

BIS101 F2013 Lecture 13: Bacterial Gene Expression

Gene regulation possibilities

Why regulate gene expression?

- * respond to changing environment
- * save resources -- don't waste amino acids, nucleotides, ATP making stuff that isn't necessary
- * development -- in eukaryotes different tissues, different time periods, need different sets of genes.

organisms have a number of methods to regulate gene expression. today we'll focus on some ways it's done in prokaryotes, that get across many of the main concepts.

**for general knowledge of operon Ch. 5 not needed, but strongly recommended in order to do HW and information on test."

constitutive? (always on) vs regulated:

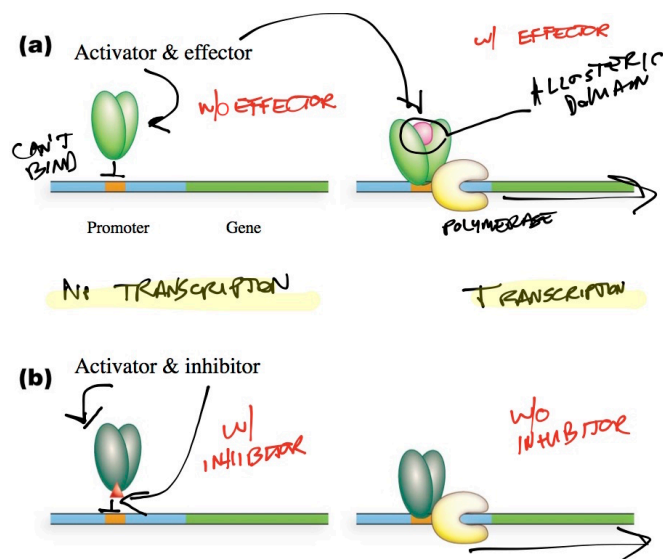
transcriptional regulation can affect 1) initiation 2) amount

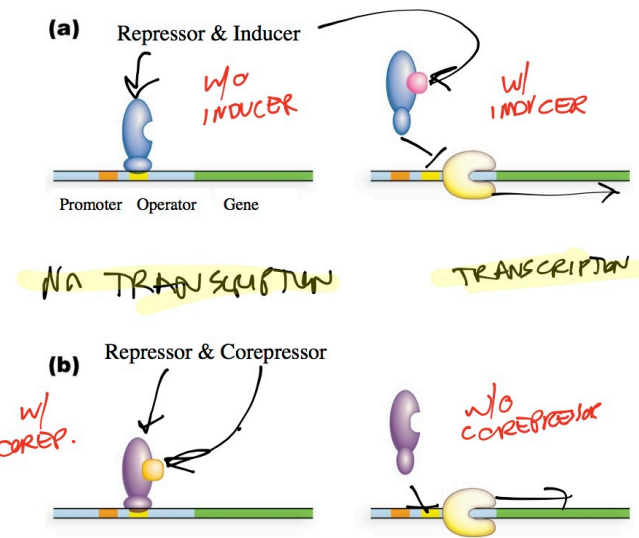
- **Activator**

- binds to activator binding site on DNA (on or near promoter region, or in enhancer sites in eukaryotes)
- facilitate binding of RNA polymerase and initiate transcription
- can require binding of an **effector**
- **allosteric domain** and conformation change
- **inhibitor** can prevent from binding

- **Repressor**

- binds to control sequence called an **operator** and prevents transcription
- **corepressor** molecule can be required for binding
- **inducer** molecule can prevent from binding





cis regulation vs. **trans** regulation cis means "on this side" ? -- a sequence that affects gene regulation of nearby, linked genes. trans means affects distant or unlinked genes.

Gene: What is a gene?

Fundamental physical and functional unit of heredity which carries information from one generation to the next

A segment of DNA, composed of a transcribed region and usually a regulatory sequence that makes possible transcription.

