Mechanism of Oxidative Stress and Synapse Dysfunction in the Pathogenesis of Alzheimer's Disease: Understanding the Therapeutics Strategies

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Abstract Synapses are formed by interneuronal connections that permit a neuronal cell to pass an electrical or chemical signal to another cell. This passage usually gets damaged or lost in most of the neurodegenerative diseases. It is widely believed that the synaptic dysfunction and synapse loss contribute to the cognitive deficits in patients with Alzheimer's disease (AD). Although pathological hallmarks of AD are senile plaques, neurofibrillary tangles, and neuronal degeneration which are associated with increased oxidative stress, synaptic loss is an early event in the pathogenesis of AD. The involvement of major kinases such as mitogen-activated protein kinase (MAPK), extracellular receptor kinase (ERK), calmodulin-dependent protein kinase (CaMKII), glycogen synthase-3β (GSK-3β), cAMP response element-binding protein (CREB), and calcineurin is dynamically associated with oxidative stress-mediated abnormal hyperphosphorylation of tau and suggests that alteration of these kinases could exclusively be involved in the pathogenesis of AD. N-methyl-Daspartate (NMDA) receptor (NMDAR) activation and beta amyloid (Aβ) toxicity alter the synapse function, which is also

hyperphosphorylation (two main events of AD). However, the involvement of oxidative stress in synapse dysfunction is poorly understood. Oxidative stress and free radical generation in the brain along with excitotoxicity leads to neuronal cell death. It is inferred from several studies that excitotoxicity, free radical generation, and altered synaptic function encouraged by oxidative stress are associated with AD pathology. NMDARs maintain neuronal excitability, Ca2+ influx, and memory formation through mechanisms of synaptic plasticity. Recently, we have reported the mechanism of the synapse redox stress associated with NMDARs altered expression. We suggest that oxidative stress mediated through NMDAR and their interaction with other molecules might be a driving force for tau hyperphosphorylation and synapse dysfunction. Thus, understanding the oxidative stress mechanism and degenerating synapses is crucial for the development of therapeutic strategies designed to prevent AD pathogenesis.

associated with protein phosphatase (PP) inhibition and tau

Keywords NMDA receptor · Oxidative stress · Kinases · Tau protein · Synaptic function · Alzheimer's disease

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Abbreviations

AD Alzheimer's disease
CNS Central nervous system
RNS Reactive nitrogen species
ROS Reactive oxygen species
NMDA receptor N-methyl-D-aspartate receptor

Aβ Beta amyloid

APP Amyloid precursor protein

CaMKII Calmodulin-dependent protein kinase

NFT Neurofibrillary tangle

NO Nitric oxide

MAPK Mitogen-activated protein kinase

PKA Protein kinase

ERK Extracellular receptor kinase

ECD Excitotoxic cell death

LTD Depression

LTP Long-term potentiation

CREB cAMP response element-binding protein

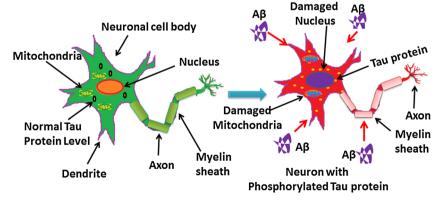
GSK-3β Glycogen synthase-3β

Introduction

Alzheimer's disease (AD), the most common neurodegenerative disorder, is characterized by deposition of amyloid-beta plaques (AB), neurofibrillary tangles (NFTs), and hyperphosphorylated tau (a microtubule binding protein) [1]. It has been reported that impairment of amyloid precursor protein (APP) metabolism in AD leads to increased production of A\beta. A high level of A\beta production is directly correlated with other critical events such as formation of tangles, neuron loss, synapse loss, and neurotransmission dysfunction [2] (Fig. 1). Interestingly, these changes are associated with Nmethyl-D-aspartate receptor (NMDA) receptor activation and oxidative stress which ultimately results in AD pathology. Besides, Aβ is also reported to trigger NMDA-mediated Ca²⁺ influx, excitotoxicity, and stress-related signaling pathways in neurons which may exacerbate aging-related increases in oxidative stress, impaired energy metabolism, and defective Ca²⁺ homeostasis [3]. The NMDA receptors (NMDARs) are cationic channels gated by the neurotransmitter glutamate having critical roles in excitatory synaptic transmission, plasticity, as well as in excitotoxicity in the central nervous system (CNS). The activation of NMDAR glutamate release leads to massive Ca²⁺ fluxes into the postsynaptic cells. Previous reports suggest that oligomeric A\beta-induced Ca2+ influx occurs through formation of reactive oxygen species (ROS) and oxidative stress [4]. Synapses are formed by connections between two neurons that allow a neuronal cell to pass a signal to another cell. This channel usually gets damaged or lost in most neurodegenerative diseases (Fig. 2). Accumulating evidence suggests that dysfunction and loss of synaptic connections may be an important early event underlying AD progression. Insightful synapse degeneration in AD is characterized by the worsening of cognitive function, synapse loss, and neuronal cell death [5]. Synaptic function and plasticity have also been extensively studied in the transgenic mouse models that show abnormal synaptic transmission and impaired long-term potentiation (LTP) which are often well associated with Aß plaque formation [6]. Neurodegenerative disorders are characterized by progressive cell loss in specific neuronal populations and mechanisms that have been put forward to account for AD with aging including inflammation and oxidative stress [7, 8]. Recently, Rai et al. [9] also proved that NMDAR activation, excessive Ca²⁺ fluxes, and free radical generation are associated with synaptic dysfunction and tau phosphorylation [10]. Excessive amounts of glutamate are associated with intense transient influx of Ca2+, leading to mitochondrial functional impairments characterized by activation of the permeability transition pores in the inner mitochondrial membrane, cytochrome c release and depletion of ATP, and simultaneous formation of ROS [11]. In addition, an increase in cytoplasmic Ca²⁺ triggers intracellular cascades which lead to increased levels of ROS and oxidative stress [12]. Analysis of AD brains revealed that the extensive synapse loss is strongly correlated with cognitive impairment [13]. Cognitive function in AD patients is also closely interrelated to the density of presynaptic glutamatergic neurons and postsynaptic neurotoxicity [8, 9, 14]. A previous report by Arendt [15] suggests that synaptic decline occurs early in disease progression and neuronal death alone is not sufficient for disease progression. Cooper and Bear [16] have reported that synapses are selectively removed prior

postsynaptic NMDAR. Furthermore, this can lead to excessive

Fig. 1 Comparative changes in healthy and diseased neuron implicated in AD pathogenesis. The abnormal function of $A\beta$ activates several stress-related kinases that results in damaged nucleus and mitochondria in diseased neurons in AD pathogenesis

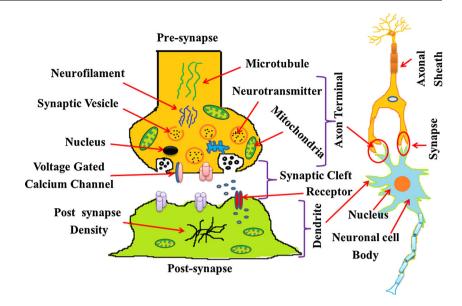


Healthy Neuron

Diseased Neuron



Fig. 2 Structure of synapse: Synapse is a specialized communication junction between two cells comprised of two major units: a presynaptic cell (usually a neuron) that sends out a signal and a postsynaptic cell that receives the signal. Neurotransmitter molecules diffuse across the synaptic cleft and bind to their specific receptors on the postsynaptic cell. This recreates the action potential in the postsynaptic cell. This channel usually gets damaged or lost in neurodegenerative diseases



to cell death and dementia in AD patient may therefore be attributed to progressive reduction in synaptic integrity [17]. Thus, proper synaptic function is crucial for learning and memory. Therefore, from previous observations, it seems that proper NMDAR and synapse function are necessary for learning and memory, and any improper in NMDAR and synapse function may lead to progression of AD pathogenesis. In this review, we connect the link between NMDAR-mediated oxidative stress, Ca²⁺ dysregulation, and kinases on synapse function.

NMDA Receptor-Mediated Oxidative Stress

NMDA type of glutamate receptor (NMDAR) plays an important role in learning and memory formation and also considered crucial for brain development and function in the central nervous system. Activation of NMDAR leads to cytosolic free intracellular Ca²⁺ increase [10] required for LTP and long-term depression (LTD) [18] and, more likely, for synaptic plasticity [19]. NMDA receptor subunit such as NR1, NR2A, and NR2B maintains the synaptic plasticity and neuronal function. The NR2A subunit is involved in the induction of LTP, whereas the NR2B subunit contributes to the formation of LTD and, thus, memory function. Elevation of cytosolic free Ca²⁺ leads to derangement of many intracellular processes that normally regulate Ca²⁺ sequestration and energy metabolism [7]. Modulations of Ca²⁺, glutamate, and NMDAR also induce some other biochemical mechanisms such as oxidative stress which further compromise with cell death [8]. Oxidative stress is one of the most important mechanisms involved in toxic events observed in neuronal cells in different neurodegenerative disorders as measured by free radical generation and lipid peroxidation [20]. Under normal conditions,

ROS act as signaling molecules in many physiological processes including redox homeostasis and cellular signal transduction [21]. By activating proteins such as tyrosine kinases, mitogen-activated protein kinases and ROS are important mediators of signal transduction pathways [22]. Increased production of cellular ROS and oxidative stress has been reported to induce autophagy, a homeostatic process that enables cells to degrade cytoplasmic proteins and organelles [23]. Nitric oxide (NO) released by NO synthase may induce synaptic changes by causing neurotoxicity. Recent studies suggest that NO may be acting as a neuronal messenger in the central nervous system and is involved in the pathophysiology of neurodegenerative disorders [24]. Liu et al. [25] suggested that generation of reactive nitrogen species (RNS) and ROS triggers oxidative stress and eventually leads to neuronal damage. Thus, the increased free radical generation may ultimately lead to synaptic dysfunction which is an important pathophysiological component of AD [26]. Moreover, Kamat et al. [10] also suggest that oxidative stress causes cognitive deficiency, neurofibrillary tangle (NFT)-like pathological changes, and oxidative stress as seen in AD pathology via tau hyperphosphorylation. AD includes a variety of risk factors such as extracellular deposition of β-amyloid, accumulation of intracellular neurofibrillary tangles, oxidative neuronal damage, and inflammatory cascades [27]. Therefore, NMDAR activation, free radical generation, apoptosis, and their consequence on synapse function may lead to AD progression.

