



University
of Glasgow

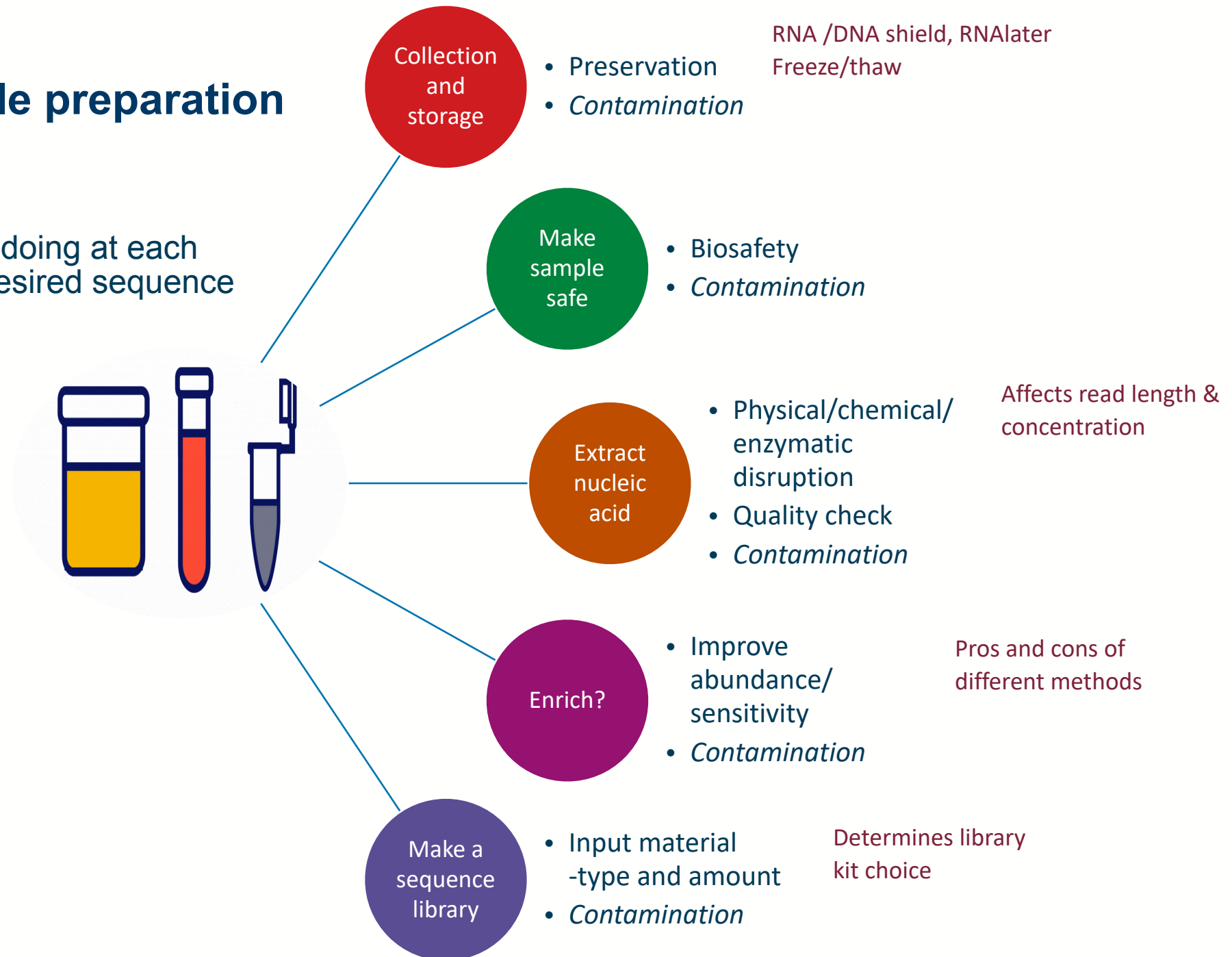
Sample preparation

Kirstyn Brunker
NCDC, Abuja
12-16th Feb 2024



Sample preparation

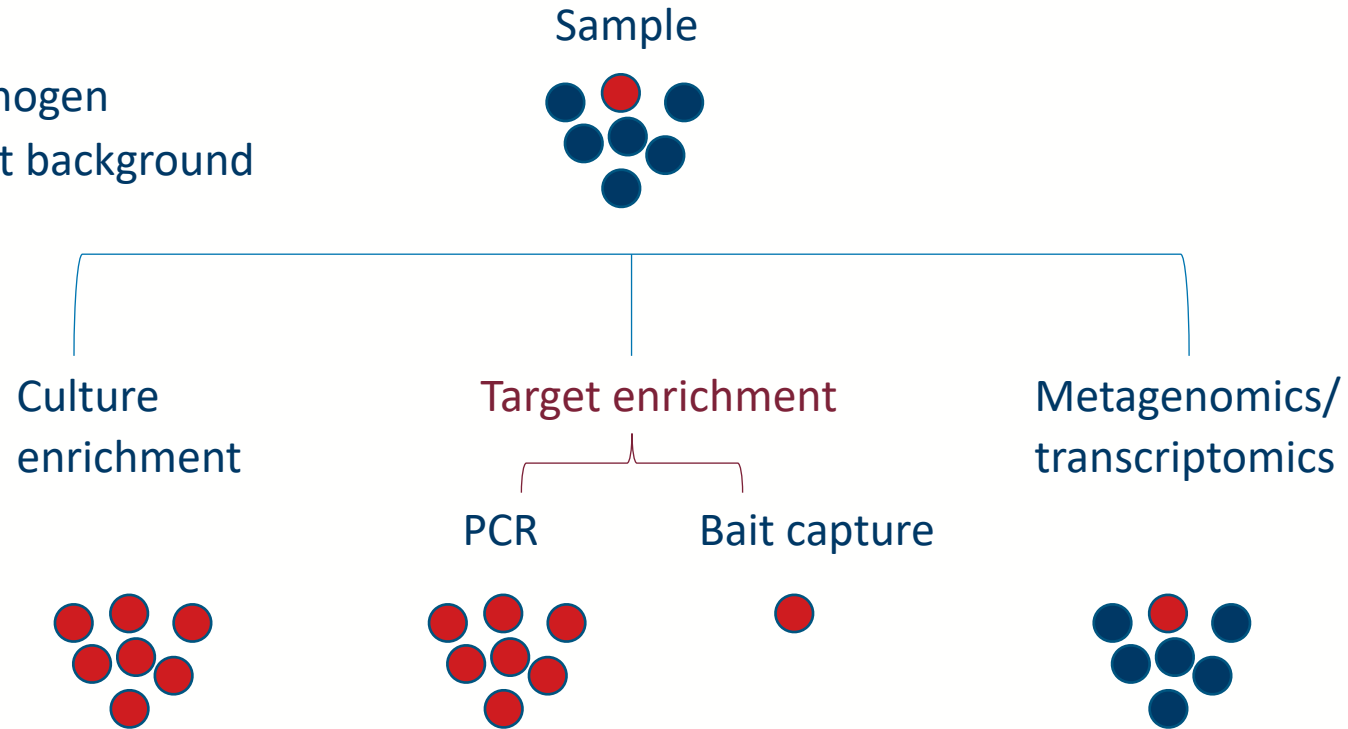
- Consider what you are doing at each step to optimize your desired sequence data output!





Approaches

- Pathogen
- Host background

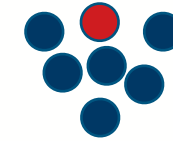
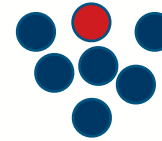


