Scripps's Murrelet Egg Size Prediction

Amelia J. DuVall & Marcela Todd

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Introduction

This document details steps taken to predict Scripps's Murrelet (*Synthliboramphus scrippsi*) egg size at Santa Barbara Island within Channel Islands National Park using top model results.

This is v.2021-07-24

Prediction

Model prediction for the 2 top models

Run the top models and create a table of AIC scores.

[2,] 10559.64 1.372339 0.3348858

```
topmod1 <- lmer(Size ~ EggOrder + BEUTI + NPGO + (1 | Plot), data = SCMUdf, REML = TRUE)
topmod2 <- lmer(Size ~ EggOrder + NPGO + ONI + (1 | Plot), data = SCMUdf, REML = TRUE)

# AIC tab for 2 top models
AIC.tab2 <- matrix(NA, nrow = 2, ncol = 3)
AIC.tab2[1,1] <- AIC(topmod1)
AIC.tab2[2,1] <- AIC(topmod2)
AIC.tab2[,2] <- AIC.tab2[,1] - min(AIC.tab2[,1])
AIC.tab2[,3] <- exp(-0.5*AIC.tab2[,2])/(sum(exp(-0.5*AIC.tab2[,2])))
print(AIC.tab2)

## [,1] [,2] [,3]
## [1,] 10558.27 0.000000 0.6651142</pre>
```

We want predictions with all predictors but one held constant, and then we'll do this one-at-a-time for each of the predictors. This will allow us to produce plots looking at the marginal effect of each predictor. Set-up a dataset with everything at its mean value (except Egg Order).

```
set.seed(1)
SCMU.null <- SCMUdf[c(1:200),]
SCMU.null$EggOrder <- c(rep("Egg1",100),rep("Egg2",100)) # the first 100 eggs with be Egg
SCMU.null$BEUTI <- mean(SCMUdf$BEUTI)
SCMU.null$NPGO <- mean(SCMUdf$NPGO)
SCMU.null$ONI <- mean(SCMUdf$ONI)
SCMU.null$ANCHL <- mean(SCMUdf$ANCHL)
SCMU.null$PDO <- mean(SCMUdf$PDO)
SCMU.null$PDO <- mean(SCMUdf$PDO)
SCMU.null$PDO <- mean(SCMUdf$PDO)</pre>
```

```
## look at min/max covariate values
summary(SCMUdf$BEUTI)
##
      Min. 1st Qu.
                      Median
                                 Mean 3rd Qu.
                                                   Max.
## -1.76150 -0.48909 0.02571 -0.12028 0.39502 1.51698
summary(SCMUdf$NPGO)
      Min. 1st Qu.
                      Median
                                 Mean 3rd Qu.
                                                   Max.
## -1.61193 -0.76135
                     0.52847 0.02653 1.10971
                                               1.39094
summary(SCMUdf$ONI)
##
      Min. 1st Qu.
                      Median
                                 Mean 3rd Qu.
                                                   Max.
## -1.34238 -0.49482 -0.35178 0.07779 0.78612 1.72334
```

