

Laboratory Diagnosis of Infection

The Role of Microbiology Laboratory

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Learning objectives

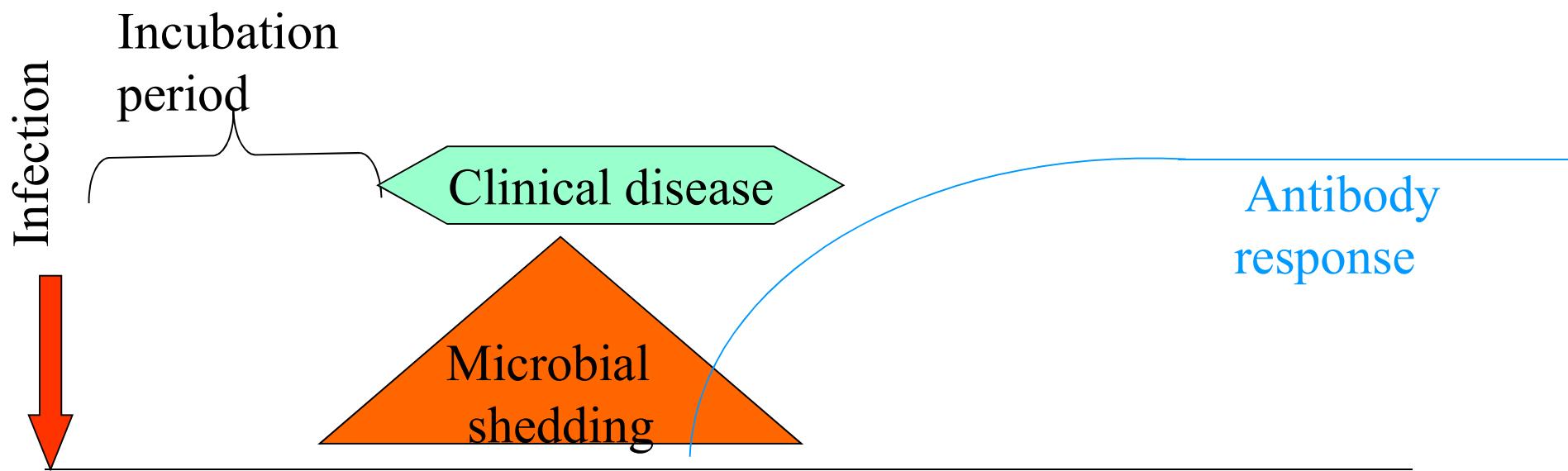
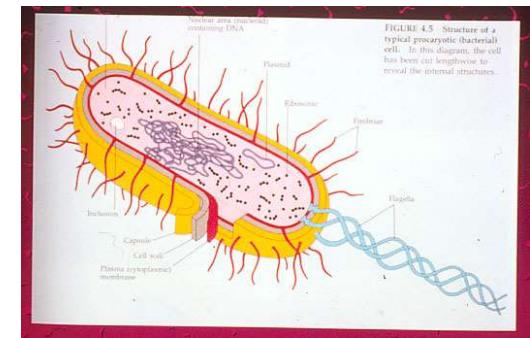
- List the reasons why microbiological laboratory diagnosis is important for good clinical care?
- Describe the factors that need to be considered in collection and transport of specimens for microbiological laboratory diagnosis?
- Describe the precautions to be followed when collecting blood for bacteriological culture?
- Describe the precautions to be followed when collecting urine for diagnosis of urinary tract infection?
- Describe the available options for diagnosis of viral infections?
- Describe how serological results are interpreted to arrive at a microbiological diagnosis?

Microbiological diagnosis: Why?

- Anatomical (clinical) diagnosis vs. microbiological diagnosis.
 - *A disease may be caused different pathogens*
 - e.g. pyogenic meningitis in a child is most often caused by *Streptococcus pneumoniae*, *Haemophilus influenzae*, *Neisseriae meningitidis*
- Antibiotic sensitivity test: *Accurately target antimicrobial therapy – specific vs. broad spectrum or combination therapy*
- Monitor effectiveness of therapy
- Infection control
- Public health and epidemiology

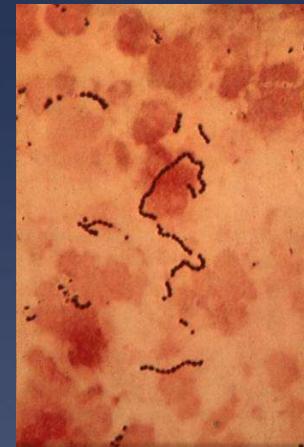
Microbial diagnosis: How?

- # Detect the microbe
- # Detect antibody response - serology



Detect the microbe: How

- see - microscopy, *minutes*
- grow (culture) – *days*
- Microbial antigen detection – *hours*
- Microbial nucleic acid detection (e.g. Polymerase chain reaction) (PCR) - *hours / days*



Collection and transport of clinical specimens for microbiological examination

- What specimen? representation of the infectious disease process
- Well collected specimen
- Appropriate transport medium or procedure
- Appropriate storage, packing and transport to the laboratory.
- Specimen request form: Relevant clinical details
 - *Date of onset of illness*
 - *Suspected clinical diagnosis*
 - *Prior antibiotic therapy*
 - *Known biological hazards (e.g. HIV positive) clearly stated on the request form and biohazard labels be put on the specimen bag.*

Queen Mary Hospital Department of Microbiology
 Bacteriology & Virology Investigation Form

Date of collection: _____ Time: _____ AM/PM
 (Day/Month/Year)

2nd Label

Biohazard: unknown/known (specify) _____	Date of admission: _____
H.K.I.D. No. _____	Hosp Name & No. _____
AE HN	
Surname _____	Sex/Age: _____
Firstname _____	Ward/Bed: _____
Physician in charge/unit _____	OUTPATIENT

Date received: _____	Lab no.: _____
Diagnosis/Clinical history (Specimen without relevant details may be rejected)	

NATURE OF SPECIMEN

(For Virus Culture: Must be in Virus transport medium)

1. ASPIRATE

- BAL 120
- BA 27
- ETA 24
- NPA 14
- Gastric 102
- Tracheal 21
- Other site

2. BILE (please specify)

Site: GB/PTBD/CBD
 /T-tube

3. BLOOD: (culture) 30
 (clotted) 31
 EDTA 106
 Heparin 121

4. C.S.F 32
 V.P. Shunt 109

5. FLUID:
 Pleural 38
 Peritoneal 42
 Peritoneal dial 122
 Joint 40
 Pericardial 39
 Other : _____

6. IV CATHETER

- Broviac/Hickman 91
- Other site: (please specify) _____

7. PUS

8. SPUTUM: 20

9. SWAB:

- Endocervical 88
- Ear Rt. 55 Lt. 56
- Eye Rt. 57 Lt. 58
- High/vaginal 85
- Nasal 17
- Nasopharyngeal 18
- Oral 16
- Perineal 80
- Peritoneal 128
- Placental 130
- Rectal 97
- Throat 19
- Umbilical 79
- Urethral 86
- Ulcer 131

- Superficial wound swab 63
- Other site swab
- (please specify) _____

10. SALIVA

- 133
- 11. Skin Scraping 78
- 12. Skin vesicle fluid 134

13. STOOL

14. STERILITY TEST:

- Milk 98
- Spore Strip 99

15. TISSUE

Site 69

16. URINE

- Bag 4
- CSU (indwelling) 3
- CSU (once) 142
- Early morning (AFB) 10
- Midstream 1
- Suprapubic 5
- Nephrostomy urine 12

17. OTHERS (please specify)

Sputum for AFB : Day 1 / Day 2

Chest x-ray abnormal

If CXR normal, test must be endorsed by ward
 specialist in-charge

(Specialist name / staff no.)

Ward ordering on CMS (Clinical Management System)

Clinical Management System [CMS] Last successful logon: 14-Aug-2018 09:40 (QMHCIT1)

File Clinical Investigation Enquiry Booking DT Report Doc./Print Other System Info Admin.

Logoff Close PSP Endoscopy Ix Request cPRad Med Cert Referrals OT Record Immunisation Form A/B HONDS AED Patient Elective OT Next Pat Time Out

GCR Ix Request

Unknown Details Alert

譚某某 TAM, XX HKID XXXXX

F 77y DOB: 25-Jan-1941 A173599(6) MED A2-10 Adm: 07-Oct-2003 HN06048935(7)

Request Date Admission / Current Dx Requested By Requesting Location Report To Copy To

14/08/2018 23234 GCRS GCRS REQUEST *DF* (DEFAULT)

Discipline Department Personal History Discipline M Microbiology Ix Search

LAB

- 1 Obsolete
- 3 DKCH Lab
- 5 Biochemistry (GH)
- A Histopathology
- B Blood Bank
- C Biochemistry
- H Haematology
- I Immunology
- M Microbiology**
- S Sendout Microbiology
- T T&I
- TX Toxicology RefLab
- Z TWH Lab

DM / ENDO

DM & Endo Services

RADIOLOGY

Angiography&Vas IR

Breast Imaging

CT Scan

Fluoro/Contrast XRay

General IR

MRI

Nuclear Medicine

PET-CT (PYN)

Plain X-Ray

Private (Non-HA)

Ultrasound

DH

Specimen

- Abscess Wall
- Aspirate
- Bile CBD
- Bile GB
- Bile PTBD
- Bile T-Tube
- Biopsy
- Blood (Culture)
- Blood Clot
- Bone
- Bone Marrow Aspirate
- CSF
- Cat. IV Brow/Hick
- Cat. IV Central line
- Cat. IV Peripheral line
- Cat. V-P Shunt
- Clotted blood
- Cord Blood
- Diarrhoea

Test

Blood (Culture), Bacterial culture

Site:

- Peripheral
- Broviac
- Hickman(red)
- Hickman(white)
- Central line

Selected Ix / Services

Save Cancel

Save and Print Reminder

Save and Print Label/Job Sheet

Pay Code: NE9

Add Ix

Specimen number / Staff ref.)

