k$. Let PART(S) be the set of partitions of S. For $\pi, \sigma \in \mathsf{PART}(S)$, we write $\pi \leq \sigma$ if for every block B of π there exists a block C of σ such that $B \subseteq C$. It is easy to verify that the relation " \leq " is a partial order on PART(S). For example, if $S = \{1, 2, \ldots, 5\}$, then $\{\{1, 2\}, \{3\}, \{4, 5\}\} \leq \{\{1, 2, 3\}, \{4, 5\}\}$.

Consider two partitions $\pi = \{B_1, \dots, B_l\}$ and $\sigma = \{C_1, \dots, C_k\}$ in PART(S). Define the Goodman–Kruskal coefficient of these partitions $\mathsf{GK}(\pi, \sigma)$ as the number

$$\mathsf{GK}(\pi,\sigma) = 1 - \sum_{i=1}^{l} \max_{1 \le j \le k} \frac{|C_j \cap B_i|}{|S|}.$$

The partitions π , σ define two random variables

$$X: \begin{pmatrix} 1 & \cdots & l \\ \frac{|B_1|}{|S|} & \cdots & \frac{|B_l|}{|S|} \end{pmatrix} \quad \text{and} \quad Y: \begin{pmatrix} 1 & \cdots & k \\ \frac{|C_1|}{|S|} & \cdots & \frac{|C_k|}{|S|} \end{pmatrix}$$

such that conditional probability P(Y = j | X = i) is given by

$$P(Y = j \mid X = i) = \frac{P(Y = j \land X = i)}{P(X = i)} = \frac{|C_j \cap B_i|}{|B_i|}.$$

Thus, for a fixed i, the largest error in predicting Y is

$$1 - \max_{1 \le j \le k} P(Y = j \mid X = i) = 1 - \max_{1 \le j \le k} \frac{|C_j \cap B_i|}{|B_i|}.$$

The expected value of this error is

$$\sum_{i=1}^{l} \frac{|B_i|}{|S|} \cdot \left(1 - \max_{1 \le j \le k} \frac{|C_j \cap B_i|}{|B_i|}\right)$$

$$= 1 - \sum_{i=1}^{l} \max_{1 \le j \le k} \frac{|C_j \cap B_i|}{|S|}$$

which is exactly the Goodman–Kruskal coefficient GK(X,Y).

Several properties of GK important from the point of view of classification are given next. The proofs can be found in the Appendix.

Theorem 2.1: Let S be a finite set, and let $\pi, \sigma \in \mathsf{PART}(S)$, where $\sigma = \{C_1, \dots, C_k\}$. We have

$$\mathsf{GK}(\pi,\sigma) \le 1 - \max_{1 \le j \le k} \frac{|C_j|}{|S|}.$$

Proof: See the Appendix, Section A. Theorem 2.2: Let S be a finite set and let $\pi = \{B_1, \dots, B_l\}$ and $\sigma = \{C_1, \dots, C_k\}$ be two partitions of the set S. We have $\mathsf{GK}(\pi, \sigma) = 0$ if and only if $\pi < \sigma$.

Proof: See the Appendix, Section B.

Theorem 2.3: The function GK is monotonic in its first argument and dually monotonic in its second. In other words, if

T				
	A_1	A_2		A_n
t_1	a_{11}	a_{12}		a_{1n}
t_2	a_{21}	a_{22}		a_{2n}
:	:		:	:
t_m	a_{m1}	a_{m2}		a_{mn}

Fig. 1. Structure of a table.

 π, π', σ are three partitions of the set S such that $\pi \leq \pi'$, then $\mathsf{GK}(\pi, \sigma) \leq \mathsf{GK}(\pi', \sigma)$, and if π, σ', σ are three partitions of the set S such that $\sigma \leq \sigma'$, then $\mathsf{GK}(\pi, \sigma) \geq \mathsf{GK}(\pi, \sigma')$.

Proof: See the Appendix, Section C.

The Goodman–Kruskal coefficient allows us to define a metric on $\mathsf{PART}(S)$. Consider the function $d_{\mathsf{GK}}: \mathsf{PART}(S) \times \mathsf{PART}(S) \longrightarrow \mathbb{R}$ given by

$$d_{GK}(\pi, \sigma) = GK(\pi, \sigma) + GK(\sigma, \pi)$$

for $\pi, \sigma \in \mathsf{PART}(S)$.

Theorem 2.4: The function d_{GK} is a metric on the set PART(S).