Model prediciton for 2 top models

boundary (singular) fit: see ?isSingular

BEUTI

Vary just one predictor at a time for prediction. For example, create the dataset SCMU. BEUTI that will be used for predictions where the only thing that values is BEUTI.

```
SCMU.BEUTI <- SCMU.null
SCMU.BEUTI$BEUTI <- rep(seq(from = -2, to = 2, length.out = 100),2)
# SCMU.BEUTI$BEUTI <- rep(seq(from = min(SCMUdf$BEUTI), to = max(SCMUdf$BEUTI), length.out = 100),2)
#PREDICT FOR SCMU.BEUTI
plot.sd.1 <- as.data.frame(VarCorr(topmod1))$sdcor[1]</pre>
resid.sd.1 <- as.data.frame(VarCorr(topmod1))$sdcor[2]</pre>
plot.sd.2 <- as.data.frame(VarCorr(topmod2))$sdcor[1]</pre>
resid.sd.2 <- as.data.frame(VarCorr(topmod2))$sdcor[2]</pre>
sims <- 10000 # run 100 to check, otherwise update to 10,000
## model prediction for 2 top models
pv.BEUTI <- matrix(nrow = sims, ncol = nrow(SCMU.BEUTI))</pre>
for(i in 1:sims){
  #choose a model
  model \leftarrow which(rmultinom(n = 1, size = 1, prob = c(AIC.tab2[,3]))==1)
  if(model == 1){
    #we simulate conditioning on no specific random effects levels
    y <- unlist(simulate(topmod1))</pre>
    bmod <- refit(topmod1,y)</pre>
    pv.BEUTI[i,] <- predict(bmod, re.form = ~0, newdata = SCMU.BEUTI) + rnorm(1,0,sd=plot.sd.1) + rnorm
  }
  if(model == 2){
    #we simulate conditioning on no specific random effects levels
    y <- unlist(simulate(topmod2))</pre>
    bmod <- refit(topmod2,y)</pre>
    pv.BEUTI[i,] <- predict(bmod, re.form = ~0, newdata = SCMU.BEUTI) + rnorm(1,0,sd=plot.sd.2) + rnorm
  }
}
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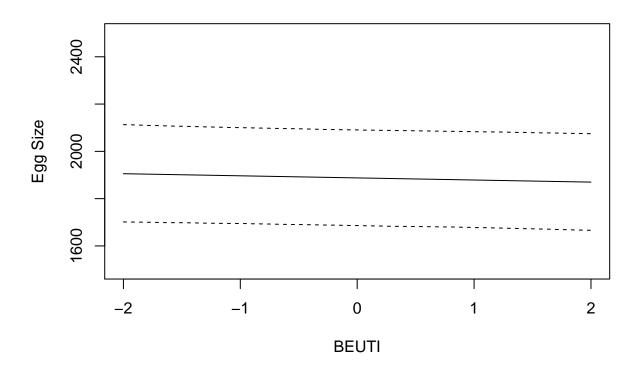
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```
## boundary (singular) fit: see ?isSingular
## check plot
plot(SCMU.BEUTI$BEUTI[1:100],y = apply(pv.BEUTI,2,mean)[1:100],xlab = "BEUTI",ylab = "Egg Size",type = lines(SCMU.BEUTI$BEUTI[1:100],y = apply(pv.BEUTI,2,function(x)quantile(x,probs= 0.025))[1:100],lty=2)
lines(SCMU.BEUTI$BEUTI[1:100],y = apply(pv.BEUTI,2,function(x)quantile(x,probs= 0.975))[1:100],lty=2)
```