Name routine DB
Name Doyle DB

ESSAY

Initial level: Creatinine
Age & Frequency:
Date of Adm.:
Section: 3 months before last dose
 (Previous) (Present)
 1 month before
 3 months before
 Time:

Specimen date: 10/10/2018

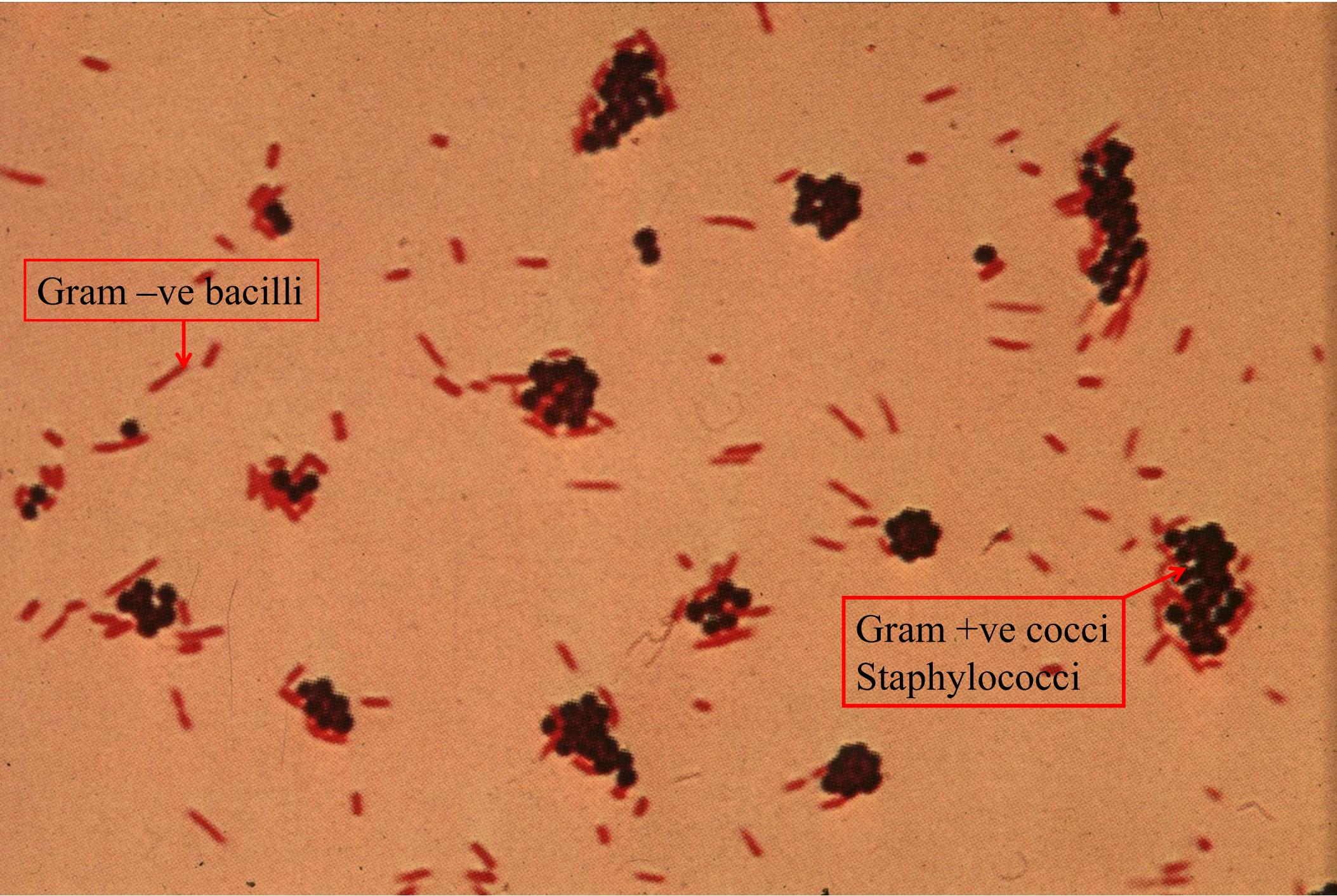
SIZE
<input type="checkbox"/> Large
<input type="checkbox"/> Medium
<input checked="" type="checkbox"/> Small

A close-up photograph showing a box of "Latex Examination Gloves" with the word "POWDERED" printed on it, and a pink plastic lid with handwritten markings "18 x 2" and "S" on it.

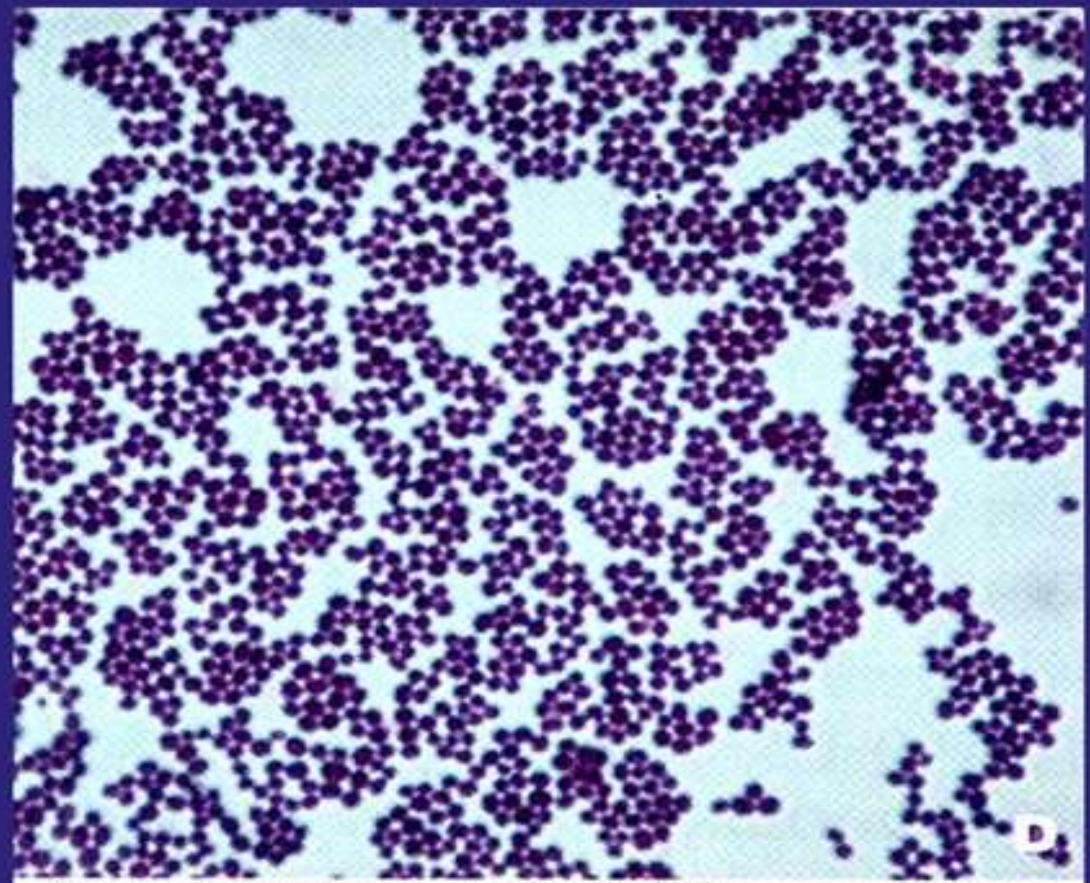
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M01589707