III. ALGORITHM FOR PREDICTOR IDENTIFICATION

We use standard relational database terminology, as it appears, for example, in [26]. A database table τ is a triple $\tau =$ (T, H, ρ) , where T is the table's name, H is its header, and ρ represents its contents. The header is a set of symbols H = $A_1 \dots A_n$ called attributes, which serve as labels of the columns of the table (see Fig. 1). For each attribute A_i , we have a set called the domain of A_i that is denoted by $Dom(A_i)$. Only values of $Dom(A_i)$ may appear in the column labeled A_i . The content of the table consists of a set of rows, $\{t_1, \ldots, t_m\}$ that belong to the set product $Dom(A_1) \times \cdots \times Dom(A_n)$. Every attribute set $K \subseteq H$ induces a partition π_K on the contents of the table. Namely, two tuples t and t' belong to the same block of the partition π_K if they have equal values on the attributes of K, that is, t[K] = t'[K] (see Fig. 2). Thus, the tuples t_1, t_2, t_3 belong to the same block of the partition π_K because they have equal values for the attributes of K, and so on. Therefore, the terms attribute set and partition are used interchangeably.

Let $\tau=(T,H,\rho)$ be a table and let K,L be two sets of attributes, $K,L\subseteq H$. The Goodman–Kruskal coefficient $\mathsf{GK}(K,L)$ of the sets of attributes K,L is defined as $\mathsf{GK}(\pi_K,\pi_L)$ and can be interpreted as the expected error that occurs when we try to predict the value of t[L] from the value of t[K]. If $K_1\subseteq K_2$, then, by Theorem 2.3, $\pi_{K_2}\leq \pi_{K_1}$, hence $\mathsf{GK}(K_2,L)\leq \mathsf{GK}(K_1,L)$, which has an intuitive interpretation: the expected error of a larger set of attributes is smaller than the expected error of a small set of attributes.

Using a similar mechanism, the metric $d_{\rm GK}$ can be transferred to the collection of sets of attributes of a table by defining $d_{\rm GK}(K,L)=d_{\rm GK}(\pi_K,\pi_L)$ for any two sets of attributes K,L. The new metric can be used for constructing classifiers, for performing discretization of continuous attributes, and for attribute clustering.

		T	
		$\longleftarrow K \longrightarrow$	
t_1		k_1	
t_2		k_1	
t_3		k_1	
:		:	:
t_l		$egin{array}{c} k_p \ k_p \end{array}$	
t_{l+1}		k_p	
t_{l+2}		k_p	
:	:	:	:
t_{n-1}		k_r k_r	
t_n		k_r	

Fig. 2. Partition of the content induced by a set of rows.

Definition 3.1: An ϵ -predictor for a set of attributes Y is a set of attributes K such that $\mathsf{GK}(K,Y) \leq \epsilon$.

If K is an ϵ -predictor for Y, then any superset K' of K is also a ϵ -predictor for Y. An ϵ -predictor such that none of its proper subsets is an ϵ -predictor is called *minimal*.

To find ϵ -predictors, we use the following algorithm:

Input: A set of attributes H, a target attribute $Y,Y \not\in H$ and an error level ϵ .

Output: Set P of all minimal ϵ -predictors from H.

- (1) Cand = $\{\{A\} : A \in H\};$
- (2) $P = \emptyset$;
- (3) $P = P \cup \{K \in Cand : GK(K, Y) \le \epsilon\};$
- (4) Cand = Cand \setminus P;
- (5) Cand = $\{L \subseteq H : \text{ for all } K \subset L, |K| = |L| 1 \text{ we have } K \in \text{Cand}\};$
- (6) goto (3);

The algorithm works in a manner very similar to the well-known data-mining Apriori algorithm [10], using the fact that, if a set is a nonminimal predictor, so are all of its supersets, which can, thus, be skipped. It begins by constructing a candidate set of predictors Cand in which all one-set attributes are initially included. The set of minimal predictors P is constructed starting from Cand. Initially, we include in P all one-attribute predictors whose error is below the threshold ϵ . These attributes are removed from P and the search for minimal two-attribute predictors makes use of the remaining candidate attributes, and so on. The stopping condition could be exceeding the maximum predictor size or finding a predictor with desired prediction error.

IV. EXPERIMENTAL RESULTS

As experimental data, we used the blood cell tumor microarray data of Khan *et al.* [14] and the leukemia data of Golub *et al.* [8]. In both datasets, gene expression levels have been normalized.

Full descriptions of experimental procedures and normalization methods used can be found in the original paper [14]; details on normalization methods applied to the second set of data are available at the web site, which is a companion of [8].

¹Available. [Online]. www-genome.wi.mit.edu/mpr/publications/projects/ Leukemia

TABLE I

original attribute	computed attributes			
Cancer type	NB	RMS	BL	EWS
NB	1	0	0	0
RMS	0	1	0	0
BL	0	0	1	0
EWS	0	0	0	1
other	0	0	0	0

The first data set (in [14]) involves microarray data that is used for differential diagnosis of four round blue cell tumors of childhood (SRBCTs): neuroblastoma (NB), rhabdomyosarcoma (RMS), Burkitt lymphoma (BL), and the Ewing family of sarcomas (EWS). The expression levels of 2308 genes were measured using cDNA microarrays and the results of a predictive model based on a single-layer neural network architecture [14] and a logistic regression model [28] were previously reported.

