import packages

import numpy as np import pandas as pd import sys import os import time as tm import pickle from functools import partial import scipy.stats as st from scipy.stats import wasserstein distance import scipy.stats import copy from sklearn.model_selection import KFold import pandas as pd import multiprocessing import matplotlib as mpl import matplotlib.pyplot as plt import scanpy as sc import warnings import subprocess import seaborn as sns from sklearn.metrics import mean squared error from scipy.spatial.distance import jensenshannon from scipy.stats import pearsonr,ttest ind,mannwhitneyu import matplotlib import time

###please make sure that you have changed the directory to 'SpatialBenchmarking'.

os.chdir('../')

###Predict gene spatial distribution of undetected genes ###Prepared files

We sssume that you have these files:

#1): scRNA-seg count files (eg. scRNA count.txt);

genes X cells, each row is a gene and each col is a cell.

#2): spatial transcriptomics count files (eg. Insitu count.txt);

spots X genes, each col is a gene. Please note that the file has no index

#3): spatial location files (eg. Locations.txt);

each col is a spot coordinates. Please note that the file has no index

default: None. It is necessary when you use SpaOTsc or novoSpaRc to integrate datasets.

#4): count files containing the number of cells in each spot for Tangram(option) (eg. count.txt).

- # The col represents a spot coordinates. Please note that has no index and the file columns must be 'cell counts'
- # default: None. It is necessary when you use Tangram seg functions to integrate datasets.
- # Please note that Tangram can be used in two ways: Tangram image or Tangram seq.
- # Only when you have information file that containing the number of cells in each spot, you can use Tangram seq.

```
import numpy as np
import pandas as pd
#Import the package "SpatialGenes" to directly predict the gene spatial distribution for any spatial datasets.
import Benchmarking. Spatial Genes as Spatial Genes
### input data directory
PATH = 'FigureData/Figure2/Dataset2 osmFISH/Rawdata/'
### scRNA-seq count files (genes X cells, each row is a gene and each col is a cell)
RNA path = PATH + 'scRNA count.txt'
### spatial transcriptomics count files (each col is a gene)
Spatial path = PATH + 'Insitu count.txt'
### spatial location files (each col is a spot coordinate)
location_path = PATH + 'Locations.txt'
### Read data
RNA_data = pd.read_table(RNA_path,header=0,index_col = 0)
Spatial_data = pd.read_table(Spatial_path,sep='\t',header=0)
### genes for integration and prediction. Please note it must be a list.
train_list = list(RNA_data.index&Spatial_data.columns)
test_list = list(set(RNA_data.index) - set(Spatial_data.columns))[:20]
### Outfile directory
outdir = 'FigureData/Figure2/Dataset2_osmFISH/Test/'
if not os.path.exists(outdir):
     os.mkdir(outdir)
### Running
test = SpatialGenes.GenePrediction(RNA path, Spatial path, location path, train list = train list, test list = test list,
outdir = outdir)
### choose tools you you want use for prediction
Methods = ['SpaGE','novoSpaRc','SpaOTsc','gimVI','Tangram image','Seurat','LIGER']
### Prediction
Result = test.Imputing(Methods)
Spatial_path = PATH + 'Insitu_count.txt'
### Read data
RNA_data = pd.read_table(RNA_path,header=0,index_col = 0)
Spatial data = pd.read table(Spatial path,sep='\t',header=0)
### genes for integration and prediction. Please note it must be a list.
train list = list(RNA data.index&Spatial data.columns)
test list = list(set(RNA data.index) - set(Spatial data.columns))[:20]
### Outfile directory
outdir = 'FigureData/Figure2/Dataset2 osmFISH/Test/'
if not os.path.exists(outdir):
     os.mkdir(outdir)
### Running
test = SpatialGenes.GenePrediction(RNA_path, Spatial_path, location_path, train_list = train_list, test_list = test_list,
outdir = outdir)
### choose tools you you want use for prediction
Methods = ['SpaGE','novoSpaRc','SpaOTsc','gimVI','Tangram_image','Seurat','LIGER']
### Prediction
```

Result = test.Imputing(Methods)

```
###Prediction Cell Locations
###Prepared files:
 #1): scRNA-seq count files (eg. scRNA count.txt);
     # genes X cells, each row is a gene and each col is a cell.
 #2): spatial transcriptomics count files (eg. combined spatial count.txt);
     # spots X genes, each col is a gene. Please note that the file has no index.
 #3): spatial location files (eg. combined Locations.txt);
     # spots X coordinates, each col is a spot coordinates. Please note that the file has no index.
     # default: None. It is necessary when you use SpaOTsc or novoSpaRc to integrate datasets.
 #4): scRNA cell annotation files (scRNA annotate.txt)
     # cells X celltype, each row is a cell and each col is a cell annotation.
     # Please note that the file must have a columns named 'celltype'
 #5): count files containing the number of cells in each spot for Tangram (option) (eg. combined
cell counts.txt);
     # spots X numbers. each row is a spot index and the col represents the cell count in each spot.
     # Please note that the file columns must be 'cell count'
     # default: None. It is necessary when you use Tangram to integrate datasets.
#import the package "CellAssigment" to directly predict the cell locations.
import Benchmarking. Cell Assignment as Cell Assignment
time start=time.time()
### input data directory
PATH = 'FigureData/Figure4/Dataset7 STARmap/'
### scRNA-seq count files (genes X cells, each row is a gene and each col is a cell)
scRNA = PATH + 'Rawdata/scRNA count.txt'
### spatial transcriptomics count files (each col is a gene)
spatial count = PATH + 'Simulated STARmap/combined spatial count.txt'
###spatial location file (spots X coordinates, each col is a spot coordinates. Please note that the file has no index).
location = PATH + 'Simulated_STARmap/combined_Locations.txt'
### count files containing the number of cells in each spot for Tangram (each row is a spot index and each col is a
cell)
cell_counts = PATH + 'Simulated_STARmap/combined_cell_counts.txt'
### scRNA cell annotation files (each row is a cell and each col is a cell annotation)
scrna meta = PATH + 'Rawdata/scRNA annotate.txt'
### In scRNA cell annotation files, for one cell you may have different cell annotation, choose a columns as a annota-
tion for input
annotatetype = 'subclass'
### Outfile directory
outdir = PATH + 'Test/'
if not os.path.exists(outdir):
  os.mkdir(outdir)
### Running
MC = CellAssigment.MappingCell(RNA path = scRNA, Spatial path = spatial count, location path = location,
                   count path = cell counts, scrna annotation = scrna meta,
                   annotatetype = annotatetype,outdir = outdir)
### choose tools you you want use for prediction
Tools = ['novoSpaRc', 'SpaOTsc', 'Seurat', 'Tangram']
### prediction
MC.workstart(Tools)
time end=time.time()
print('STARmap datasets prediction time cost', (time end-time start)/60, 'minutes')
```