NMDA Receptor-Mediated Apoptotic Cell Death

Excitotoxic cell death (ECD) is a characteristic of mammalian brains in several types of neuronal apoptosis. A key event in



ECD is a massive increase in intracellular Ca²⁺ by overstimulation of NMDAR. Excitotoxicity is defined as a toxic process characterized by a sustained stimulation of excitatory amino acid receptors mainly involving NMDARs [28, 29]. Different toxic events derived from excitotoxicity have been characterized in experimental models including upregulation of detrimental signaling pathways, disrupted Ca2+ homeostasis, and ROS/RNS with further oxidative/nitrosative stress ultimately leading to cell death [9]. Apoptosis plays a significant role in cell death during neurodegenerative disorders such as AD [30]. The activation of caspases, a major apoptotic pathway, is also characterized by mitochondrial dysfunction with the release of cytochrome c and activation of caspase-9 and, subsequently, of caspase-3 [7]. Lines of evidence suggest that caspase-3 activations were also found in the brain of AD patients [31]. Most neurons in the mammalian central nervous system possess receptors for apoptosis, the excitatory neurotransmitter glutamate. Overactivation of glutamate receptors can induce apoptosis by a mechanism involving Ca²⁺ influx [32]. Neuronal cell death is also triggered in response to increased oxidative stress, in which free radicals such as the superoxide anion radical and the hydroxyl radical damage cellular lipids, proteins, and nucleic acids [33]. So, collective information suggests that NMDAR-mediated oxidative stress and neuronal apoptosis directly or indirectly influences synapse function.

Biochemical Mechanism of Apoptosis and Synaptic Dysfunction

Biochemical mechanisms involved in apoptosis can be activated at synapse which can alter synaptic function and promote degeneration of synapses [34]. Apoptosis can be induced in synaptosome preparations and neuritis of cultured brain neurons by insults that induce apoptosis in intact neurons. Caspase-mediated cleavage of synaptic proteins may control the process of neuronal apoptosis. α-Amino-3-hydroxy-5methyl-4-isoxazolepropionic acid (AMPA) receptor subunits are selectively degraded in hippocampal neurons after exposure to an apoptotic dose of glutamate, resulting in decreased Ca²⁺ influx and, thereby, preventing excitotoxicity [32]. Apoptotic pathways may also function in synaptic plasticity, particularly under conditions of stress and injury. The elevation of caspase-3 activity correlated temporally with memory impairment, reduced spine density and size, altered excitatory synaptic transmission, and enhanced LTD. Remarkably, pharmacologic inhibition of caspase-3 ameliorated the synaptic transmission, spine size, and memory deficits in these AD transgenic mice [35]. Increased caspase-3 activity is also reported in human AD brain, and elevated levels of caspase-3 are observed in the postsynaptic density fraction of AD brain [36]. Inhibition of caspase-3 activity is beneficial to reverse cognitive decline in APP mice possibly due to a requirement for caspase-3 activity in normal synaptic function [37, 38]. Thus, apoptotic executes by caspase-3 activation may contribute to neuronal apoptosis and synapse dysfunction and further provoke AD progression.

Mitochondria as a Regulator of Apoptosis and Oxidative Stress

Since most neurodegenerative disorders are associated with mitochondrial abnormalities, AD is associated with similar fashion of mitochondrial dysfunction. The mitochondria play a critical role in the regulation of apoptosis which is implicated in the aging process. Age-related mitochondrial oxidative stress may contribute to apoptosis [39]. The mitochondria are significantly reduced in various types of cells and its dysfunction is one of the causative pathophysiology of AD [40]. The most regular defect in mitochondrial electron transport enzymes in AD is a deficiency in cytochrome c oxidase which leads to an increase in ROS production, a reduction in energy stores, and disturbance in energy metabolism [41]. Indeed, ROS generation is an important mechanism accounting for cellular injury in many neurodegenerative disorders [42]. Such selective oxidative modification may cause the cells to be more vulnerable to apoptotic inducers [43]. Thus, the mitochondria appear to influence the aging process via modifying the regulatory machinery of apoptosis and that the mitochondria have a central role in aging-related neurodegenerative diseases like AD [7]. Consistently, oxidative stress-induced accumulation of Aβ-protein in AD causes lysosome membrane degradation and ultimately leads to neuronal cell death [44]. The mitochondria damaged by oxidative stress in pyramidal neurons of AD are subjected to neurodegeneration [45]. Thus, inhibition of the mitochondrial complexes leads to diminished ATP production and resulted in impaired energy metabolism. Several lines of evidence support that NO impairs mitochondrial/cellular respiration and other functions by inhibiting the activities of several key enzymes, particularly cytochrome c oxidase, and thereby causing ATP depletion [46]. In cultured neuroblastoma cells, overexpression of tau results in mitochondria with decreased ATP levels and increased susceptibility to oxidative stress [47]. Furthermore, the activity and composition of mitochondrial enzymes are disrupted in the mouse model of tauopathy [48]. Thus, tau may also directly influence mitochondrial function. It appears that mitochondria-mediated oxidative stress influences tau function, and the abnormal function of tau also influences the mitochondria. Hence, the mitochondria



and tau influence each other's function, which is profoundly associated with AD pathogenesis.

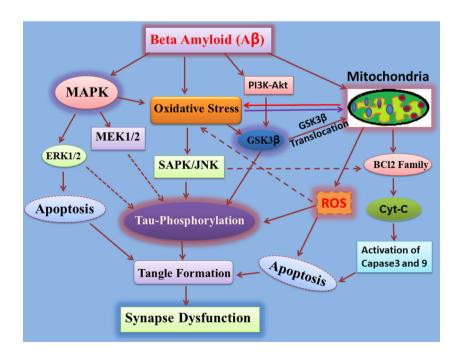
Molecular Mechanism of Oxidative Stress-Mediated Tau Phosphorylation

Inactivation of protein phosphatase 1/protein phosphatase 2A (PP1/PP2A) via oxidative stress has been shown in vitro and in vivo to be involved in hyperphosphorylation of tau and prolonged phosphorylation of extracellular receptor kinase (ERK) 1/2 [49]. Thus, it is intriguing to postulate that oxidative stress-mediated PP1 and PP2A inhibition in AD may account for enhanced ERK1/2 activity and subsequent tau hyperphosphorylation and neurofibrillary tangle formation. Li et al. [50] suggested that a decrease in PP2A activity causes the activation of ERK1/2, MEK1/2, and several other kinases and the abnormal hyperphosphorylation of tau both via an increase in its phosphorylation and a decrease in its dephosphorylation in AD brain. Stress-activated protein kinase (SAPK) and p38 MAPKs are members of the complex superfamily of MAP serine/threonine protein kinases. This superfamily also includes the ERKs (also referred to as MAPKs), which are typically activated by mitogens, c-Jun N-terminal kinase (JNK)/SAPK, and p38 mitogen-activated protein kinase (MAPK), which are known as stress-activated kinases [51]. This can be attributed to the fact that the activities of these enzymes are stimulated by a variety of exogenous and endogenous stress-inducing stimuli including ROS and oxidative stress (Fig. 3). The kinases that phosphorylate tau can be activated by NMDA-mediated oxidative stress such as hyperactivation of Cdk5 signaling pathway, MAPK, and several stress-activated protein kinases [52]. Thus, induction of this protein by JNK/SAPK could serve as a potential marker for pathologies associated with chronic oxidative stress. Among them, cyclic AMP-dependent protein kinase (PKA) and calcium-calmodulin-dependent protein kinase II (CaMKII) are associated with NMDAR remodeling. The phosphorylation of tau by these kinases inhibits the ability of tau to promote microtubule assembly and facilitates the polymerization of tau into paired helical filament (PHF) [53]. Protein phosphatase mediates the regulation of protein kinase C during long-term depression in the adult hippocampus in vivo. The neural substrates of learning and memory are thought to involve long-term changes in synaptic function, including LTD of synaptic strength. All these studies signify that inhibition of phosphates activates the number of kinases which actively participate in the activation of oxidative stress. Activation of kinases also stimulates the NMDAR and CaMKII remodeling. Moreover, inhibition of phosphatase also causes tau hyperphosphorylation, a critical event of AD pathology. Thus, phosphatase inhibition, kinase activation, NMDAR remodeling, and oxidative stress are strongly correlated with each other and, hence, influence synaptic function.

