Package 'RLumModel'

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Type Package

Title Modelling Ordinary Differential Equations Leading to Luminescence

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Description A collection of functions to simulate luminescence signals in the mineral quartz based on published models.

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License GPL-3

Depends R (>= 3.2.3), utils, Luminescence (>= 0.5.1)

Imports deSolve (>= 1.12), FME, methods, Rcpp

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RoxygenNote 5.0.1

Suggests R.rsp, testthat

VignetteBuilder R.rsp

LinkingTo Rcpp, RcppArmadillo

NeedsCompilation yes

R **topics documented:**

RLumModel-package	*Modelling Ordinary Differential Equations Leading to Luminescence*

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A collection of function to simulate luminescence signals in the mineral quartz based on published models.

Details

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Author(s)

Authors

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Project source code repository

https://github.com/R-Lum

Related projects

http://www.r-luminescence.de
http://cran.r-project.org/package=Luminescence
http://shiny.r-luminescence.de
http://cran.r-project.org/package=RLumShiny

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Acknowledgement

ExampleData.FittingTL *Example data with TL curves extracted from a TL-SAR protocol*

Description

Example data with TL curves extracted from a TL-SAR protocol

Format

A RLum.Analysis object containing measured TL curves.

Function version

0.1.0

Author(s)

Johannes Friedrich, University of Bayreuth (Germany)

See Also

modCost

Examples

```
data("ExampleData.FittingTL", envir = environment())
```

ExampleData.ModelOutput

Example data (TL curve) simulated from Bailey (2001, fig. 1)

Description

Example data (TL curve) simulated from Bailey (2001, fig. 1)

Format

A RLum.Analysis object containing one TL curve as RLum.Data.Curve.

Function version

0.1.0

Note

This example has only one record (TL). The used sequence was sequence <- list(IRR = c(temp = 20, dose = 10, DoseRate = 1), TL = c(temp_begin = 20, temp_end = 400, heating_rate = 5))

Author(s)

Johannes Friedrich, University of Bayreuth (Germany)

Source

model_LuminescenceSignals()

References

Bailey, R.M., 2001. Towards a general kinetic model for optically and thermally stimulated luminescence of quartz. Radiation Measurements 33, 17-45.

Examples

```
data("ExampleData.ModelOutput", envir = environment())

TL_curve <- get_RLum(model.output, recordType = "TL$", drop = FALSE)

##plot TL curve
plot_RLum(TL_curve)

TL_concentrations <- get_RLum(model.output, recordType = "(TL)", drop = FALSE)
plot_RLum(TL_concentrations)
```

---

extract_pars2FME   *Prepare parameters for use with R package FME*

Description

Prepare parameters for use with R package FME

Usage

```
extract_pars2FME(model)
```

Arguments

model   Name of the quartz luminescence model. Possible choices are: "Bailey2001",
                  "Bailey2002, "Bailey2004", "Pagonis2007" and "Pagonis2008".

Value

This function returns a named vector for use with R package FME

Function version

0.1.0

Author(s)

Johannes Friedrich, University of Bayreuth (Germany),

References

Bailey, R.M., 2001. Towards a general kinetic model for optically and thermally stimulated luminescence of quartz. Radiation Measurements 33, 17-45.

Bailey, R.M., 2002. Simulations of variability in the luminescence characteristics of natural quartz and its implications for estimates of absorbed dose. Radiation Protection Dosimetry 100, 33-38.

Bailey, R.M., 2004. Paper I-simulation of dose absorption in quartz over geological timescales and it simplications for the precision and accuracy of optical dating. Radiation Measurements 38, 299-310.

Pagonis, V., Chen, R., Wintle, A.G., 2007: Modelling thermal transfer in optically stimulated luminescence of quartz. Journal of Physics D: Applied Physics 40, 998-1006.

Pagonis, V., Wintle, A.G., Chen, R., Wang, X.L., 2008. A theoretical model for a new dating protocol for quartz based on thermally transferred OSL (TT-OSL). Radiation Measurements 43, 704-708.

Soetaert K., Petzoldt T., 2010: Inverse Modelling, Sensitivity and Monte Carlo Analysis in R Using Package FME. Journal of Statistical Software, 33, 1-28.