There are 63 training (12 NB, 20 RMS, 8 BL, and 23 EWS) and 25 test cases (6 EWS, 5 RMS, 6 NB, 3 BL, and 5 non-SRBCTs) in the dataset, which includes samples obtained from cell lines and biopsies. The test cases include five cases which do not belong to any of the predicted SRBCT types. Such cases are not present in the training set.

In the initial preprocessing step, we replace each class attribute with four binary attributes, one for each cancer type. Each of the four binary attributes has a value of 1 if and only if the corresponding cancer type is present. Table I illustrates this encoding.

Then a separate predictor is built for each of the new binary attributes, i.e., we try to predict every tumor type separately. The main reason for this is to allow for handling of cases of type "other" present in the test set but absent in the training set. We expect that for "other" cancer type all of the predictors will give the value of 0, thus indicating that none of the four cancer types is present.

A disadvantage is that predictors may contradict each other; however, the experiments have shown that such cases are infrequent due to a low error rate of individual classifiers. If, however, the presence of more than one cancer type is predicted, we assume that the prediction cannot be made in such a case and consider it misclassified.

Since the number of training cases is very small, there is a serious risk of so-called overfitting, that is, building a complex classifier that fits particular training data very well but does not generalize to unseen cases. Building a separate predictor for each cancer type makes the predictors very simple and, thus, less likely to overfit the data.

Next, every gene expression level X attribute is discretized into two intervals: $X \leq T$ and X > T for some threshold T, thus changing the continuous gene expression level X into a binary attribute X'. The threshold T is chosen such that the Shannon entropy $H(Y \mid X')$ of the target Y conditional on the discretized attribute X' is minimal. A separate discretization has been performed with respect to each cancer type. To identify the thresholds used in discretization, we applied the Fayyad–Irani algorithm presented in [5] and [6].

Afterwards, the algorithm given above is used to find all predictors with one or two attributes, allowing up to one misclassified instance on the training set. Thus, the stopping rule for the learning algorithm is reaching the maximum prescribed size of the predictor or obtaining an error rate less than to (1/t), where t is the size of the training set. Building classifiers with more attributes would incur a large risk of overfitting due to small number of training cases (see [19]). Since the number of attributes is huge, there is an extra attribute selection step involved; namely, before the algorithm is applied, all but 30 of the most predictive attributes are discarded.

For each class (cancer type), the first predictor with minimal training error is manually picked at random (without looking at its test set performance to avoid bias in the choice). The results are summarized in Table II.

The results show that a fairly large number (12–30) of very simple predictors have been found for each cancer type. Each of those predictors has very good classification rate on the training set: up to one misclassified case is allowed.

For each cancer type, the number of errors on the test set is also given for selected predictors. The predictions on the test are slightly worse than on the training set, but still fairly accurate. This suggests that overfitting was not a serious issue here.

The results show that there are many genes, based on which a diagnosis can be made for each cancer type.

Of note, all genes except for the one that predicts BL were reported among the 96 selected in [14]. The probability that a single gene expression predicts BL perfectly in a training set when there is no correlation between the gene and the tumor type is given as follows:

$$2 \cdot \frac{55! \cdot 8!}{63!} = 5.16 \cdot 10^{-10}$$

which is much less than $0.05/2308 = 2.16 \cdot 10^{-5}$, the 5% significance level after Bonferroni correction. This shows that the selected gene is very likely related to BL. Similar statements hold for other cancer types. Bonferroni correction has been used since the number of attributes is large and there could be a possibility that one of them passes the statistical test due to chance rather than true correlation. In the absence of the attribute independence, the Bonferroni correction is overly conservative. However, the inferences using this correction are correct.

We combined the classifiers for each tumor type in the following way: if a classifier for only one type of tumor gave a positive prediction, then the instance was classified as this type of tumor. If none of them gave a positive prediction, we declared the case to be "other tumor type." If more than one classifier was active, the case was considered a prediction error.

The combined classifier used a total of 6 genes and classified correctly 18 out of 25 cases. Out of the 6 misclassified cases, two gave classifications when the real outcome was "other," three SRBCT cases were undetected, and there was one conflict.

The leukemia training data set contains 38 cases (27 acute lymphoblastic leukemia and 11 acute myelocytic leukemia) and the test set contains 34 cases (20 ALL and 14 AML); the data refer to 6817 genes. We discretized the gene expression levels using the same procedure as for the data given in [14]. We re-

TABLE II
PREDICTORS FOR ROUND BLUE CELL TUMORS

Cancer	selected predictor	image ids	mtr	mte	1GP	2GP
type						
BL	$WAS \le 0.69 \Rightarrow BL$	236282	0	1	15	5
EWS	$FCGRT \le 1.59 \Rightarrow EWS$	770394	1	3	2	10
NB	$MAP1B > 2.17 \text{ or } RCV1 > 1.98 \Rightarrow NB$	629896 - 383188	0	0	2	28
RMS	TNNT2 > 0.55 or SGCA > 0.44 \Rightarrow RMS	298062 - 796258	0	2	0	25