Modulation of NMDA Receptor and Kinases on Oxidative Stress

NMDA receptors influence death and survival pathways for synapse function. Activation of NMDAR induces the ERKs, which promote a signaling cascade important for neuronal survival. Thus, the synaptic NMDAR activates ERK, promoting cell survival [54], whereas the extra synaptic pool of

Fig. 3 Synapse dysfunction: as a consequence of MAPK and mitochondrial oxidative stress-mediated tau hyperphosphorylation. β-Amyloid activates several stress-related kinases that causes oxidative stress. The phosphorylation/activation of ERK1/2 and MEK1/2 via MAPK results in apoptosis. Consequently, abnormal hyperphosphorylation of tau leads to synapse dysfunction. Aß also results in mitochondrial dysfunction by affecting the prosurvival protein Bcl-2. The mitochondrial-mediated caspase pathway gets triggered by the Bcl-2 family of proteins, then cytochrome c release, and finally, apoptosis. This ultimately leads to synapse dysfunction





NMDARs trigger mitochondrial membrane potential breakdown, as well as cell body and dendritic damage [55]. Moreover, activation of synaptic NMDAR leads to activation of the cAMP response element-binding protein (CREB), a transcription factor also related to cell survival pathways [56, 57], and phosphorylates CREB [58]. Several studies have shown that oligomeric Aß induces partial blockade of NMDAR currents, which leads to reduction of calcium influx that limits CaMKII function [59, 60]. In fact, oligomeric Aβmediated LTP impairment is believed to involve a decrease in the activation of MAPK, CaMKII, and Akt/protein kinase B, but not protein kinases A and C [61, 62]. AB has also been shown to induce synaptic depression by activating mGluRs, which triggers a series of downstream molecular events involving MAPK and calcineurin, which ultimately promotes internalization of AMPA receptors and synapse collapse [63]. Kinases that are important for synaptic function such as PKC, PKA, and CaMKII have reduced basal activity and stimulation-induced activation is impaired [64]. Phosphatase activity contributes to changes in phosphorylation state of bcl-2 family member protein BAD, and the CREB and dephosphorylation of BAD and CREB is associated with impaired memory [65]. This regulatory mechanism of Aß influences NMDAR and CaMKII function and impairs learning and memory function. Abnormal function of AB also activates

several stress-related kinases that cause oxidative stress and apoptosis, which are heavily implicated in AD pathogenesis (Fig. 4). In the later stage, all these disparities result into neuronal dysfunction and synaptic loss.

Glycogen Synthase Kinase-3β

Glycogen synthase kinase-3 (GSK-3) is a pivotal molecule in the development of AD. Inhibition of GSK-3 reduces the production of AB peptides in amyloid plagues and the hyperphosphorylation of tau protein in neurofibrillary tangles [66]. GSK-3beta is involved in the formation of PHF-tau, which is an integral component of the NFT deposits that disrupt neuronal function and a marker of neurodegeneration in AD [10, 67]. GSK-3 also phosphorylates and inhibits cAMP responsive element-binding protein [68], a universal modulator of memory and intermediate molecule of the NMDAR pathway. Moreover, GSK-3\beta promotes actin and tubulin assembly [69], processes required for synaptic reorganization during memory formation. Additionally, within the brain, MAPK inactivates GSK-3\beta by direct phosphorylation at its C-terminus [70]. Dephosphorylation of GSK-3 at inhibitory sites is coordinated by PP1, PP2A, and protein phosphatase 2B (PP2B, calcineurin) [71]. PP1 preferentially acts as a

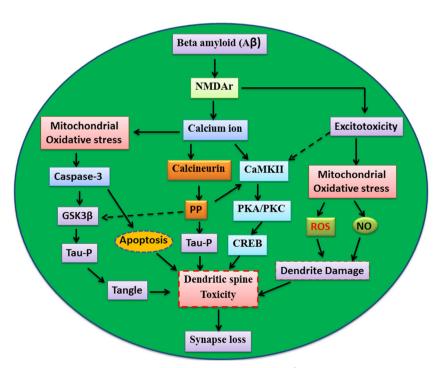


Fig. 4 Molecular mechanism of tau phosphorylation mediated synaptic dysfunction: High level of Aβ production is directly correlated with critical event for synaptic dysfunction, i.e., formation of tangles. Excitotoxic cell death (ECD) is an event due to increased intracellular Ca^{2+} by overstimulation of NMDA receptor. Overstimulation of NMDAR leads to upregulation of detrimental signaling pathways,

disrupting Ca²⁺ homeostasis and oxidative/nitrosative stress ultimately toward apoptosis. The mitochondrial oxidative stress and the release of cytochrome c, activation of caspase-9, and subsequently of caspase-3 cause neuronal damage. Mitochondria-mediated oxidative stress influences tau function resulting in the hyperphosphorylation of tau which governs the major synaptic dysfunction by forming tangles



phosphatase for GSK-3 β , while PP2A favors GSK-3 α [72, 73]. On the other hand, the overexpression of GSK-3ß inhibits PP2A, which may serve as a negative feedback mechanism for GSK-3\beta activity [74]. GSK-3\beta activity is negatively regulated by several signal transduction cascades that protect neurons against apoptosis suggesting the interesting possibility that activation of GSK-3β may contribute to neuronal apoptosis [75]. In response to oxidant stress, GSK-3β translocates to the mitochondria in a kinase activity-dependent manner and enhances production of cytotoxic ROS from mitochondria. This study identifies GSK-3\beta, a kinase known to participate in oxidative stress, cell death, and neurodegeneration, as a fundamental element in the downregulation of the antioxidant cell defense [76]. Collectively, it seems that AB activates GSK-3\beta which induces oxidative stress, hyperphosphorylation of tau, NFT formation, neuronal death, and synaptic loss that can induce memory deficits.

NMDA Receptor and Calcineurin (PP2B) Activity

Since intracellular Ca²⁺ coming from several sources and mostly through glutamate-mediated contribute to calcineurin (CaN) activation and may be preferentially increased by NMDAR activity in animals [77]. Furthermore, mild oxidative stress is thought to increase CaN activity through the release of Ca²⁺ from intracellular stores [78] or a decrease in effectiveness of inhibitory proteins [79]. This would suggest that the release of Ca²⁺ from intracellular stores could decrease NMDAR function through CaN activation and increased oxidative stress that can influence synapse function [80]. Chen et al. [81] have reported that β-amyloid reduces NMDAR function and impairs LTP through enhanced CaN activity. As the disease progression starts, increased expression of CaN inhibitory proteins may shift the balance of Ca²⁺activated kinases/phosphatases. Therefore, calcineurin alteration promotes tau phosphorylation, neurodegeneration, tangle formation [82, 83] and, subsequently, synapse dysfunction.

Phosphatase Inhibition, NMDA Receptor, Tau Hyperphosphorylation, and Synapse Function

Protein phosphatase inhibition leads to the induction of synaptic plasticity in the form of LTP involved in memory formation. Inhibition of protein phosphatases reduces postsynaptic signaling as a major mechanism for basal synaptic transmission and memory formation [84]. It has been reported that inhibition of protein phosphatases increases tau phosphorylation and initiates neuronal cell death which includes altered Ca²⁺ homeostasis and glutamate excitotoxicity that alter the memory formation [85, 86]. Previous reports suggested that

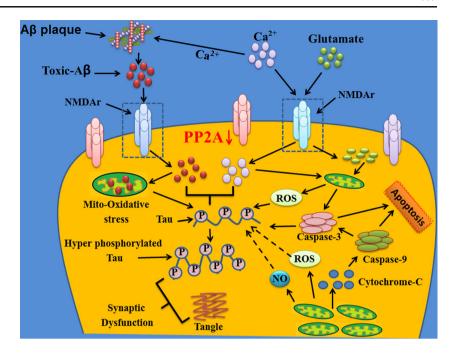
accumulation and mislocalization of hyperphosphorylated tau in the somatodendritic compartment of neurons in AD disrupts glutamate receptor trafficking and synaptic function [87–89]. Not many available reports show how tau hyperphosphorylation and aggregation contributes to the synaptic deficits and neuronal death in AD. Hoover et al. [88] suggest that abnormal tau phosphorylation may also affect postsynaptic receptor targeting and display disrupted targeting of excitatory glutamate receptor and dendritic spines (Fig. 5). Impaired NMDAR can lead to dendritic spine dysfunction and removal of dendritic spine [90]. Tau protein also binds to a postsynaptic protein complex which includes PSD-95, the scaffold for synaptic NMDARs which is a critical regulator of synaptic plasticity [91]. Endogenous tau is typically present in dendrites in the postsynapse, where it interacts with the PSD-95/NMDAR complex [92]. Furthermore, microinjection of human tau into the presynaptic terminal of the squid axon has blocked synaptic transmission possibly by preventing proper docking of synaptic vesicles [93]. Therefore, tau may play an important role in regulating synaptic function and targeting neurotransmitter receptors to the synapse [94]. Zhang et al. [95, 96] also showed that glutamate release during stimulus and intracellular Ca²⁺ release from presynaptic terminals led to abnormal synapse function. These reports suggest that presynaptic dysfunction might be an early component of synaptic dysfunction in AD. Thus, lines of evidence supported that the tau protein is actively engaged in synapse dysfunction and associated with NMDAR function. In conclusion, the glutamate-induced excitotoxicity and synaptic dysfunction could be an excellent target for the therapy of AD.

