

2020 SISBID Graphical Models Lab

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Load packages

```
library("igraph")
library("XMRF")
library("huge")
library("glasso")
library("WGCNA")
library("glmnet")
```

Read the Sachs et al data: Flow cytometry proteomics in single cells, $p = 11$ proteins measured in $n = 6466$ cells.

```
sachscov <- as.matrix(read.table("sachscov.txt"))
sachscor <- cov2cor(sachscov)
sachsdatt <- as.matrix(read.table("sachs.data.txt"))
ps <- c("praf", "pmek", "plcg", "PIP2", "PIP3", "P44", "pakts", "PKA", "PKC", "P38", "pjnk")
colnames(sachsdatt) <- ps
p <- ncol(sachsdatt)
n <- nrow(sachsdatt)
dim(sachsdatt)
```

```
## [1] 7466 11
```

```
dim(sachscov)
```

```
## [1] 11 11
```

```
head(sachsdatt)
```

```
##      praf      pmek      plcg      PIP2      PIP3      P44      pakts
## [1,] -97.67193 -132.18100 -46.03364 -132.8207  31.765040 -20.021190 -64.16721
## [2,] -88.17193 -128.88100 -42.55364 -134.3207 -18.904960  -8.031193 -48.66721
## [3,] -64.67193 -101.28100 -40.25364 -140.9207 -14.034960 -11.731190 -48.66721
## [4,] -51.07193  -62.58096 -31.75364 -137.6207 -25.744960 -20.801190 -69.36721
## [5,] -90.37193 -125.58100 -49.66364 -141.3907  -2.234962  -5.531193 -35.06721
## [6,] -105.27190 -141.63100 -37.25364 -129.0207 -16.134960 -14.731190 -55.46721
##      PKA      PKC      P38      pjnk
## [1,] -211.75860 -13.34166  -90.1145 -33.267500
## [2,] -273.75860 -26.97166 -118.5145 -11.767500
## [3,] -222.75860 -18.94166 -103.1145 -53.767500
## [4,]  -97.75859 -16.64166 -106.4145 -50.167500
## [5,] -320.75860 -25.68166 -109.3145   8.032497
## [6,]  -15.75859 -16.64166  -85.9145 -15.467500
```

```
load("UnsupL_SISBID_2020.Rdata")
```

Coexpression network

simple thresholding of correlations, at a cutoff chosen to give similar number of edges to partial correlation methods

- a randomly chosen threshold

```
tau <- 0.1
A1 <- abs(sachscor) > tau
diag(A1) <- 0
sum(A1)/2
```

```
## [1] 43
```

testing for nonzero correlations

- testing for nonzero correlation, using Fisher Z-transform

```
fisherzs <- atanh(sachscor)
fisherps <- 2*pnorm(abs(fisherzs), 0, 1/sqrt(n-3), lower.tail=FALSE)
A2 <- fisherps < (0.01/(p*(p-1)/2))
diag(A2) <- 0
sum(A2)/2
```

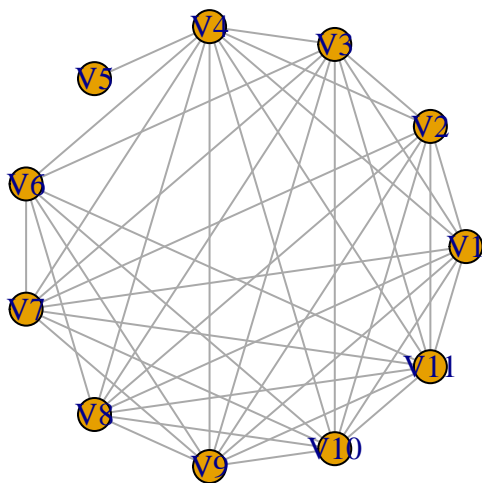
```
## [1] 45
```

plot the three networks

```
g1 <- graph.adjacency(A1, mode="undirected")
g2 <- graph.adjacency(A2, mode="undirected")
g0 <- g2
```

