Fluorometer-based nucleic acid quantification is a commonly used method in molecular biology and biotechnology laboratories for measuring the concentration of DNA, RNA, or other nucleic acids in a sample.

**Principle:** Fluorometer-based nucleic acid quantification works by mixing a sample containing DNA or RNA with a fluorescent dye that specifically binds to nucleic acids. When excited by light of a specific wavelength, the dye emits fluorescence, and its intensity is directly proportional to the concentration of nucleic acids in the sample. A fluorometer detects this fluorescence and calculates the nucleic acid concentration. It's a rapid and sensitive method used widely in molecular biology research and diagnostics.

**Examples of Fluorometers:** Quantus™ Fluorometer (Promega); Qubit Fluorometer (Thermo Fisher Scientific) [tube or multi-tube readers]. Promega GloMax; Tecan Infinite Series [plate readers] etc

**Specificity:** Different fluorometric assays use specific dyes that bind to nucleic acids, such as SYBR Green, PicoGreen, or Hoechst dyes. These dyes are chosen based on their specificity for DNA or RNA and their ability to emit fluorescence upon binding. Fluorometers are generally less susceptible to interference, as the specific binding of fluorophores to nucleic acids minimizes the impact of contaminants. This makes them more accurate than nanodrop for targeted quantification of specific nucleic acids.

**Sensitivity:** Fluorometric assays offer high sensitivity, allowing the detection of low concentrations of nucleic acids in samples. This makes them suitable for applications where accurate quantification of nucleic acids is required, such as PCR, qPCR, sequencing, and cloning.

**Dynamic Range:** Fluorometer-based assays typically have a wide dynamic range, allowing for the quantification of nucleic acids across a broad concentration range. This enables the measurement of both high and low concentrations of nucleic acids in a single assay. For example, the Qubit machine has both high sensitivity and broad range dsDNA dyes.

**Accuracy and Precision:** Proper calibration and optimization of fluorometer settings are essential to ensure accurate and precise quantification of nucleic acids. Standard curves generated using known concentrations of DNA or RNA standards are used to calibrate the fluorometer and establish the linear range of detection.

**Sample Handling:** Careful sample handling and preparation are critical for accurate nucleic acid quantification. It is important to take care when pipetting volumes (usually working with small volumes, a few uLs) for quantification. Sloppy pipetting will result in inaccurate quantification.

**Quality Control:** While fluorometers provide accurate measurements of nucleic acid concentration, they do not directly assess the quality or purity of the nucleic acid sample. This must be done by other methods such as spectrophotometry (e.g. nanodrop) or gel electrophoresis (to see the size and integrity of nucleic acid).

**Data Interpretation:** Proper data interpretation involves understanding the limitations of fluorometric assays and considering factors that may influence the accuracy of measurements, such as sample volume, assay conditions, and instrument settings.

**Normalization:** Quantification is important is to allow for the normalization of nucleic acid concentrations to account for variations in sample volume, input material, or extraction efficiency and it is important for comparing data across different samples or experiments.

**Applications:** Fluorometer-based nucleic acid quantification is widely used in molecular biology research, diagnostics, and biotechnology applications, including gene expression analysis, nucleic acid quantification for PCR and qPCR, next-generation sequencing library preparation, and RNA interference studies.

**Key differences between Nanodrop and Fluorometer:** NanoDrop measures absorbance, while fluorometers detect fluorescence. Fluorometers offer higher specificity, wider quantification range, and lower interference sensitivity compared to NanoDrop. NanoDrop requires small sample volumes, while fluorometers can accommodate larger volumes. NanoDrop is more affordable and versatile, while fluorometers are specialized for fluorescence-based assays.

**Activity**

* Each group will measure the DNA concentrations of a set of “samples” (each person should just 2 samples + negative RABV08 from their set, otherwise will take too long!)
  + Group 1: use provided PCR products
  + Groups 2-5: will use the PCR products they prepared at the mPCR station

**Instruction:**

* Instructor will show how to calibrate machine
* Participants will prepare tubes for measurement
  + Add 200µl of QuantiFluor® dsDNA Dye working solution to 0.5ml PCR tubes
  + Add 1 µl of sample
  + Vortex well, and incubate for 5mins (protect tube from light)
* Measure each sample on the Quantus and record values. Compare to expected values and discuss results.
* Expected concentrations:

|  |  |
| --- | --- |
| **Sample ID** | **Desired Concentration (ng/µL)** |
| RABV01 | 100 |
| RABV02 | 65 |
| RABV03 | 45 |
| RABV04 | 35 |
| RABV05 | 25 |
| RABV06 | 15 |
| RABV07 | 5 |
| RABV08 | 0 |

**Expected results/Learning outcomes:**

* An understanding of quantification- how it works and importance
* Emphasis of good practice and quality control