Gram stained smear of *Staphylococcus aureus*
on blood agar plate

Staphylococcus aureus

Antibiotic sensitivity test



NAME : XXXXX XXXX XXX
SEX/AGE : F/65
WARD/BED : K8N/24
HOSP.NO : 9456877
REQ. DATE : 1/4/2014

LAB.NO : 94156788
SPECIMEN : Blood
INVEST : C&ST

Culture : Pure growth of *E. coli*

Ampicillin
Cephalothin
Gentamicin
Co-trimoxazole

R
S
S
R

Bacterium is Resistant
Cannot use for treatment

Bacterium is Sensitive
to antibiotic.
Can use for treatment

Blood Culture

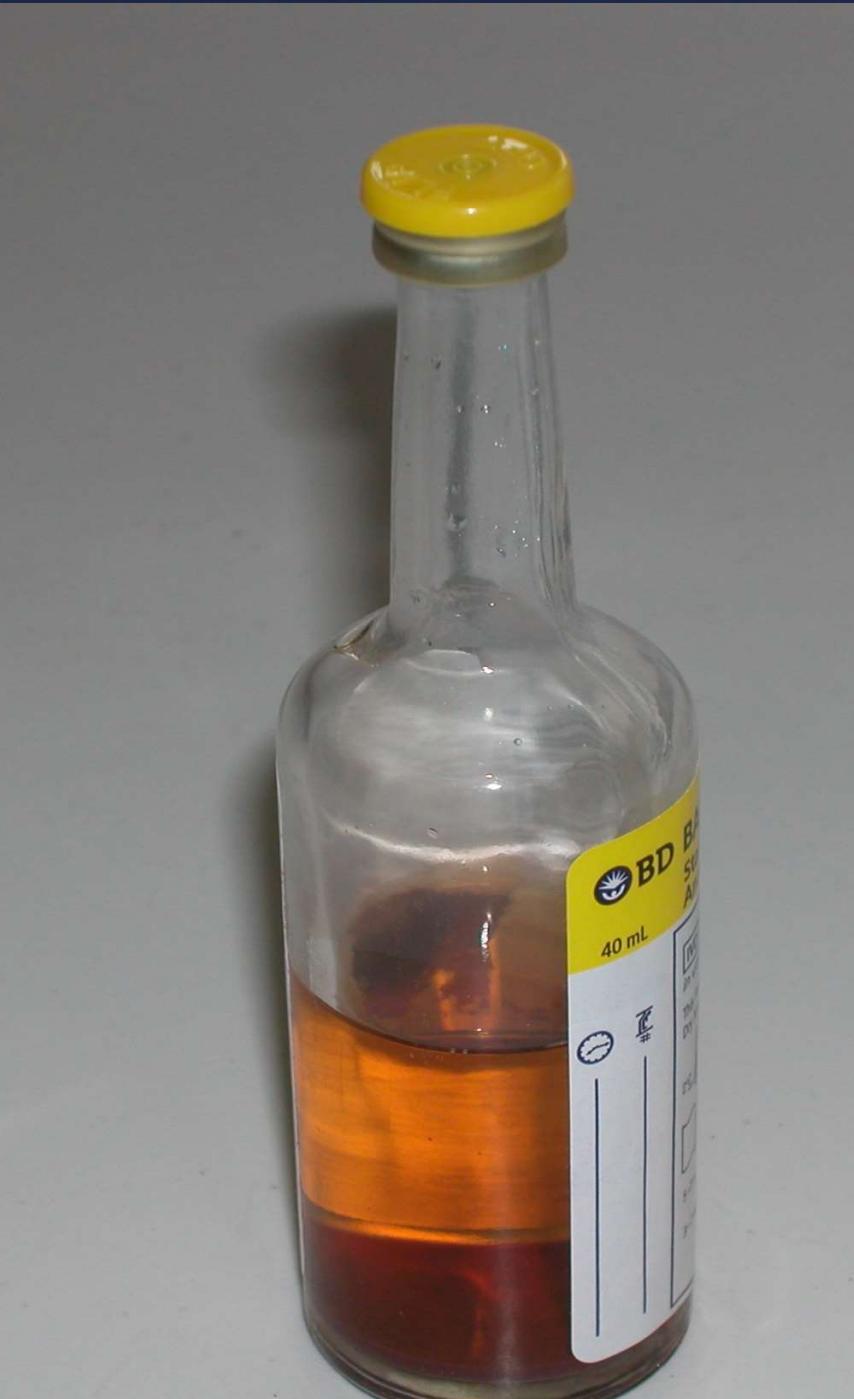
Common Indications: endocarditis, suspected sepsis

- Unexplained fever
- Sudden relative increase in pulse rate, temperature;
- change in consciousness, confusion
- onset of chills, prostration and hypotension;
- prolonged, mild, intermittent fever without/with a heart murmur.









Blood Culture

- At least 2 blood cultures should be taken; in febrile patients with a central venous catheter, one set should be taken through the catheter and another set by peripheral venous puncture
- Aerobic / Anaerobic micro-organisms;
- Aseptic technique;
- Adequate volume (~20 ml)
- Sample collection before antibiotic administration
- Take specimens to the laboratory promptly, or place them in an incubator at 37 C.

Blood Culture

- Broth (BACTEC) with resins to adsorb (inactivate) antibiotics.



RESINS

Cerebrospinal Fluid (CSF);
Sterile Body Fluid

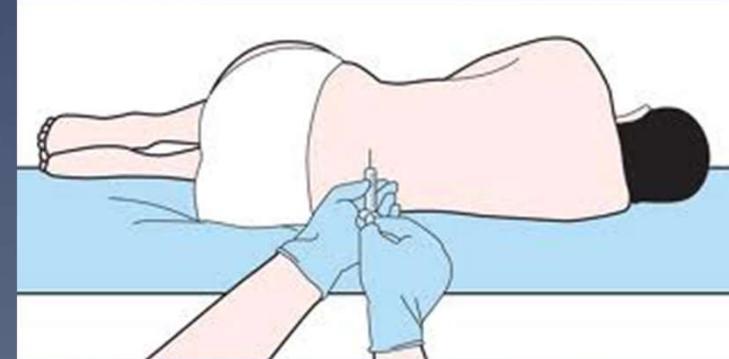
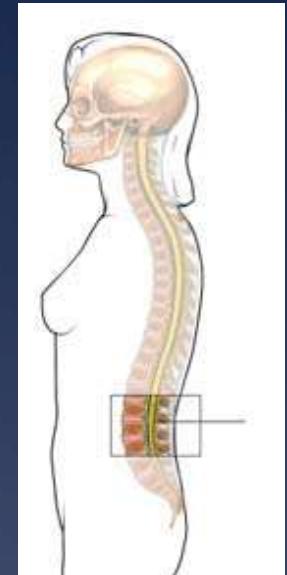
Cerebro-spinal fluid (CSF)

The representative specimen for meningitis.

Lumbar puncture should be undertaken whenever meningitis is suspected after a intracranial space occupying lesion is excluded.

Common indications :

- Fever, headache and meningeal irritation
- unexplained febrile illness in an irritable infant who is feeding poorly.



CSF

- Strict aseptic technique
- Use of new (Non-reused) sterile glass bottles (labeled: microbiology only) to prevent falsely positive gram smear results.
- Aliquots for sample bottles
- Immediate transport to the laboratory.

Sterile Body Fluid

- Including pleural, peritoneal (dialysis), pericardial and synovial (joint) fluid.
- Strict aseptic technique : per-cutaneous needle aspiration.
- Use of bottles containing citrate or heparin may be necessary if the specimen is prone to clotting (e.g bone marrow).

Sterile Bottle Suitable
for Culture only
WITH TRANSPORT MEDIUM

Sex/Age: _____

Ward/Bed No.: _____

Specimen taken: - 1 JUL 2003

Department of Microbiology, Q.M.

10 ml
8
6
4
2

Wound Swabs

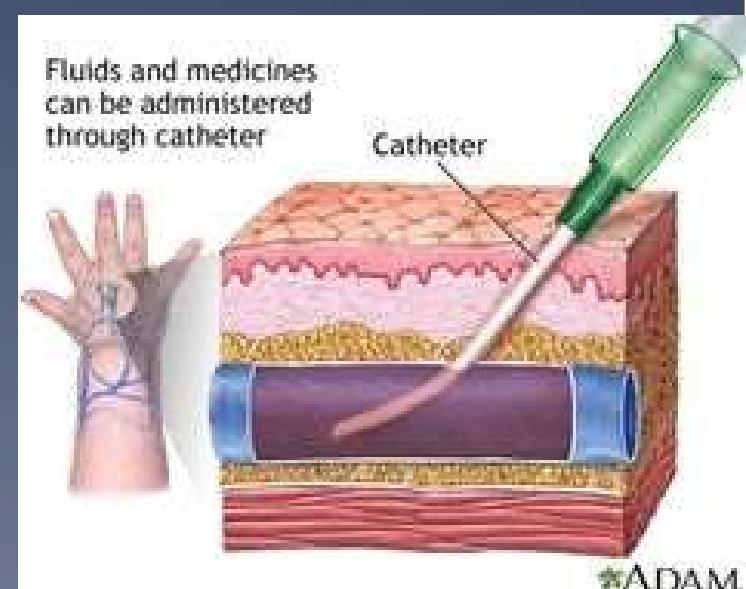
Swabs

- **Wound swabs, ear and eye swabs, drainage fluid, abscess (pus), sterile aspirate, tissue**
- **Tissue biopsy specimens should be put into sterile, wide mouth, screw-cap containers e.g. universal bottles.**

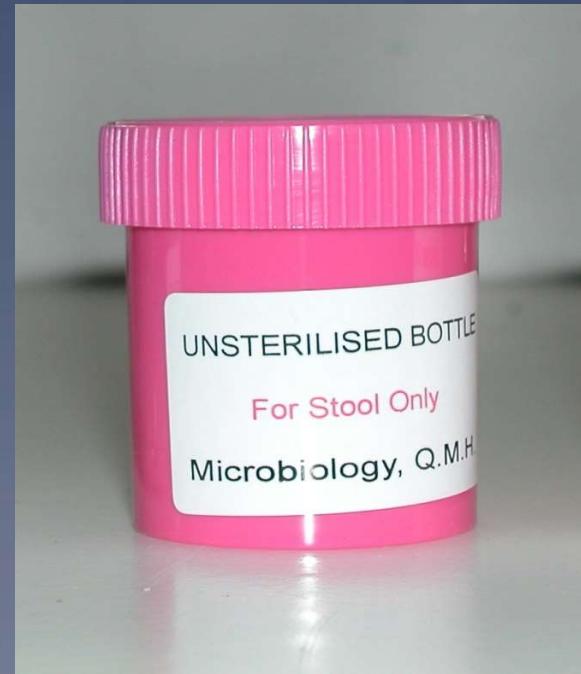


Intravascular catheter

- The exit site of the catheter is disinfected; after the area is dry, the catheter is removed and the distal 5 cm segment is aseptically cut off and directly dropped into a dry screw-cap container.
- The catheter segment is rolled over the blood agar plate 4 times.
- The presence of ≥ 15 colony forming units of a single organism after 48 hours of incubation is associated with an increased risk of catheter-related infections.