Legend:

mte misclassified cases in test set
mtr misclassified cases in training set
1GP number of one-gene predictors
2GP number of two-gene predictors

TABLE III

Error Interval	Number of 2-attribute
	predictors
[0.0, 0.05]	2
(0.05, 0.10]	9
(0.10, 0.15]	10
(0.15, 0.20]	7
(0.20, 0.25]	13
(0.25, 0.30]	14
(0.30, 0.35]	3
(0.35, 0.40]	4
(0.40, 0.45]	3

tained 20 genes for which the Goodman–Kruskal coefficient was below 0.04. Five single-genes predictors and 66 two-gene predictors were identified.

It is worth noting that our technique identified two two-gene predictors (MGST1, APLP2 and CD33, CystatinA) for which the errors on the test set are 0 and 0.029 411 8, respectively. CD33 was among the 50 genes selected in [8]. The distribution of the errors on the test set for the remaining set of minimal two-gene predictors is shown in Table III.

To obtain a robust classifier, we used a voting mechanism on the second data set. Initially, we extracted 19 one-attribute predictors whose prediction error on the training set did not exceed 5.3% (that is, two errors out of the 38 training cases). A vote was taken, and the instance was classified according to the majority vote. We obtained 3 errors on the test set of 34 cases. Namely, the errors occurred on the 57th, 60th, and 66th cases of the original Golub test set. It is worth mentioning that all errors occurred on cases that had low prediction strengths and were considered "unclassifiable" in [8].

V. CONCLUSION AND FUTURE RESEARCH

The Goodman–Kruskal dissimilarity GK is a simple but powerful measure of predictive power that can be used to produce robust classifiers that are comparable with those obtained using more complex techniques. In our opinion, the small number of training cases makes reliable construction of more complex models like Bayesian networks or C4.5 trees very difficult or even impossible. On the other hand, methods related to Naive Bayesian classifiers suffer from independence assumptions which may not be satisfied in the microarray

setting where most gene measurements are correlated with each other. As we mentioned earlier, the use of the Bonferroni correction, although conservative, yields valid results.

It would be interesting to see how more sophisticated attribute selection techniques influence the quality of classification for microarray data, see, e.g., [11], [12], and [17].

The fact that GK generates a metric suggests that one could use this metric to cluster attributes such that those that belong to a cluster have similar predictive power and, thus, are interchangeable in classifiers. The clustering structure could be used in forming the voting committees and in simplifying and increasing the robustness of predictive algorithms. This approach dovetails with other metrics on partitions that we explored in [24].

APPENDIX

Lemma 1.1: Let $\{x_{ij} \mid 1 \le i \le l, 1 \le j \le k\}$ be a collection of lk real numbers. We have

$$\max_{1 \le j \le k} \sum_{i=1}^{l} x_{ij} \le \sum_{i=1}^{l} \max_{1 \le j \le k} x_{ij}.$$

Proof: Observe that $x_{ij} \leq \max_{1 \leq j \leq k} x_{ij}$ for every i, j, which implies $\sum_{i=1}^{l} x_{ij} \leq \sum_{i=1}^{l} \max_{1 \leq j \leq k} x_{ij}$ for every j. Thus, we obtain

$$\max_{1 \le j \le k} \sum_{i=1}^{l} x_{ij} \le \sum_{i=1}^{l} \max_{1 \le j \le k} x_{ij}.$$

A. Proof of Theorem 2.1

The definition of $GK(\pi, \sigma)$ gives

$$\mathsf{GK}(\pi,\sigma) = 1 - \sum_{i=1}^{l} \max_{1 \leq j \leq k} \frac{|C_j \cap B_i|}{|S|}$$

where $\pi = \{B_1, \dots, B_l\}$. By Lemma 1.1 we can write

$$\begin{aligned} \mathsf{GK}(\pi,\sigma) &\leq 1 - \max_{1 \leq j \leq k} \sum_{i=1}^{l} \frac{|C_j \cap B_i|}{|S|} \\ &= 1 - \max_{1 \leq j \leq k} \frac{|C_j|}{|S|} \end{aligned}$$

which is the desired inequality.

B. Proof of Theorem 2.2

Suppose that $GK(\pi, \sigma) = 0$, which means that

$$\sum_{i=1}^{l} \max_{1 \le j \le k} \frac{|C_j \cap B_i|}{|S|} = 1.$$

This is possible only if for every B_i there exists C_j such that $B_i \subseteq C_j$, that is, if $\pi \le \sigma$. The reverse implication is clear.

C. Proof of Theorem 2.3

To prove the monotonicity of GK in its first argument, it suffices to show that, if π' is obtained from π by fusing two blocks B_i and B_h , then $GK(\pi, \sigma) \leq GK(\pi', \sigma)$.