NMDA Receptor and Synapse Function

NMDARs are located in neuronal cell membranes at synaptic and extrasynaptic locations, where they are believed to mediate distinct physiological and pathological processes. NMDARs are glutamate-gated ion channels that are highly permeable to Ca²⁺ and Ca²⁺ influx through NMDAR. This is essential for synaptogenesis. Synaptic remodeling leads to long-lasting changes in synaptic efficacy such as long-term potentiation and long-term depression [97]. The NMDARs are involved in cellular mechanism for learning and memory function [9]. NMDARs are involved in a wide array of biological processes which are crucial for brain development and function [98]. NMDAR activation might also play an important role in extracellular adenosine regulation, with important consequences for the regulation of excitatory synaptic transmission, plasticity, and excitotoxicity [99]. NMDAR activation also led to influx of Ca²⁺ through a ligand- and voltagesensitive calcium channel [100], which triggered significant advances in understanding the cellular cascades initiated as a result of tetanic stimulation. The glutamatergic hypothesis of



Fig. 5 Beta amyloid-mediated synapse loss: Deposition of amyloid plaques (Aβ) is characterized in Alzheimer's disease (AD) which affects NMDAR resulting in dendritic damage caused due to mitochondrial oxidative stress as a consequence of excitotoxicity. Aβ also trigger NMDA mediated Ca²⁺ influx which results in dendritic spine toxicity mediated by CREB and hyperphosphorylation of Tau. Ca²⁺ influx can also lead to mitochondrial oxidative stress along with caspase-3-mediated apoptosis and tau hyperphosphorylation and ultimately leads to synapse loss



AD states that glutamate-related excitotoxic mechanisms involving the NMDAR lead to neurodegeneration and cell death [101]. The activation of glutamate receptors has also been found to induce the release of glutamate and induce a massive accumulation of Ca²⁺. This influx of Ca²⁺ contributes to an alteration of cell function, leading to cell death either through free radicals or through overload of the mitochondria, resulting in free radical formation, caspase activation, and the release of apoptosis-inducing factors [7, 102]. Therefore, synaptic stimulation through NMDARs is important for learning and memory functions, but excess glutamate can overstimulate these receptors resulting in excitotoxicity and neurodegeneration.

Glutamate and $A\beta$ -Mediated Ca^{2+} Dysregulation and Synapse Function

Glutamate plays an essential role in learning and memory by triggering NMDAR to allow a controlled amount of calcium into a nerve cell. Ca²⁺ helps to create the chemical environment required for information storage. Excess glutamate, on the other hand, overstimulates NMDARs so that they allow even more calcium into nerve cells and the process leads to disruption and death of the cells [20]. It has been reported that inhibition of protein phosphatases increases tau phosphorylation and initiates neuronal cell death which include altered Ca²⁺ homeostasis and glutamate excitotoxicity [86]. In contrast to most disease models, aged animals exhibit neurologic changes that usually include synaptic dysfunction and Ca²⁺ dysregulation [103]. Aβ plaques, a pathological feature of AD, induce extracellular accumulation of glutamate and

intracellular deposition of calcium ion Ca²⁺. In vivo Ca²⁺ imaging studies corroborate the idea that different subsets of neurons in AD transgenic mice can be found exclusively near Aβ plagues and appeared to result from a relative decrease in synaptic inhibition [104]. In vivo imaging of aged APP transgenic mouse brain shows elevated intracellular Ca²⁺ and aberrant Ca²⁺ homeostasis in a subset of neurites in close proximity of Aβ plaques [105]. The abnormal Ca²⁺ handling of neurons affected by Aß is associated with loss of dendritic spines and neuritic dystrophy, mediated in part by the Ca²⁺dependent protein phosphatase calcineurin [106]. It is notable that calcineurin is also required for apoptosis and LTD, as well as for Aβ-induced spine loss and endocytosis of NMDARs [107]. Thus, a large buildup of glutamate can occur and induce a massive accumulation of Ca²⁺, leading to apoptosis. It has also been noted that Aß plaques increase neurons' vulnerability to excitotoxicity and loss of synaptic protein.

NMDA Receptor, CaMKII, and CREB

The transcription factor CREB plays an essential role in the maintenance of LTP. CREB signaling has been recently involved in several brain pathological conditions including cognitive and neurodegenerative disorders. Altered hippocampal-dependent synaptic plasticity and memory mediates synapse loss through the CREB signaling pathway [108]. The activation of CREB by phosphorylation is triggered in neurons by a wide variety of signaling processes, from increases in intracellular Ca²⁺ through activation of voltage- or ligand-gated channels to changes in cAMP levels. The phosphorylation of CREB by kinases from several signaling pathways may be a



mechanism for memory formation [109]. Activation of synaptic NMDARs toward CaMKII, which may also phosphorylate and activate CREB in neurons, is associated with increased Ca²⁺ [110]. Interestingly, the increase in nuclear Ca²⁺ can also activate CREB, indicating that nuclear kinases may have a direct role in the modulation of CREB activity [111]. Loss of excitatory synapses in the hippocampus induced by Aß requires functional NMDARs for proper synapse maintenance [61]. Aß also influences CREB activation, which is crucial for the maintenance of LTP. LTP plays a crucial role in memory formation. Vitolo et al. [112] showed that AB decreases the activity of CREB and thus reduces the expression of genes encoding proteins that are essential for LTP. Another study found that excessive activation of extrasynaptic NR2Bcontaining NMDARs, which leads to downregulation of CREB, underlies oligomeric A\beta-mediated LTP inhibition [113]. Thus, it may be inferred from the above reports that Aß also modulates CaMKII, CREB, and NMDAR essential for maintenance of LTP.

Therapeutic Strategies

The therapies applied till date for the treatment of AD include the following: antibody vaccination and immunization therapies; gene therapy; enzyme-based therapies such as βsecretase inhibitors, γ -secretase inhibitors and modulators, and cholinesterase inhibitors; receptor-based therapies such as NMDA receptor antagonists, AMPA receptor modulators, peroxisome proliferator-activated receptor agonists, M1 muscarinic agonists, receptor for advanced glycation end productrelated mechanisms, ad nicotine acetylcholine receptor agonists; redox stress-based therapies such as antioxidants and anti-inflammatory and neuroprotective approaches; and taurelated therapies. Moreover, several other therapies were also designed to treat AD by targeting cholesterol biosynthesis, astrocyte-modulating agents, homocysteine-lowering strategies, and caspase inhibitors. Most of the abovementioned therapies also passed through clinical trials but results were not successful so far. Among all the therapies which are currently in the market, only two classes of drugs are available for commercial purpose such as the cholinesterase inhibitor and NMDA receptor antagonist. However, these classes of drugs are also not successful either to give complete remedy from AD. There may be some significant gaps which are necessary to understand and improve the current therapies.

Significant Gap in Research

Oxidative stress-induced damage in the brain is associated with aging and is usually involved in the development of AD pathology in a clinical set of condition. Some observational studies have suggested that antioxidant therapy could overcome the disease progression of AD. Much research has been carried out with antioxidant therapy, but antioxidant randomized clinical trials in AD have had mixed results. Oxidative stress is a well-established pathophysiological factor in AD but till now the use of antioxidants in the prevention or therapy gave conflicting results. Some of the reports have shown antioxidant therapy for the betterment of AD pathology (Table 1), but still therapy is not so promising. In our opinion, antioxidant therapy alone is not looking so hopeful. We should use some more specific targets with combinational therapy rather than a single antioxidant such as NMDAR antagonist, neuronal kinases, and antioxidant as well as flavonoids.

Future Direction

In conclusion, through this review, we have tried to give our perspective on the wide variety of interaction between NMDAR-mediated oxidative stresses with the etiology of

 Table 1
 List of pharmacologically active antioxidants, flavonoids, and vitamins used in AD pathology

S. nos.	Antioxidants/flavanoids	References
1	Vitamin C and vitamin E	[114, 115]
2	Vitamin C	[116]
3	Vitamin B ₁₂	[117, 118]
4	Vitamin E	[119–123]
5	α-Lipoic acid, CoQ10	[124, 125]
6	Selenium and vitamin E	[126]
7	MnSOD	[127]
8	Curcumin	[128, 129]
9	Memory XL (folic acid, vitamin B ₁₂ , vitamin E, acetyl-L-carnitine, SAM, NAC)	[130, 131]
10	Ebenone/idebenone	[132–134]
11	Estrogen	[135, 136]
12	Colostrinin	[137–140]
13	Epigallocatechine gallate (EGCG)	[141]
14	Resveratrol	[142, 143]
15	Pramipexole	[144]
16	Latrepirdine	[145, 146]
17	Ubiquinone, vitamin E, or lipoic acid	[147, 148]
18	Lipoic acid	[149]
19	Silibinin/quercitin	[150-152]
19	Melatonin	[153-155]
20	Caffeine	[156]
21	Selegiline (L-deprenyl)	[119]
22	Ginkgo biloba	[157–159]
23	Gugulipid	[160, 161]



Alzheimer's disease. NMDAR-mediated oxidative stress mechanisms are likely to play an important role in the synapse dysfunction in the pathogenesis of AD. Moreover, mitochondrial-mediated oxidative stress and apoptosis are also suggested to be contributing factors in AD pathogenesis. Furthermore, oxidative stress-mediated kinase and tau phosphorylation provides a connection of synapse dysfunction in AD. As we are not getting complete remedies from antioxidant therapy or known NMDAR antagonist drug used for AD pathology, should we go for combinational therapy? Or are there so many intermediate molecules between NMDAR to neurodegeneration? Should we go for target intermediate molecules? Therefore, understanding the role of oxidative stress-associated molecule and kinases in synapse dysfunction during AD pathogenesis may also lead to the development of mechanism-based therapeutics and better constructive strategies.