NPGO

```
SCMU.NPGO <- SCMU.null
SCMU.NPGO$NPGO <- rep(seq(from = -2, to = 2, length.out = 100),2)
# SCMU.NPGO$NPGO <- rep(seq(from = min(SCMUdf$NPGO), to = max(SCMUdf$NPGO), length.out = 100),2)

plot.sd.1 <- as.data.frame(VarCorr(topmod1))$sdcor[1]
resid.sd.1 <- as.data.frame(VarCorr(topmod1))$sdcor[2]

plot.sd.2 <- as.data.frame(VarCorr(topmod2))$sdcor[1]
resid.sd.2 <- as.data.frame(VarCorr(topmod2))$sdcor[2]

sims <- 10000 # run 100 to check, otherwise update to 10,000
```

```
pv.NPGO <- matrix(nrow = sims, ncol = nrow(SCMU.NPGO))</pre>
for(i in 1:sims){
  #choose a model
  model \leftarrow which(rmultinom(n = 1, size = 1, prob = c(AIC.tab2[,3])) == 1)
  if(model == 1){
    #we simulate conditioning on no specific random effects levels
    y <- unlist(simulate(topmod1))</pre>
    bmod <- refit(topmod1,y)</pre>
    pv.NPGO[i,] <- predict(bmod, re.form = ~0, newdata = SCMU.NPGO) + rnorm(1,0,sd=plot.sd.1) + rnorm(1
  if(model == 2){
    #we simulate conditioning on no specific random effects levels
    y <- unlist(simulate(topmod2))</pre>
    bmod <- refit(topmod2,y)</pre>
    pv.NPGO[i,] <- predict(bmod, re.form = ~0, newdata = SCMU.NPGO) + rnorm(1,0,sd=plot.sd.2) + rnorm(1
  }
}
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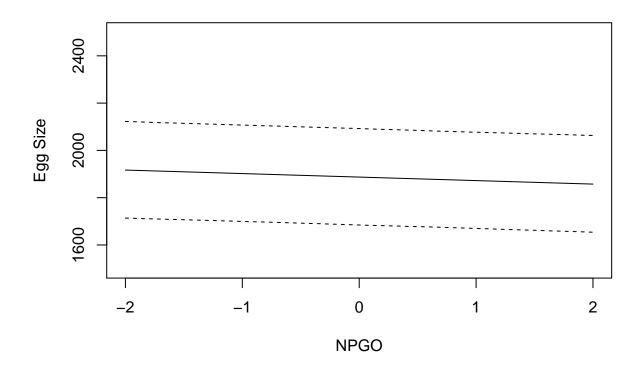
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## boundary (singular) fit: see ?isSingular
## check plots
plot(SCMU.NPGO$NPGO[1:100],y = apply(pv.NPGO,2,mean)[1:100],xlab = "NPGO",ylab = "Egg Size",type =
lines(SCMU.NPGO$NPGO[1:100],y = apply(pv.NPGO,2,function(x)quantile(x,probs= 0.025))[1:100],lty=2)
lines(SCMU.NPGO$NPGO[1:100],y = apply(pv.NPGO,2,function(x)quantile(x,probs= 0.975))[1:100],lty=2)
```



ONI

```
SCMU.ONI <- SCMU.null
SCMU.ONI\$ONI <- rep(seq(from = -2, to = 2, length.out = 100),2)
# SCMU.ONI$ONI <- rep(seq(from = min(SCMUdf$ONI), to = max(SCMUdf$ONI), length.out = 100),2)
plot.sd.1 <- as.data.frame(VarCorr(topmod1))$sdcor[1]</pre>
resid.sd.1 <- as.data.frame(VarCorr(topmod1))$sdcor[2]</pre>
plot.sd.2 <- as.data.frame(VarCorr(topmod2))$sdcor[1]</pre>
resid.sd.2 <- as.data.frame(VarCorr(topmod2))$sdcor[2]</pre>
sims <- 10000 # run 100 to check, otherwise update to 10,000
pv.ONI <- matrix(nrow = sims, ncol = nrow(SCMU.ONI))</pre>
for(i in 1:sims){
  #choose a model
  model \leftarrow which(rmultinom(n = 1, size = 1, prob = c(AIC.tab2[,3]))==1)
  if(model == 1){
    #we simulate conditioning on no specific random effects levels
    y <- unlist(simulate(topmod1))</pre>
    bmod <- refit(topmod1,y)</pre>
    pv.ONI[i,] <- predict(bmod, re.form = ~0, newdata = SCMU.ONI) + rnorm(1,0,sd=plot.sd.1) + rnorm(1,0
  if(model == 2){
```

```
#we simulate conditioning on no specific random effects levels
    y <- unlist(simulate(topmod2))</pre>
    bmod <- refit(topmod2,y)</pre>
    pv.ONI[i,] <- predict(bmod, re.form = ~0, newdata = SCMU.ONI) + rnorm(1,0,sd=plot.sd.2) + rnorm(1,0
  }
}
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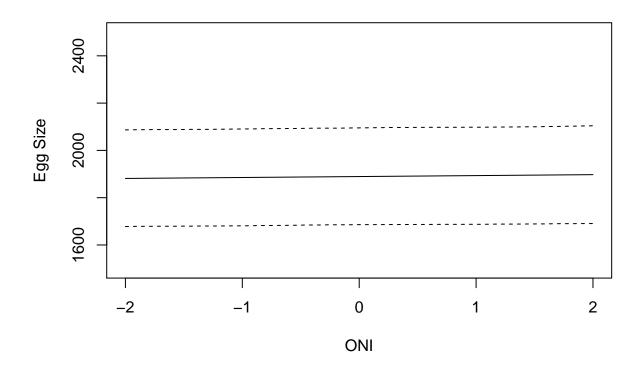
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## boundary (singular) fit: see ?isSingular
## check plots
plot(SCMU.ONI$ONI[1:100],y = apply(pv.ONI,2,mean)[1:100],xlab = "ONI",ylab = "Egg Size",type = "l",ylim
lines(SCMU.ONI$ONI[1:100],y = apply(pv.ONI,2,function(x)quantile(x,probs= 0.025))[1:100],lty=2)
lines(SCMU.ONI$ONI[1:100], y = apply(pv.ONI,2,function(x)quantile(x,probs= 0.975))[1:100], lty=2)
```