Faeces (Stool/Rectal swab)



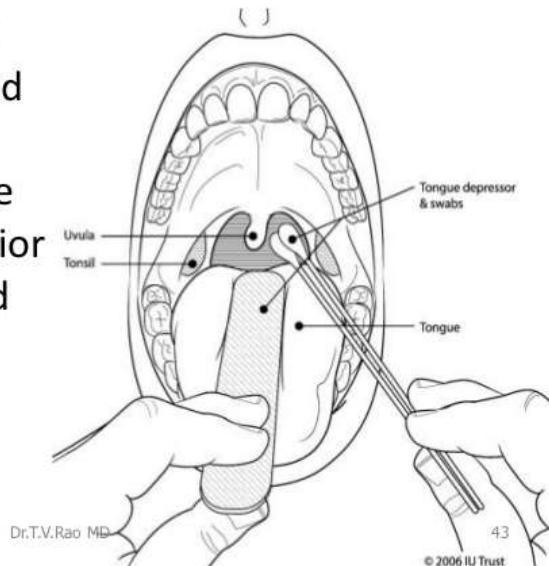
Upper Respiratory Tract

Throat Swab



Specimen collection in Throat Infections

- A plain cotton wool swab should be used to collect as much exudates as possible from tonsils, posterior pharyngeal wall and other area that is inflamed or bears exudates



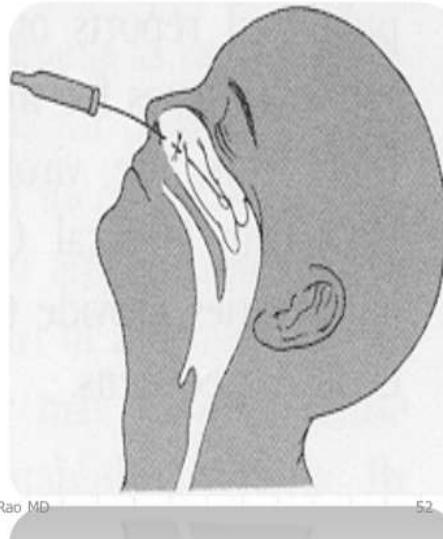
Nasopharyngeal Swab for *Bordetella pertussis* (*Whooping cough*)

Virus diagnosis



Nasopharyngeal swab

- A deep nasal swab generally yields the same information as throat swab.
- Nasal swabs are taken to detect healthy carriers than diagnose deep infection
- Deep nasal are taken to diagnose S.pyogenes and Diphtheria bacillus.



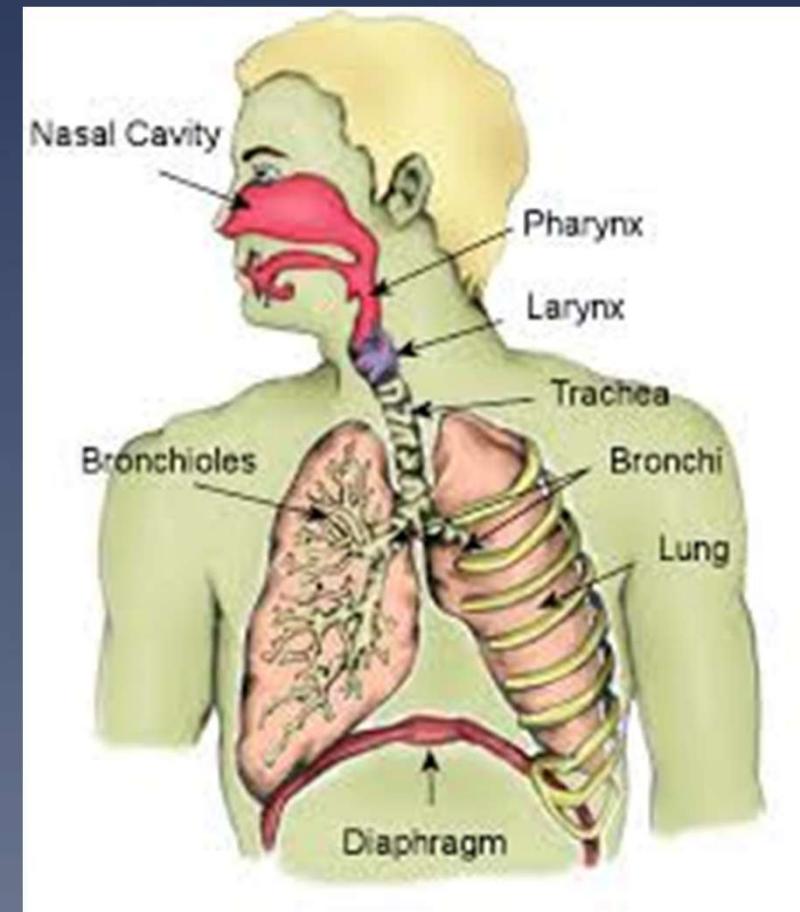
Dr.T.V.Rao MD

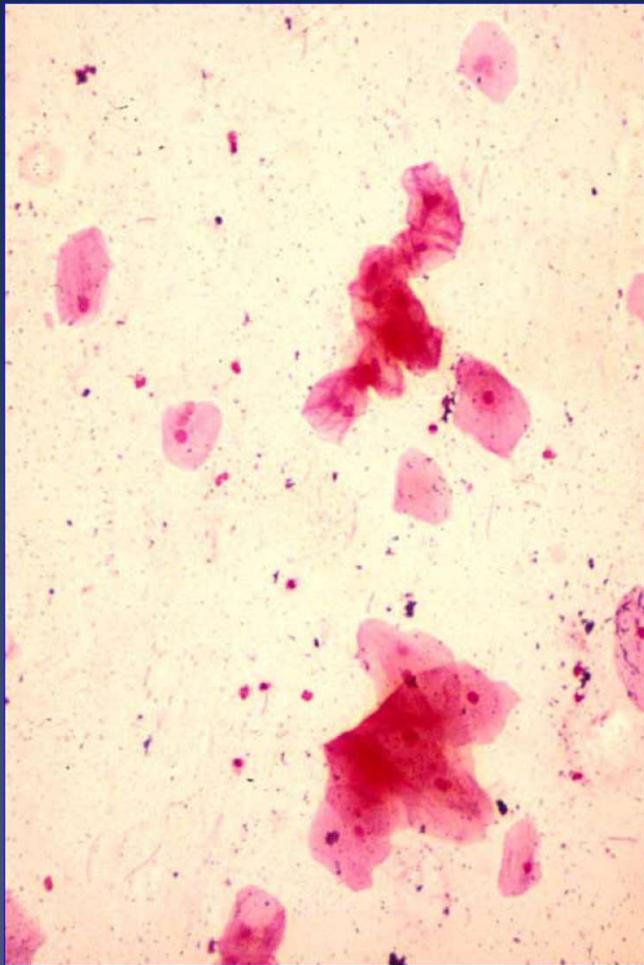
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Lower Respiratory Tract Infections

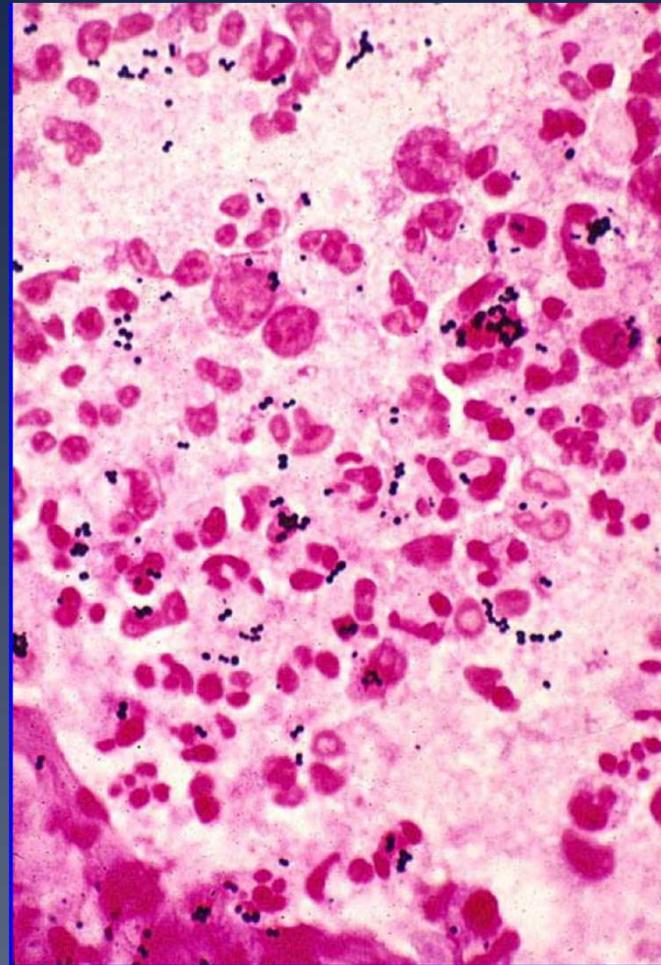
Expectorated sputum :

- Cough from lower respiratory tract. Not saliva.
- A high squamous epithelial cell content (from the buccal mucosa) together with a low white cell count on gram staining suggest oral-pharyngeal contamination).





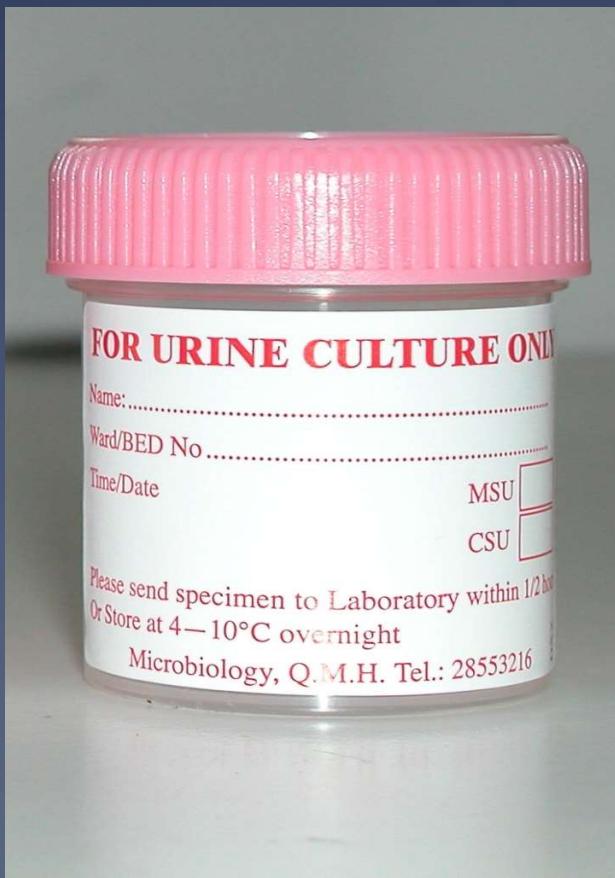
Squamous epithelial cells from the buccal mucosa



Mainly neutrophils (pus cells) indicative of specimen from the site of infection.