Considerations

- Time
- Cost
- Sensitivity
- Detection power
- Readiness/portability



Metagenomics

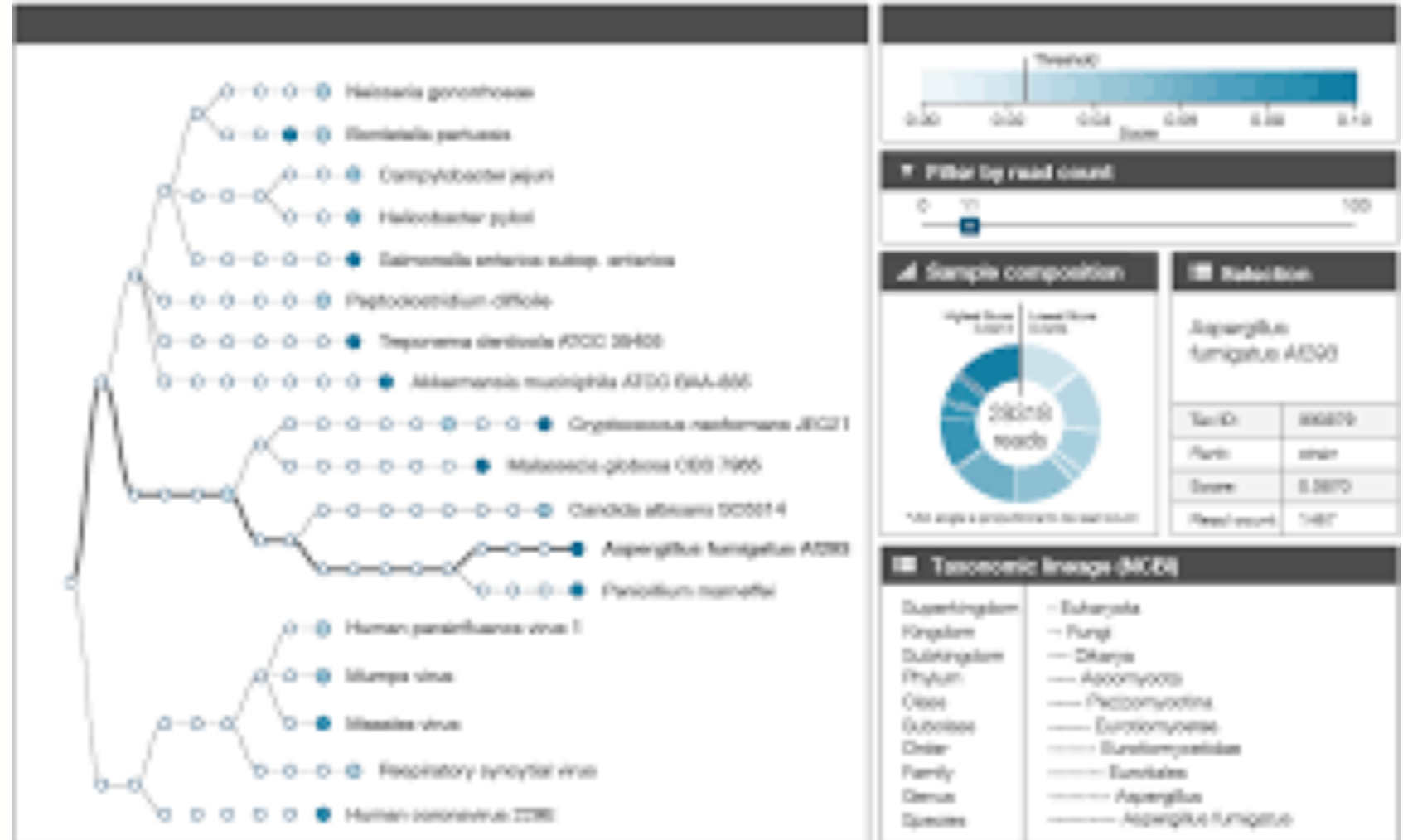


Pros

- Gold standard
- Pathogen agnostic
- Unbiased

Cons

- *Lower sensitivity*
- *More expensive*
- *Complex*





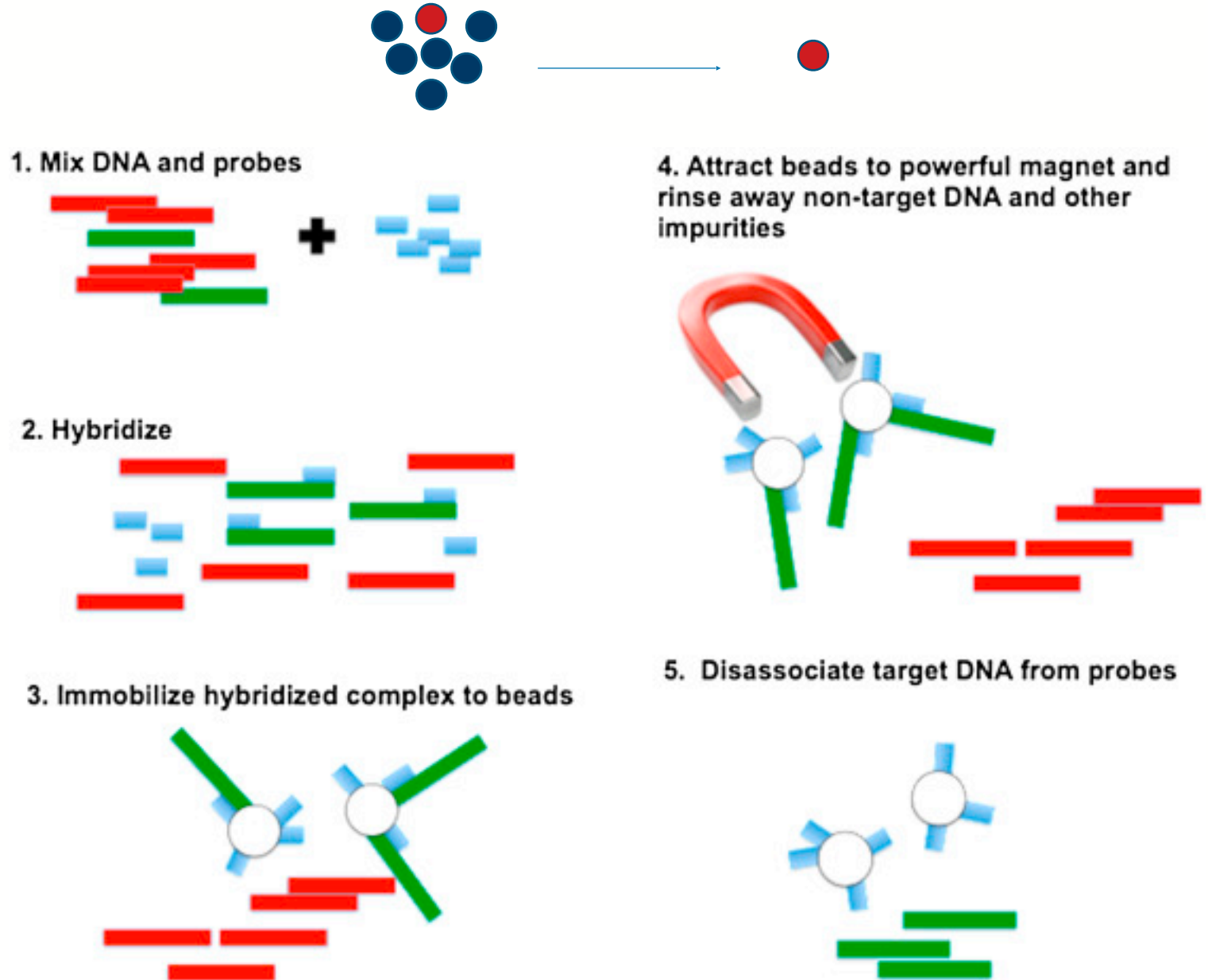
Bait capture

Pros

- Tolerant of diversity

Cons

- *A priori knowledge*
- *Expensive*
- *Slow & complex*





Culture enrichment



Pros

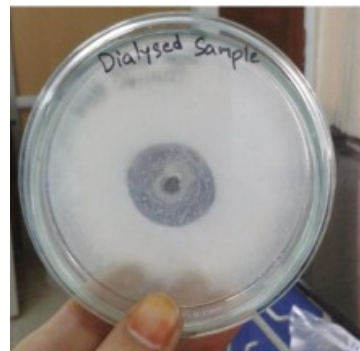
- Cheap
- Sensitive

Cons

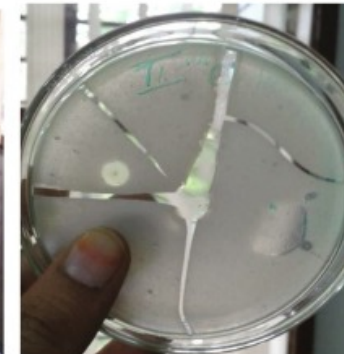
- *A priori knowledge*
- *Slow*
- *Specific expertise*



(A)



(B)