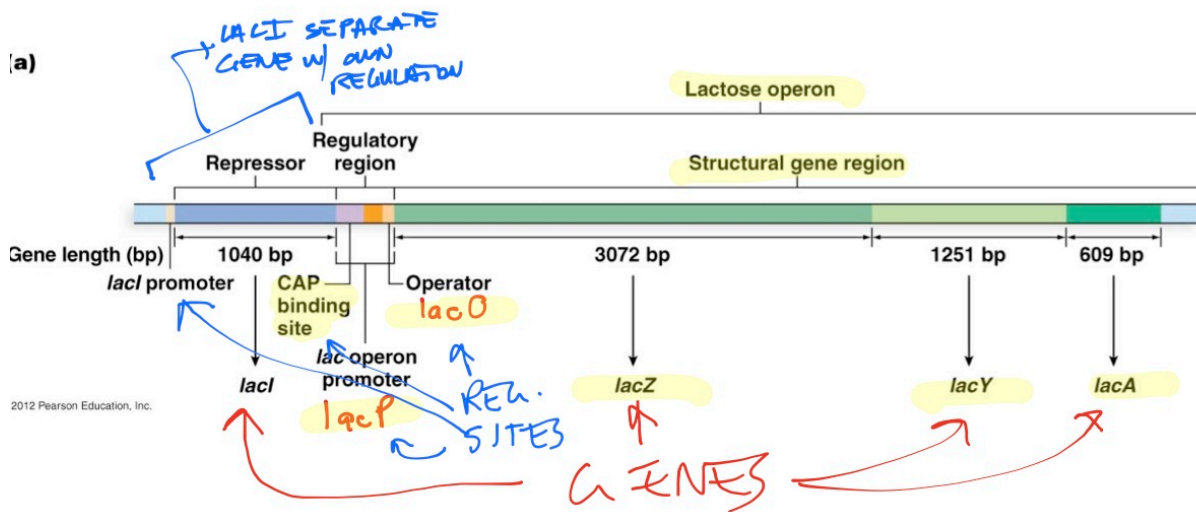
What is an operon ? a group of genes that are coregulated. Each codes for a separate product, but all share common transcription and regulation.

makes for efficient regulation of a system or pathway.

polycistronic transcription (vs. monocistronic): makes multiple proteins from same transcript; common in bacteria, rare in eukaryotes

lac operon

- *lac* operon



normally *E. coli* uses glucose for metabolism

lac operon is **inducible** -- normally not highly expressed, doesn't take up or use lactose in absence of glucose, *lac*⁺ (wild type) can induce expression of operon to use lactose

confusing naming

genes:

lacZ *lacY* *lacA* are protein-coding DNA

- permease ***lacY*** allows lactose in
- lactose → galactose + glucose by enzyme beta-galactosidase (***lacZ***)
 - occasionally produces allolactose
- ***lacA*** transacetylase → not well known but perhaps transfer acetyl group from nonmetabolizable sugars so as to prevent reentry into cell?

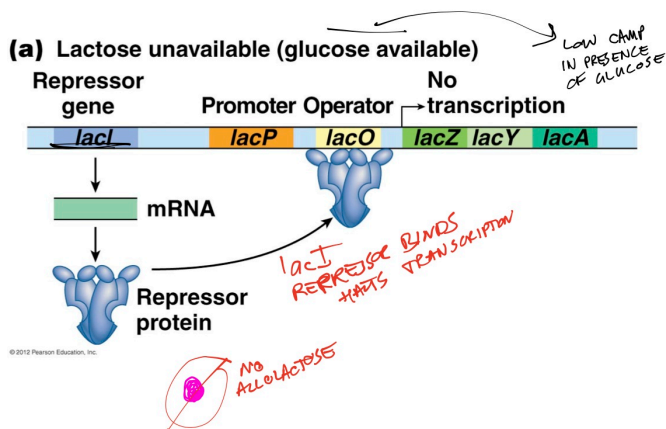
regulatory regions

lacP promoter

CAP binding site binds CAP-CAMP complex: catabolite activator protein (CAP) and cyclic adenosine monophosphate (cAMP) (signal transduction). bends DNA to open for RNA pol and ++ expression

glycolysis **?** (metabolic pathway get energy from glucose) reduces availability of cAMP, keeping expression low