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Conflict of Interest The authors declare that they have no conflict of interest.

References

- Gustafson DR, Skoog I, Rosengren L, Zetterberg H, Blennow K (2007) Cerebrospinal fluid beta-amyloid 1-42 concentration may predict cognitive decline in older women. J Neurol Neurosurg Psychiatry 78:461–464
- Wang Z, Yang L, Zheng H (2012) Role of APP and Abeta in synaptic physiology. Curr Alzheimer Res 9:217–226
- Bezprozvanny I, Mattson MP (2008) Neuronal calcium mishandling and the pathogenesis of Alzheimer's disease. Trends Neurosci 31:454–463
- De Felice FG, Velasco PT, Lambert MP, Viola K, Fernandez SJ, Ferreira ST, Klein WL (2007) Abeta oligomers induce neuronal oxidative stress through an N-methyl-D-aspartate receptordependent mechanism that is blocked by the Alzheimer drug memantine. J Biol Chem 282:11590–11601
- Rai S, Kamat PK, Nath C, Shukla R (2014) Glial activation and post-synaptic neurotoxicity: the key events in streptozotocin (ICV) induced memory impairment in rats. Pharmacol Biochem Behav 117:104–117
- Roberson ED, Halabisky B, Yoo JW, Yao J, Chin J, Yan F, Wu T, Hamto P, Devidze N, Yu GQ, Palop JJ, Noebels JL, Mucke L (2011) Amyloid-beta/Fyn-induced synaptic, network, and cognitive impairments depend on tau levels in multiple mouse models of Alzheimer's disease. J Neurosci 31:700–711
- Kamat PK, Tota S, Shukla R, Ali S, Najmi AK, Nath C (2012) Mitochondrial dysfunction: a crucial event in okadaic acid (ICV) induced memory impairment and apoptotic cell death in rat brain. Pharmacol Biochem Behav 100:311–319
- Kamat PK, Rai S, Swarnkar S, Shukla R, Nath C (2014) Mechanism of synapse redox stress in okadaic acid (ICV) induced memory impairment: role of NMDA receptor. Neurochem Int. doi:10.1016/ j.neuint.2014.06.012

- Rai S, Kamat PK, Nath C, Shukla R (2013) A study on neuroinflammation and NMDA receptor function in STZ (ICV) induced memory impaired rats. J Neuroimmunol 254:1–9
- Kamat PK, Rai S, Swarnkar S, Shukla R, Ali S, Najmi AK, Nath C (2013) Okadaic acid-induced tau phosphorylation in rat brain: role of NMDA receptor. Neuroscience 238:97–113
- Nicholls DG (2002) Mitochondrial function and dysfunction in the cell: its relevance to aging and aging-related disease. Int J Biochem Cell Biol 34:1372–1381
- Farooqui AA, Yi Ong W, Lu XR, Halliwell B, Horrocks LA (2001) Neurochemical consequences of kainate-induced toxicity in brain: involvement of arachidonic acid release and prevention of toxicity by phospholipase A(2) inhibitors. Brain Res Brain Res Rev 38:61– 78
- Serrano-Pozo A, Frosch MP, Masliah E, Hyman BT (2011) Neuropathological alterations in Alzheimer disease. Cold Spring Harb Perspect Med 1(1):a006189
- Bell KF, Bennett DA, Cuello AC (2007) Paradoxical upregulation of glutamatergic presynaptic boutons during mild cognitive impairment. J Neurosci 27:10810–10817
- Arendt T (2009) Synaptic degeneration in Alzheimer's disease. Acta Neuropathol 118:167–179
- Cooper LN, Bear MF (2011) The BCM theory of synapse modification at 30: interaction of theory with experiment. Nat Rev Neurosci 13:798–810
- Selkoe DJ (2002) Alzheimer's disease is a synaptic failure. Science 298:789–791
- Fetterolf F, Foster KA (2011) Regulation of long-term plasticity induction by the channel and C-terminal domains of GluN2 subunits. Mol Neurobiol 44:71–82
- Lau CG, Takeuchi K, Rodenas-Ruano A, Takayasu Y, Murphy J, Bennett MV, Zukin RS (2009) Regulation of NMDA receptor Ca2+ signalling and synaptic plasticity. Biochem Soc Trans 37:1369– 1374
- Kamat PK, Tota S, Saxena G, Shukla R, Nath C (2010) Okadaic acid (ICV) induced memory impairment in rats: a suitable experimental model to test anti-dementia activity. Brain Res 1309:66–74
- Droge W (2002) Aging-related changes in the thiol/disulfide redox state: implications for the use of thiol antioxidants. Exp Gerontol 37: 1333–1345
- Droge W (2002) Free radicals in the physiological control of cell function. Physiol Rev 82:47–95
- 23. Kissova I, Plamondon LT, Brisson L, Priault M, Renouf V, Schaeffer J, Camougrand N, Manon S (2006) Evaluation of the roles of apoptosis, autophagy, and mitophagy in the loss of plating efficiency induced by Bax expression in yeast. J Biol Chem 281: 36187–36197
- Prast H, Philippu A (2001) Nitric oxide as modulator of neuronal function. Prog Neurobiol 64:51–68
- Liu D, Liu J, Sun D, Alcock NW, Wen J (2003) Spinal cord injury increases iron levels: catalytic production of hydroxyl radicals. Free Radic Biol Med 3:64–71
- Sachdeva R, Babbar R, Puri V, Agarwal S, Krishana B (2011)
 Correlation between cognitive functions and nitric oxide levels in patients with dementia. Clin EEG Neurosci 42:190–194
- Chopra K, Misra S, Kuhad A (2011) Neurobiological aspects of Alzheimer's disease. Expert Opin Ther Targets 15:535–555
- Shin JH, Linden DJ (2005) An NMDA receptor/nitric oxide cascade is involved in cerebellar LTD but is not localized to the parallel fiber terminal. J Neurophysiol 94:4281–4289
- Nicholls DG, Johnson-Cadwell L, Vesce S, Jekabsons M, Yadava N (2007) Bioenergetics of mitochondria in cultured neurons and their role in glutamate excitotoxicity. J Neurosci Res 85:3206-3212
- 30. Loh KP, Huang SH, De Silva R, Tan BK, Zhu YZ (2006) Oxidative stress: apoptosis in neuronal injury. Curr Alzheimer Res 3:327–337



- Engidawork E, Gulesserian T, Yoo BC, Cairns N, Lubec G (2001)
 Alteration of caspases and apoptosis-related proteins in brains of patients with Alzheimer's disease. Biochem Biophys Res Commun 281:84–93
- Glazner GW, Chan SL, Lu C, Mattson MP (2000) Caspasemediated degradation of AMPA receptor subunits: a mechanism for preventing excitotoxic necrosis and ensuring apoptosis. J Neurosci 20:3641–3649
- Sastry PS, Rao KS (2000) Apoptosis and the nervous system. J Neurochem 74:1–20
- Mattson MP, Duan W (1999) "Apoptotic" biochemical cascades in synaptic compartments: roles in adaptive plasticity and neurodegenerative disorders. J Neurosci Res 58:152–166
- 35. D'Amelio M, Cavallucci V, Middei S, Marchetti C, Pacioni S, Ferri A, Diamantini A, De Zio D, Carrara P, Battistini L, Moreno S, Bacci A, Ammassari-Teule M, Marie H, Cecconi F (2011) Caspase-3 triggers early synaptic dysfunction in a mouse model of Alzheimer's disease. Nat Neurosci 14:69–76
- Louneva N, Cohen JW, Han LY, Talbot K, Wilson RS, Bennett DA, Trojanowski JQ, Arnold SE (2008) Caspase-3 is enriched in postsynaptic densities and increased in Alzheimer's disease. Am J Pathol 173:1488–1495
- 37. Jo J, Whitcomb DJ, Olsen KM, Kerrigan TL, Lo SC, Bru-Mercier G, Dickinson B, Scullion S, Sheng M, Collingridge G, Cho K (2011) Abeta(1-42) inhibition of LTP is mediated by a signaling pathway involving caspase-3, Akt1 and GSK-3beta. Nat Neurosci 14:545–547
- Xie H, Guan J, Borrelli LA, Xu J, Serrano-Pozo A, Bacskai BJ (2013) Mitochondrial alterations near amyloid plaques in an Alzheimer's disease mouse model. J Neurosci 33:17042–17051
- Yamaguchi R, Perkins G (2009) Dynamics of mitochondrial structure during apoptosis and the enigma of Opa1. Biochim Biophys Acta 1787:963–972
- Hirai K, Aliev G, Nunomura A, Fujioka H, Russell RL, Atwood CS, Johnson AB, Kress Y, Vinters HV, Tabaton M, Shimohama S, Cash AD, Siedlak SL, Harris PL, Jones PK, Petersen RB, Perry G, Smith MA (2001) Mitochondrial abnormalities in Alzheimer's disease. J Neurosci 21:3017–3023
- Cardoso SM, Rego AC, Penacho N, Oliveira CR (2004) Apoptotic cell death induced by hydrogen peroxide in NT2 parental and mitochondrial DNA depleted cells. Neurochem Int 45:693–698
- Valko M, Leibfritz D, Moncol J, Cronin MT, Mazur M, Telser J (2007) Free radicals and antioxidants in normal physiological functions and human disease. Int J Biochem Cell Biol 39:44–84
- Zhang YQ, Herman B (2008) Expression and modification of ARC (apoptosis repressor with a CARD domain) is distinctly regulated by oxidative stress in cancer cells. J Cell Biochem 104:818–825
- 44. Zhang XD, Wang Y, Wu JC, Lin F, Han R, Han F, Fukunaga K, Qin ZH (2009) Down-regulation of Bcl-2 enhances autophagy activation and cell death induced by mitochondrial dysfunction in rat striatum. J Neurosci Res 87:3600–3610
- Moreira PI, Zhu X, Wang X, Lee HG, Nunomura A, Petersen RB, Perry G, Smith MA (2009) Mitochondria: a therapeutic target in neurodegeneration. Biochim Biophys Acta 1802:212–220
- 46. Cui H, Chen B, Chicoine LG, Nelin LD (2011) Overexpression of cationic amino acid transporter-1 increases nitric oxide production in hypoxic human pulmonary microvascular endothelial cells. Clin Exp Pharmacol Physiol 38:796–803
- 47. Dumont M, Stack C, Elipenahli C, Jainuddin S, Gerges M, Starkova NN, Yang L, Starkov AA, Beal F (2011) Behavioral deficit, oxidative stress, and mitochondrial dysfunction precede tau pathology in P301S transgenic mice. FASEB J 25:4063–4072
- 48. Ho AK, Terriff DL, Price DM, Wloka MT, Chik CL (2007)
 The role of inducible repressor proteins in the adrenergic induction of arylalkylamine-N-acetyltransferase and mitogen-

