import packages

import numpy as np import pandas as pd import sys import os import time as tm import pickle from functools import partial import scipy.stats as st from scipy.stats import wasserstein distance import scipy.stats import copy from sklearn.model selection import KFold import pandas as pd import multiprocessing import matplotlib as mpl import matplotlib.pyplot as plt import scanpy as sc import warnings import subprocess import seaborn as sns from sklearn.metrics import mean squared error from scipy.spatial.distance import jensenshannon from scipy.stats import pearsonr,ttest ind,mannwhitneyu import matplotlib import time

###please make sure that you have changed the directory to 'SpatialBenchmarking'.

os.chdir('../')
please add the your SpatialBenchmarking dir into the pythn path sys.path.append('~/SpatialBenchmarking/')

###Predict gene spatial distribution of undetected genes ###Prepared files

We sssume that you have these files:

#1): scRNA-seq count files (eg. scRNA_count.txt);

genes X cells, each row is a gene and each col is a cell.

#2): spatial transcriptomics count files (eg. Insitu_count.txt);

spots X genes, each col is a gene. Please note that the file has no index

#3): spatial location files (eq. Locations.txt);

each col is a spot coordinates. Please note that the file has no index

default: None. It is necessary when you use SpaOTsc or novoSpaRc to integrate datasets.

#4): CellTypeAnnotate file for scRNA-seq data, when using Tangram for prediction. (Option)

if this CellTypeAnnotate is supported, you must set the annotate != None and modes = 'clusters'

```
import numpy as np
import pandas as pd
###Import the package "SpatialGenes" to directly predict the gene spatial distribution for any spatial datasets.
import Benchmarking. Spatial Genes as Spatial Genes
### input data directory
PATH = 'DataUpload/Dataset15/'
### scRNA-seq count files (genes X cells, each row is a gene and each col is a cell)
RNA path = PATH + 'scRNA count.txt'
### spatial transcriptomics count files (each col is a gene)
Spatial path = PATH + 'Insitu count.txt'
### spatial location files (each col is a spot coordinate)
location_path = PATH + 'Locations.txt'
### Read data
RNA_data = pd.read_table(RNA_path,header = 0,index_col = 0)
Spatial_data = pd.read_table(Spatial_path,sep = '\t',header=0)
### genes for integration and prediction. Please note it must be a list.
train_list = list(RNA_data.index&Spatial_data.columns)
test_list = list(set(RNA_data.index) - set(Spatial_data.columns))[:20]
### Outfile directory
outdir = 'Dataset15/'
if not os.path.exists(outdir):
  os.mkdir(outdir)
### Running
test = SpatialGenes.GenePrediction(RNA path, Spatial path, location path, train list = train list, test list = test list,
outdir = outdir)
### choose tools you you want use for prediction
Methods = ['SpaGE','novoSpaRc','SpaOTsc','Seurat','LIGER']
### Prediction
Result = test.Imputing(Methods)
###GPU Platform gimVI, Tangram, and stPlus
import os
import numpy as np
import pandas as pd
from stPlus import *
import Benchmarking. Spatial Genes as Spatial Genes
import os
PATH = 'DataUpload/Dataset15/'
RNA_path = PATH + 'scRNA_count.txt'
Spatial_path = PATH + 'Insitu_count.txt'
location_path = PATH + 'Locations.txt'
RNA_data = pd.read_table(RNA_path,header = 0,index_col = 0)
Spatial_data = pd.read_table(Spatial_path,sep = '\t',header=0)
train_list = list(RNA_data.index&Spatial_data.columns)
print (train_list)
test_list = list(set(RNA_data.index) - set(Spatial_data.columns))[:20]
outdir = 'Dataset15/'
if not os.path.exists(outdir):
  os.mkdir(outdir)
test = SpatialGenes.GenePrediction(RNA_path, Spatial_path, location_path, train_list = train_list, test_list = test_list,
outdir = outdir)
Methods = ['Tangram', 'gimVI', 'stPlus']
```

Result = test.Imputing(Methods)

```
###Prediction Celltype deconvolution
###Prepared files:
```

#You can import the package "DeconvolutionSpot" to directly predict the cell locations for any spatial datasets.

```
# Before forecasting, please prepare the following files:
#1): scRNA count files, h5ad file or h5seurat file;
```

#2): spatial count files, h5ad file or h5seurat file;

#3): scRNA cell annotation files;

#4): output dir.

For more details, please see the Benchmarking/DeconvolutionSpot.py and Figure Data.

time start=time.time() ### please add the your SpatialBenchmarking dir into the pythn path sys.path.append('~/SpatialBenchmarking/') import Benchmarking. Deconvolution Spot as Deconvolution Spot

Celltype deconvolution Prediction

for Cell2location, Stereoscope, Tangram, DestVI, you must have .h5ad files as input.

RNA h5ad = 'ExampleData/Simulated STARmap/starmap sc rna.h5ad' Spatial h5ad = 'ExampleData/Simulated STARmap/starmap spatial.h5ad' celltype_key = 'celltype' output_path = 'FigureData/Figure4/Dataset10_STARmap/Result_STARmap/' if not os.path.exists(output path): os.mkdir(output path) test = DeconvolutionSpot.Deconvolutions(RNA_h5ad = RNA_h5ad, Spatial_h5ad = Spatial_h5ad, celltype_key = celltype_key, output_path = output_path)

Methods = ['Cell2location', 'Stereoscope', 'Tangram', 'DestVI']

Result = test.Dencon(Methods)

time end=time.time()

print('STARmap datasets prediction time cost',(time_end-time_start)/60,'minutes')

sys.path.append('~/SpatialBenchmarking/')

import Benchmarking. Deconvolution Spot as Deconvolution Spot

for SpatialDWLS, RCTD, Seurat, SPOTlight, you must have .h5seurat files as input. ### for SpatialDWLS, you must add my python path.