We claim that for every i, h we have

$$\max_{1 \le j \le k} |C_j \cap (B_i \cup B_h)| \le \max_{1 \le j \le k} |C_j \cap B_i| + \max_{1 \le j \le k} |C_j \cap B_h|.$$

Indeed, observe that $|C_j \cap (B_i \cup B_h)| = |C_j \cap B_i| + |C_j \cap B_h| \le \max_{1 \le j \le k} |C_j \cap B_i| + \max_{1 \le j \le k} |C_j \cap B_h|$. This gives the desired inequality. Thus

$$\begin{split} \mathsf{GK}(\pi,\sigma) &= 1 - \left(\sum_{m=1,m \neq i,h}^{l} \max_{1 \leq j \leq k} \frac{|C_j \cap B_m|}{|S|} \right. \\ &+ \max_{1 \leq j \leq k} \frac{|C_j \cap B_i|}{|S|} + \max_{1 \leq j \leq k} \frac{|C_j \cap B_h|}{|S|} \right) \\ &\leq 1 - \left(\sum_{m=1,m \neq i,h}^{l} \max_{1 \leq j \leq k} \frac{|C_j \cap B_m|}{|S|} \right. \\ &+ \max_{1 \leq j \leq k} \frac{|C_j \cap (B_i \cup B_h)|}{|S|} \right) = \mathsf{GK}(\pi',\sigma). \end{split}$$

For the second part of the theorem, it suffices to show that, if σ' is obtained from σ by fusing two blocks C_i and C_h , then the above inequality holds.

For a block B_i of π , we have

$$\max\{|B_i \cap C_1|, \dots, |B_i \cap C_i|, \dots, |B_i \cap C_h|, \dots, |B_i \cap C_k|\}$$

$$\leq \max\{|B_i \cap C_1|, \dots, |B_i \cap (C_i \cup C_h)|, \dots, |B_i \cap C_k|\}.$$

Therefore

$$\sum_{i=1}^{k} \max_{1 \le j \le k} |B_i \cap C_j| \le \sum_{i=1}^{k} \max\{|B_i \cap C_j| | C_j \in \sigma'\}$$

which gives the inequality of the theorem.

D. Proof of Theorem 2.4

It is obvious that the function $d_{\rm GK}$ is nonnegative and symmetric.

Suppose that $d_{\mathrm{GK}}(\pi,\sigma)=0$, which implies $\mathrm{GK}(\pi,\sigma)=\mathrm{GK}(\sigma,\pi)=0$. Thus, by Theorem 2.2, we have both $\pi\leq\sigma$ and $\sigma\leq\pi$, which implies $\pi=\sigma$. Since $d_{\mathrm{GK}}(\pi,\pi)=0$ for any partition $\pi\in\mathsf{PART}(S)$, it follows that $d_{\mathrm{GK}}(\pi,\sigma)=0$ if and only if $\pi=\sigma$.

Let (b_{jh}) be a $k \times g$ matrix where $b_{jh} \ge 0$ for $1 \le j \le k$ and $1 \le h \le g$. Let j_0 be an integer between 1 and k and let h_0 be the least integer in $\{1,\ldots,g\}$ such that $b_{j_0h_0} = \max_{1 \le h \le g} b_{j_0h}$. We claim that

$$\sum_{h=1}^{g} b_{j_0 h} + \sum_{j=1}^{k} \max_{1 \le h \le g} b_{j h} \le \sum_{j=1}^{k} \sum_{h=1}^{g} b_{j h} + \sum_{j=1}^{k} b_{j h_0}.$$
 (1)

Inequality (1) can be written as

or

$$\sum_{j=1}^{k} \max_{1 \le h \le g} b_{jh} \le \sum_{j=1, j \ne j_0}^{k} \sum_{h=1}^{g} b_{jh} + \sum_{j=1}^{k} b_{jh_0},$$

$$\sum_{j=1, j \neq j_0}^{k} \max_{1 \le h \le g} b_{jh} + \max_{1 \le h \le g} b_{j_0 h}$$

$$\leq \sum_{k=1}^{k} \sum_{j=1}^{g} b_{jh} + \sum_{k=1}^{k} b_{jh_0}$$

which follows from the fact that $\max_{1 \le h \le g} b_{j_0 h}$ is one of the members of the sum $\sum_{j=1}^k b_{jh_0}$ in view of the definition of h_0 . This proves (1). Consequently, we have

$$\sum_{h=1}^{g} b_{j_0 h} + \sum_{j=1}^{k} \max_{1 \le h \le g} b_{jh} \le \sum_{j=1}^{k} \sum_{h=1}^{g} b_{jh} + \max_{1 \le h \le g} \sum_{j=1}^{k} b_{jh}$$

for every j_0 which, in turn, implies

$$\max_{1 \le j \le k} \sum_{h=1}^{g} b_{jh} + \sum_{j=1}^{k} \max_{1 \le h \le g} b_{jh} \le \sum_{j=1}^{k} \sum_{h=1}^{g} b_{jh} + \max_{1 \le h \le g} \sum_{j=1}^{l} b_{jh}.$$
(2)