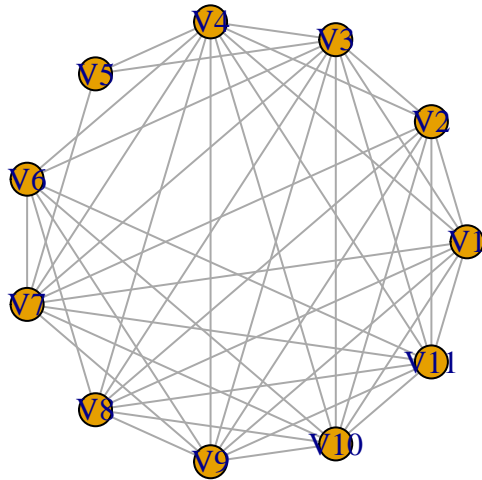
```
plot(g1,layout=layout.circle(g1), main='simple thresholding of correlations')
```

simple thresholding of correlations



```
plot(g2,layout=layout.circle(g2), main='testing for nonzero correlations')
```

testing for nonzero correlations



Partial correlation networks (under gaussian graphical models assumption)

inverse covariance matrix

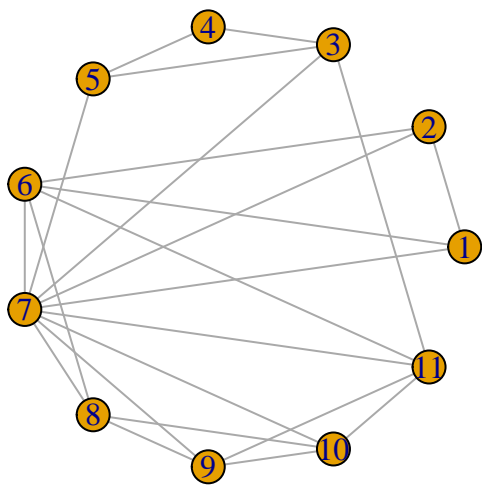
```
invcov <- abs(round(solve(sachscor),3))
invcor <- cov2cor(invcov)
A1 <- 1*(invcor > 0.05)
diag(A1) <- 0
sum(A1)/2
```

```
## [1] 23
```

```
g1 <- graph.adjacency(A1, mode="undirected")
```

```
plot(g1,layout=layout.circle(g1),main = "Partial correlation networks")
```

Partial correlation networks



Graphical lasso

Calculate lambda, based on formula in the slides (the third method)

```
alpha = 0.01
num = qt(p=alpha/(2*(p^2)),df=n-2, lower.tail=F)
lambda = num / sqrt(n-2 + num)
```

Apply glasso

```
glasso.est <- glasso(s=sachscor,rho=lambda*4.2,approx=FALSE,
                    penalize.diagonal=FALSE)
A2 <- abs(glasso.est$wi) > 1E-16
diag(A2) <- 0
g2 <- graph.adjacency(A2, mode="undirected")
```

Neighborhood selection

```
ns.est <- glasso(s=sachscor, rho=lambda, approx=TRUE, penalize.diagonal=FALSE)
A3 <- abs(ns.est$wi) > 1E-16; diag(A3) <- 0
g3 <- graph.adjacency(A3, mode="undirected")
```

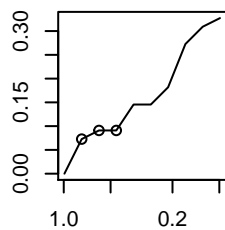
Neighborhood selection estimate with huge (Stability selection for the value of λ)

```
X <- data.matrix(scale(sachsdatt))
neth = huge(X,method="mb")
```

```
## Conducting Meinshausen & Buhlmann graph estimation (mb)....done
```