Plots

```
# Updated plot - include both egg order
\# plot(SCMU.BEUTI$BEUTI[1:100], y = apply(pv.BEUTI,2,mean)[1:100], xlab = "BEUTI",ylab = "Egg Size",type
\#\ lines(SCMU.BEUTI \$BEUTI [101:200], y = apply(pv.BEUTI, 2, mean)[101:200], col = "red")
# lines(SCMU.BEUTI$BEUTI[101:200], y = apply(pv.BEUTI, 2, function(x) quantile(x, probs=0.975))[101:200], lt
## ggplot
color_pal <- c("#593d9cff", "#f68f46ff")</pre>
### BEUTI
y1 <- as.data.frame(apply(pv.BEUTI,2,mean))</pre>
quantile.low1 <- as.data.frame(apply(pv.BEUTI,2,function(x)quantile(x,probs= 0.025)))
quantile.high1 <- as.data.frame(apply(pv.BEUTI,2,function(x)quantile(x,probs= 0.975)))</pre>
BEUTI.pred <- cbind(y1, quantile.low1, quantile.high1, SCMU.BEUTI$BEUTI) %>%
  mutate(EggOrder = c(rep("Egg1",100),rep("Egg2",100)))
colnames(BEUTI.pred) <- c("mean", "low", "high", "BEUTI", "egg_order")</pre>
p1 <- ggplot(data = BEUTI.pred) +</pre>
  geom_line(aes(x = BEUTI, y = mean, color = egg_order)) +
  geom_ribbon(aes(ymin = low, ymax = high, x = BEUTI, fill = egg_order), alpha = 0.25, linetype = "dash
 ylim(c(1500, 2500)) +
  scale_color_manual(values = color_pal) +
  scale_fill_manual(values = color_pal) +
```

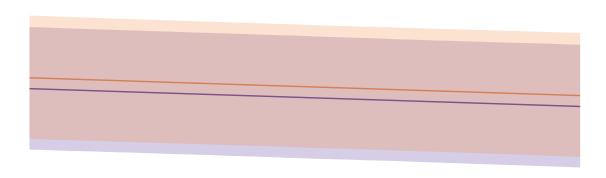
```
ylab("Egg Size") +
  theme_minimal() +
  theme(panel.grid.major = element_blank(),
        panel.grid.minor = element_blank(),
        legend.position = "none")
р1
   2500
   2250
   2000
   1750
   1500
           -2
                                                                                       2
                                                                    1
                              -1
                                              BEUTI
### NPGO
y2 <- as.data.frame(apply(pv.NPGO,2,mean))</pre>
quantile.low2 <- as.data.frame(apply(pv.NPGO,2,function(x)quantile(x,probs= 0.025)))
quantile.high2 <- as.data.frame(apply(pv.NPGO,2,function(x)quantile(x,probs= 0.975)))
NPGO.pred <- cbind(y2, quantile.low2, quantile.high2, SCMU.NPGO$NPGO) %>%
  mutate(EggOrder = c(rep("Egg1",100),rep("Egg2",100)))
colnames(NPGO.pred) <- c("mean", "low", "high", "NPGO", "egg_order")</pre>
p2 <- ggplot(data = NPGO.pred) +
  geom_line(aes(x = NPGO, y = mean, color = egg_order)) +
  geom_ribbon(aes(ymin = low, ymax = high, x = NPGO, fill = egg_order), alpha = 0.25, linetype = "dashe
```