Consider the partitions $\pi = \{B_1, \ldots, B_i, \ldots, B_l\}, \sigma = \{C_1, \ldots, C_j, \ldots, C_k\}$, and $\tau = \{D_1, \ldots, D_h, \ldots, D_g\}$ on the set S and define $a_{ijh} = |B_i \cap C_j \cap D_h|$ for $1 \le i \le l, 1 \le j \le k$, and $1 \le h \le g$. Inequality (2) implies

$$\max_{1 \le j \le k} \sum_{h=1}^{g} a_{ijh} + \sum_{j=1}^{k} \max_{1 \le h \le g} a_{ijh}$$

$$\leq \sum_{i=1}^{k} \sum_{h=1}^{g} a_{ijh} + \max_{1 \le h \le g} \sum_{i=1}^{k} a_{ijh}$$

for $1 \le i \le l$. Summing on i, we have

$$\sum_{i=1}^{l} \max_{1 \le j \le k} \sum_{h=1}^{g} a_{ijh} + \sum_{i=1}^{l} \sum_{j=1}^{k} \max_{1 \le h \le g} a_{ijh}$$

$$\leq \sum_{i=1}^{l} \sum_{j=1}^{k} \sum_{h=1}^{g} a_{ijh} + \sum_{i=1}^{l} \max_{1 \le h \le g} \sum_{i=1}^{l} a_{ijh}.$$

Equivalently,

$$\sum_{i=1}^{l} \max_{1 \le j \le k} |B_i \cap C_j| + \sum_{i=1}^{l} \sum_{j=1}^{k} \max_{1 \le h \le g} a_{ijh}$$

$$\leq |S| + \sum_{i=1}^{l} \max_{1 \le h \le g} |B_i \cap D_h|. \quad (3)$$

Observe that

$$\sum_{i=1}^{l} \max_{1 \le j \le k} |B_i \cap C_j| + \sum_{j=1}^{k} \max_{1 \le h \le g} |C_j \cap D_h|$$

$$= \sum_{i=1}^{l} \max_{1 \le j \le k} |B_i \cap C_j| + \sum_{j=1}^{k} \max_{1 \le h \le g} \sum_{i=1}^{l} |B_i \cap C_j \cap D_h|$$

$$\leq \sum_{i=1}^{l} \max_{1 \le j \le k} |B_i \cap C_j| + \sum_{j=1}^{k} \sum_{i=1}^{l} \max_{1 \le h \le g} |B_i \cap C_j \cap D_h|$$
(by Lemma 1.1)
$$= \sum_{i=1}^{l} \max_{1 \le j \le k} |B_i \cap C_j| + \sum_{j=1}^{k} \sum_{i=1}^{l} \max_{1 \le h \le g} a_{ijk}$$

$$\leq |S| + \sum_{i=1}^{l} \max_{1 \le h \le g} |B_i \cap D_h| \quad \text{[due to Inequality (3)]}.$$

After an elementary transformation, the last inequality can be written as

$$1 - \frac{1}{|S|} \sum_{i=1}^{l} \max_{1 \le j \le k} |B_i \cap C_j|$$

$$+ 1 - \frac{1}{|S|} \sum_{j=1}^{k} \max_{1 \le h \le g} |C_j \cap D_h|$$

$$\ge 1 - \frac{1}{|S|} \sum_{i=1}^{l} \max_{1 \le h \le g} |B_i \cap D_h|$$

which is equivalent to

$$GK(\pi, \sigma) + GK(\sigma, \tau) \ge GK(\pi, \tau).$$
 (4)

Similarly, we can write

$$GK(\tau, \sigma) + GK(\sigma, \pi) \ge GK(\tau, \pi).$$
 (5)

Adding the inequalities (4) and (5) yields the triangular inequality $d_{\rm GK}(\pi,\sigma)+d_{\rm GK}(\sigma,\tau)\geq d_{\rm GK}(\pi,\tau)$. This allows us to conclude that $d_{\rm GK}$ is a metric.

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REFERENCES

- [1] U. Alon, N. Barkai, D. A. Notterman, K. Gish, S. Ybarra, D. Mack, and A. J. Levine, "Broad patterns of gene expression revealed by clustering analysis of tumor and normal colon tissues probed by oligonucleotide arrays," in *Proc. Nat. Acad. Sci.*, vol. 96, 1999, pp. 6745–6750.
- [2] A. A. Alizadeh, M. B. Eisen, R. E. Davis, C. Ma, I. S. Lossos, A. Rosenwald, J. C. Boldrick, H. Sabet, T. Tran, X. Yu, J. I. Powell, L. Yang, G. E. Marti, T. Moore, J. Hudson Jr., L. Lu, D. B. Lewis, R. Tibshirani, G. Sherlock, W. C. Chan, T. C. Greiner, D. D. Weisenburger, J. O. Armitage, R. Warnke, and L. M. Staudt, "Distinct types of diffuse large b-cell lymphoma identified by gene expression profiling," *Nature*, vol. 403, no. 6769, pp. 503–511, 2000.
- [3] M. D. Adams, J. M. Kelley, J. D. Gocayne, M. Dubnick, M. H. Polymeropoulos, H. Xiao, C. R. Merril, A. Wu, B. Olde, and R. F. Moreno, "Complementary dna sequencing: Expressed sequence tags and human genome project," *Science*, vol. 252, no. 5013, pp. 1651–1656, 1991.