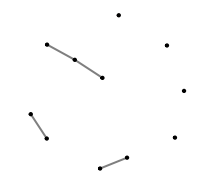
```
plot(neth)
```

arsity vs. Regularizati

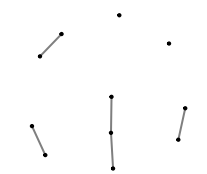


Regularization Parameter

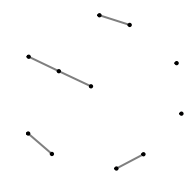
lambda = 0.767



lambda = 0.594



lambda = 0.46

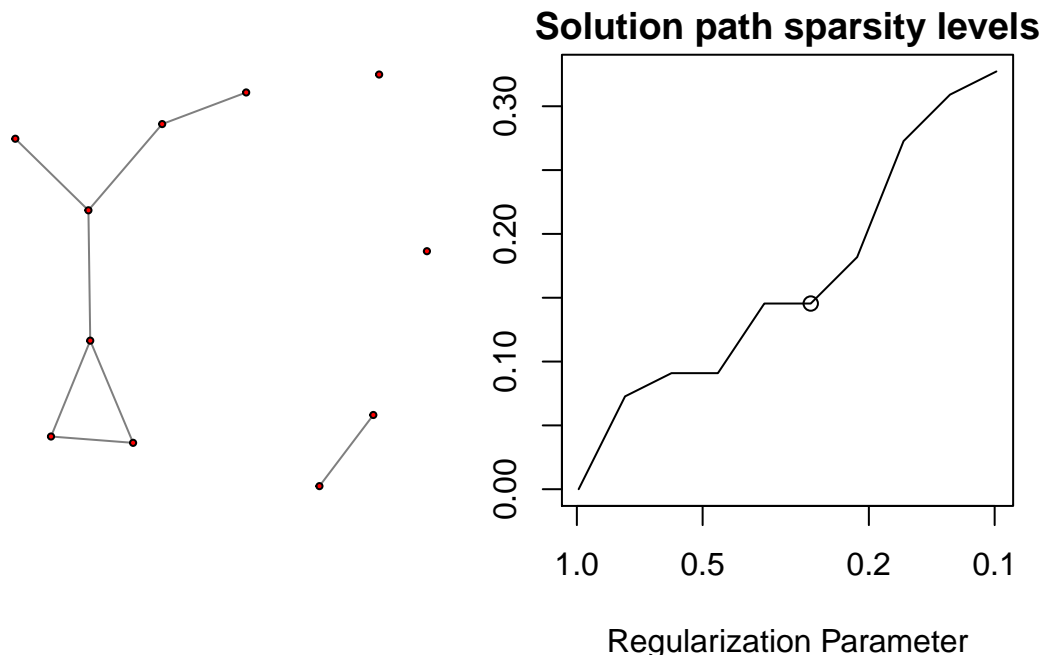


```
## stability selection with huge
```

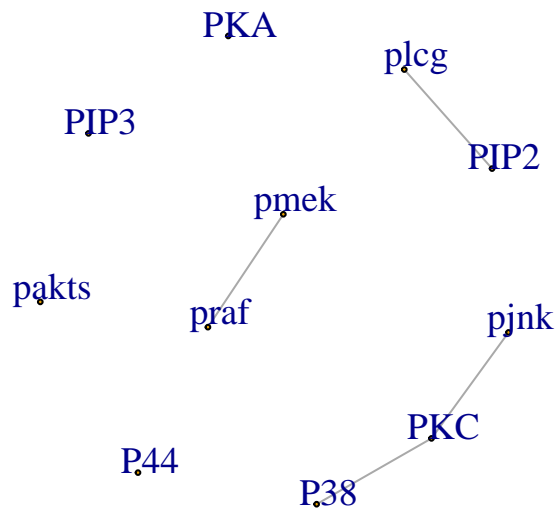
```
net.s <- huge.select(neth, criterion="stars")
```

```
## Conducting Subsampling....in progress:5% Conducting Subsampling....in progress:10% Conducting Subsam
```

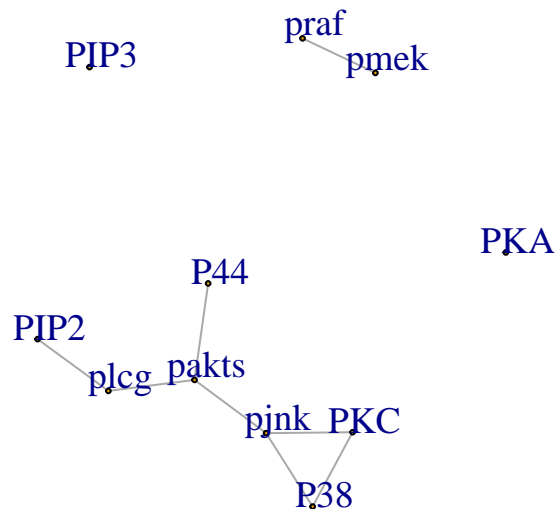
```
plot(net.s)
```



```
#larger lambda
mat = neth$path[[2]]
neti = as.undirected(graph_from_adjacency_matrix(mat))
plot(neti,vertex.label=colnames(X),vertex.size=2,vertex.label.cex=1.2,vertex.label.dist=1,layout=layout,
```



```
#smaller lambda
mat = neth$path[[5]]
neti = as.undirected(graph_from_adjacency_matrix(mat))
plot(neti,vertex.label=colnames(X),vertex.size=2,vertex.label.cex=1.2,vertex.label.dist=1,layout=layout,
```