Glucose, no lactose



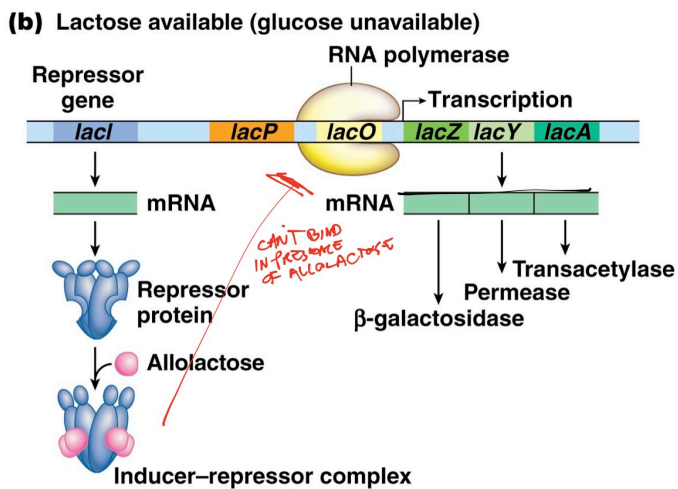
lacI repressor protein binds to *lacO* operator

lacO actually overlaps promoter and transcript; RNA pol can't bind

lacI operate cis or trans ? (trans)

mutations in *lacO* cis or trans ? (cis)

Lactose, no glucose



RNA pol binds to *lacP* promoter (not operator as in figure)

cAMP abundant, CAP-cAMP open DNA → lots of transcription

Problem ? need lactose to get started, but how's it get in with no permease ?

repressor falls off occasionally!

Glucose and lactose?

cAMP very limited, so low expression of operon

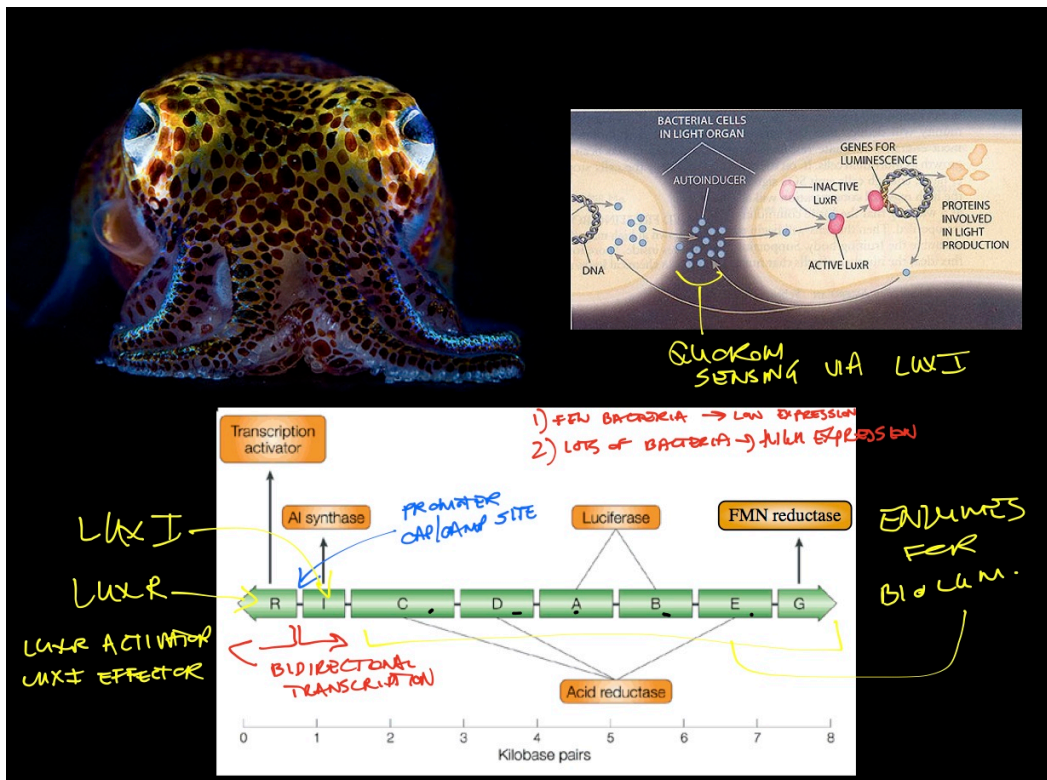
allolactose inducer may be present, no binding of repressor