RNA_h5Seurat = 'ExampleData/Simulated_STARmap/starmap_sc_rna.h5seurat' Spatial h5Seurat = 'ExampleData/Simulated STARmap/starmap spatial.h5seurat' celltype key = 'celltype' my python path = '~/miniconda2/envs/Deconvolution/bin/python' output_path = 'FigureData/Figure4/Dataset10_STARmap/Result_STARmap/' if not os.path.exists(output_path): os.mkdir(output path)

test = DeconvolutionSpot.Deconvolutions(RNA h5Seurat= RNA h5Seurat, Spatial_h5Seurat = Spatial_h5Seurat, celltype_key = celltype_key, python_path = my_python_path, output_path = output_path) Methods = ['SpatialDWLS', 'RCTD', 'Seurat', 'SPOTlight'] Result = test.Dencon(Methods)

sys.path.append('~/SpatialBenchmarking/') import Benchmarking. Deconvolution Spot as Deconvolution Spot

for STRIDE, have count matrix files as input.

RNA file = 'ExampleData/Simulated STARmap/starmap sc rna.tsv' Spatial file = 'ExampleData/Simulated STARmap/starmap spatial.tsv' celltype_file = 'ExampleData/Simulated_STARmap/starmap_sc_rna_celltype.tsv' output path = 'FigureData/Figure4/Dataset10 STARmap/Result STARmap/' if not os.path.exists(output_path): os.mkdir(output path)

test = DeconvolutionSpot.Deconvolutions(RNA file = RNA file, Spatial file = Spatial file, celltype file = celltype file, output_path = output path) Methods = ['STRIDE'] Result = test.Dencon(Methods)

```
###Prediction Celltype deconvolution
###Prepared files:
 #You can import the package "DeconvolutionSpot" to directly predict the cell locations for any spatial datasets.
 # Before forecasting, please prepare the following files:
 #1): scRNA count files, h5ad file or h5seurat file;
 #2): spatial count files, h5ad file or h5seurat file;
 #3): scRNA cell annotation files;
 #4): output dir.
 # For more details, please see the Benchmarking/DeconvolutionSpot.py and Figure Data.
time start=time.time()
### please add the your SpatialBenchmarking dir into the pythn path
sys.path.append('~/SpatialBenchmarking/')
import Benchmarking.DeconvolutionSpot as DeconvolutionSpot
### Celltype deconvolution Prediction
### for Cell2location, Stereoscope, Tangram, DestVI, you must have .h5ad files as input.
RNA_h5ad = 'ExampleData/Simulated_STARmap/starmap_sc_rna.h5ad'
Spatial_h5ad = 'ExampleData/Simulated_STARmap/starmap_spatial.h5ad'
celltype key = 'celltype'
output_path = 'FigureData/Figure4/Dataset10_STARmap/Result_STARmap/'
if not os.path.exists(output_path):
 os.mkdir(output_path)
test = DeconvolutionSpot.Deconvolutions(RNA h5ad = RNA h5ad, Spatial h5ad = Spatial h5ad, celltype key =
celltype_key, output_path = output_path)
Methods = ['Cell2location', 'Stereoscope', 'Tangram', 'DestVI']
Result = test.Dencon(Methods)
time_end=time.time()
print('STARmap datasets prediction time cost',(time end-time start)/60,'minutes')
sys.path.append('~/SpatialBenchmarking/')
import Benchmarking.DeconvolutionSpot as DeconvolutionSpot
### for SpatialDWLS, RCTD, Seurat, SPOTlight, you must have .h5seurat files as input.
### for SpatialDWLS, you must add my python path.
RNA_h5Seurat = 'ExampleData/Simulated_STARmap/starmap_sc_rna.h5seurat'
Spatial_h5Seurat = 'ExampleData/Simulated_STARmap/starmap_spatial.h5seurat'
celltype_key = 'celltype'
my python path = '~/miniconda2/envs/Deconvolution/bin/python'
output_path = 'FigureData/Figure4/Dataset10 STARmap/Result STARmap/'
if not os.path.exists(output_path):
 os.mkdir(output_path)
test = DeconvolutionSpot.Deconvolutions(RNA h5Seurat= RNA h5Seurat, Spatial h5Seurat = Spatial h5Seurat,
celltype_key = celltype_key, python_path = my_python_path, output_path = output_path)
Methods = ['SpatialDWLS','RCTD', 'Seurat', 'SPOTlight']
Result = test.Dencon(Methods)
sys.path.append('~/SpatialBenchmarking/')
import Benchmarking.DeconvolutionSpot as DeconvolutionSpot
### for STRIDE, have count matrix files as input.
RNA_file = 'ExampleData/Simulated_STARmap/starmap_sc_rna.tsv'
Spatial_file = 'ExampleData/Simulated_STARmap/starmap_spatial.tsv
celltype_file = 'ExampleData/Simulated_STARmap/starmap_sc_rna_celltype.tsv'
output_path = 'FigureData/Figure4/Dataset10_STARmap/Result STARmap/'
if not os.path.exists(output_path):
 os.mkdir(output_path)
test = DeconvolutionSpot.Deconvolutions(RNA file = RNA file, Spatial file = Spatial file, celltype file = celltype file,
output path = output path)
Methods = ['STRIDE']
Result = test.Dencon(Methods)
```