ylim(c(1500, 2500)) +

ylab("Egg Size") +
theme_minimal() +
guides(size = FALSE) +

scale_color_manual(values = color_pal) +
scale_fill_manual(values = color_pal) +

```
axis.title.y = element_blank(),
axis.text.y = element_blank(),
axis.ticks.y = element_blank(),
legend.title = element_blank(),
legend.position = "bottom")
```

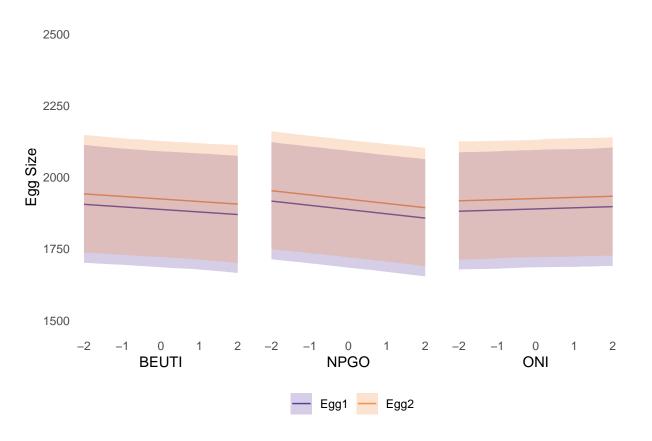


```
### ONI
y3 <- as.data.frame(apply(pv.ONI,2,mean))
quantile.low3 <- as.data.frame(apply(pv.ONI,2,function(x)quantile(x,probs= 0.025)))
quantile.high3 <- as.data.frame(apply(pv.ONI,2,function(x)quantile(x,probs= 0.975)))</pre>
ONI.pred <- cbind(y3, quantile.low3, quantile.high3, SCMU.ONI$ONI) %>%
  mutate(EggOrder = c(rep("Egg1",100),rep("Egg2",100)))
colnames(ONI.pred) <- c("mean", "low", "high", "ONI", "egg_order")</pre>
p3 <- ggplot(data = ONI.pred) +
  geom_line(aes(x = ONI, y = mean, color = egg_order)) +
  geom_ribbon(aes(ymin = low, ymax = high, x = ONI, fill = egg_order), alpha = 0.25, linetype = "dashed
  ylim(c(1500, 2500)) +
  scale_color_manual(values = color_pal) +
  scale_fill_manual(values = color_pal) +
  ylab("Egg Size") +
  theme_minimal() +
  guides(size = FALSE) +
  theme(panel.grid.major = element_blank(),
        panel.grid.minor = element_blank(),
```

```
axis.title.y = element_blank(),
    axis.text.y = element_blank(),
    axis.ticks.y = element_blank(),
    legend.position = "none")
p3
```

-2 -1 0 1 2 ONI

```
## plot together
library(patchwork)
all <- p1 + p2 + p3
all</pre>
```



ggsave(here("results", "pred_plots.pdf"), width = 10, height = 7)