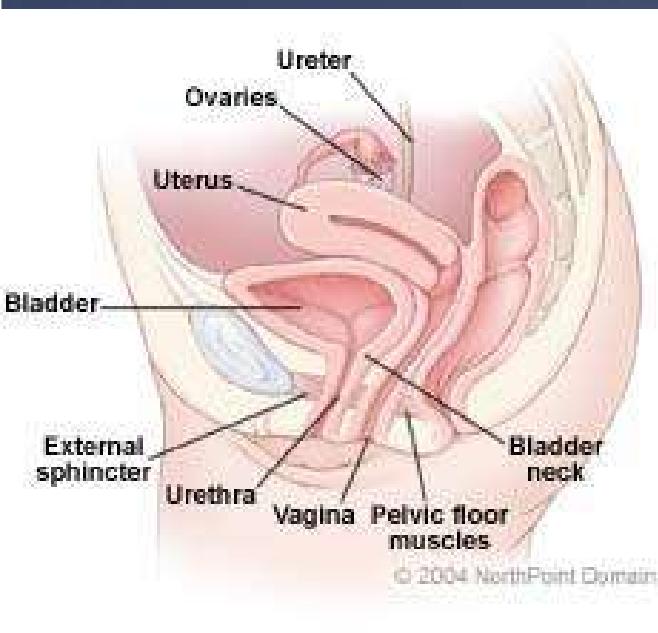
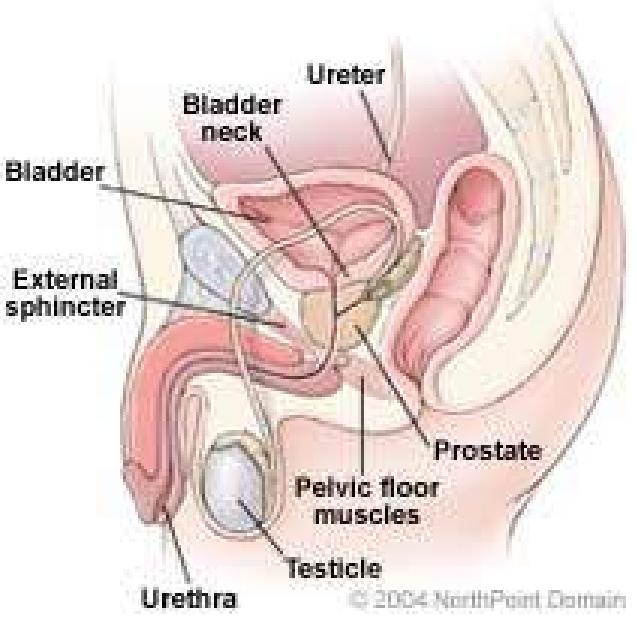
http://www.fletcherallen.org/upload/photos/6088Sputum_Gram_Stain.pdf

Urine



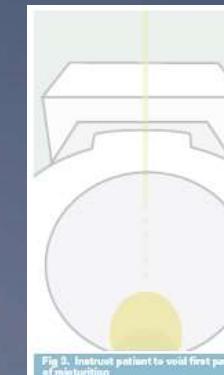
Urine

- Proper instruction regarding the collection of clean-voided (catch), mid-stream urine (MSU) is critical to ensure accuracy of results.
- All urine specimens must be transported to the laboratory for processing within an hour of collection unless refrigerated at 4°C.
- Quantitative culture



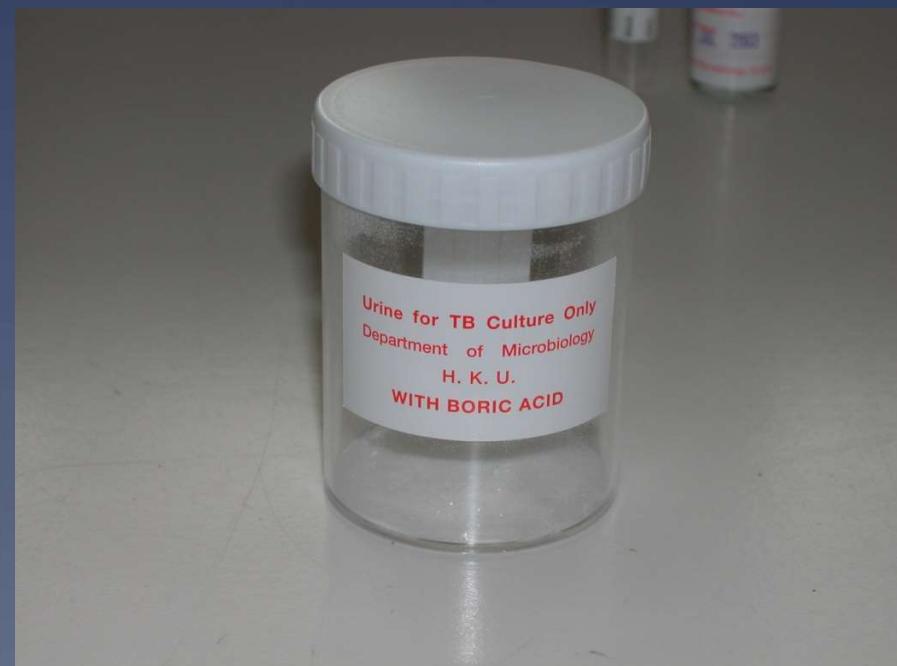
Mid-stream Urine

- Uncircumcised men → instructed to retract the foreskin before micturition. Women should be instructed to part the labia.
- Clean with sterile water.
- Strong urine flow is important in clearing bacteria from the urethral meatus. Obtaining the sample when bladder is full will result in the least contaminated sample.

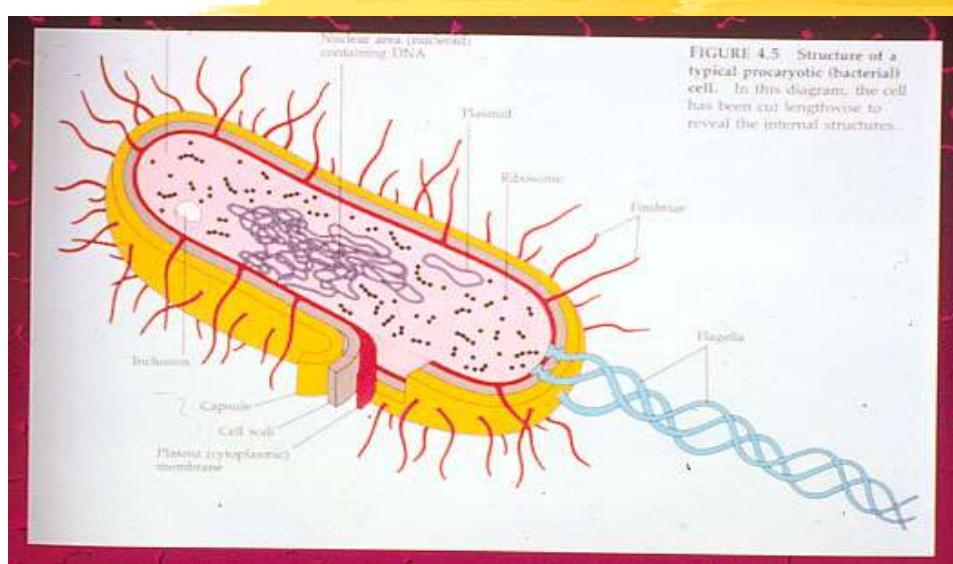


Urine for *Mycobacterium tuberculosis* (TB) culture

- Three consecutive early morning urine should be collected in patients with suspected renal tuberculosis

