

## Package 'dalycare'

October 17, 2023

**Title** Danish Lymphoid Cancer Research

**Priority** NA

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**Depends** R ( $\geq 3.5.0$ ), dplyr,

**Imports** Codes\_NPU

**LazyData** ?

**LazyDataCompression** ?

**ByteCompile** ?

**Description** Contains definitions and grouping of Danish electronic health data from SDS, RKKP, and SP.

**License** ?

**URL** NA

**NeedsCompilation** NA

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**Repository** ?

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## Index

Package “dalycare” is loaded on our NGC cloud as:

```
source("/ngc/projects2/dalyca_r/clean_r/load_data.R")
```

### Cleaning

#### `clear_ram`

##### Description

Cleans Global environment and frees RAM from NGC/dalycare

##### Usage

```
clear_ram()
```

#### `clean_RKKP_LYFO`

##### Description

Cleans the dataset RKKP\_LYFO. Works only for LYFO version 20 or higher, please see [rkkp-documentation](#)

##### Usage

```
RKKP_LYFO_CLEAN = RKKP_LYFO %>% clean_RKKP_LYFO()
```

#### `clean_RKKP_LYFO_SNOMED`

##### Description

Cleans SNOMED codes in RKKP\_LYFO. Works only for LYFO version 20 or higher, please see [rkkp-documentation](#)

##### Usage

```
RKKP_LYFO %>%  
  mutate(icd10 = clean_RKKP_LYFO_SNOMED(snomed = Reg_WHOHisto))
```

#### `clean_RKKP_CLL`

##### Description

Cleans the dataset RKKP\_CLL. Works only for CLL registry version 15 or higher, please see [rkkp-documentation](#)

##### Usage

```
RKKP_CLL_CLEAN = RKKP_CLL %>% clean_RKKP_CLL()
```

#### `clean_RKKP_DAMYDA`

##### Description

Cleans (or translates) the dataset RKKP\_DAMYDA. Works only for DAMYDA version 18 or higher, please see [rkkp-documentation](#)

##### Usage

```
RKKP_DAMYDA_CLEAN = RKKP_DAMYDA %>% clean_RKKP_DAMYDA()
```

#### `clean_RKKP_DAMYDA_SNOMED`

##### Description

Cleans SNOMED (or translates) codes in RKKP\_DAMYDA. Works only for DAMYDA version 20 or higher, please see [rkkp-documentation](#)

##### Usage

```
RKKP_DAMYDA %>%  
  mutate(icd10 = clean_RKKP_DAMYDA_SNOMED(snomed = Reg_WHOHisto))
```

### clean\_abbreviations

#### Description

Replaces commonly used Danish abbreviations containing punctuation to allow for better separation of free text into complete sentences.

E.g. 'f.eks. ' to 'f\_eks\_' pattern.

Caveat: time lapse with large datasets: subset data before use.

#### Usage

```
SP_Journalnotater_del1 %>%
  mutate(notat_text = clean_abbreviations(notat_text))
```

### clean\_lab\_values

#### Description

Cleans and converts common laboratory values with correct units based on NPU codes.

E.g. B2M nmol/l converts to mg/l.

#### Usage

```
load_npu_common()
LAB_clean = load_biochemistry(NPU.GROUP.INFECTION) %>%
  clean_lab_values()
```

### clean\_SDS\_t\_mikro

#### Description

Cleans and aggregates pathology free-text descriptions from the t\_mikro dataset.

E.g. lines 1 “Biopsy examined”, 2 “by immunohistochemistry”, and 3 “shows follicular lymphoma” convert to “Biopsy examined by immunohistochemistry shows follicular lymphoma”.

#### Usage

```
t_mikro_clean = SDS_t_mikro %>%
  clean_SDS_t_mirko()
```

### Load data

#### load\_dataset

#### Description

Loads data directly from the DALY-CARE database when specifying dataset(s).

*Dataset* may be specified as a vector of datasets.

Returns a complete list of dataset options when *dataset* is NULL (default).

Imports subset of dataset(s) when specifying *sample* as a vector of *patientid(s)*.

Also imports subset of dataset on other existing variables specifying *filter* argument.

