Introduction to hierarchical (mixed-effects) models

FW 891

Christopher Cahill 25 September 2023



Purpose

- Introduce an important extention of all previous models
 - Will have several lectures on mixed effects models
- An example
 - both frequentist and Bayesian
 - Explain in words and algebra what mixed effects models are
- Pros and cons of mixed effects modeling
- Why these models likely deserve to be the default approach to many problems

The polyonomous model (misery)

- These models are known by many names:
 - Mixed effects models (fixed effects + random effects)
 - Multilevel models
 - Hierarchical models
 - Partial-pooling models
- Our example model might also be called:
 - Random-intercepts model

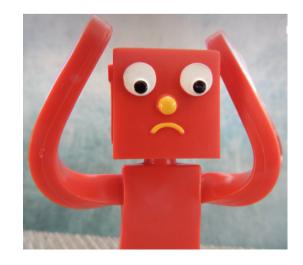
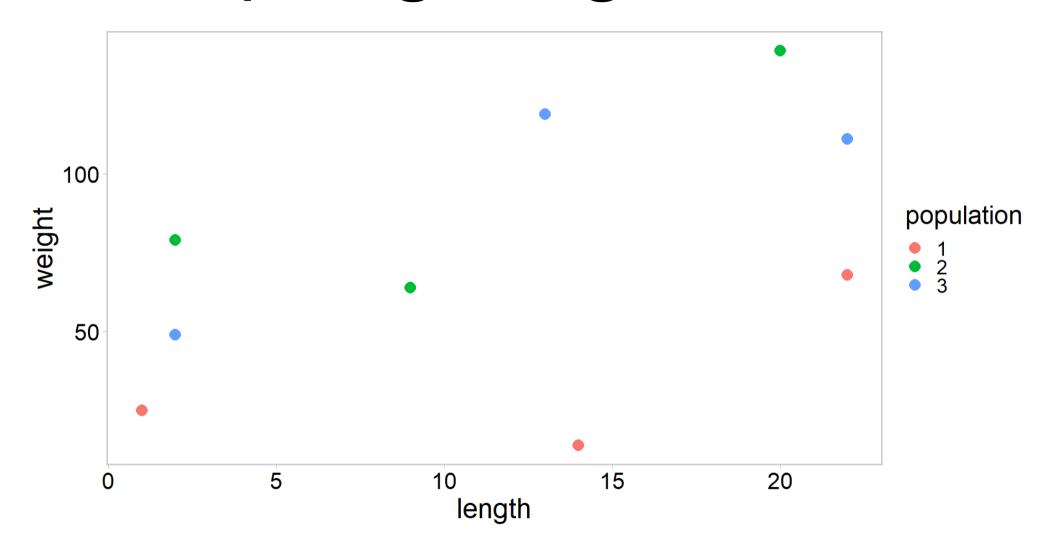




image credit

- Revisiting the ANCOVA example, but let's change the variable names
 - Plot the weight of emperor scorpions as a function of length

```
1 library(tidyverse)
2 library(ggqfc)
3 weight <- c(25, 14, 68, 79, 64, 139, 49, 119, 111) # obs
4 population <- factor(c(1, 1, 1, 2, 2, 2, 3, 3, 3)) # group
5 length <- c(1, 14, 22, 2, 9, 20, 2, 13, 22) # covariate
6 my_df <- data.frame(weight, population, length)
7
8 my_df %>%
9 ggplot(aes(length, weight, color = population)) +
10 geom_point(pch = 16, size = 3.5) + theme_qfc()
```



What does this plot suggest?