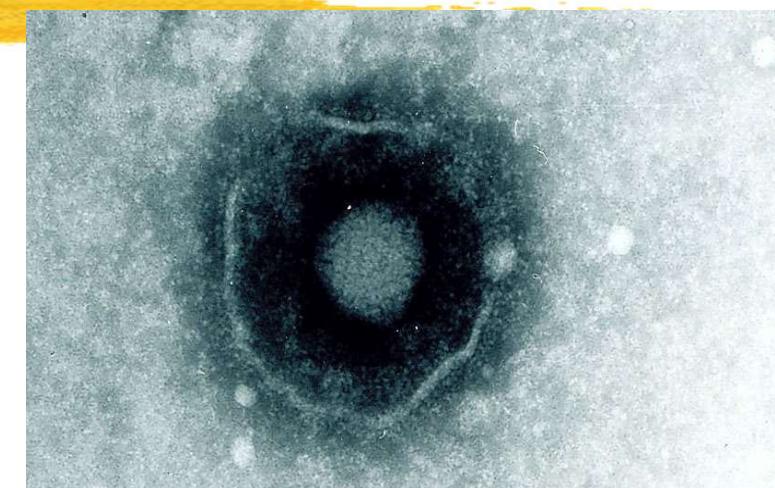


Bacteria

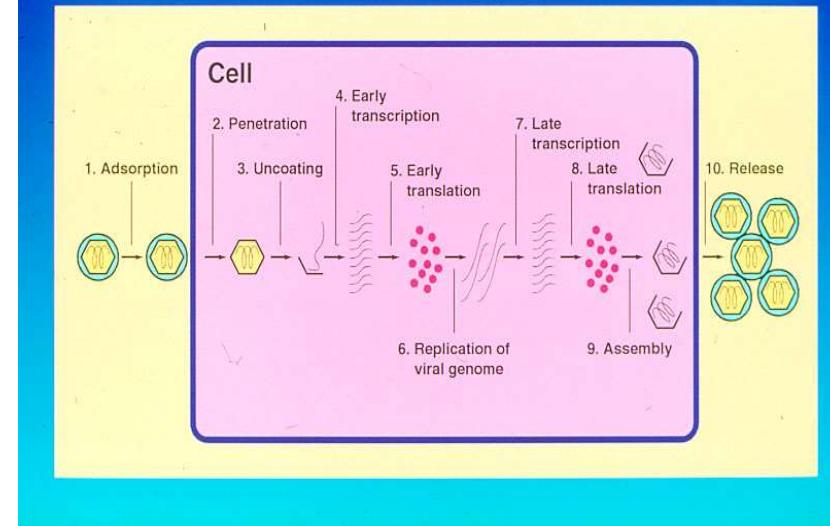


- ⌘ Human >20,000
- ⌘ Bacterium ~ 4000
- ⌘ Herpesvirus ~ 100
- ⌘ Hepatitis B v <10

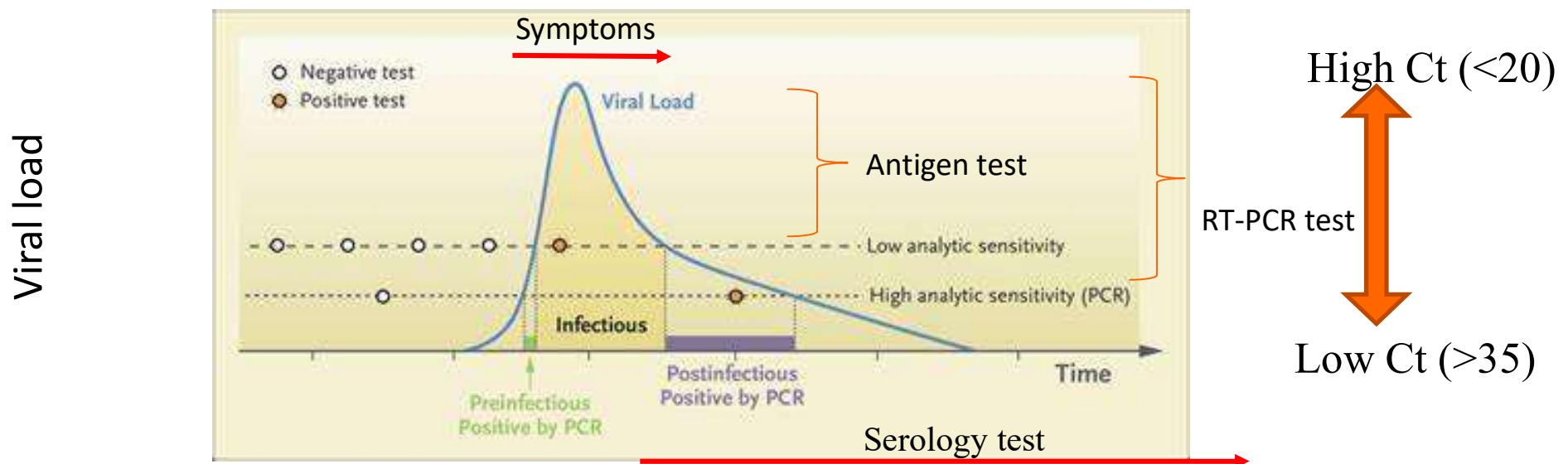
Virus



Mechanism of virus replication



Infectiousness and testing strategy in COVID-19



Modified from Mina et al NEJM Oct 2020

RT-PCR positive does not necessarily mean the person is infectious

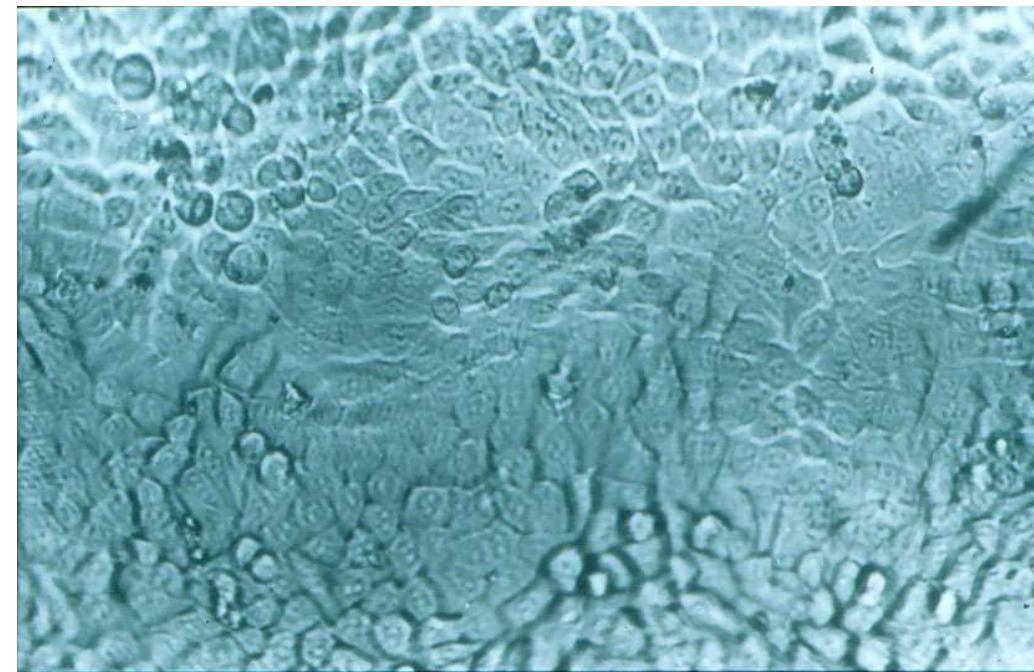
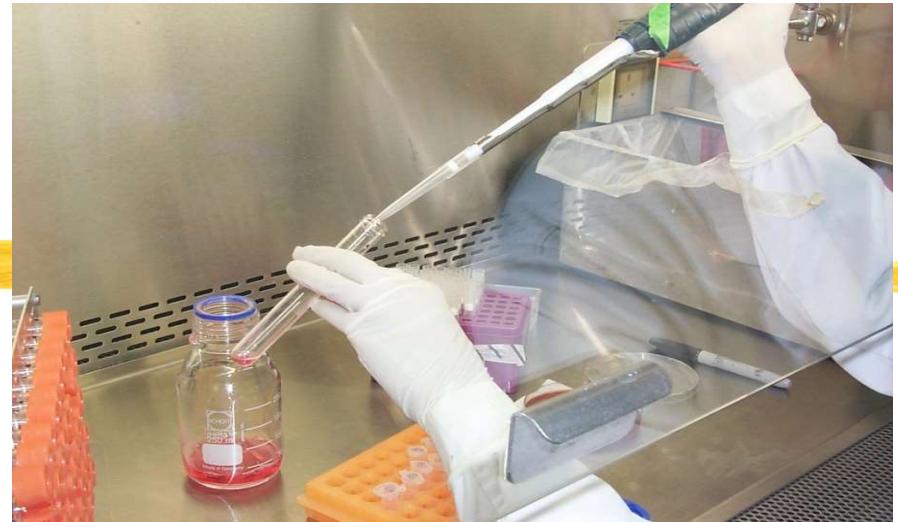
Use the test appropriate for the question you are trying to answer

Viral diagnosis: Detect virus

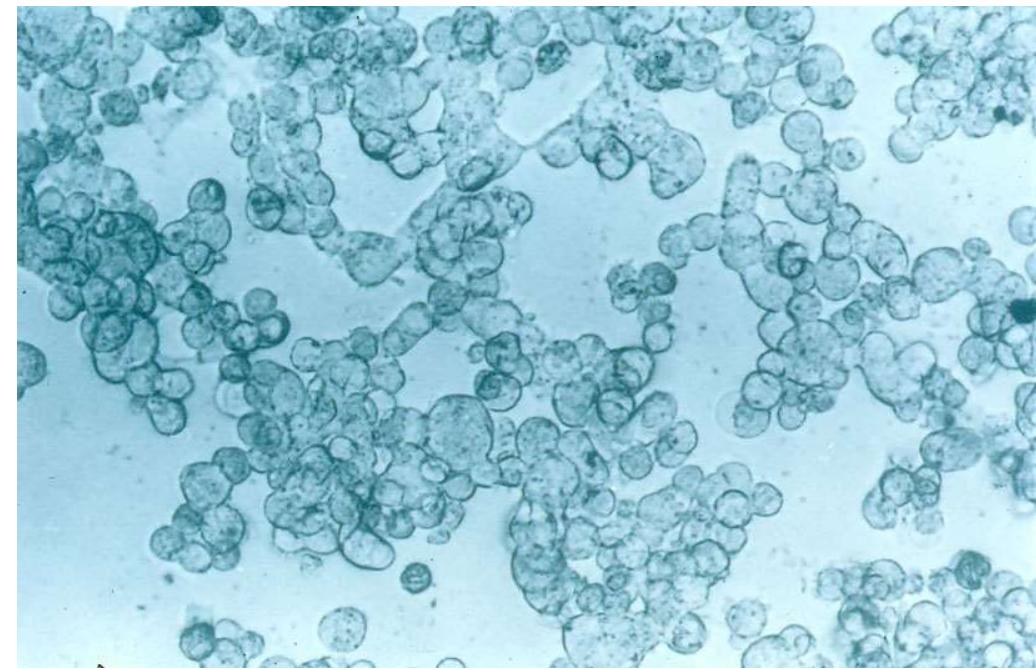


- ⌘ grow (virus culture in cell lines) - *days / weeks*
- ⌘ viral antigen detection - *hours*
- ⌘ viral nucleic acid detection (e.g. Polymerase chain reaction) (PCR) - *hours / days*