#### Usage

```
load_dataset() #Returns a list of available datasets
load_dataset(c('patient', 'RKKP_CLL_CLEAN')) # loads both
load_dataset('RKKP_DAMYDA', value = sample(PATIENT$patientid, 100)) #only sample
load_dataset('SP_OrdineretMedicin', value = c('J06BA02', 'J01CE01'), column = 'atc')
```

### load\_datasets\_head

#### Description

Loads the head of all PERSIUNE, RKKP, SDS and SP datasets to get an overview of data structure.

Specify nrows in the head argument (e.g. head = 20).

## Usage

`load_datasets_head()` #Returns the header of all *datasets* to your global environment

## load\_all\_variables

### Description

Loads all variables of all DALY-CARE datasets: Please see Table S2 and Appendix3

### Usage

`load_all_variables() %>% print_data()`

## load\_dalycare\_icd10

### Description

Loads definitions of DALY-CARE entities based on ICD10 diagnoses into vectors located in your Global Environment in R: Please see Table S7

### Usage

```
load_dalycare_icd()
FL = t_dalycare_diagnoses %>%
  filter(diagnosis %in% ICD10.FL)
```

## load\_blood\_culture\_SP

### Description

Loads blood cultures from SP\_AlleProvesvar

### Usage

`BC = load_blood_culture_SP()`

## load\_npu\_common

### Description

Loads a list of vectors containing common NPU codes to Global Environment. You may also specify individual codes such as NPU.LYM (ie. lymphocytes) or groups of NPU codes such as GROUP.NPU.CBC (i.e. complete blood count) or NPU.GROUP.MYELOMA (i.e. standard myeloma blood test set).

### Usage

```
load_npu_common()
NPU.HGB # returns NPU02319
# Use NPUs to load_dataset() subset
load_dataset('SDS_lab_forsker', c(NPU.B2M, NPU.LYM), 'analysiscode')
SDS_lab_forsker_subset$analysiscode %>% table()
```

## load\_biochemistry

### Description

Loads dataset containing biochemistry from SDS\_lab\_forsker. 'labs' must contain NPU codes, e.g. from lists from load\_npu\_common()

### Usage

```
LAB_df = load_biochemistry(c(NPU.B2M, NPU.LDH))
BSI_df = load_biochemistry(NPU.BSI) #Blood cultures

#assign data as SDS_lab_forsker_subset into Global Environment
load_biochemistry(labs = NPU.GROUP.MSPIKE, assign = TRUE)
```

## go\_live

### Description

Loads SP (EPIC) go live dates for the three hospitals HGH, Herlev; Rigshospitalet; and SUH, Roskilde.

### Usage

```
go_live()
```

## Definitions

### filter\_first\_diagnosis

#### Description

Defines first DALY-CARE diagnosis from 't\_dalycare\_diagnoses' as the earliest occurrence, and calculates KM years from table 'patient'.

#### Usage

```
load_dataset('t_dalycare_diagnoses', 'patient') #loads all DALY-CARE diagnoses
PCD = t_dalycare_diagnoses %>%
  filter_first_diagnosis(c('DC90')) #includes any DC90.x

MZL = t_dalycare_diagnoses %>%
  filter_first_diagnosis(c('DC830C', 'DC830D', 'DC884', 'DC884A', 'DC884B', 'DC884C'))

SLL = t_dalycare_diagnoses %>%
  filter_first_diagnosis('DC830', str_contains = FALSE) #matches 'DC830'

RICHTER = t_dalycare_diagnoses %>%
  filter_first_diagnosis(c('DC833', 'DC911'), multiple = 'both') #matches both
```

### first\_diagnosis

#### Description

Defines first DALY-CARE diagnosis from view\_dalycare\_diagnoses as the earliest occurrence, regardless of source/tablename.

#### Usage

```
load_dataset('t_dalycare_diagnoses') #loads all DALY-CARE diagnoses
CLL = t_dalycare_diagnoses %>%
  first_diagnosis('DC911')

MZL = t_dalycare_diagnoses %>%
  first_diagnosis(c('DC830C', 'DC830D', 'DC884', 'DC884A', 'DC884B', 'DC884C'))
```

### right\_truncation (aka truncate\_time\_to\_event)

#### Description

Right truncates date of last follow-up as date\_event\_death\_fu and calculates time\_to\_event and event\_competing as 0 (cens), 1 (event), and 2 (comepeting). This

should always be checked in time-to-event analyses to avoid immortality bias, especially when linking data.