- Linear relationship between weight and length
 - Perhaps with a different intercept for each population?
 - Regression model should account for population differences
- The simplest way to account for population differences:

$$egin{aligned} ext{weight}_i &= lpha_{j(i)} + eta \cdot ext{ length }_i + arepsilon_i \ & arepsilon_i \sim ext{Normal}ig(0, \sigma^2ig) \end{aligned}$$

 Weight of scorpion i depends on length in a linear way, where each population j has a different intercept, and residuals come from a zero-mean normal distribution

Motivating random effects

- Independent intercepts for each population represents one assumption that we might make about the data
- Assumption 1: No Pooling: these are the only three populations we are interested in (no pooling; estimate each α_i separately)

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- Assumption 2: Complete Pooling: we think these actually represent one population (complete pooling; estimate one α across all three populations)
- Assumption 3: Partial Pooling: these populations merely represent a sample from a larger number of scorpion populations that we could have studied and we want our conclusions to generalize to this larger universe of statistical populations (hierarchical, partial pooling; model parameters are viewed as a sample from a population distribution)

Translating assumption 1 to math

No pooling:

$$egin{aligned} ext{weight}_i &= lpha_{j(i)} + eta \cdot ext{ length }_i + arepsilon_i \ & arepsilon_i \sim ext{Normal}ig(0, \sigma^2ig) \end{aligned}$$

Translating assumption 2 to math

Complete pooling:

$$egin{aligned} \operatorname{weight}_i &= lpha + eta \cdot \ \operatorname{length}_i + arepsilon_i \ &\sim \operatorname{Normal}ig(0,\sigma^2ig) \end{aligned}$$

• This is literally just a linear regression with one intercept and one slope

Translating assumption 3 to math

Hierarchical, partial pooling:

```
egin{aligned} \operatorname{weight}_i &= lpha_{j(i)} + eta \cdot \operatorname{length}_i + arepsilon_i \ &\sim \operatorname{Normal}ig(0,\sigma^2ig) \ &lpha_j \sim \operatorname{Normal}ig(\mu_lpha,\sigma_lpha^2ig). \end{aligned} This line makes lpha_j random!
```

• Third equation is the *only* difference between a model that treats α_j as a fixed effect (assumption 1) and one where α_j is a random effect (assumption 3)

Translating assumptions 1-3 to R code (Frequentist)

```
1 # ass. 1 - no pooling
2 fit_no <- lm(my_df$weight ~ my_df$population - 1 + my_df$length)
3
4 # ass. 2 - complete pooling:
5 fit_complete <- lm(my_df$weight ~ 1 + my_df$length)
6
7 # ass. 3 - partial pooling:
8 library(lme4)
9 fit_partial <- lmer(my_df$weight ~ 1 + my_df$length + (1|my_df$population))</pre>
```

 Don't worry about the computational details underlying how 1mer() is estimating the random effects

Compare the models (Frequentist)

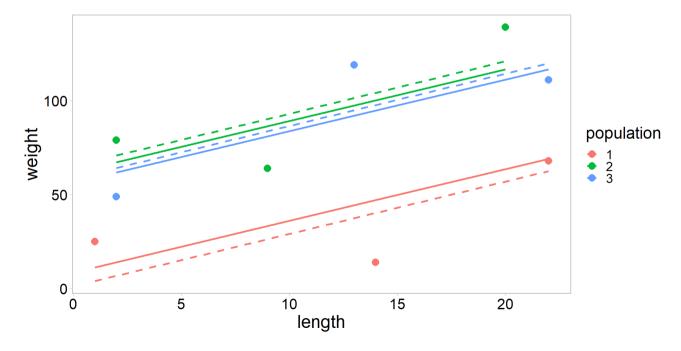
```
1 AIC(fit_no)
[1] 88.22643

1 AIC(fit_complete)
[1] 95.11368

1 AIC(fit_partial)
[1] 85.08788
```