- activated protein kinase phosphatase-1 in rat pinealocytes. Endocrinology 148:743–751
- Li X, Schulz E, Wenzel P, Munzel T, Daiber A (2014) Mitochondrial redox signaling: interaction of mitochondrial reactive oxygen species with other sources of oxidative stress. Antioxid Redox Signal 20:308–324
- Bhandari A, Holmes CP, Szardenings AK (2004) Alpha, alphadifluoro-beta-ketophosphonates as potent inhibitors of protein tyrosine phosphatase 1B. Bioorg Med Chem Lett 14:4301–4306
- Tibbles LA, Woodgett JR (1999) The stress-activated protein kinase pathways. Cell Mol Life Sci 55:1230–1254
- Fuentes F, Zimmer D, Atienza M, Schottenfeld J, Penkala I, Bale T, Bence KK, Arregui CO (2012) Protein tyrosine phosphatase PTP1B is involved in hippocampal synapse formation and learning. PLoS ONE 7:e41536
- 53. Evans DB, Rank KB, Bhattacharya K, Thomsen DR, Gurney ME, Sharma SK (2000) Tau phosphorylation at serine 396 and serine 404 by human recombinant tau protein kinase II inhibits tau's ability to promote microtubule assembly. J Biol Chem 275:24977–24983
- 54. Ivanov A, Pellegrino C, Rama S, Dumalska I, Salyha Y, Ben-Ari Y, Medina I (2006) Opposing role of synaptic and extrasynaptic NMDA receptors in regulation of the extracellular signal-regulated kinases (ERK) activity in cultured rat hippocampal neurons. J Physiol 572:789–798
- Leveille F, El Gaamouch F, Gouix E, Lecocq M, Lobner D, Nicole O, Buisson A (2008) Neuronal viability is controlled by a functional relation between synaptic and extrasynaptic NMDA receptors. FASEB J 22:4258–4271
- Kaufman AM, Milnerwood AJ, Sepers MD, Coquinco A, She K, Wang L, Lee H, Craig AM, Cynader M, Raymond LA (2012) Opposing roles of synaptic and extrasynaptic NMDA receptor signaling in cocultured striatal and cortical neurons. J Neurosci 32: 3992–4003
- 57. Zhou Q, Sheng M (2013) NMDA receptors in nervous system diseases. Neuropharmacology 74:69–75
- Ramirez M, Lamas M (2009) NMDA receptor mediates proliferation and CREB phosphorylation in postnatal Muller glia-derived retinal progenitors. Mol Vis 15:713

 –721
- Gomperts SN, Carroll R, Malenka RC, Nicoll RA (2000) Distinct roles for ionotropic and metabotropic glutamate receptors in the maturation of excitatory synapses. J Neurosci 20:2229–2237
- Bellone C, Nicoll RA (2007) Rapid bidirectional switching of synaptic NMDA receptors. Neuron 55:779–785
- Shankar GM, Bloodgood BL, Townsend M, Walsh DM, Selkoe DJ, Sabatini BL (2007) Natural oligomers of the Alzheimer amyloidbeta protein induce reversible synapse loss by modulating an NMDA-type glutamate receptor-dependent signaling pathway. J Neurosci 27:2866–2875
- Townsend M, Mehta T, Selkoe DJ (2007) Soluble Abeta inhibits specific signal transduction cascades common to the insulin receptor pathway. J Biol Chem 282:33305–33312
- 63. Wang Q, Rowan MJ, Anwyl R (2004) Beta-amyloid-mediated inhibition of NMDA receptor-dependent long-term potentiation induction involves activation of microglia and stimulation of inducible nitric oxide synthase and superoxide. J Neurosci 24:6049–6056
- Kumar A, Foster TC (2004) Enhanced long-term potentiation during aging is masked by processes involving intracellular calcium stores. J Neurophysiol 91:2437–2444
- Reese LC, Zhang W, Dineley KT, Kayed R, Taglialatela G (2008) Selective induction of calcineurin activity and signaling by oligomeric amyloid beta. Aging Cell 7:824

 –835
- Phiel CJ, Wilson CA, Lee VM, Klein PS (2003) GSK-3alpha regulates production of Alzheimer's disease amyloid-beta peptides. Nature 423:435–439
- Hooper C, Killick R, Lovestone S (2008) The GSK3 hypothesis of Alzheimer's disease. J Neurochem 104:1433–1439



- Hansen T, Rehfeld JF, Nielsen FC (2004) GSK-3beta reduces cAMP-induced cholecystokinin gene expression by inhibiting CREB binding. Neuroreport 15:841–845
- Koivisto L, Hakkinen L, Matsumoto K, McCulloch CA, Yamada KM, Larjava H (2004) Glycogen synthase kinase-3 regulates cytoskeleton and translocation of Rac1 in long cellular extensions of human keratinocytes. Exp Cell Res 293:68–80
- Thornton TM, Pedraza-Alva G, Deng B, Wood CD, Aronshtam A, Clements JL, Sabio G, Davis RJ, Matthews DE, Doble B, Rincon M (2008) Phosphorylation by p38 MAPK as an alternative pathway for GSK3beta inactivation. Science 320:667–670
- Peineau S, Taghibiglou C, Bradley C, Wong TP, Liu L, Lu J, Lo E, Wu D, Saule E, Bouschet T, Matthews P, Isaac JT, Bortolotto ZA, Wang YT, Collingridge GL (2007) LTP inhibits LTD in the hippocampus via regulation of GSK3beta. Neuron 53:703–717
- Hernandez F, Gomez de Barreda E, Fuster-Matanzo A, Lucas JJ, Avila J (2010) GSK3: a possible link between beta amyloid peptide and tau protein. Exp Neurol 223:322–325
- Kim WY, Wang X, Wu Y, Doble BW, Patel S, Woodgett JR, Snider WD (2009) GSK-3 is a master regulator of neural progenitor homeostasis. Nat Neurosci 12:1390–1397
- 74. Liu GP, Zhang Y, Yao XQ, Zhang CE, Fang J, Wang Q, Wang JZ (2008) Activation of glycogen synthase kinase-3 inhibits protein phosphatase-2A and the underlying mechanisms. Neurobiol Aging 29:1348–1358
- Hetman M, Cavanaugh JE, Kimelman D, Xia Z (2000) Role of glycogen synthase kinase-3beta in neuronal apoptosis induced by trophic withdrawal. J Neurosci 20:2567–2574
- Rojo AI, Sagarra MR, Cuadrado A (2008) GSK-3beta down-regulates the transcription factor Nrf2 after oxidant damage: relevance to exposure of neuronal cells to oxidative stress. J Neurochem 105: 192–202
- Jouvenceau A, Dutar P (2006) A role for the protein phosphatase 2B in altered hippocampal synaptic plasticity in the aged rat. J Physiol Paris 99:154–161
- Kamsler A, Segal M (2004) Hydrogen peroxide as a diffusible signal molecule in synaptic plasticity. Mol Neurobiol 29:167–178
- Lin CH, Yeh SH, Leu TH, Chang WC, Wang ST, Gean PW (2003) Identification of calcineurin as a key signal in the extinction of fear memory. J Neurosci 23:1574–1579
- Serrano F, Klann E (2004) Reactive oxygen species and synaptic plasticity in the aging hippocampus. Ageing Res Rev 3:431

