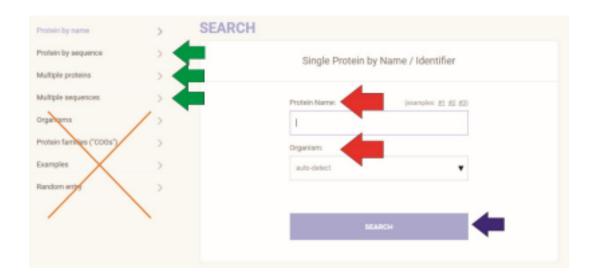
1)Data aquisition

1.1.) If we dont have our gene list, we can generate with the metasearch tool STRING 10 [https://string-db.org/]. We can choose one gene or multiple gene lists (or protein names). After that, we need to choose organism (*Mus musculus*, *Homo sapiens*,...). O



Given the gene list

IDUA

IDS

NAGLU

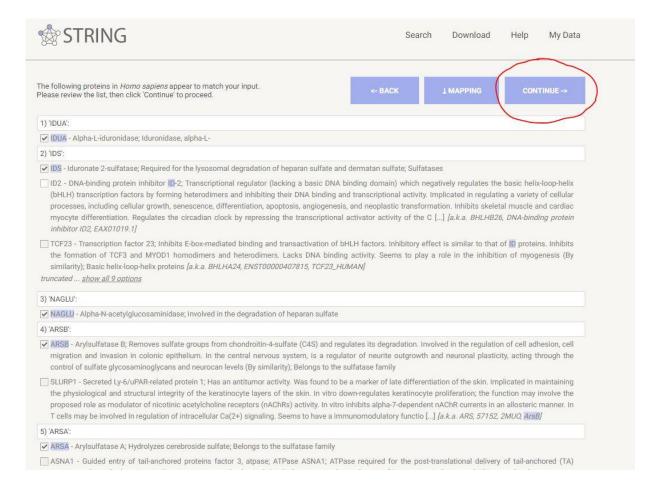
ARSB

ARSA

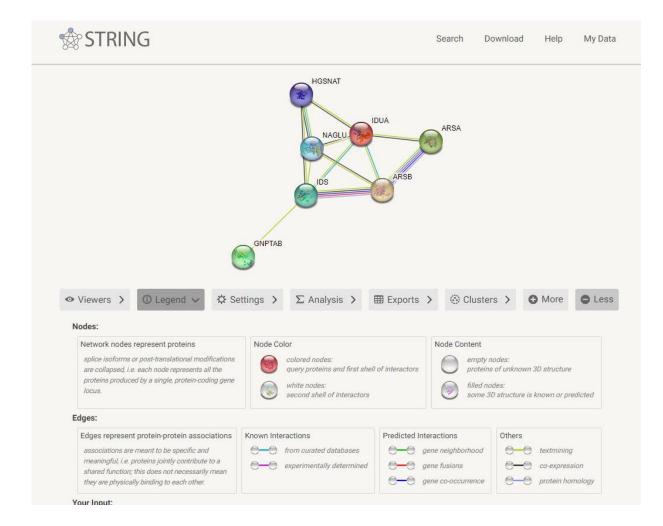
HGSNAT

GNPTAB

Select the organism - Homo sapiens and press enter. The followed output was generate:



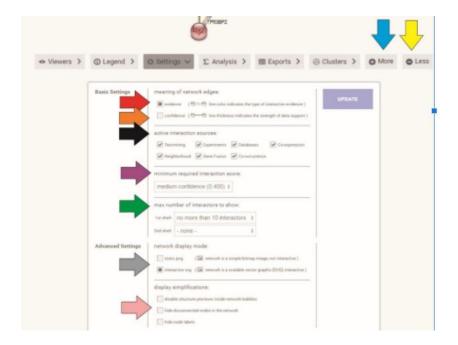
The following proteins in *Homo sapiens* appear to match your input. Please review the list, then click 'Continue' to proceed.



1.2.) Configuration to generate the network

In SETTINGS below the network we can see the available connector view types. We have two options to describe edges.

- "Confidence" the most reliable (trust) associations will be illustrated by thicker connectors, and thinner ones by less reliable ones;
- "Evidence" string will place several connectors between pairs of proteins that indicate where the information about the interaction of these proteins was taken from. This will be discussed later



We have more two options:

- "More", we can add more than 10 nodes (genes or proteins). This is not random, its based on confidence.
- "Less", remove the nodes add by "More" option.