Soetaert, K., Cash, J., Mazzia, F., 2012: Solving differential equations in R. Springer Science & Business Media.

See Also

[model_LuminescenceSignals](), [sensFun](), [sensRange](), [fit_RLumModel2data]()

Examples

```
parms <- extract_pars2FME(model = "Bailey2001")
```

---

fit_RLumModel2data   *Fit model parameters to experimental data*

Description

Fit model parameters to experimental data

Usage

```
fit_RLumModel2data(sequence, model, seq.step2fit, lab.dose_rate = 1,
  simulate_sample_history = FALSE, verbose = FALSE, plot = FALSE, ...)
```

Arguments

sequence	[list]() (**required**): set sequence to model as [list]() or as *.seq file from the Riso sequence editor. To simulate SAR measurements there is an extra option to set the sequence list (cf. details). See [model_LuminescenceSignals]() for a detailed description.
model	[character]() (**required**): set model to be used. Available models are: "Bailey2001", "Bailey2002", "Bailey2004", "Pagonis2007", "Pagonis2008"
seq.step2fit	[numeric](): Choose the sequence step number to fit. **Note: Only sequence steps with an signal output are allowed! This sequence steps are: RF, TL, OSL, LM-OSL**
lab.dose_rate	[numeric]() (with default): laboratory dose rate in XXX Gy/s for calculating seconds into Gray in the *.seq file.
simulate_sample_history	
	[logical]() (with default): FALSE (with default): simulation begins at laboratory conditions, TRUE: simulations begins at crystallization (all levels 0) process
verbose	[logical]() (with default): Verbose mode on/off.
plot	[logical]() (with default): Enables or disables plot output. Recommended: No plot output, because fitting function will run a lot of times.
...	further arguments and graphical parameters passed to [plot.default](). See details for further information.

Value

This function returns a function, which is necessary for further calculations with the package "FME".

Function version

0.1.0

Author(s)

Johannes Friedrich, University of Bayreuth (Germany)

References

Soetaert K., Petzoldt T., 2010: Inverse Modelling, Sensitivity and Monte Carlo Analysis in R Using Package FME. Journal of Statistical Software, 33, 1-28.

See Also

sensFun, sensRange, model_LuminescenceSignals, extract_pars2FME

Examples

```
sequence <- list(
  TL = c(20, 450, 5),
  IRR = c(20, 5, 0.5),
  TL = c(0, 450, 5))

model <- "Pagonis2007"

parms <- extract_pars2FME(model = model)

func_FME <- fit_RLumModel2data(
  sequence = sequence,
  model = model,
  seq.step2fit = 3)

SensR <- FME::sensFun(
  func = func_FME,
  parms = parms,
  sensvar = "signal",
  senspar = c("N1","s1","E1"))

plot(
  SensR,
  legpos = "bottomleft",
  xlab = "Temperature [\u00B0C]",
  main = "Local Sensitivity Analysis TL")
```

---

model_LuminescenceSignals

Model Luminescence Signals

Description

This function models luminescence signals for quartz based on published physical models. It is possible to simulate TL, (CW-) OSL, RF measurements in a arbitrary sequence. This sequence is definded as a [list](#) of certain abrivations. Furthermore it is possible to load a sequence direct from the Riso Sequence Editor. The output is an [RLum.Analysis](#)object and so the plots are done by the [plot_RLum.Analysis](#) function. If a SAR sequence is simulated the plot output can be disabled and SAR analyse functions can be used.

Usage

```
model_LuminescenceSignals(model, sequence, lab.dose_rate = 1,
  simulate_sample_history = FALSE, plot = TRUE, verbose = TRUE,
  show.structure = FALSE, parms = NULL, ...)
```

Arguments

model	[character](#) (**required**): set model to be used. Available models are: "Bailey2001", "Bailey2002", "Bailey2004", "Pagonis2007", "Pagonis2008"
sequence	[list](#) (**required**): set sequence to model as [list](#) or as *.seq file from the Riso sequence editor. To simulate SAR measurements there is an extra option to set the sequence list (cf. details).
lab.dose_rate	[numeric](#) (with default): laboratory dose rate in XXX Gy/s for calculating seconds into Gray in the *.seq file.
simulate_sample_history	
	[logical](#) (with default): FALSE (with default): simulation begins at laboratory conditions, TRUE: simulations begins at crystallization (all levels 0) process
plot	[logical](#) (with default): Enables or disables plot output
verbose	[logical](#) (with default): Verbose mode on/off
show.structure	[logical](#) (with default): Shows the structure of the result. Recommended to show record.id to analyse concentrations.
parms	[list](#) or [numeric](#) (with default): This argument is only necessary, if fit_data2RLumModel is used. There is no need to change this parameter per hand, all is done automatically. Nevertheless is it necessary for the package "FME" to have the parameters direct in the function call.
...	further arguments and graphical parameters passed to [plot.default](#). See details for further information.