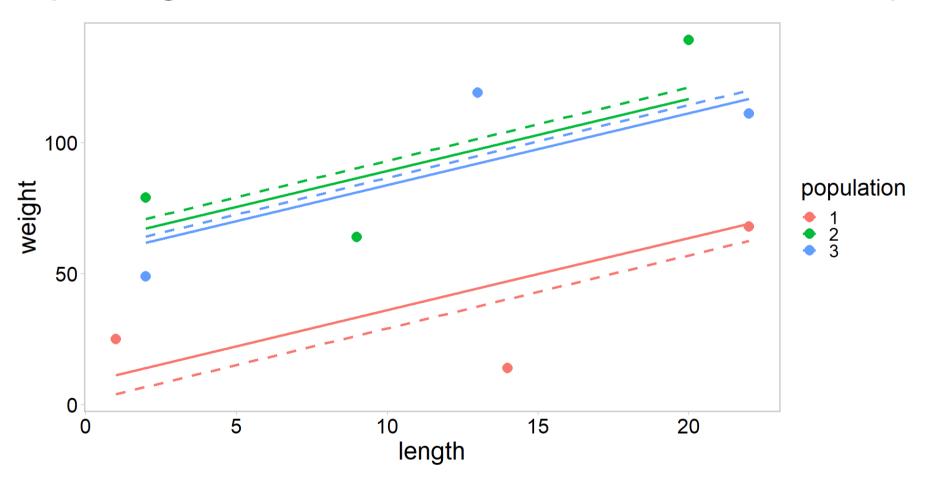
Plot no pooling vs. partial pooling

```
1 my_df$pred_no <- predict(fit_no, newdata = my_df)
2 my_df$pred_partial <- predict(fit_partial, newdata = my_df, level = 0)
3 my_df %>%
4 ggplot(aes(length, weight, color = population)) +
5 geom_point(pch = 16, size = 3.5) +
6 geom_line(data = my_df, aes(y = pred_no), size = 1, lty = 2) +
7 geom_line(data = my_df, aes(y = pred_partial), size = 1) +
8 theme_qfc()
```



What is going on here?

 Solid lines are from mixed-effects model with partial pooling, dashed lines are from ANCOVA with no pooling



Another example of shrinkage

county-level radon levels

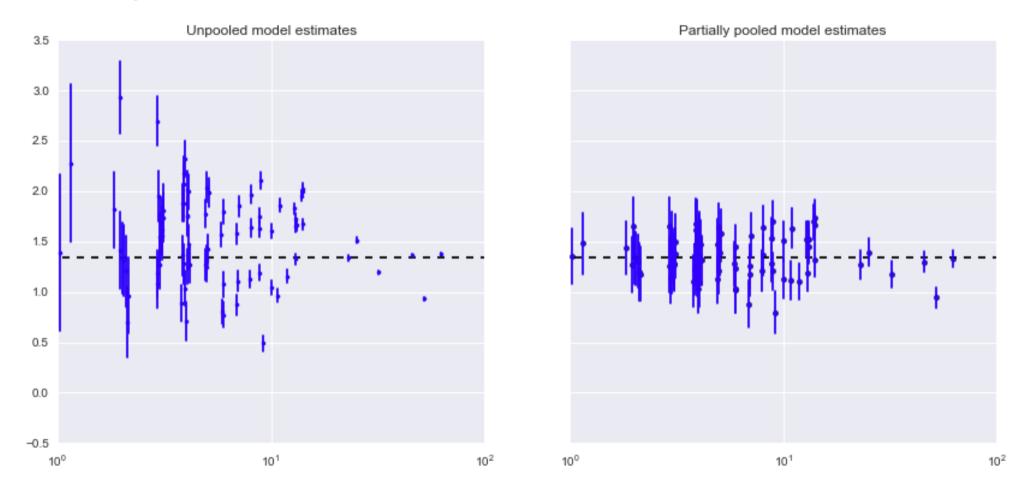


image credit

Let's build the scorpion model in Stan

Math for the random intercepts model

Hierarchical, partial pooling:

```
egin{aligned} \operatorname{weight}_i &= lpha_{j(i)} + eta \cdot \operatorname{length}_i + arepsilon_i \ &\sim \operatorname{Normal}ig(0,\sigma^2ig) \ &lpha_j \sim \operatorname{Normal}ig(\mu_lpha,\sigma_lpha^2ig). \end{aligned} This line makes lpha_j random!
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• Third equation is the *only* difference between a model that treats α_j as a fixed effect (assumption 1) and one where α_j is a random effect (assumption 3)

Stan code for the random intercepts model