Model prediction for the top model only BEUTI

```
# model prediction for one model only (top model)
pv.BEUTIv2 <- matrix(nrow = sims, ncol = nrow(SCMU.BEUTI))</pre>
for(i in 1:sims){
    #we simulate conditioning on no specific random effects levels
    y <- unlist(simulate(topmod1))</pre>
    bmod <- refit(topmod1,y)</pre>
    pv.BEUTIv2[i,] <- predict(bmod, re.form = ~0, newdata = SCMU.BEUTI) + rnorm(1,0,sd=plot.sd.1) + rno
}
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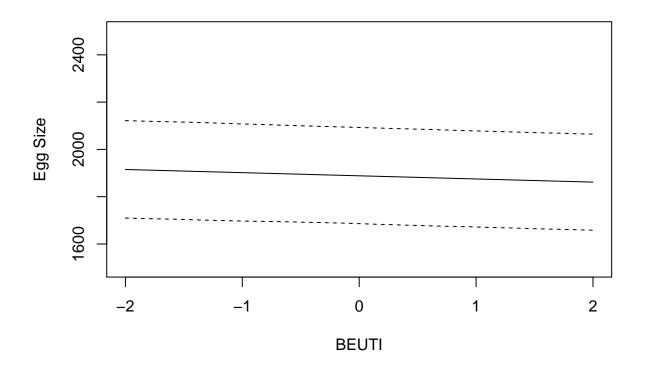
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## boundary (singular) fit: see ?isSingular
## check plots
plot(SCMU.BEUTI$BEUTI[1:100],y = apply(pv.BEUTIv2,2,mean)[1:100],xlab = "BEUTI",ylab = "Egg Size",type
lines(SCMU.BEUTI$BEUTI[1:100], y = apply(pv.BEUTIv2,2,function(x)quantile(x,probs= 0.025))[1:100], lty=2)
lines(SCMU.BEUTI$BEUTI[1:100],y = apply(pv.BEUTIv2,2,function(x)quantile(x,probs= 0.975))[1:100],lty=2)
```



NPGO

```
# model prediction for one model only (top model)
pv.NPGOv2 <- matrix(nrow = sims, ncol = nrow(SCMU.NPGO))</pre>
for(i in 1:sims){
    #we simulate conditioning on no specific random effects levels
    y <- unlist(simulate(topmod1))</pre>
    bmod <- refit(topmod1,y)</pre>
    pv.NPGOv2[i,] <- predict(bmod, re.form = ~0, newdata = SCMU.NPGO) + rnorm(1,0,sd=plot.sd.1) + rnorm
}
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## boundary (singular) fit: see ?isSingular
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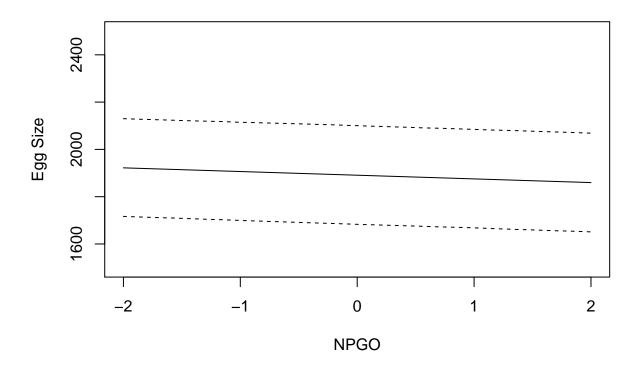
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## boundary (singular) fit: see ?isSingular
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## boundary (singular) fit: see ?isSingular
## check plots
plot(SCMU.NPGO$NPGO[1:100],y = apply(pv.NPGOv2,2,mean)[1:100],xlab = "NPGO",ylab = "Egg Size",type = "l
lines(SCMU.NPGO$NPGO[1:100],y = apply(pv.NPGOv2,2,function(x)quantile(x,probs= 0.025))[1:100],lty=2)
lines(SCMU.NPGO$NPGO[1:100],y = apply(pv.NPGOv2,2,function(x)quantile(x,probs= 0.975))[1:100],lty=2)
```