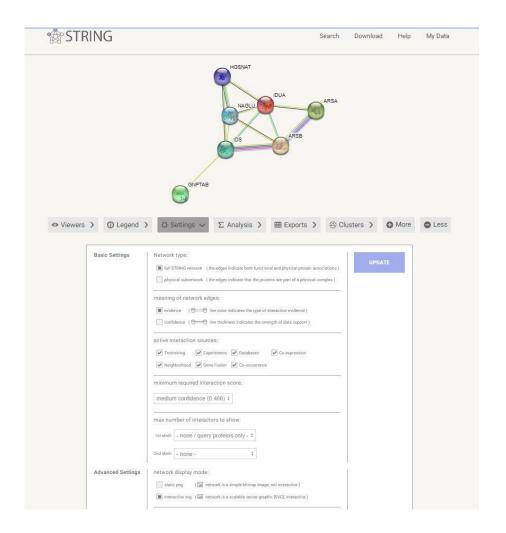
In "Active Interaction Sources" it is possible to choose which of the types of connection information is desired. Information describing what each type does can be seen in Figure 4. It is recommended to always exclude the "text mining" option, as it is the least reliable. The "gene-fusion" option is normally only used when the target organism is a prokaryote, if not, it is advisable to exclude it from the search. Information about each type of interaction can be seen on the LEGEND tab.

In "Minimum required interaction score" it is possible to choose the degree of confidence of the connections, ranging from 0 to 1 (the closer to 1, the greater the confidence). To follow the protocol presented here, leave it at the default of 0.700.

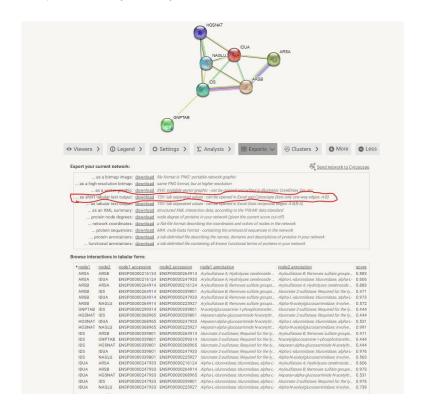
In the option "Max number of interactions to shown" we can determine how many interactions per protein is desired. The more you choose, the more loaded the network becomes, so in the protocol, choose "no more than 10 interactions" and nothing in the second bottom option.

In "Network display mode", we can use "static png", to generate the network image or "interactive svg" to browse into the interactive network display.

After this, click on UPDATE.



1.3.) Save: Exports option

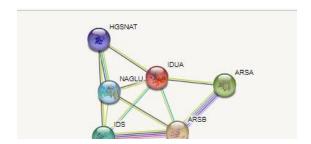


To open this generated network into Cytoscape, choose "save as simple tabular output" and Download.

The followed table be generated:

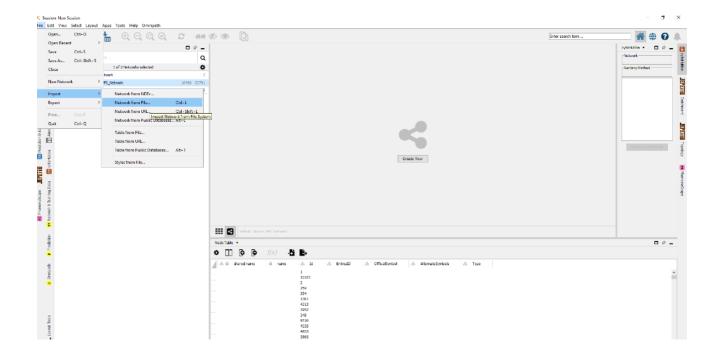
Node 1 and Node 2 columns are the most important. Others are metadata generated by string.