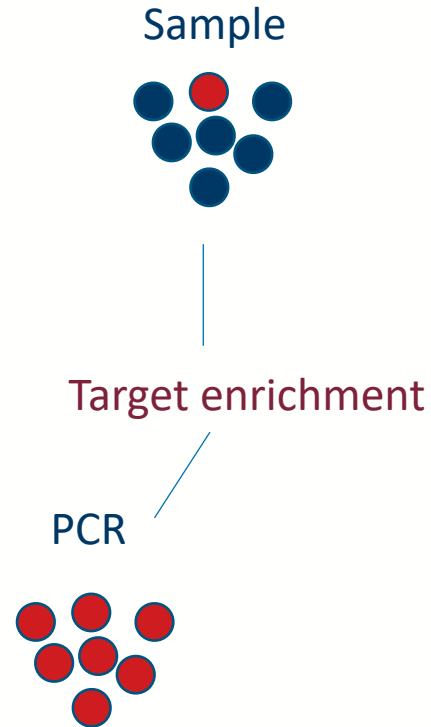


(C)



PCR enrichment

- Pathogen
- Host background



Why did we choose this?

- Enriches samples with low viral material
- Helps with poor quality sampled (fragmented RNA)
- Cost-effective
- High sensitivity

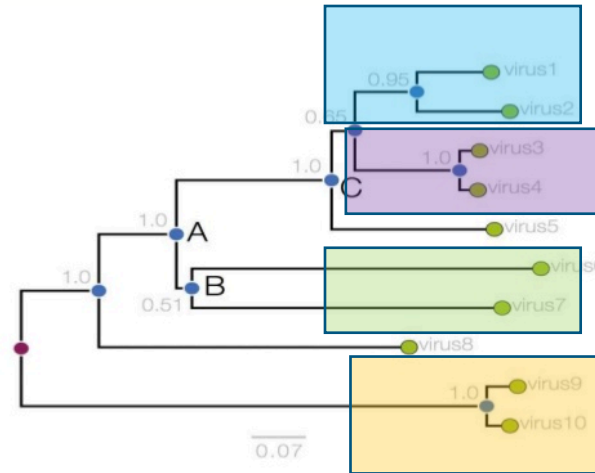
Caveats:

- Requires *a priori* knowledge of pathogen
- Tolerant of limited diversity
- Potential for contamination



Designing primers

- What do I want to capture?
 - What pathogen
 - Location
 - Host variant
 - Whole or partial genome
- What *a priori* knowledge is available?





Challenges & potential solutions

- There is no existing data for study area

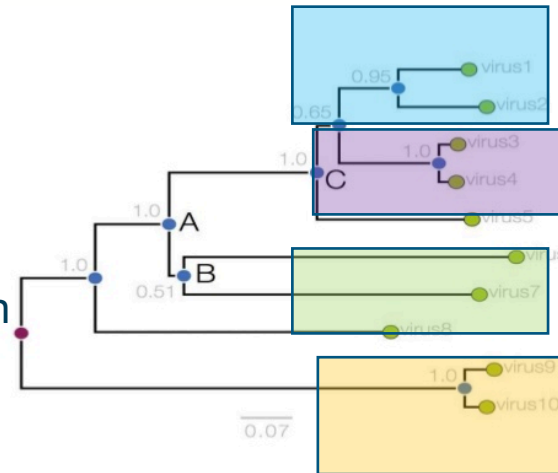
Solution: do some preliminary metagenomic sequencing to get a reference

- There are only partial genomes

Solution: use to get minor clade assignments, then use most closely related public sequences as a reference

- Diversity is too great

Solution: create multiple primer sets or try probes



Note: This is just advice not a hard set of rules!



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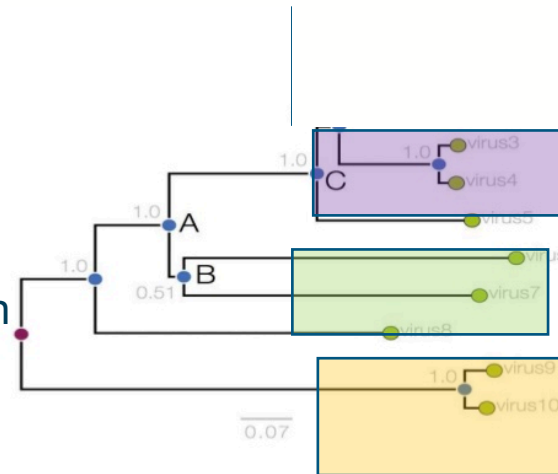
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