#### Usage

```
load_dataset('t_dalycare_diagnoses', 'patient') #loads all DALY-CARE diagnoses
CLL = t_dalycare_diagnoses %>%
  filter_first_diagnosis('DC911', string_contains = FALSE) %>%
  left_join(your_event_data %>% select(patientid, date_event), by = 'patientid') %>%
#expects your_event_data in wide format
  right_truncation(date_event, #date of event. Expects NA for non-events
    date_start = date_diagnosis, # prediction date, e.g. date_diagnosis
    date_truncation = '2023-1-1') #last event as character

library(cmprsk) #for competing risk analyses
fit = cuminc(ftime = CLL$time_to_event,
  fstatus = CLL$event_competing,
  group = 1) #group stratifies
timepoints(fit, c(1,5,10))

library(Publish) #for plotting competing risk analyses with no. at risk.
aj = prodlim(Hist(time_to_event, event_competing)~1, data=CLL)
plot(aj)
#"Error in plot.new()" may be rectified by: par(mar=c(1,1,1,1))
```

#### [scr\\_low\\_48h](#)

##### Description

Defines lowest serum creatinine (scr) within 48 hours using lab\_forsker data. SDS\_lab\_forsker data should be filtered to contain creatinine only (NPU.KREA) to avoid time-lapse. Used to define acute kidney injury (AKI).

#### Usage

```
load_npu_common()
load_dataset('SDS_lab_forsker', c(NPU.KREA), 'analysiscode') #loads creatinine
DATA_scr_low_48h = SDS_labforsker_subset %>%
mutate(
  cpr_enc = patientid,
  date_time = as.numeric(seconds(as.POSIXct(paste(samplingdate, samplingtime)))),
  i.scr_inhos = 0
) %>%
scr_low_48h()
```

#### [scr\\_low\\_7d](#)

##### Description

Defines lowest serum creatinine (scr) within 7 days using lab\_forsker data. SDS\_lab\_forsker data should be filtered to contain creatinine only (NPU.KREA) to avoid time-lapse.

#### Usage

```
load_npu_common()
load_dataset('SDS_lab_forsker', c(NPU.KREA), 'analysiscode') #loads creatinine
DATA_scr_low_48h = SDS_labforsker_subset %>%
mutate(
```

```
cpr_enc = patientid,  
date_time = as.numeric(seconds(as.POSIXct(paste(samplingdate, samplingtime)))),  
i.scr_inhos = 0  
) %>%  
scr_low_7d()
```

#### scr\_base\_median

##### Description

Defines baseline serum creatinine (BL scr) a rolling median using lab\_forsker data. SDS\_lab\_forsker data should be filtered to contain creatinine only (NPU.KREA) to avoid time-lapse.

##### Usage

```
load_npu_common()  
load_dataset('SDS_lab_forsker', c(NPU.KREA), 'analysiscode') #loads creatinine  
DATA_scr_low_48h = SDS_labforsker_subset %>%  
mutate(  
  cpr_enc = patientid,  
  date_time = as.numeric(seconds(as.POSIXct(paste(samplingdate, samplingtime)))),  
  i.scr_inhos = 0  
) %>%  
scr_base_median()
```

#### AE\_AKI

##### Description

Defines acute kidney injury based on a 1.5x increase from the baseline serum creatinine (scr\_base\_median) within 7 days (scr\_low\_7d) or an absolute scr increase of 26.5  $\mu\text{mol/L}$  within 48 hours (scr\_low\_48h) using lab\_forsker data. SDS\_lab\_forsker data should be filtered to contain creatinine only (NPU.KREA) to avoid time-lapse.

##### Usage

```
load_dataset('SDS_lab_forsker', c(NPU.KREA), 'analysiscode') #loads creatinine  
CREATININE_clean = SDS_labforsker_subset %>% clean_lab_values()  
AKI = CREATININE_clean %>% AE_AKI(value = value2)
```

##### Citation

Carrero JJ et al. Kidney Int. 2023 Jan;103(1):53-69.