 –443
- Chen QS, Wei WZ, Shimahara T, Xie CW (2002) Alzheimer amyloid beta-peptide inhibits the late phase of long-term potentiation through calcineurin-dependent mechanisms in the hippocampal dentate gyrus. Neurobiol Learn Mem 77:354–371
- Ermak G, Morgan TE, Davies KJ (2001) Chronic overexpression of the calcineurin inhibitory gene DSCR1 (Adapt78) is associated with Alzheimer's disease. J Biol Chem 276:38787–38794
- 83. Cook CN, Hejna MJ, Magnuson DJ, Lee JM (2005) Expression of calcipressin1, an inhibitor of the phosphatase calcineurin, is altered with aging and Alzheimer's disease. J Alzheimers Dis 8:63–73
- Koss DJ, Hindley KP, Riedel G, Platt B (2007) Modulation of hippocampal calcium signalling and plasticity by serine/threonine protein phosphatases. J Neurochem 102:1009–1023
- Lu Y, Rosenberg PA (2007) NMDA receptor-mediated extracellular adenosine accumulation is blocked by phosphatase 1/2A inhibitors. Brain Res 1155:116–124
- Butterfield DA, Pocernich CB (2003) The glutamatergic system and Alzheimer's disease: therapeutic implications. CNS Drugs 17:641– 652
- Ballatore C, Lee VM, Trojanowski JQ (2007) Tau-mediated neurodegeneration in Alzheimer's disease and related disorders. Nat Rev Neurosci 8:663–672
- Hoover BR, Reed MN, Su J, Penrod RD, Kotilinek LA, Grant MK,
 Pitstick R, Carlson GA, Lanier LM, Yuan LL, Ashe KH, Liao D

- (2012) Tau mislocalization to dendritic spines mediates synaptic dysfunction independently of neurodegeneration. Neuron 68:1067–1081
- Li YY, Cui JG, Hill JM, Bhattacharjee S, Zhao Y and Lukiw WJ (2011) Increased expression of miRNA-146a in Alzheimer's disease transgenic mouse models. Neurosci Lett 487:94–98.
- McKinney RA (2010) Excitatory amino acid involvement in dendritic spine formation, maintenance and remodelling. J Physiol 588: 107–116
- El-Husseini AE, Schnell E, Chetkovich DM, Nicoll RA, Bredt DS (2000) PSD-95 involvement in maturation of excitatory synapses. Science 290:1364–1368
- Mondragon-Rodriguez S, Trillaud-Doppia E, Dudilot A, Bourgeois C, Lauzon M, Leclerc N, Boehm J (2012) Interaction of endogenous tau protein with synaptic proteins is regulated by N-methyl-D-aspartate receptor-dependent tau phosphorylation. J Biol Chem 287:32040–32053
- Moreno H, Choi S, Yu E, Brusco J, Avila J, Moreira JE, Sugimori M, Llinas RR (2011) Blocking effects of human tau on squid giant synapse transmission and its prevention by T-817 MA. Front Synaptic Neurosci 3:3
- Ittner LM, Gotz J (2011) Amyloid-beta and tau—a toxic pas de deux in Alzheimer's disease. Nat Rev Neurosci 12:65–72
- 95. Zhang L, Li YH, Meng K, Xie W (2010) The expressions of AMPAR/GluR2 in hippocampal CA1 area of rats before and after late-phase long-term potentiation reversal. Sheng Li Xue Bao 62: 23–29
- Zhang H, Wu CY, Wang W, Harrington MA (2011) Interneuronal synapses formed by motor neurons appear to be glutamatergic. Neuroreport 22:809–813
- Massey PV, Johnson BE, Moult PR, Auberson YP, Brown MW, Molnar E, Collingridge GL, Bashir ZI (2004) Differential roles of NR2A and NR2B-containing NMDA receptors in cortical longterm potentiation and long-term depression. J Neurosci 24:7821– 7828
- Perez-Otano I, Ehlers MD (2004) Learning from NMDA receptor trafficking: clues to the development and maturation of glutamatergic synapses. Neurosignals 13:175–189
- Desilva TM, Kinney HC, Borenstein NS, Trachtenberg FL, Irwin N, Volpe JJ, Rosenberg PA (2007) The glutamate transporter EAAT2 is transiently expressed in developing human cerebral white matter. J Comp Neurol 501:879–890
- Ascher P, Nowak L (2009) Early biophysics of the NMDA receptor channel. J Physiol 587:4563

 –4564
- Bleich S, Sperling W, Wiltfang J, Maler JM, Kornhuber J (2003)
 Excitatory neurotransmission in alcoholism. Fortschr Neurol Psychiatr 71:S36–S44
- 102. Adams JP, Sweatt JD (2002) Molecular psychology: roles for the ERK MAP kinase cascade in memory. Annu Rev Pharmacol Toxicol 42:135–163
- Foster TC, Kumar A (2002) Calcium dysregulation in the aging brain. Neuroscientist 8:297–301
- 104. Busche MA, Eichhoff G, Adelsberger H, Abramowski D, Wiederhold KH, Haass C, Staufenbiel M, Konnerth A, Garaschuk O (2008) Clusters of hyperactive neurons near amyloid plaques in a mouse model of Alzheimer's disease. Science 321:1686–1689
- 105. Kuchibhotla KV, Goldman ST, Lattarulo CR, Wu HY, Hyman BT, Bacskai BJ (2008) Abeta plaques lead to aberrant regulation of calcium homeostasis in vivo resulting in structural and functional disruption of neuronal networks. Neuron 59:214–225
- 106. Snyder EM, Nong Y, Almeida CG, Paul S, Moran T, Choi EY, Nairn AC, Salter MW, Lombroso PJ, Gouras GK, Greengard P (2005) Regulation of NMDA receptor trafficking by amyloid-beta. Nat Neurosci 8:1051–1058
- 107. Hsieh H, Boehm J, Sato C, Iwatsubo T, Tomita T, Sisodia S, Malinow R (2006) AMPAR removal underlies Abeta-induced synaptic depression and dendritic spine loss. Neuron 52:831–843



- 108. Souza MA, Magni DV, Guerra GP, Oliveira MS, Furian AF, Pereira L, Marquez SV, Ferreira J, Fighera MR, Royes LF (2012) Involvement of hippocampal CAMKII/CREB signaling in the spatial memory retention induced by creatine. Amino Acids 43:2491–2503
- 109. Vierci G, Oliveira CS, Perera LR, Bornia N, Leal RB, Rossi FM (2013) CREB is modulated in the mouse superior colliculus in developmental and experimentally-induced models of plasticity. Int J Dev Neurosci 31:46–52
- Bito H, Deisseroth K, Tsien RW (1996) CREB phosphorylation and dephosphorylation: a Ca(2+)- and stimulus duration-dependent switch for hippocampal gene expression. Cell 87:1203–1214
- Hardingham GE, Chawla S, Johnson CM, Bading H (1997) Distinct functions of nuclear and cytoplasmic calcium in the control of gene expression. Nature 385:260–265
- 112. Vitolo OV, Sant'Angelo A, Costanzo V, Battaglia F, Arancio O, Shelanski M (2002) Amyloid beta-peptide inhibition of the PKA/CREB pathway and long-term potentiation: reversibility by drugs that enhance cAMP signaling. Proc Natl Acad Sci U S A 99:13217–13221
- 113. Li XY, Zhan XR, Liu XM, Wang XC (2011) CREB is a regulatory target for the protein kinase Akt/PKB in the differentiation of pancreatic ductal cells into islet beta-cells mediated by hepatocyte growth factor. Biochem Biophys Res Commun 404:711–716
- 114. Arlt S, Müller-Thomsen TM, Beisiegel U, Kontush A (2012) Effect of one-year vitamin C- and E-supplementation on cerebrospinal fluid oxidation parameters and clinical course in Alzheimer's disease. Neurochem Res 37:2706–2714
- 115. Galasko DR, Peskind E, Clark CM, Quinn JF, Ringman JM, Jicha GA, Cotman C, Cottrell B, Montine TJ, Thomas RG et al (2012) Antioxidants for Alzheimer disease: a randomized clinical trial with cerebrospinal fluid biomarker measures. Arch Neurol 69:836–841
- 116. Bagi Z, Cseko C, Toth E, Koller A (2003) Oxidative stress-induced dysregulation of arteriolar wall shear stress and blood pressure in hyperhomocysteinemia is prevented by chronic vitamin C treatment. Am J Physiol Heart Circ Physiol 285:2277–2283
- Nadeau A, Roberge AG (1988) Effects of vitamin B12 supplementation on choline acetyltransferase activity in cat brain. Int J Vitam Nutr Res 58:402–406
- Ikeda T, Yamamoto K, Takahashi K (1992) Treatment of Alzheimertype dementia with intravenous methylcobalamin. Clin Ther 14: 426–437
- Sano M, Ernesto C, Thomas RG et al (1997) A controlled trial of selegiline, alpha-tocopherol, or both as treatment for Alzheimer's disease. N Engl J Med 336:1216–1222
- 120. Sung S, Yao Y, Uryu K et al (2004) Early vitamin E supplementation in young but not aged mice reduces Abeta levels and amyloid deposition in a transgenic model of Alzheimer's disease. FASEB J 18:323–325
- 121. Nakashima H, Ishihara T, Yokota O et al (2004) Effects of α -tocopherol on an animal model of tauopathies. Free Radic Biol Med 37:176–186
- Dias-Santagata D, Fulga TA, Duttaroy A, Feany MB (2007)
 Oxidative stress mediates tau-induced neurodegeneration in Drosophila. J Clin Investig 117:236–245
- 123. Pavlik VN, Doody RS, Rountree SD, Darby EJ (2009) Vitamin E use is associated with improved survival in an Alzheimer's disease cohort. Dement Geriatr Cogn Disord 28:536–540
- 124. Bolognesi ML, Bergamini C, Fato R, Oiry J, Vasseur JJ, Smietana M (2014) Synthesis of new lipoic acid conjugates and evaluation of their free radical scavenging and neuroprotective activities. Chem Biol Drug Des 83:688–696
- 125. Sancheti H, Kanamori K, Patil I, Díaz Brinton R, Ross BD, Cadenas E (2014) Reversal of metabolic deficits by lipoic acid in a triple transgenic mouse model of Alzheimer's disease: a 13C NMR study. J Cereb Blood Flow Metab 34:288–296