ONI

```
# model prediction for one model only (top model)
pv.ONIv2 <- matrix(nrow = sims, ncol = nrow(SCMU.ONI))</pre>
for(i in 1:sims){
    #we simulate conditioning on no specific random effects levels
    y <- unlist(simulate(topmod1))</pre>
    bmod <- refit(topmod1,y)</pre>
    pv.ONIv2[i,] <- predict(bmod, re.form = ~0, newdata = SCMU.ONI) + rnorm(1,0,sd=plot.sd.1) + rnorm(1
}
## boundary (singular) fit: see ?isSingular
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boundary (singular) fit: see ?isSingular

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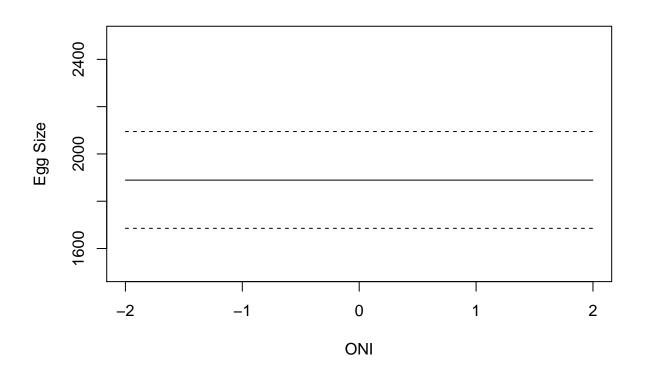
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## boundary (singular) fit: see ?isSingular
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## boundary (singular) fit: see ?isSingular
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## boundary (singular) fit: see ?isSingular
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## boundary (singular) fit: see ?isSingular
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```
## boundary (singular) fit: see ?isSingular
## check plots
plot(SCMU.ONI$ONI[1:100],y = apply(pv.ONIv2,2,mean)[1:100],xlab = "ONI",ylab = "Egg Size",type = "l",yl
lines(SCMU.ONI$ONI[1:100],y = apply(pv.ONIv2,2,function(x)quantile(x,probs= 0.025))[1:100],lty=2)
lines(SCMU.ONI$ONI[1:100],y = apply(pv.ONIv2,2,function(x)quantile(x,probs= 0.975))[1:100],lty=2)
```



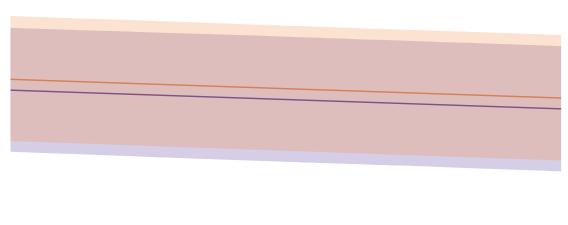
Plots

```
## ggplot
color_pal <- c("#593d9cff", "#f68f46ff")

### BEUTI
y4 <- as.data.frame(apply(pv.BEUTIv2,2,mean))</pre>
```

```
quantile.low4 <- as.data.frame(apply(pv.BEUTIv2,2,function(x)quantile(x,probs= 0.025)))
quantile.high4 <- as.data.frame(apply(pv.BEUTIv2,2,function(x)quantile(x,probs= 0.975)))
BEUTI.pred4 <- cbind(y4, quantile.low4, quantile.high4, SCMU.BEUTI$BEUTI) %>%
  mutate(EggOrder = c(rep("Egg1",100),rep("Egg2",100)))
colnames(BEUTI.pred4) <- c("mean", "low", "high", "BEUTI", "egg_order")</pre>
p4 <- ggplot(data = BEUTI.pred4) +
  geom line(aes(x = BEUTI, y = mean, color = egg order)) +
  geom_ribbon(aes(ymin = low, ymax = high, x = BEUTI, fill = egg_order), alpha = 0.25, linetype = "dash
  ylim(c(1500, 2500)) +
  scale_color_manual(values = color_pal) +
  scale_fill_manual(values = color_pal) +
  ylab("Egg Size") +
  theme_minimal() +
  theme(panel.grid.major = element_blank(),
        panel.grid.minor = element_blank(),
        legend.position = "none")
p4
   2500
   2250
   2000
   1750
   1500
                                                                                      2
           -2
                                              BEUTI
### NPGO
y5 <- as.data.frame(apply(pv.NPGOv2,2,mean))
quantile.low5 <- as.data.frame(apply(pv.NPGOv2,2,function(x)quantile(x,probs= 0.025)))
quantile.high5 <- as.data.frame(apply(pv.NPGOv2,2,function(x)quantile(x,probs= 0.975)))
NPGO.pred5 <- cbind(y5, quantile.low5, quantile.high5, SCMU.NPGO$NPGO) %>%
  mutate(EggOrder = c(rep("Egg1",100),rep("Egg2",100)))
colnames(NPGO.pred5) <- c("mean", "low", "high", "NPGO", "egg_order")</pre>
```

```
p5 <- ggplot(data = NPGO.pred5) +
  geom\_line(aes(x = NPGO, y = mean, color = egg\_order)) +
  geom_ribbon(aes(ymin = low, ymax = high, x = NPGO, fill = egg_order), alpha = 0.25, linetype = "dashe
  ylim(c(1500, 2500)) +
  scale_color_manual(values = color_pal) +
  scale_fill_manual(values = color_pal) +
  ylab("Egg Size") +
  theme minimal() +
  guides(size = FALSE) +
  theme(panel.grid.major = element_blank(),
        panel.grid.minor = element_blank(),
        axis.title.y = element_blank(),
        axis.text.y = element_blank(),
        axis.ticks.y = element_blank(),
        legend.title = element_blank(),
        legend.position = "bottom")
p5
```