Details

Defining a **sequence**

Arguments	Description	Sub-arguments
TL	thermally stimulated luminescence	'temp begin', 'temp end', 'heating rate'
OSL	optically stimulated luminescence	'temp', 'duration','optical_power'
ILL	illumination	'temp', 'duration','optical_power'
LM_OSL	linear modulated OSL	'temp', 'duration', optional: 'start_power', 'end_power'
RL/RF	radioluminescence	'temp','dose', 'dose_rate'
IRR	irradiation	'temp','dose', 'dose_rate'
CH	cutheat	'temp', optional: 'duration', 'heating_rate'
PH	preheat	'temp', 'duration' optional: 'heating_rate'
PAUSE	pause	'temp', 'duration'

Note: 100 % illumination power equates to 20 mW/cm^2

Defining a **SAR-sequence**

Abrivation	Description	examples
RegDose	Dose points of the regenerative cycles	c(0, 80, 140, 260, 320, 0, 80)
TestDose	Test dose for the SAR cycles	50
PH	Temperature of the preheat	240
CH	Temperature of the cutheat	200
OSL_temp	Temperature of OSL read out	125
OSL_duration	Duration of OSL read out	default: 40
Irr_temp	Temperature of irradiation	default: 20
PH_duration	Duration of the preheat	default: 10
dose_rate	Dose rate of the laboratory irradiation source	default: 1
optical_power	Percentage of the full illumination power	default: 90
Irr_2recover	Dose to be recovered in a dose-recovery-test	20

Value

This function returns an `RLum.Analysis` object with all TL, (LM-) OSL and RF/RL steps in the sequence. Every entry is an `RLum.Data.Curve` object and can be plotted, analysed etc. with further RLum-functions.

Function version

0.1.1

Author(s)

Johannes Friedrich, University of Bayreuth (Germany), Sebastian Kreutzer, IRAMAT-CRP2A, Universite Bordeaux Montaigne (France)

References

Bailey, R.M., 2001. Towards a general kinetic model for optically and thermally stimulated luminescence of quartz. Radiation Measurements 33, 17-45.

Bailey, R.M., 2002. Simulations of variability in the luminescence characteristics of natural quartz and its implications for estimates of absorbed dose. Radiation Protection Dosimetry 100, 33-38.

Bailey, R.M., 2004. Paper I-simulation of dose absorption in quartz over geological timescales and it simplications for the precision and accuracy of optical dating. Radiation Measurements 38, 299-310.

Pagonis, V., Chen, R., Wintle, A.G., 2007: Modelling thermal transfer in optically stimulated luminescence of quartz. Journal of Physics D: Applied Physics 40, 998-1006.

Pagonis, V., Wintle, A.G., Chen, R., Wang, X.L., 2008. A theoretical model for a new dating protocol for quartz based on thermally transferred OSL (TT-OSL). Radiation Measurements 43, 704-708.

Soetaert, K., Cash, J., Mazzia, F., 2012. Solving differential equations in R. Springer Science & Business Media.