#### CTCAE\_lab

##### Description

Defines CTC adverse events (AE) from biochemistry. Works only with lab\_forsker data. SDS\_lab\_forsker data should be filtered to contain NPU of interest to avoid time-lapse. E.g. May calculate 'ANEMIA', 'THROMBOCYTOPENIA', 'DIC', and 'HEMOLYSIS'.

##### Usage

```
load_npu_common()  
HGB = load_biochemistry(NPU.HGB) %>% clean_lab_values()  
ANEMIA_AE = HGB %>%  
  CTCAE_lab() %>%  
  select(patientid, ANEMIA.GRADE, everything())
```

```
HEMOLYSIS = load_biochemistry(NPU.GROUP.HEMOLYSIS) %>%
  clean_lab_values()
# expect time-lapse for large samples, consider down sampling
HEMOLYSIS_AE = HEMOLYSIS %>%
  CTCAE_lab() %>%
  select(patientid, HEM.GRADE, everything())
```

### TX\_group

#### Description

Groups treatment protocols into meaningful groups as class characters.

#### Usage

```
SP_Behandlingsplaner_del1 %>% TX_group() %>% pull(TX_group)
```

### filter\_virus

#### Description

Subsets RSV, SARS-CoV-2 (SARS) and seasonal influenza (FLU) into class character (type) and result.

#### Usage

```
SP_Bloddyrkning_del1 %>% filter_virus() %>% select(patientid, type, result)
```

#### Citation

Niemann et al. *Blood*. Aug 4 2022;140(5):445-450.

### filter\_sentence

#### Description

Subsets all free-text sentences (i.e. from \\. to \\.) containing pattern.  
 Caveat: Free text often contains punctuation such as abbreviation causing separation; please see clean\_abbreviations()

#### Usage

```
SP_Journalnotater_del1 %>% filter_sentence(notat_text, 'SAGM')
SDS_t_mikro_ny %>% filter_sentence(v_fritekst, 'EBER')
```

### ATC\_polypharmacy

#### Description

Calculates number of 1<sup>st</sup> to 5<sup>th</sup> level ATC codes per patient and defines polypharmacy as ≥5 drug classes.

#### Usage

```
SDS_epikur %>% ATC_polypharmacy(level = 3) %>% pull(Polypharmacy)
```

#### Citation

Brieghel et al. ASH annual meeting 2023. P5133

### COD2

#### Description

Groups cause of death (COD) ICD10 codes into meaningful groups. Prioritizes infections.

#### Usage

```
SDS_t_dodsaarsag_2 %>% COD2()
```



#### Citation

Rotbain et al. *Leukemia*. 2021;35(9):2570-2580.

#### CCI

##### Description

Calculates Charlson comorbidity index (CCI) scores from ICD10 codes.  
 exclude\_CLL\_score = FALSE (default) includes the DC911 score, if present.  
 include\_LC\_score = FALSE (default) calculates the LC score only if present.

##### Usage

```
SDS_t_adm %>% CCI(icd10 = c_adia) %>% select(patientid, CCI.score, CCI.2011.update)
view_diagnoses_all %>% CCI() %>% select(patientid, CCI.score, CCI.2011.update)
```

#### Citation

Quan et al. *Med Care*. 2005;45:1130-9 as CCI.score

Quan et al. *Am J Epidemiol*. 2011;173:676-82 for CCI.2011.update

#### CLL\_CI

##### Description

Calculates CLL comorbidity index (CLL-CI) scores from vascular, GI, and endocrinology defined SKS codes (LPR), ATC codes (EPIKUR), and ICD10 codes (diagnoses\_all).

##### Usage

```
CLL_cohort = t_dalycare_diagnoses %>%
  filter_first_diagnosis('DC911', str_contains = FALSE) #create CLL cohort
CLL_CI_cohort = CLL_cohort %>%
  CCI_CI() # Input only requires variable with 'patientid'
```

#### Citation

Rotbain et al. *Blood Adv*. 2022;6(8):2701-6

#### NMI

##### Description

Calculates Nordic Multimorbidity Index (NMI) scores from ICD10 and ATC codes before date of diagnosis (as date\_diagnosis).