```
1 data {
2   int<lower=0> n;
3   vector[n] weights;
4   array[n] int population;
5   vector[n] lengths;
6 }
7 parameters {
8   real mu_alpha;
9   vector[3] alpha_j;
10   real<lower=0> sd_alpha;
11   real b1;
12   real<lower=0> sd_obs;
13 }
14 // more code below
```

Stan code for the random intercepts model

```
1 //... see code on previous slide
 2 transformed parameters {
   vector[3] alpha pop;
    alpha pop = mu alpha + alpha j; // group int is global int + random effect
6 model {
    vector[n] w preds;
    mu alpha \sim normal(50, 15); // hyper prior
    sd alpha ~ normal(0, 100); // hyper prior
    alpha_j ~ normal(mu_alpha, sd_alpha); // hyper distribution
    b1 \sim normal(0, 10);
                                    // slope
    sd obs \sim normal(0, 25);
12
                                   // likelihood error term
13
    for(i in 1:n){
14
    w preds[i] = alpha pop[population[i]] + b1*lengths[i];
15
16
    17 }
```

Compiling the Stan model

```
library(cmdstanr)
 2 mod <- cmdstan model("src/random ints.stan")</pre>
   stan data <- list(
   n = length(weight),
    weights = weight,
   population = population,
    lengths = length
 8
 9
   inits <- function() {</pre>
11
   list(
12 mu alpha = 50,
13 \operatorname{sd}_{\operatorname{alpha}} = 30,
14 alpha_j = rep(0, 3),
15 b1 = 10,
16 \qquad \text{sd obs} = 10
17 )
18 }
```

Running that hawg 😇 🐂 🐽







```
1 fit <- mod$sample(</pre>
   data = stan data,
   init = inits,
  seed = 1,
   chains = 4,
   iter warmup = 1000,
   iter sampling = 1000,
   parallel chains = 4,
   refresh = 0,
  adapt delta = 0.9999, # look here
11 step size = 1e-3 # look here
12 )
```

Running MCMC with 4 parallel chains...

```
Chain 4 finished in 0.5 seconds.
Chain 2 finished in 0.6 seconds.
Chain 1 finished in 0.7 seconds.
Chain 3 finished in 0.8 seconds.
All 4 chains finished successfully.
Mean chain execution time: 0.6 seconds.
Total execution time: 0.9 seconds.
```

Check the diagnostics (quick and dirty)

fit\$cmdstan diagnose() Processing csv files: C:/Users/Chris/AppData/Local/Temp/RtmpOil1G0/random ints-202309040845-1-282d3e.csv, C:/Users/Chris/AppData/Local/Temp/RtmpOil1G0/random ints-202309040845-2-282d3e.csv, C:/Users/Chris/AppData/Local/Temp/RtmpOil1G0/random ints-202309040845-3-282d3e.csv, C:/Users/Chris/AppData/Local/Temp/RtmpOil1G0/random ints-202309040845-4-282d3e.csv Checking sampler transitions treedepth. Treedepth satisfactory for all transitions. Checking sampler transitions for divergences. No divergent transitions found. Checking E-BFMI - sampler transitions HMC potential energy. E-BFMI satisfactory. Effective sample size satisfactory. Split R-hat values satisfactory all parameters. Processing complete, no problems detected.

Pluck out the posterior summaries and compare to lmer()