Nonparanormal Models: rank-based correlation

```

scor <- cor(sachsdat,method='spearman')
scor <- 2*sin(scor*pi/6)
nnp.est <- glasso(s=scor, rho=lambda, approx=FALSE, penalize.diagonal=FALSE)
A4 <- abs(nnp.est$wi) > 1E-16
diag(A4) <- 0
g4 <- graph.adjacency(A4, mode="undirected")

```

Nonparanormal Models – alternative estimation

```

nnp.cor <- huge.nnp(x=sachsdat, nnp.func="skeptical", nnp.thresh=NULL, verbose=FALSE)
nnp.est <- glasso(s=nnp.cor, rho=lambda, penalize.diagonal=FALSE)
A5 <- abs(nnp.est$wi) > 1E-16
diag(A5) <- 0
g5 <- graph.adjacency(A5, mode="undirected")

```

binary network estimation

```

sachsbin <- 1*(sachsdat > 0) + -1*(sachsdat <= 0)
head(sachsbin)

```

```

##      praf pmek plc PIP2 PIP3 P44 pakts PKA PKC P38 pjnk
## [1,]  -1  -1  -1  -1   1  -1  -1  -1  -1  -1  -1
## [2,]  -1  -1  -1  -1  -1  -1  -1  -1  -1  -1  -1
## [3,]  -1  -1  -1  -1  -1  -1  -1  -1  -1  -1  -1
## [4,]  -1  -1  -1  -1  -1  -1  -1  -1  -1  -1  -1
## [5,]  -1  -1  -1  -1  -1  -1  -1  -1  -1  -1   1
## [6,]  -1  -1  -1  -1  -1  -1  -1  -1  -1  -1  -1

```

```

bin.est <- matrix(0,p,p)
## estimate the neighborhood for each node
for(j in 1:p){
  ## this is the same method used in neighborhood selection, the only difference is 'family'
  nbr <- glmnet(x=sachsbin[,-j], y=sachsbin[,j], family='binomial', lambda=lambda)
  bin.est[j,-j] <- 1*(abs(as(nbr$beta,"matrix")) > 0) #store the estimates in jth row of matrix
}

```

```

}
A6 <- bin.est
diag(A6) <- 0
sum(A6)/2

```

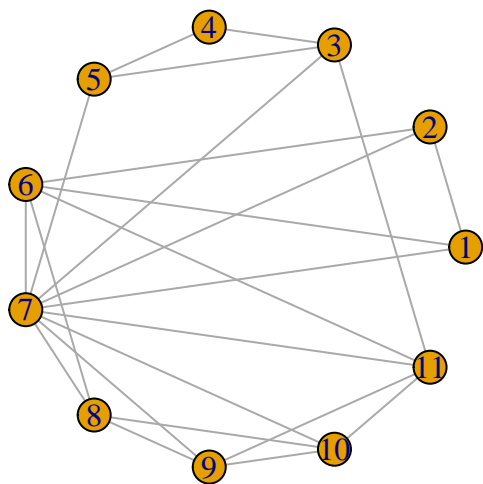
```
## [1] 20
```

```
g6 <- graph.adjacency(A6, mode="undirected")
```

plot the networks

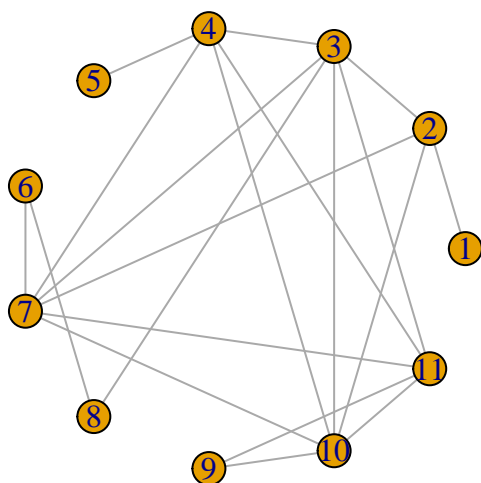
```
plot(g1,layout=layout.circle(g1), main='Partial correlation networks')
```