##### Usage

```
your_cohort = t_dalycare_diagnoses %>%
  filter_first_diagnosis(ICD10.CLL) %>%
  NMI() %>%
  select(patientid, NMI_score)
```

#### Citation

Kristensen et al. *CLEP*. 2022;14:567-79

#### ATC\_AB

##### Description

Subsets and groups all antimicrobials.

##### Usage

```
SDS_epikur %>% ATC_AB()
SP_Administreret_Medicin %>% ATC_AB()
```

### ATC\_hypertensives

#### Description

Subsets and groups all antihypertensive drugs.

#### Usage

```
SDS_epikur %>% ATC_hypertensives()
SP_Administreret_Medicin %>% ATC_hypertensives ()
```

### ATC\_opioids

#### Description

Subsets and groups all opioids.

#### Usage

```
SDS_epikur %>% ATC_opioids()
SP_Administreret_Medicin %>% ATC_opioids()
```

### qSOFA

#### Description

Calculates qSOFA scores from vital values assuming that AVPU less than alert (A) replaces GCS < 15.

#### Usage

```
SP_VitaleVaerdier %>% qSOFA() %>% pull(qSOFA)
```

### BMI

#### Description

Calculates body mass index (BMI) and body surface area (BSA) from vital values.

#### Usage

```
SP_VitaleVaerdier %>% BMI() %>%
  select(patientid, BMI, BSA_DuBois, BSA_Mosteller)
```

### BSA

#### Description

Calculates body mass index (BMI) and body surface area (BSA) from vital values.

#### Usage

```
SP_VitaleVaerdier %>% BSA() %>%
  select(patientid, BMI, BSA_DuBois, BSA_Mosteller)
```

### transform\_2\_ERIC

#### Description

Transforms DALY-CARE data to the standard format for submission of data to projects within European Research Initiative on CLL (ERIC) and the ERIC database. Data defined from RKKP\_CLL, RKKP\_LYFO (for SLL cases), LAB\_IGHVIMGT, patient, t\_dalycare\_diagnoses, diagnoses\_all (comorbidity and second malignancy), SDS\_t\_doedsaarsag and CLL\_TREAT\_CLEAN (only if 2<sup>nd</sup> line treatment is missing from RKKP).

write\_xlsx = TRUE. Writes an Excel file with 2 sheets and appreciated by ERIC

pseudonymize = TRUE. Creates pseudonymized `Patient Lab id`.

NB! Always deselect the DALY-CARE patientid before sharing data with Thomas Chatzikonstantinou via secure warehouse: [thomas.chatzikonstantinou@certh.gr](mailto:thomas.chatzikonstantinou@certh.gr)

#### Usage

```
CLL_SLL_cohort = t_dalycare_diagnoses %>%
```

```

filter_first_diagnosis(c('DC911', 'DC833'), str_contains = FALSE) #create CLL/SLL
#cohort
ERIC_data = CLL_SLL_cohort %>%
  transform_2_ERIC() # Input only requires variable with 'patientid'
ERIC_data = CLL_SLL_cohort %>%
  transform_2_ERIC(write_xlsx= TRUE, pseudonymize = TRUE) # writes Excel file with
#pseudo IDs

```

#### Citation

Chatzidimitrou et al. *Hemasphere*. 2020;4(5):e425

#### CLL\_IPI

##### Description

Calculates CLL-IPI risk as class factor.

##### Usage

```
RKKP_CLL_CLEAN %>% CLL_IPI() %>% pull(CLL.IPI) %>% table()
```

#### Citation

da Cunha-Bang et al. *Blood*. 2016;128(17):2181-3.

#### CLL\_WONT

##### Description

Calculates CLL-WONT risk as class factor. Needs ALC (NPU02636) and LDH (NPU19658; NPU19978; NPU19975) from e.g. SDS\_lab\_forsker. Consider skipping data preparation.