Cell culture

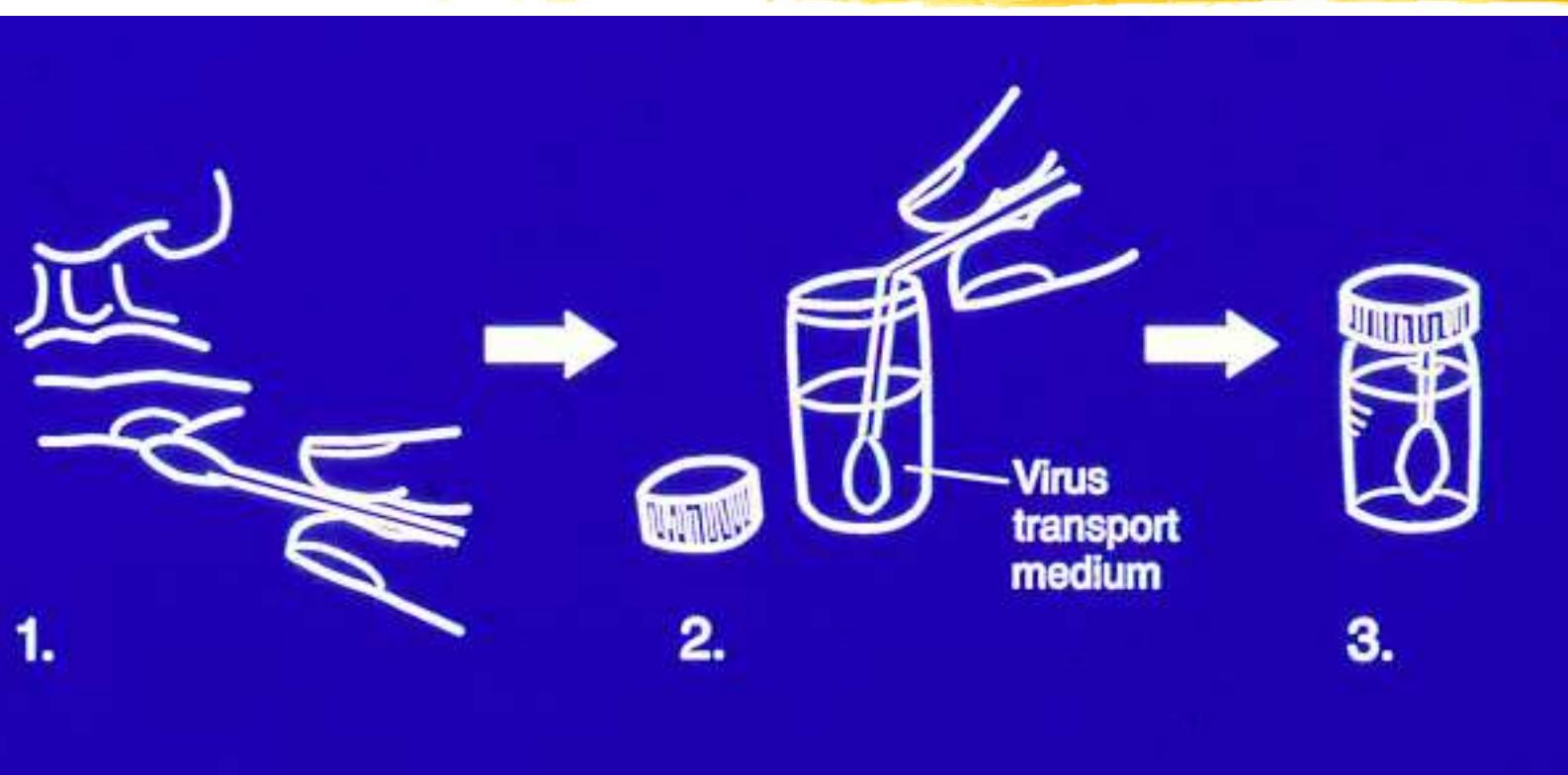


Normal cells



Virus cytopathic effect

Virus transport medium



Buffer – pH
pH indicator

Protein – maintain virus stability
Antibiotics – prevent bacterial overgrowth

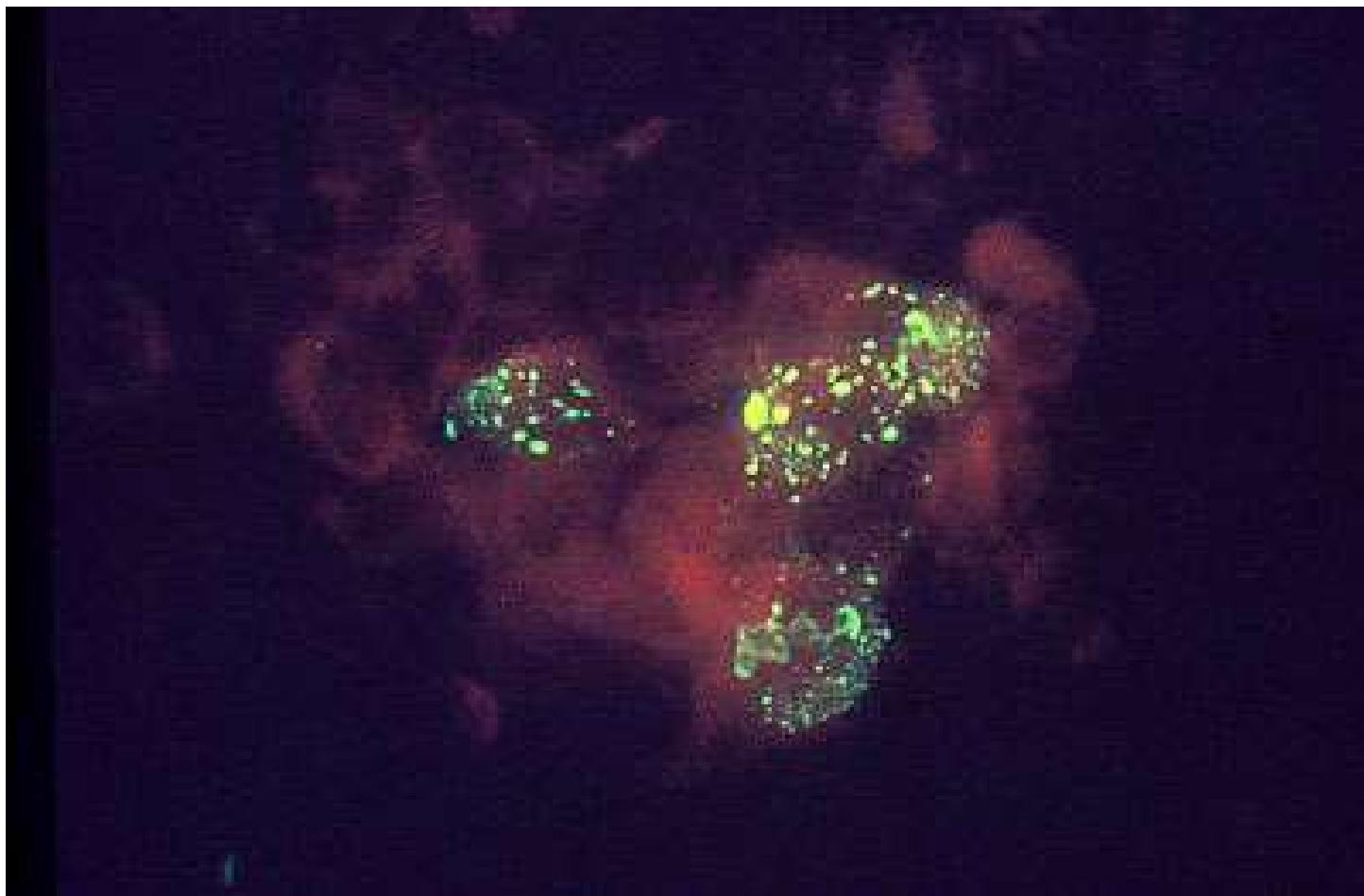
Virus
transport
medium
CANNOT be
used for
culture of
bacteria

Viral diagnosis: Detect virus



- ⌘ grow (virus culture in cell lines) - *days / weeks*
- ⌘ viral antigen detection - *hours*
- ⌘ viral nucleic acid detection (e.g. Polymerase chain reaction) (PCR) - *hours / days*

Viral antigen detection by immunofluorescence



Viral diagnosis: Detect virus



- ⌘ grow (virus culture in cell lines) - *days / weeks*
- ⌘ viral antigen detection - *hours*
- ⌘ viral nucleic acid detection (e.g. Polymerase chain reaction) (PCR) - *hours / days*