```
1 fit$summary("alpha pop")
                                        # Bavesian
# A tibble: 3 \times 10
 variable mean median
                                        q95
                                            rhat ess bulk ess tail
                          sd
                              mad
                                    q5
 <chr>
      <n11m>
                                                           <num>
1 alpha_pop[1] 9.75 8.79 22.9 21.3 -25.3
                                                   1284.
                                       49.3 1.00
                                                          1097.
2 alpha pop[2] 61.5 62.3 20.3 18.7 28.3 92.4 1.00 1627. 1153.
3 alpha pop[3] 56.5
                        21.5 19.8 22.2 90.4
                 56.8
                                            1.00
                                                   1930.
                                                           1829.
 1 coef(fit partial) $ `my df$population`[,1]  # frequentist
    8.650009 61.667157 56.276072
```

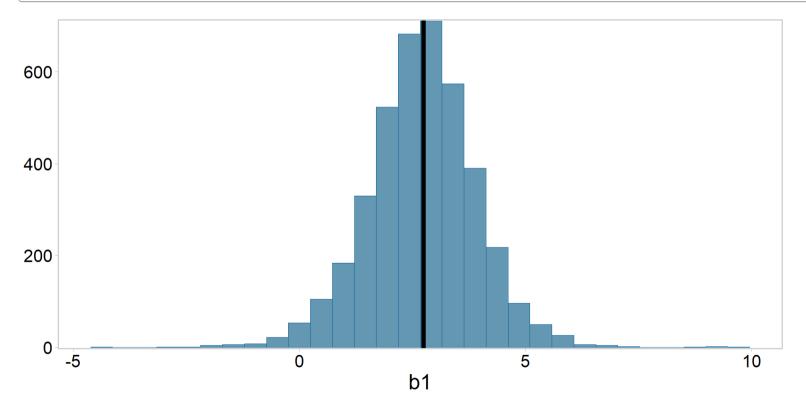
- Pretty similar estimates (small sample size)
- You have now fitted a Bayesian hierarchical model

Pluck out the posterior summaries and compare to lmer()

```
1 fit$summary("b1")
                       # Bavesian
# A tibble: 1 \times 10
 variable mean median
                                       rhat ess bulk ess tail
                     sd
                         mad
                               q5 q95
 <num>
                                                     <num>
         2.71
               2.73 1.19 1.07 0.722 4.58 1.00
                                                     1321.
                                             1400.
1 b1
 1 fixef(fit partial)[2] # frequentist
my df$length
   2.744955
```

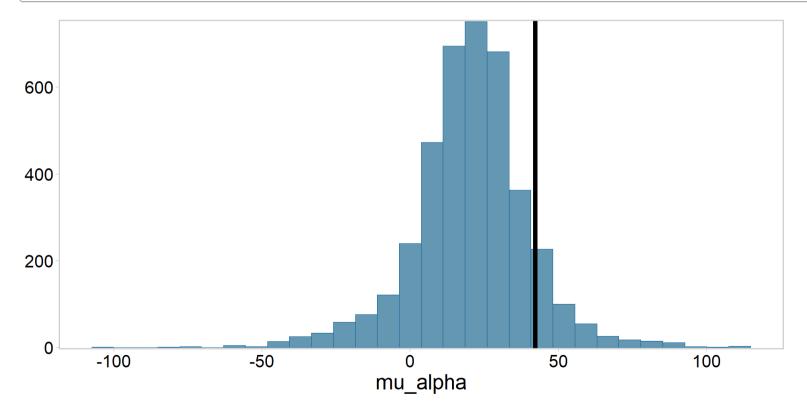
Posterior for β_1 vs. frequentist estimate

```
1 library(bayesplot)
2 posterior <- fit$draws(format = "df")
3 color_scheme_set("blue")
4 p <- mcmc_hist(posterior, par = "b1")
5 p + geom_vline(xintercept = fixef(fit_partial)[2], lwd = 2) + theme_qfc()</pre>
```



Posterior for μ_{α} vs. frequentist estimate