##### Usage

```

# Data preparation
load_npu_common()
LAB = load_biochemistry (labs = c(NPU.LYM, NPU.LDH)) %>%
  clean_lab_values()
ALC = LAB %>%
  filter(NPU %in% NPU.LYM) %>%
  transmute(patientid, date_ALC = samplingdate, ALC = value2)
LDH = LAB %>%
  filter(NPU %in% NPU.LDH) %>%
  transmute(patientid, date_LDH = samplingdate, LDH = value2)

# Data preparation continued...
RKKP_CLL_WITH_ALC_AND_LDH = RKKP_CLL_CLEAN %>%
  left_join(ALC, by = 'patientid') %>%
  left_join(LDH, by = 'patientid') %>%
  mutate(time_ALC = diff_days(Date_diagnosis, date_ALC),
         time_LDH = diff_days(Date_diagnosis, date_LDH)) %>%
  filter(time_ALC <= 0, time_ALC >= -90,
         time_LDH <= 0, time_LDH >= -90) %>%
  group_by(patientid) %>%
  arrange(patientid, desc(time_ALC), desc(time_LDH)) %>%
  slice(1) %>%
  ungroup()

# CLLWONT calculation
RKKP_CLL_WITH_ALC_AND_LDH %>% CLL_WONT() %>%
  pull(CLLWONT) %>% table()

```

#### Citation

Brieghel et al. *Eur J Haematol*. May 2022;108(5):369-378.  
 Brieghel et al. *Blood Adv*. 2024;8(16):4449-56.

#### NCCN\_IPI

##### Description

Calculates NCCN-IPI risk for DLBCL as class factor.  
 NB! Input is complex and not generalizable.

##### Usage

```
RKKP_LYFO %>% clean_RKKP_LYFO() %>%  
  NCCN_IPI() %>% pull(NCCN_IPI) %>% table()
```

##### Citation

Zhou et al. *Blood*. Feb 6 2014;123(6):837-42.  
 Jelacic et al. *BJC*. 2023;13(1):157.

#### MIPI

##### Description

Calculates MIPI risk for Mantle cell lymphoma as class factor

##### Usage

```
RKKP_LYFO %>% clean_RKKP_LYFO() %>%  
  MIPI() %>% pull(MIPI) %>% table()
```

##### Citation

Hoster et al. *Blood*. Jan 15 2008;111(2):558-65.

#### IPS

##### Description

Calculates IPS risk for Hodgkin lymphoma as class factor

##### Usage

```
RKKP_LYFO %>% clean_RKKP_LYFO() %>%  
  IPS() %>% pull(IPS) %>% table()
```

##### Citation

Hasenclever et al. *NEJM*. 1998;339:1506-14.

#### rIPSSWM

##### Description

Calculates rIPSSWM risk for Waldenström macroglobulinemia (WM) and LPL as class factor.

##### Usage

```
RKKP_LYFO %>% clean_RKKP_LYFO() %>%  
  rIPSSWM() %>% pull(rIPSSWM) %>% table()
```

##### Citation

Kastritis et al. *Leukemia*. Nov 2019;33(11):2654-2661.

#### MAYO\_20\_20\_20

##### Description

Calculates Mayo Institute 20-20-20 risk for progression of smoldering myeloma as class factor.

##### Usage

```
RKKP_DAMYDA_CLEAN %>%  
  MAYO_20_20_20() %>%  
  pull(MAYO_20_20_20) %>%
```

table()

#### Citation

Mateos et al. *Blood cancer journal*. Oct 16 2020;10(10):102

#### R\_ISS

##### Description

Calculates revised ISS (R-ISS) risk for multiple myeloma as class factor.

##### Usage

RKKP\_DAMYDA\_CLEAN %>% R\_ISS() %>% pull(R\_ISS) %>% table()

#### Citation

Palumbo et al. *J Clin Oncol*. Sep 10 2015;33(26):2863-9.

#### R2\_ISS

##### Description

Calculates second revised ISS (R2-ISS) risk for multiple myeloma as class factor.

##### Usage

RKKP\_DAMYDA\_CLEAN %>% R2\_ISS() %>% pull(R2\_ISS) %>% table()

#### Citation

D'Agostino et al. *J Clin Oncol*. Oct 10 2022;40(29):3406-3418.

#### RW\_ISS

##### Description

Calculates revised-world ISS (RW-ISS) risk for multiple myeloma as class factor.