Partial correlation networks



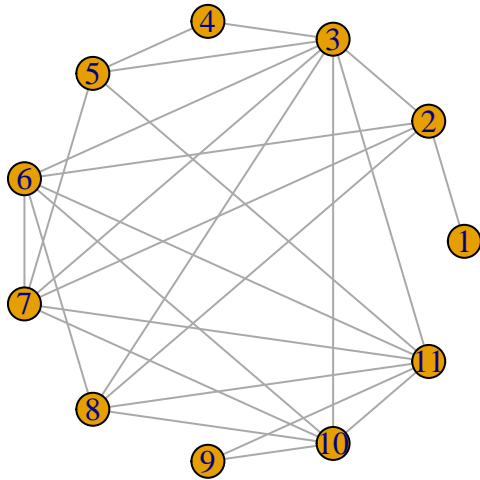
```
plot(g2,layout=layout.circle(g2), main='Glasso')
```

Glasso



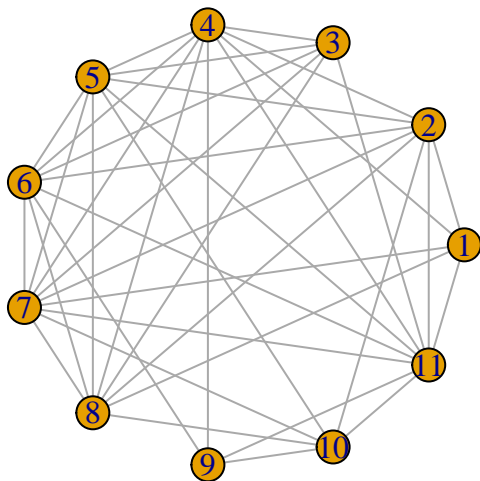
```
plot(g3,layout=layout.circle(g3), main='Neighborhood selection')
```

Neighborhood selection



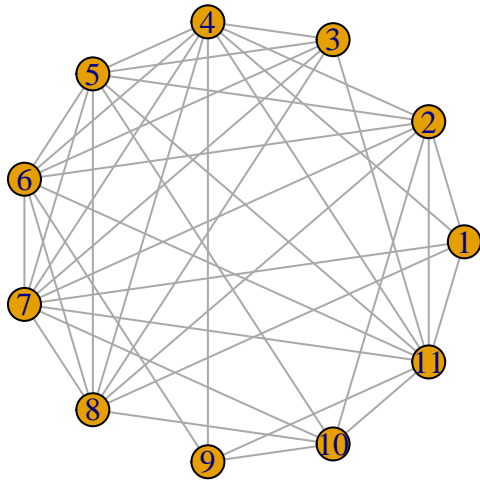
```
plot(g4,layout=layout.circle(g4), main='nonparanormal')
```

nonparanormal



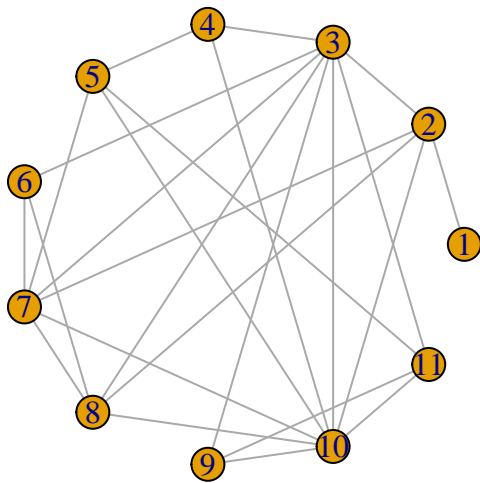
```
plot(g5,layout=layout.circle(g5), main='nonparanormal - v2')
```


nonparanormal – v2



```
plot(g6,layout=layout.circle(g6), main='Binary')
```

Binary



Poisson Graphical Models via XMRF package

Data set - Author Data. This data set consists of word counts from chapters written by four British authors.

```
TrueAuth = as.factor(rownames(author))
X2 = log(1 + author[,1:69]) #log transform data

lam = lambdaMax(X2)*sqrt(log(ncol(X2))/nrow(X2))*0.02
net = XMRF(t(X2),method="LPGM",lambda.path=lam,N=1,th=.001)

## LPGM ::: Conducting sampling ... in progress: 100 %
## LPGM Completed.

mat = net$network[[1]]
neti = as.undirected(graph_from_adjacency_matrix(mat))
```

```
plot(neti, vertex.label=colnames(X2), vertex.size=2, vertex.label.cex=1.2, vertex.label.dist=1, layout=layou
```



```
plot(neti, vertex.label=colnames(X2), vertex.size=2, vertex.label.cex=1.2, vertex.label.dist=1, layout=layou
```