```
1 library(bayesplot)
2 posterior <- fit$draws(format = "df")
3 color_scheme_set("blue")
4 p <- mcmc_hist(posterior, par = "mu_alpha")
5 p + geom_vline(xintercept = fixef(fit_partial)[1], lwd = 2) + theme_qfc()</pre>
```



This was meant to be an 'easy' example for a reason

- Probably too few groups (usually want 5-10)
 - Obviously we can do it, but likely means our priors are more influential
- I had to fiddle with this a bit to get it to behave
 - Also low sample size within groups
- This likely explains at least some of the discrepancy among methods

What is a random effect (words)

- Two or more effects or parameters that "belong together" in some way
 - e.g., originating from some common distribution
 - sometimes called "latent variables," may not be observable
- Bayesians put prior probabilities on all unknown quantities, MCMC
- Frequentists remove random effects from the model via numerical integration
- Exchangeability as a key assumption
 - Ordering of the random effects doesn't matter

The distribution of random effects

- Called the hyperdistribution
- The parameters of the random effects distribution are called hyperparameters
 - The priors of the hypperparameters are called hyperpriors
- No rule against more than one level (i.e., we could have hyper-hyperparameters etc.)
- Key point is to note the hierarchy of effects
- Typically random effects are continuous in ecology
 - however see occupancy models or binomial mixture models

Benefits of hierarchical modeling

- Sounds dope, also scares a lot of people
- de facto standard for modern ecological analyses
- Helps you draw (statistical) universe-level inferences if you have a good experimental design
- Borrowing information via a hyperprior
 - Robin-Hooding (see Punt et al. 2011)
- Helps safeguard against overfitting

Some reasons we might use mixed effects models

- To adjust for repeat sampling
- To adjust estimates for imbalanced sampling
- To study variation among groups
- To partition variation among groups
- Borrow information

Some reasons we might use mixed effects models

- Avoiding pseudoreplication
- Improving scope of inference
- To avoid averaging
- Improve understanding
 - Many common classes of models can be reformulated as hierarchical models
- McElreath (2023) argues mixed effects methods should be the default for many modeling problems

Drawbacks of hierarchical modeling?



Why treat any parameter as fixed?

- Treating a factor with very few levels as random results in very imprecise estimates of the hyperparameter
 - Rarely treat factors with fewer than 5-10 levels as random (however see Gelman 2005)
- Assumption of exchangeability may not hold, particularly if groups differ in some systematic way

Hierarchical modeling as a philosophical middle ground

• Berliner (1966) offered a definition of hierarchical models based on sub-models and inference from a joint probability distribution:

[data|process, parameters][process|parameters][parameters]

- observation model component
- process model component
- assumptions about parameters
- A key point:

Hierarchical modeling as a philosophical middle ground

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- observation model component
- process model component
- assumptions about parameters
- A key point:
 - Hierarchical models admit observation error, conditioned on some (underlying) ecological process of interest and your prior assumptions
- Whenever possible, prefer a hiearchical model based on an explicit ecological process

Summary and outlook

- Many names for the same general type of model
- Shrinkage as a key concept, which is related to the phrase "partial pooling"
- The difference between fixed effects and random effects, at least for this simple example
- A powerful framework for decomposing complex problems into manageable subcomponents/models
- Many important extentions to this simple model

References

- Berliner, L. M. 1996, Hierarchical Bayesian time series models", Maximum Entropy and Bayesian Methods, 15-22.
- Gelman and Hill 2007. Data analysis using regression and multilevel models
- Royle and Dorazio 2008. Hierarchical modeling and inference in ecology.
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- McElreath 2023. Statistical Rethinking. Second Edition, Chapters 2 and 9.
- Punt et al. 2011. Among-stock comparisons for improving stock assessments of data-poor stocks: the "Robin Hood" approach. ICES Journal of Marine Sci.