##### Usage

RKKP\_DAMYDA\_CLEAN %>% RW\_ISS() %>% pull(RW\_ISS) %>% table()

#### House keeping

##### is\_odd

##### Description

Logical output from numeric values .

##### Usage

sample(1:10, 5) %>% is\_odd()

##### diff\_days

##### Description

Calculates numeric date intervals in days.

##### Usage

diff\_days(date\_start, date\_end)

##### diff\_years

##### Description

Calculates numeric date intervals in years.

##### Usage

diff\_years(date\_start, date\_end)

### filter\_str\_detect

#### Description

Subsets data with strings containing vector of patterns.

#### Usage

```
CLL = t_dalycare_diagnoses %>%
  filter_first_diagnosis('DC911')
load_dataset('SP_Behandlingsplaner_del1', CLL$patientid, 500)

SP_Behandlingsplaner_del1_subset %>%
  filter_str_detect(protocol_navn, c('OBI', 'VEN'))
```

### str\_between

#### Description

Subsets string character between two patterns for text-mining purposes.

#### Usage

```
load_dataset('SP_Journalnotater_del1', patient$patientid, 500)

SP_Journalnotater_del1_subset %>%
  filter(notat_type=='AOP') %>%
  mutate(sex = str_between(notat_text, 'årig', c('henvist|møder|kendt|indlægges')))%>%
  pull(sex)
[1] "mand "
[2] "mand "
[3] " mand "
[4] ""
[5] "kvinde "
[6] " kvinde "
[7] " kvinde "
[8] "kvinde "
[9] "kvinde "
[10] "kvinde."
```

### censor\_med\_keep\_first

#### Description

Subsets dates x days apart. Useful for censoring medication in grace period.

#### Usage

```
censor_med_keep_first(date, days_karens = 14)
```

#### Citation

Packness et al. EHA annual meeting 2022. P1596

### cut\_year

#### Description

Cuts year-time into monthly intervals (e.g. 3-month intervals, by = 0.25) and outputs class factor.

#### Usage

```
Data %>% censor_med_keep_first(year_cut = cut_year(time = Time, by = 0.25))
```

### n\_patients

#### Description

Counts distinct patients in a dataset assuming that patients are found in 'patientid'.

#### Usage

```
patient %>% n_patients()
```

### nrow\_npatients

#### Description

Counts distinct patients and number of rows in a dataset assuming that patients are found in 'patientid'.

#### Usage

```
patient %>% nrow_npatients()
```

### slice\_closest\_value

#### Description

Slices the absolute closest value to a baseline date (date\_baseline) within time interval (interval\_days, c(-90, 0) default). Useful when adding lab values to wide format data.

#### Usage

```
load_dataset('SP_AlleProvesvar', NPU.HGB, 'component')  
load_dataset('patient')
```

```
patient %>%  
  left_join(SP_AlleProvesvar_subset %>%  
    transmute( patientid,  
               date_lab = as_date(specimn_taken_time),  
               HGB = as.numeric(ord_value))) %>%  
  slice_closest_value(date_baseline = date_diagnosis, date_value = date_lab)
```

## ggplots

### KM\_plot

#### Description

Depends on library('ggplot') and library('survminer').  
Plots survminer::ggsurvplot with really nice aesthetics.

#### Usage

```
CLL = t_dalycare_diagnoses %>%  
filter_first_diagnosis('DC911')  
  
fit = survfit(Surv(time_dx_death, status) ~ sex, data = CLL)  
KM_plot(fit)
```

### tile\_pairwise\_survdif

#### Description

Depends on library('ggplot') and library('survminer').  
Tiles pairwise log-rank tests from survminer::pairwise\_survdif for visual purposes.

#### Usage

```
CLL = t_dalycare_diagnoses %>%  
filter_first_diagnosis('DC911') %>%  
left_join(RKKP_CLL_CLEAN, by = 'patientid')  
  
pairwise_survdif(Surv(time_dx_death, status) ~ CLL.IPI, data = CLL, p.adjust.method =  
'none') %>% tile_pairwise_survdif(position = 'LL', palette = c(1,2,3,4), labs = FALSE)
```