Mutants of lac operon

- Oc repressor can't bind: effect ? constitutive expression of lac operon (cis/trans ?)
- I- bad repressor, can't bind.
- Z- or Y- can't make functional genes
- complementation ? because can make a merodiploid by using F plasmid

Go over F' I+P+OcZ-Y+A+/ I+P+O+Z+Y-A+ F' can't make galactosidase, bacteria can't make permease. neither functions F' also constitutively expressed b/c can't bind promoter. het functions normally -> constitutive expression of permease, inducible expression of galactosidase

Hawaiian bobtail squid



Operon worked out by Joanne Engelbrecht here at UCD.

bioluminescent underbelly to be camouflaged at night against starry/moonlight sky contains special organs to capture and cultivate *Alivibrio fischeri* bacteria

- fischeri* lux operon is expressed under glucose starvation (CAMP binding site) bidirectional transcription between I & R

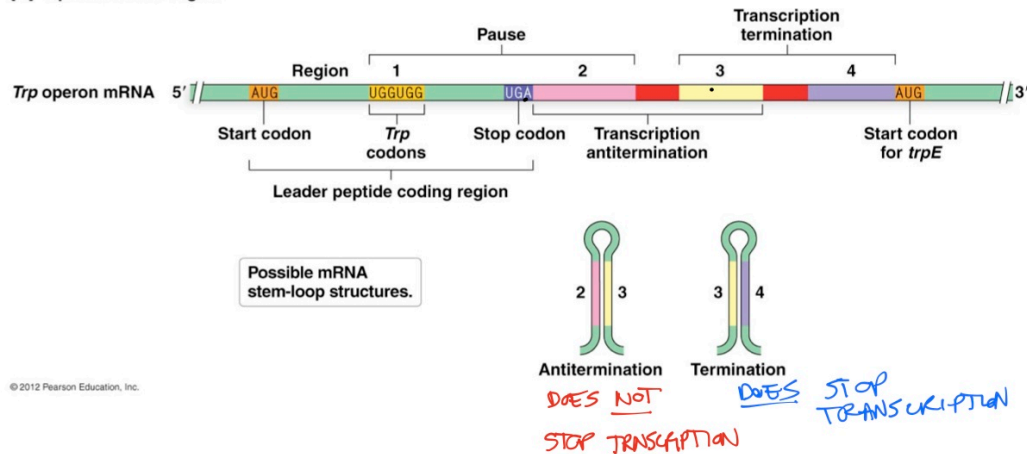
also binding site for transcription activator luxR, which only binds when bound to its effector luxI

luxI protein secreted from cell. a single cell doesn't produce enough luxI to bioluminesce (free-living *A. fischeri* don't bioluminesce!)

when cells are concentrated, enough luxI produced -> feedback loop

Attenuation of trp operon

(a) *TrpL* attenuator region



(draw "some genes to make tryptophan" downstream)

makes amino acid tryptophan

has repressor-cosuppressor model (tryptophan is corepressor, needed to bind & repress)

but instead of just on/off like lac, has dimmer switch in form of repeats in RNA

initial short polypeptide w/ repeated tryp codons

lots of tryp: ribosome stops at small stop codon and is on top of 1&2, loop 3-4 forms and terminates transcription of polymerase

little tryp: ribosome stalls in initial polypeptide b/c can't make it through tryp AAs. stalled ribosome let 2-3 mRNA loop form