The meaning is ARSA gene found in Node1 column interacts with IDUA gene

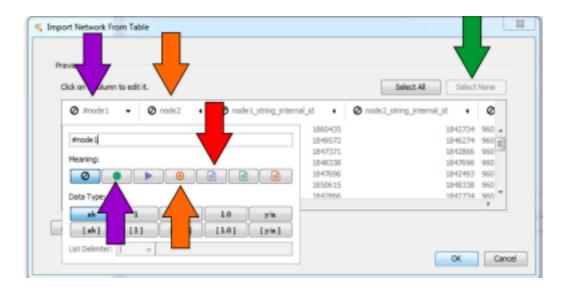


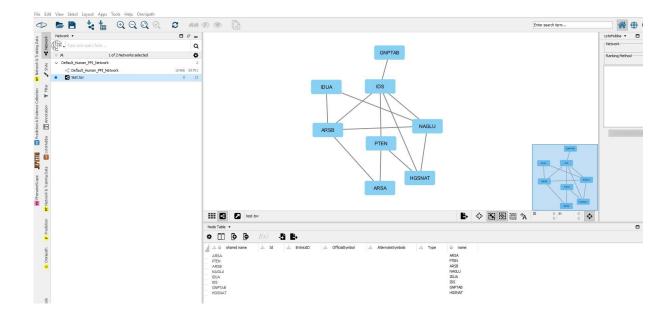
2) Importing the network file into Cytoscape

2.1.) Open cytoscape. Go to "File", "Import", "Network". "File" or press Ctrl L

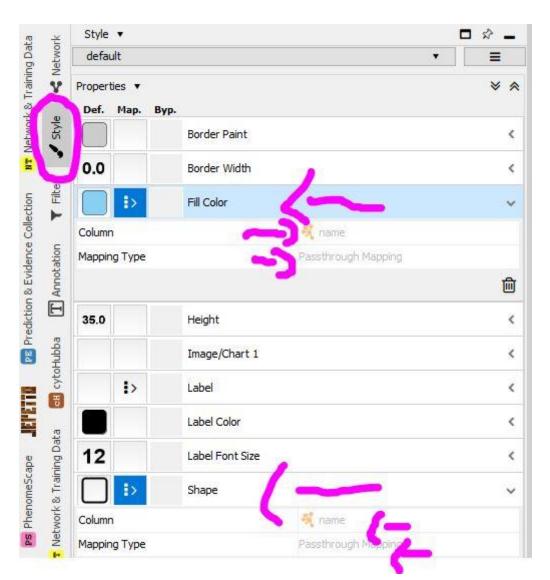


This window will be appear. To import the string network into Cytoscape, we need to click first in "Select None". After "node 1", choose "Source Node"; "Node 2", choose "Target Node". As mentioned before, Node 1 and Node means column 1 genes interact with column 2 genes. Next, "combined score" set to "Edge Attribute".

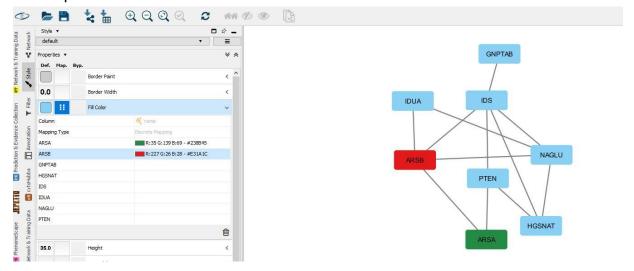




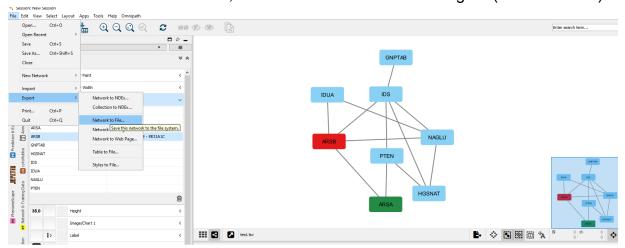
2.2.) Now we can customize the network, as follows:



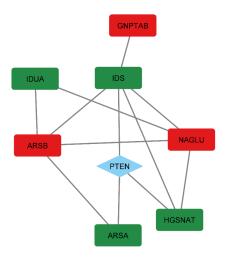
Style / Fill color / Mapping type / name / Discrete mapping / choose the colors We adopted red to upregulate genes, green to downregulated genes, and blue to transcription factors.



After finished the customization, we can save the network in a figure (PNG or JPEG)

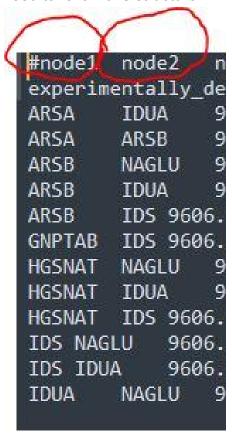


Congrats!



Using custom tables to generate networks

If we have only a gene list with Transcription factors, miRNAs, or other elements, we need to follow this structure:



And save as .tsv