See Also

plot, RLum, read_SEQ2R

Examples

```
##=================================================================##
## Example 1: Simulate sample history of Bailey2001
## (cf. Bailey, 2001, Fig. 1)
##=================================================================##

##set sequence with the following steps
## (1) Irradiation at 20 deg. C with a dose of 10 Gy and a dose rate of 1 Gy/s
## (2) TL from 20-400 deg. C with a rate of 5 K/s

sequence <-
  list(
    IRR = c(20, 10, 1),
    TL = c(20, 400, 5)
  )

##model sequence
model.output <- model_LuminescenceSignals(
  sequence = sequence,
  model = "Bailey2001"
)

##get all TL concentrations

TL_conc <- get_RLum(model.output, recordType = "(TL)", drop = FALSE)

plot_RLum(TL_conc)

##plot 110 deg. C trap concentration
```

```
TL_110 <- get_RLum(TL_conc, recordType = "conc. level 1")
plot_RLum(TL_110)

##========================================================================##
## Example 2: compare different optical powers of stimulation light
##========================================================================##

# call function "model_LuminescenceSignals", model = "Bailey2004"
# and simulate_sample_history = FALSE (default),
# because the sample history is not part of the sequence
# the optical_power of the LED is varied and then compared.

optical_power <- seq(from = 0,to = 100,by = 20)

model.output <- lapply(optical_power, function(x){

  sequence <- list(IRR = c(20, 50, 1),
                   PH = c(220, 10, 5),
                   OSL = c(125, 50, x)
                   )

  data <- model_LuminescenceSignals(
            sequence = sequence,
            model = "Bailey2004",
            plot = FALSE,
            verbose = FALSE
            )

  return(get_RLum(data, recordType = "OSL$", drop = FALSE))
})

##combine output curves
model.output.merged <- merge_RLum(model.output)

##plot
plot_RLum(
 object = model.output.merged,
 xlab = "Illumination time [s]",
 ylab = "OSL signal [a.u.]",
 main = "OSL signal dependency on optical power of stimulation light",
 legend.text = paste("Optical power density", 20*optical_power/100, "mW/cm^2"),
 combine = TRUE)

## Not run:
##========================================================================##
## Example 3: Simulate Thermal-Activation-Characteristics (TAC)
##========================================================================##

 ##set temperature
 act.temp <- seq(from = 80, to = 600, by = 20)

 ##loop over temperature
 model.output <- vapply(X = act.temp, FUN = function(x) {
```

```
##set sequence, note: sequence includes sample history
  sequence <- list(
    IRR = c(20, 1, 1e-11),
    IRR = c(20, 10, 1),
    PH = c(x, 1),
    IRR = c(20, 0.1, 1),
    TL = c(20, 150, 5)
)
##run simulation
  temp <- model_LuminescenceSignals(
    sequence = sequence,
    model = "Pagonis2007",
    simulate_sample_history = TRUE,
    plot = FALSE,
    verbose = FALSE
  )
    ## "TL$" for exact matching TL and not (TL)
  TL_curve <- get_RLum(temp, recordType = "TL$")
  ##return max value in TL curve
  return(max(get_RLum(TL_curve)[,2]))
}, FUN.VALUE = 1)

##plot resutls
plot(
  act.temp[-(1:3)],
  model.output[-(1:3)],
  type = "b",
  xlab = "Temperature [\u00B0C]",
  ylab = "TL [a.u.]"
)

##=============================================================================##
## Example 4: Simulate SAR sequence
##=============================================================================##

##set SAR sequence with the following steps
## (1) RegDose: set regenerative dose [Gy] as vector
## (2) TestDose: set test dose [Gy]
## (3) PH: set preheat temperature in deg. C
## (4) CH: Set cutheat temperature in deg. C
## (5) OSL_temp: set OSL reading temperature in deg. C
## (6) OSL_duration: set OSL reading duration in s

sequence <- list(
 RegDose = c(0,10,20,50,90,0,10),
 TestDose = 5,
 PH = 240,
 CH = 200,
 OSL_temp = 125,
 OSL_duration = 70)

# call function "model_LuminescenceSignals", set sequence = sequence,
```

```
# model = "Pagonis2007" (palaeodose = 20 Gy) and simulate_sample_history = FALSE (default),
# because the sample history is not part of the sequence

 model.output <- model_LuminescenceSignals(
   sequence = sequence,
   model = "Pagonis2007",
   plot = FALSE
 )

# in environment is a new object "model.output" with the results of
# every step of the given sequence.
# Plots are done at OSL and TL steps and the growth curve

# call "analyse_SAR.CWOSL" from RLum package
 results <- analyse_SAR.CWOSL(model.output,
                             signal.integral.min = 1,
                             signal.integral.max = 15,
                             background.integral.min = 601,
                             background.integral.max = 701,
                             fit.method = "EXP",
                             dose.points = c(0,10,20,50,90,0,10))


##=======================================================================##
## Example 5: generate sequence from *.seq file and run SAR simulation
##=======================================================================##

# load example *.SEQ file and construct a sequence.
# call function "model_LuminescenceSignals", load created sequence for sequence,
# set model = "Bailey2002" (palaeodose = 10 Gy)
# and simulate_sample_history = FALSE (default),
# because the sample history is not part of the sequence

path <- system.file("extdata", "example_SAR_cycle.SEQ", package="RLumModel")

sequence <- read_SEQ2R(file = path)

model.output <- model_LuminescenceSignals(
  sequence = sequence,
  model = "Bailey2001",
  plot = FALSE
)


## call RLum package function "analyse_SAR.CWOSL" to analyse the simulated SAR cycle

results <- analyse_SAR.CWOSL(model.output,
                             signal.integral.min = 1,
                             signal.integral.max = 10,
                             background.integral.min = 301,
                             background.integral.max = 401,
                             dose.points = c(0,8,14,26,32,0,8),
                             fit.method = "EXP")
```

```
print(get_RLum(results))


##==========================================================================##
## Example 6: Simulate sequence at labour without sample history
##==========================================================================##

##set sequence with the following steps
## (1) Irraditation at 20 deg. C with a dose of 100 Gy and a dose rate of 1 Gy/s
## (2) Preheat to 200 deg. C and hold for 10 s
## (3) LM-OSL at 125 deg. C. for 100 s
## (4) Cutheat at 200 dec. C.
## (5) Irraditation at 20 deg. C with a dose of 10 Gy and a dose rate of 1 Gy/s
## (6) Pause at 200 de. C. for 100 s
## (7) OSL at 125 deg. C for 100 s with 90 % optical power
## (8) Pause at 200 deg. C for 100 s
## (9) TL from 20-400 deg. C with a heat rate of 5 K/s
## (10) Radiofluorescence at 20 deg. C with a dose of 200 Gy and a dose rate of 0.01 Gy/s

sequence <-
 list(
   IRR = c(20, 100, 1),
   PH = c(200, 10),
   LM_OSL = c(125, 100),
   CH = c(200),
   IRR = c(20, 10, 1),
   PAUSE = c(200, 100),
   OSL = c(125, 100, 90),
   PAUSE = c(200, 100),
   TL = c(20, 400, 5),
   RF = c(20, 200, 0.01)
)

# call function "model_LuminescenceSignals", set sequence = sequence,
# model = "Pagonis2008" (palaeodose = 200 Gy) and simulate_sample_history = FALSE (default),
# because the sample history is not part of the sequence

model.output <- model_LuminescenceSignals(
   sequence = sequence,
   model = "Pagonis2008"
   )


## End(Not run)
```