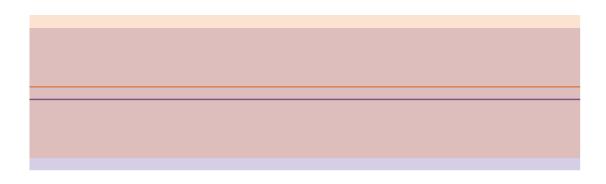


```
-2 -1 0 1 2 NPGO

-- Egg1 -- Egg2
```

```
### ONI
y6 <- as.data.frame(apply(pv.ONIv2,2,mean))
quantile.low6 <- as.data.frame(apply(pv.ONIv2,2,function(x)quantile(x,probs= 0.025)))
quantile.high6 <- as.data.frame(apply(pv.ONIv2,2,function(x)quantile(x,probs= 0.975)))
ONI.pred6 <- cbind(y6, quantile.low6, quantile.high6, SCMU.ONI$ONI) %>%
    mutate(EggOrder = c(rep("Egg1",100),rep("Egg2",100)))
colnames(ONI.pred6) <- c("mean", "low", "high", "ONI", "egg_order")</pre>
```

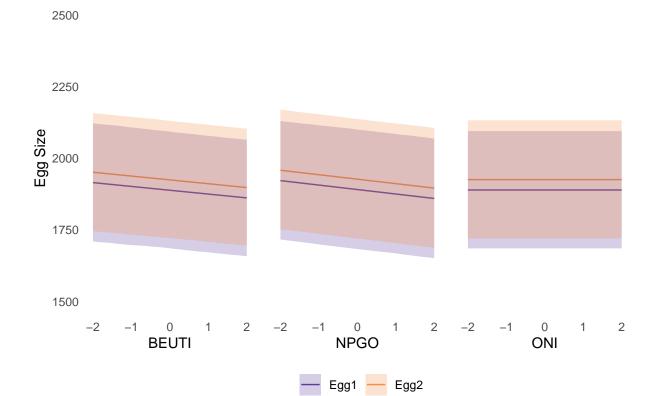
```
p6 <- ggplot(data = ONI.pred6) +
  geom\_line(aes(x = ONI, y = mean, color = egg\_order)) +
  geom_ribbon(aes(ymin = low, ymax = high, x = ONI, fill = egg_order), alpha = 0.25, linetype = "dashed
  ylim(c(1500, 2500)) +
  scale_color_manual(values = color_pal) +
  scale_fill_manual(values = color_pal) +
  ylab("Egg Size") +
  theme_minimal() +
  guides(size = FALSE) +
  theme(panel.grid.major = element_blank(),
        panel.grid.minor = element_blank(),
        axis.title.y = element_blank(),
        axis.text.y = element_blank(),
        axis.ticks.y = element_blank(),
        legend.position = "none")
p6
```



```
-2 -1 0 1 2
ONI

## plot together
library(patchwork)
all2 <- p4 + p5 + p6
```

al12



 $\# ggsave(here("results", "pred_plots_v2.pdf"), width = 10, height = 7)$