- 126. Kryscio RJ, Abner EL, Schmitt FA et al (2013) A randomized controlled Alzheimer's disease prevention trial's evolution into an exposure trial: the PREADViSE trial. J Nutr Health Aging 17:72–75
- 127. Esposito L, Raber J, Kekonius L et al (2006) Reduction in mitochondrial superoxide dismutase modulates Alzheimer's disease-like pathology and accelerates the onset of behavioral changes in human amyloid precursor protein transgenic mice. J Neurosci 26:5167– 5179
- 128. Rajasekar N, Dwivedi S, Tota SK, Kamat PK, Hanif K, Nath C, Shukla R (2013) Neuroprotective effect of curcumin on okadaic acid induced memory impairment in mice. Eur J Pharmacol 715: 381–394
- 129. Ringman M, Cole GM, Tend E (2008) Oral curcumin for the treatment of mild-to-moderate Alzheimer's disease: tolerability and clinical and biomarker efficacy results of a placebo-controlled 24-week study. Proceedings of the International Conference of Alzheimer's disease Chicago, abstract.
- 130. Remington R, Chan A, Paskavitz J, Shea TB (2009) Efficacy of a vitamin/nutriceutical formulation for moderate-stage to later-stage Alzheimer's disease: a placebo-controlled pilot study. Am J Alzheimers Dis Other Demen 24:27–33
- 131. Chan A, Remington R, Kotyla E, Lepore A, Zemianek J, Shea TB (2010) A vitamin/nutriceutical formulation improves memory and cognitive performance in community-dwelling adults without dementia. J Nutr Health Aging 14:224–230
- 132. Gutzmann H, Hadler D (1998) Sustained efficacy and safety of idebenone in the treatment of Alzheimer's disease: update on a two-year double-blind multicentre study. J Neural Transm Suppl 54:301–310
- 133. Thal LJ, Grundman M, Berg J, Ernstrom K, Margolin R, Pfeiffer E, Weiner MFF, Zamrini E, Thomas RG (2003) Idebenone treatment fails to slow cognitive decline in Alzheimer's disease. Neurology 61:1498–1502
- 134. Weyer G, Babej-Dölle RM, Hadler D, Hoffmann S, Hermann WM (1997) A controlled study of 2 doses of idebenone in the treatment of Alzheimer's disease. Neuropsychobiology 36:73–82
- 135. Mulnard RA, Cotman CW, Kawas C et al (2000) Estrogen replacement therapy for treatment of mild to moderate Alzheimer disease: a randomized controlled trial. J Am Med Assoc 283:1007–1015
- Singh S, Thakur MK (2011) Gonadal steroids do not affect apolipoprotein E expression in aging mouse cerebral cortex. Cell Mol Neurobiol 31:401–405
- Boldogh I, Liebenthal D, Hughes TK et al (2003) Modulation of 4HNE-mediated signaling by proline-rich peptides from ovine colostrum. J Mol Neurosci 20:125–134
- Zabłocka A, Janusz M (2012) Effect of the proline-rich polypeptide complex/colostrininTM on the enzymatic antioxidant system. Arch Immunol Ther Exp 60:383–390
- 139. Leszek J, Inglot AD, Janusz M et al (2002) Colostrinin proline-rich polypeptide complex fromovine colostrum—a long-term study of its efficacy in Alzheimer's disease. Med Sci Monit 8:193–196
- 140. Bilikiewicz A, Gaus W (2004) Colostrinin1 (a naturally occurring proline-rich, polypeptide mixture) in the treatment of Alzheimer's disease. J Alzheimers Dis 6:17–26
- 141. Jia N, Han K, Kong JJ, Zhang XM, Sha S, Ren GR, Cao YP (2013) (-)-Epigallocatechin-3-gallate alleviates spatial memory impairment in APP/PS1 mice by restoring IRS-1 signaling defects in the hippocampus. Mol Cell Biochem 380:211–218
- 142. Li SY, Wang XB, Kong LY (2014) Design, synthesis and biological evaluation of imine resveratrol derivatives as multi-targeted agents against Alzheimer's disease. Eur J Med Chem 71:36–45
- 143. Lu C, Guo Y, Yan J, Luo Z, Luo HB, Yan M, Huang L, Li X (2013) Design, synthesis, and evaluation of multitarget-directed resveratrol derivatives for the treatment of Alzheimer's disease. J Med Chem 56:5843–5859



- 144. Jones RW (2010) Dimebon disappointment. Alzheimers Res Ther 2:25
- 145. Steele JW, Gandy S (2013) Latrepirdine (Dimebon®), a potential Alzheimer therapeutic, regulates autophagy and neuropathology in an Alzheimer mouse model. Autophagy 9:617–818
- 146. Bharadwaj PR, Bates KA, Porter T, Teimouri E, Perry G, Steele JW, Gandy S, Groth D, Martins RN, Verdile G (2013) Latrepirdine: molecular mechanisms underlying potential therapeutic roles in Alzheimer's and other neurodegenerative diseases. Transl Psychiatry 3:e332
- 147. Shinto L, Quinn J, Montine T, Dodge HH, Woodward W, Baldauf-Wagner S, Waichunas D, Bumgarner L, Bourdette D, Silbert L, Kaye J (2014) A randomized placebo-controlled pilot trial of omega-3 fatty acids and alpha lipoic acid in Alzheimer's disease. J Alzheimers Dis 38:111–120
- 148. Hardas SS, Sultana R, Clark AM, Beckett TL, Szweda LI, Murphy MP, Butterfield DA (2013) Oxidative modification of lipoic acid by HNE in Alzheimer disease brain. Redox Biol 1:80–85
- 149. Quinn JF, Bussiere JR, Hammond RS et al (2007) Chronic dietary α-lipoic acid reduces deficits in hippocampal memory of aged Tg2576 mice. Neurobiol Aging 28:213–225
- Lu P, Mamiya T, Lu LL et al (2009) Silibinin prevents amyloid b peptide-induced memory impairment and oxidative stress in mice. Br J Pharmacol 157:1270–1277
- 151. Tota S, Kamat PK, Shukla R, Nath C (2011) Improvement of brain energy metabolism and cholinergic functions contributes to the beneficial effects of silibinin against streptozotocin induced memory impairment. Behav Brain Res 221:207–215
- 152. Tota S, Awasthi H, Kamat PK, Nath C, Hanif K (2010) Protective effect of quercetin against intracerebral streptozotocin induced reduction in cerebral blood flow and impairment of memory in mice. Behav Brain Res 209:73–79
- 153. Feng Z, Zhang JT (2004) Protective effect of melatonin on β-amyloid-induced apoptosis in rat astroglioma c6 cells and its mechanism. Free Radic Biol Med 37:1790–1801

- 154. Matsubara E, Bryant-Thomas T, Quinto JP et al (2003) Melatonin increases survival and inhibits oxidative and amyloid pathology in a transgenic model of Alzheimer's disease. J Neurochem 85:1101– 1108
- 155. Saxena G, Bharti S, Kamat PK, Sharma S, Nath C (2010) Melatonin alleviates memory deficits and neuronal degeneration induced by intracerebroventricular administration of streptozotocin in rats. Pharmacol Biochem Behav 94:397–403
- 156. Prasanthi JRP, Dasari B, Marwarha G et al (2010) Caffeine protects against oxidative stress and Alzheimer's disease-like pathology in rabbit hippocampus induced by cholesterol-enriched diet. Free Radic Biol Med 49:1212–1220
- 157. Andrieu S, Ousset PJ, Coley N, Ouzid M, Mathiex-Fortunet H, Vellas B, The GuidAge study Group (2008) GuidAge study: a 5-year double-blind, randomized trial of EGb761 for the prevention of Alzheimer's disease in elderly subjects with memory complaints rationale, design and baseline data. Curr Alzheimer Res 5:406–415
- 158. Vellas B, Coley N, Ousset PJ et al (2012) Long-term use of standardized ginkgo biloba extract for the prevention of Alzheimer' disease (GuidAge): a randomised placebo-controlled trial. Lancet Neurol 11:851–859
- 159. Snitz BE, O'Meara ES, Carlson MC, Arnold AM, Ives DG, Rapp SR, Saxton J, Lopez OL, Dunn LO, Sink KM, DeKosky ST, Ginkgo Evaluation of Memory (GEM) Study Investigators (2009) Ginkgo biloba for preventing cognitive decline in older adults: a randomized trial. JAMA 302:2663–2670
- 160. Saxena G, Singh SP, Pal R, Singh S, Pratap R, Nath C (2007) Gugulipid, an extract of Commiphora whighitii with lipidlowering properties, has protective effects against streptozotocininduced memory deficits in mice. Pharmacol Biochem Behav 86: 797–805
- 161. Niranjan R, Kamat PK, Nath C, Shukla R (2010) Evaluation of guggulipid and nimesulide on production of inflammatory mediators and GFAP expression in LPS stimulated rat astrocytoma, cell line (C6). J Ethnopharmacol 127:625–630