Serum



Haemolysis

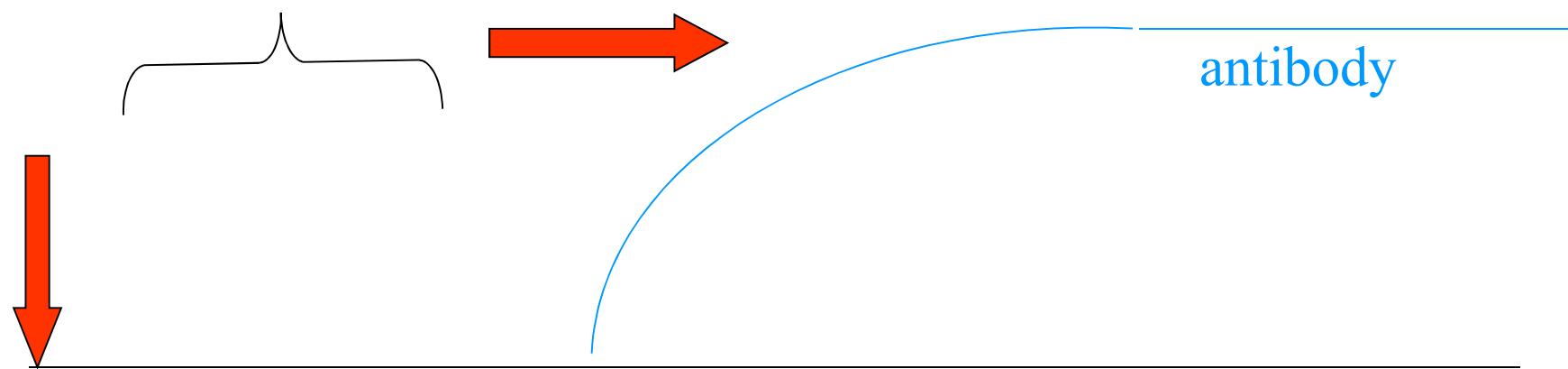
Blood

- Serum
- Plasma

Detect antibody response

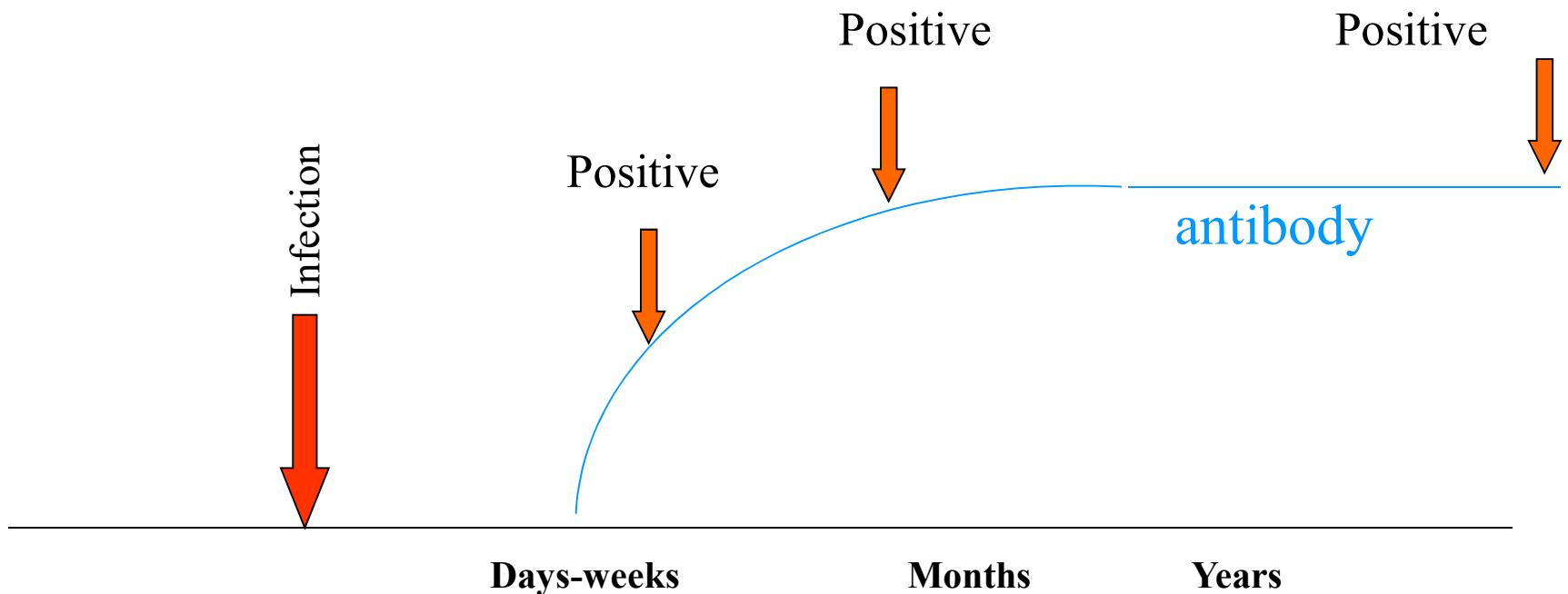


Infection Incubation Disease



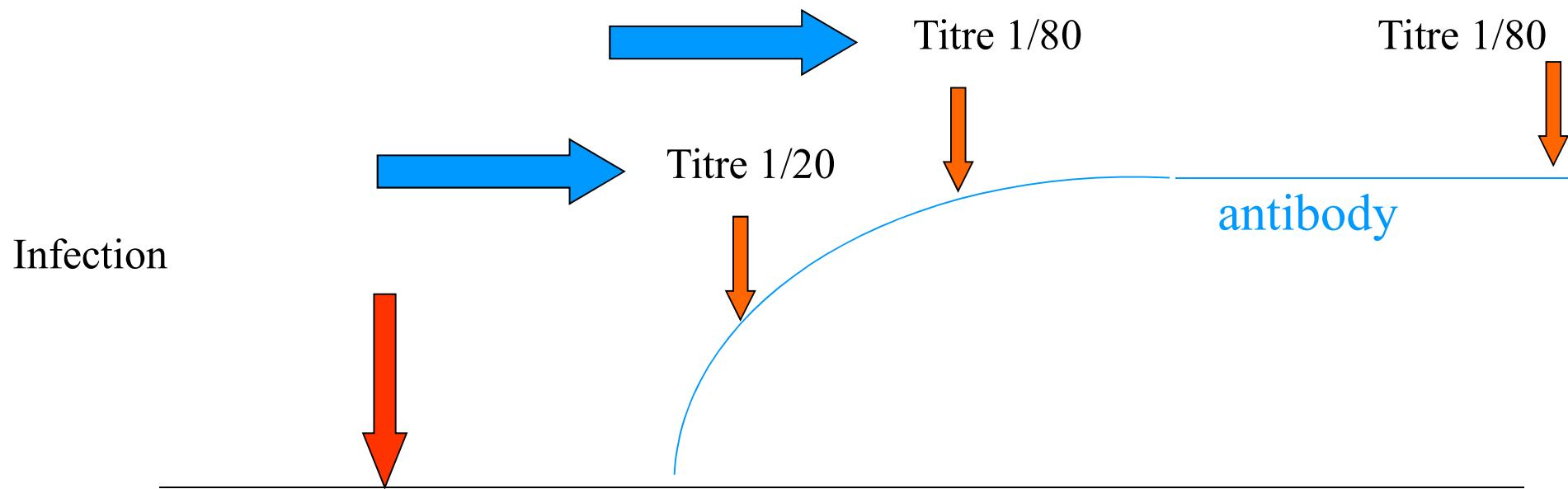
Detect antibody response

- ⌘ A single antibody test? What does it mean?
- ⌘ Infection at some time during lifetime.



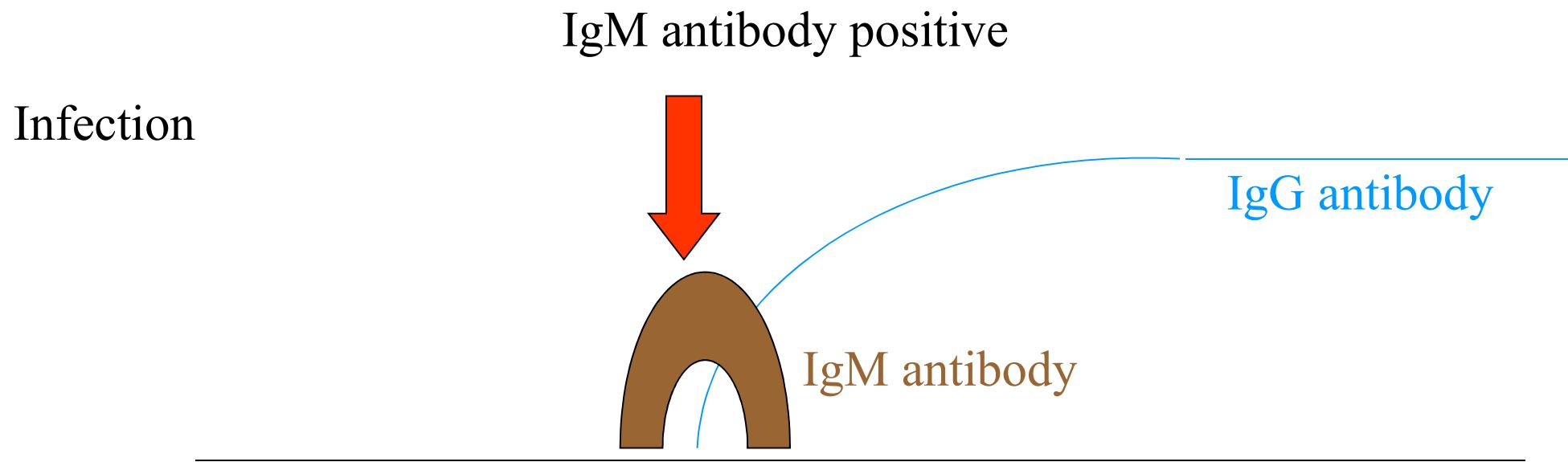
Detect antibody response

- Paired sera 10-14 days apart, tested for antibody titre (quantitation)
- Rising antibody titre: ≥ 4 fold increase



Detect antibody response

- ⌘ Antibody classes: IgM antibody / IgG antibody
- ⌘ Recent infection within last 1-3 months



Thank you



Any questions