

# BIS101 F2014 Mutation

## Point Mutations (single nucleotide/base)

We have mentioned these briefly, but now in more detail. Other forms of mutation include TEs, chromosomal aberrations

**bp substitution.** note substitution has a different connotation in popgen.

- **transition vs. transversion**
  - C-T/A-G vs. not
  - but keep track of both strands: GC->AT transition but GC->TA transversion
- transitions caused by wrong form of base.
  - Each of the bases exists in >1 form: tautomers (differ due to movement of an H and shifting of a double bond).
  - Normal "keto" form is found in DNA.
  - But shift of the bonds (ionized form) will lead to slightly different form that then pairs incorrectly. I.E. imino cytosine pairs with adenine instead of guanine during replication.
  - next round of replication the CA -> TA in one of the daughter cells.
- transversions caused by accidental pairing of pyrimidine w/ self or purine w/ self during replication

**indel** insertion/deletion

- determining ancestry hard (show with tree)
- **Replication slippage.** Draw CTGAGAGAGA and complement.
- pops out a single strand loop. loop either deleted or added next round.
- expansion of trinucleotide repeat arrays due to rep. slippage can cause diseases in humans like fragile X, huntington's disease (spinal and muscular atrophy)

sometimes adaptation/evolution occurs from new mutations that arise after selection, but often occurs as a result of **standing genetic variation** or mutations segregating in the population.

common misconception is that selection agent arises (say antibiotics) and that CAUSES adaptive mutations to arise.

Luria/Delbruck test in book is classic experiment testing this. If selective agent (antibiotics) causes mutation, all plates should show same rate. But if mutations occur spontaneously, depending on when it arose different numbers of colonies will have resistance.

## Effects on protein coding sequence

recall that DNA is transcribed to RNA, and RNA translated to protein in codons of 3bp.

**synonymous.** likely effect ? usually mild

- codon usage bias: if most of the tRNAs use one codon, then another codon even of same AA will have to wait longer, so gene is transcribed slower.
- weak or strong selection ? weak. generally only found in genes that are really highly expressed. why ? kudla suggest so doesn't mess up other genes.

**nonsynonymous** (missense). likely effect ? depends

- AA come in all sorts of categories: size, charge, hydrophobic/hydrophilic, etc.
- replacement of an AA with a similar one might be neutral. sometimes only need an AA of particular quality.
- example of protein storage gene globulin-1

**premature stop** (nonsense, truncation). likely effect ? bad

**frameshift**

- can be caused by substitution ? no. only indel
- shifts all codons -- shifts the "frame" of translation
- what happens if multiples of 3 ?

**splice-site.** what is a splice site ?

- usually bad, because turns intron -> exon and messes everything up (ex. in Yin et al. paper!)

## Effects on noncoding

Harder to predict. Some proportion of noncoding DNA likely junk, so who cares.

In Drosophila though a majority of noncoding mutations seem to be influenced by selection.

Why would noncoding mutation matter ?

All sorts of important DNA regions that don't code for protein: promoters, enhancers, etc. -- regions where proteins or RNA that regulate a gene bind either to DNA or RNA

Consequences include change in expression level, in where expression occurs. Can be mild, but can be just as deleterious as changes in protein coding regions.

## Mechanisms of spontaneous mutations

DNA replication as mentioned above

- In total, replication error is on order of  $10^{-5}$ !! In human genome that means replication causes 30,000 new mutations!

## Spontaneous lesions

Naturally occurring damage to DNA.

depurination - purine base lost when bond to deoxyribose is broken. happens spontaneously and can be exacerbated by environment. some studies suggest as much as 5K of these per day in human cells. \* resulting apurinic site has no base, so nothing to pair! wrong base likely incorporated in next round

deamination - loss of amino group cytosine → uracil

- next round of replication, uracil pairs with **?** adenine. so GC → AT what kind **?** (transition)
- especially common in methylated cytosine (and what bisulfite sequencing does! (correction from last time))

## Induced mutation

Things that can cause mutants called **mutagens**

### base damage

- UV can cause pyrimidines (CTU) on same strand to covalently bond, preventing DNA replication or transcription. these are the primary cause of skin cancer in humans. e.g. thymine dimer
  - organisms living at high altitude have to adapt to increased UV radiation. CORN!
- ionizing radiation can cause depurination, strand breaks, etc.

**base analogs.** molecules sufficiently similar that can be incorporated, but pair wrong. (2-amino-purine e.g.)

**alteration of bases** causing them to pair incorrectly

**intercalation** **?** molecules that can slip between bases and cause insertion/deletions during replication.

- how many of you worked with gel? EtBr is intercalating agent. how it binds DNA on gel, and what happens when it gets on you!

## Repair mechanisms

DNA polymerase

- polymerase adds bases 5'→3' but can remove bases 3'→5'
- recognizes errors, cuts out and repairs
- mostly due to a very tight fit in the active site.
- different polymerases have different abilities to recognize/tolerate different mutations
- This drops from  $10^{-5}$  to  $10^{-7}$ , but still 300 mutations every replication!

#### Direct reversal

- other enzymes can directly reverse some mutations (e.g. thymine dimers) but in general rare

#### Several **homology based** ? pathways to fix missing/bulky mutations

- suite of enzymes first remove altered base
- cut out surrounding DNA
- polymerase fills in, ligase seals shut
- base excision repair for nonbulky changes, nucleotide excision repair for bigger ones, etc.
- can recognize e.g. uracil which doesn't belong

#### Finally **mismatch repair**

- similarly cuts out error, fills in
- How recognize which is the wrong base ?
  - mentioned cytosine methylation, but adenine also methylated
  - since DNA replication is semiconservative, old strand will have methylation and errors always in new strand!
  - not well understood in eukaryotes!
- mutations in genes that do this in humans lead to e.g. syndromes that are high risk for cancer. not that rare: odds are good 1 person in this room has it.
  - the locus is haplosufficient. cells with one good copy fcn normal
  - so is mutation dominant or recessive ?
  - normally yes, but here what happens is anytime you accidentally lose or knock out the other copy, a cell with just the mutant copy (hemizygous) goes nuts and causes cancer. so the gene is haplosufficient but the syndrome is dominant!

Together these reduce from  $10^{-7}$  to  $10^{-9}$ , so 3 errors per rep. That's still a lot considering how often DNA is replicated.

- will all syndrome be inherited ?
- no. so if you spill EtBr on your elbow, you may get elbow cancer, but it won't effect your kids.

These repairs mechanisms are **error-free** in that don't cause new errors.

**Translesion synthesis** or SOS system is sort of emergency backup. A polymerase that can tolerate blockages that would stop normal polymerases. But these tolerant polymerases don't have error-correction, so cause new errors (but better than cell death)

## **DSB repair**

DSBs can be spontaneous, or caused by radiation.

How repair? No complementary strand to go off of.

**synthesis-dependent strand annealing.** uses strand invasion similar to what happens in recombination to repair one chromosome using the homolog as a template (instead of other strand). when can this happen **?** dividing cells only.

**NHEJ nonhomologous end joining.** enzymes that grab on to broken ends to prevent further problems, trim, and then ligate joins back together. when can this happen **?** anytime. and most cells are not undergoing division most the time.