---

read_SEQ2R   *Parse a Risoe SEQ-file to a sequence neccessary for simulating quartz
                                     luminescence*

Description

A SEQ-file created by the Risoe Sequence Editor can be imported to simulate the sequence written in the sequence editor.

Usage

```
read_SEQ2R(file, lab.dose_rate = 1, txtProgressBar = TRUE)
```

Arguments

file   [character](required): a *.seq file created by the Risoe Sequence Editor

lab.dose_rate   [character] (with default): set the dose rate of the radiation source in the laboratory [Gy/s]. Default: 1 Gy/s

txtProgressBar [logical] (with default): enables or disables the txtProgressBar for a visuell control of the progress. Default: txtProgressBar = TRUE

Details

Supported versions

Supppored and tested: version 4.36.

Value

This function returns a [list] with the parsed *.seq file and the required steps for [model_LuminescenceSignals].

Function version

0.1.0

Author(s)

Johannes Friedrich, University of Bayreuth (Germany),

References

Riso: Sequence Editor User Manual. Available at: http://www.nutech.dtu.dk/english/-/media/Andre_Universitetsenheder/Nutech/Produkter%20og%20services/Dosimetri/radiation_measurement_instruments/tl_osl_reader/Manuals/SequenceEditor.ashx?la=da

See Also

[model_LuminescenceSignals], [readLines]

Examples

```
##search "example_SAR_cycle.SEQ" in "extdata" in package "RLumModel"
path <- system.file("extdata", "example_SAR_cycle.SEQ", package="RLumModel")

sequence <- read_SEQ2R(file = path)
```

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