- [4] M. Bittner, P. Meltzer, Y. Chen, Y. Jiang, E. Seftor, M. Hendrix, M. Radmacher, R. Simon, Z. Yakhini, A. Ben-Dor, N. Sampas, E. Dougherty, E. Wang, F. Marincola, C. Gooden, J. Lueders, A. Glatfelter, P. Pollock, J. Carpten, E. Gillanders, D. Leja, K. Dietrich, C. Beaudry, M. Berens, D. Alberts, and V. Sondak, "Molecular classification of cutaneous malignant melanoma by gene expression profiling," *Nature*, vol. 406, no. 6795, pp. 536–540, 2000.
- [5] U. M. Fayyad, "On the induction of decision trees for multiple concept learning," Ph.D. dissertation, Univ. of Michigan, Ann Arbor, 1991.
- [6] U. M. Fayyad and K. Irani, "Multi-interval discretization of continuous-valued attributes for classification learning," in *Proc. 13th Int. Joint Conf. Artificial Intelligence*, 1993, pp. 1022–1027.
- [7] L. A. Goodman and W. H. Kruskal, Measures of Association for Cross-Classification. New York: Springer-Verlag, 1980, vol. 1.
- [8] T. R. Golub, D. K. Slonim, P. Tamayo, C. Huard, M. Gaasenbeek, J. P. Mesirov, H. Coller, M. L. Loh, J. R. Dowining, M. A. Caligiuri, C. D. Bloomfield, and E. S. Lander, "Molecular classification of cancer: Class discovery by gene expression monitoring," *Science*, vol. 286, pp. 531–537, 1999.
- [9] B. B. Haab, M. J. Dunham, and P. O. Brown, "Protein microarrays for highly parallel detection and quantitation of specific proteins and antibodies in complex solutions," *Genome. Biol.*, vol. 2, no. 2, pp. research0004.1–research0004.13, 2001.
- [10] J. Han and M. Kamber, Data Mining—Concepts and Techniques. San Francisco, CA: Morgan Kaufmann, 2001.
- [11] I. Inza, P. Larranaga, R. Etxeberria, and B. Sierra, "Feature subset selection by Bayesian network-based optimization," *Artif. Intell.*, vol. 123, no. 1–2, pp. 157–184, 2000.
- [12] R. Kohavi and G. H. John, "Wrappers for feature subset selection," Artif. Intell., vol. 97, no. 1–2, pp. 273–324, 1997.
- [13] W. P. Kuo, T. K. Jenssen, A. J. Butte, L. Ohno-Machado, and I. S. Kohane, "Analysis of matched mrna measurements from two different microarray technologies," *Bioinformatics*, vol. 18, no. 3, pp. 405–412, 2002.
- [14] J. Khan, J. Wei, M. Ringner, L. Saal, M. Ladanyi, F. Westerman, F. Berthold, M. Schwab, C. Antonescu, C. Peterson, and P. Meltzer, "Classification and diagnostic prediction of cancers using gene expression profiling and artificial neural networks," *Nature Medicine*, vol. 7, pp. 673–679, 2001.
- [15] R. J. Lipshutz, S. P. Fodor, T. R. Gingeras, and D. J. Lockhart, "High density synthetic oligonucleotide arrays," *Nat. Genet.*, vol. 21, pp. 20–24, 1999. (1 Suppl).
- [16] A. M. Liebtrau, *Measures of Association*. Beverly Hills, CA: SAGE,
- [17] H. Liu and H. Motoda, Feature Selection for Knowledge Discovery and Data Mining. Boston, MA: Kluwer, 1998.
- [18] J. H. Luo, Y. P. Yu, K. Cieply, F. Lin, P. Deflavia, R. Dhir, S. Finkelstein, G. Michalopoulos, and M. Becich, "Gene expression analysis of prostate cancers," *Mol. Carcinog*, vol. 33, no. 1, pp. 25–35, 2002.
- [19] T. M. Mitchell, Machine Learning. Boston, MA: McGraw-Hill, 1997.
- [20] S. Mukherjee, P. Tamayo, D. Slonim, A. Verri, T. Golub, J. Mesirov, and T. Poggio, Support Vector Machine Classification of Microarray Data: MIT Artificial Intelligence Laboratory, 1998. AI MEMO CBCL Paper 1677
- [21] L. Ohno-Machado, S. Vinterbo, and G. Weber, "Classification of gene expression data using fuzzy logic," *J. Intell. Fuzzy Syst.*, vol. 12, pp. 19–24, 2002.
- [22] C. M. Perou, S. S. Jeffrey, M. van de Rijn, C. A. Rees, M. B. Eisen, D. T. Ross, A. Pergamenschikov, C. F. Williams, S. X. Zhu, J. C. Lee, D. Lashkari, D. Shalon, P. O. Brown, and D. Botstein, "Distinctive gene expression patterns in human mammary epithelial cells and breast cancers," *Proc. Nat. Acad. Sci. USA*, vol. 96, no. 16, pp. 9212–9217, 1999.
- [23] D. Pinkel, R. Segraves, D. Sudar, S. Clark, I. Poole, D. Kowbel, C. Collins, W. L. Kuo, C. Chen, Y. Zhai, S. H. Dairkee, B. M. Ljung, J. W. Gray, and D. G. Albertson, "High resolution analysis of dna copy number variation using comparative genomic hybridization to microarrays," *Nat. Genet.*, vol. 20, no. 2, pp. 207–211, 1998.
- [24] D. A. Simovici and S. Jaroszewicz, "Generalized conditional entropy and decision trees," in *Proc. EGC* 2003, Lyon, France, pp. 369–380.
- [25] M. Schena, D. Shalon, R. W. Davis, and P. O. Brown, "Quantitative monitoring of gene expression patterns with a complementary dna microarray," *Science*, vol. 270, no. 5235, pp. 467–470, 1995.
- [26] D. A. Simovici and R. L. Tenney, Relational Database Systems. New York: Academic, 1995.
- [27] R. Todd, M. W. Lingen, and W. P. Kuo, "Gene expression profiling using laser capture microdissection," *Expert Rev. Mol. Diagn.*, vol. 2, no. 5, pp. 497–507, 2002.

- [28] G. Weber, S. Vinterbo, and L. Ohno-Machado, "Building an asynchronous web-based tool for machine learning classification," in *Proc. AMIA Symp.*, Chicago, IL, 2002, pp. 869–873.
- [29] C. Yeang, S. Ramaswami, P. Tamayo, S. Mukherjee, R. Rifkin, M. Angelo, M. Reich, E. Lander, J. Mesirov, and T. Golub, "Molecular classification of multiple tumor types," *Bioinformatics*, vol. 17, pp. 316–323, 2001.



Szymon Jaroszewicz received the M.Sc. degree from the Technical University of Szczecin, Poland, and the Ph.D. degree in computer science from the University of Massachusetts at Boston, both in computer science.

His research interests are in the applications of information-theoretical techniques in data mining, especially in the area of interestingness measures for association rules.

Dr. Jaroszewicz was the recipient of a Fulbright scholarship.



Winston P. Kuo received the D.D.S. degree from Columbia University, New York, NY, and the M.S. degree in medical informatics from the Massachusetts Institute of Technology, Cambridge, and he is currently working toward the Ph.D. degree at Harvard School of Dental Medicine, Cambridge, MA.

He is a Biomedical Informatics Research Fellow with Decision Systems Group, Children's Hospital Informatics Program, and with the Department of Genetics, all at Harvard Medical School, Cambridge. He completed his clinical specialties in Pediatric Den-

tistry and Oral Medicine at the University of Southern California, Los Angeles, and the Harvard School of Dental Medicine, respectively. His research interests include biological and computational approaches (machine learning and statistical) to explore and extract meaningful information from high-throughput approaches, such as DNA microarrays, with particular interest in the area of craniofacial development and malformation and oral cancer.



Dan A. Simovici (M'86) received the Ph.D. degree from the University of Bucharest, Bucharest, Romania

He is an author or coauthor of the books *Theory of Formal Languages with Applications* (Singapore: World Scientific, 1999), *Relational Database Systems* (New York: Academic, 1995), and *Mathematical Foundations of Computer Science* (Berlin, Germany: Springer-Verlag, 1991) and of more than 100 research papers. He is currently a Professor of Computer Science with the University of Massa-

chusetts at Boston. His main research interests are in information-theoretical methods in data mining and machine learning, and in multiple-valued logic. He is a Managing Editor of the *Journal for Multiple-Valued Logic and Soft Computing*.



Lucila Ohno-Machado received the M.D. degree from the University of Sao Paulo, Sao Paulo, Brazil and the Ph.D. degree in medical information sciences and computer science from Stanford University, Stanford, CA.

She is an Associate Professor of Radiology and Health Sciences and Technology with Harvard-MIT and Associate Director of the Decision Systems Group at Brigham and Women's Hospital, Boston, MA. Her research interests include predictive modeling using statistical and machine learning

methods, with particular emphasis on survival analysis.

Dr. Ohno-Machado is the recipient of awards for her work in biomedical informatics, including elected membership in the American College of Medical Informatics.