# Package 'Xeva'

### August 28, 2018

1108000 = 0, = 010
Type Package
Title Analysis of patient-derived xenograft (PDX) data
Version 1.0.0
Author Arvind Mer, Benjamin Haibe-Kains
Maintainer Arvind Mer <amer@uhnresearch.ca></amer@uhnresearch.ca>
<b>Description</b> Contains set of functions to perform analysis of patient-derived xenograft (PDX) data.
License Artistic-2.0
RoxygenNote 6.0.1
LazyData true
VignetteBuilder knitr
Suggests knitr
Imports methods,  BBmisc, Biobase, plyr, stringr, ggplot2, ComplexHeatmap, reshape2, grDevices, PharmacoGx, foreach, parallel, doParallel, doSNOW, acepack, mgcv
R topics documented:
ABC
addExperimentalDesign

2 ABC

	angle	4
	AUC	5
	batchNames	5
	brca	6
	creatXevaSet	6
	drugInfo	7
	drugInfo<	8
	drugSensitivitySig	8
	expDesign	9
	getExperiment	10
	getMolecularProfiles	11
	modelInfo	12
	mRECIST	12
	pdxe	13
	PDXMI	13
	PDX_MI	14
	plotBatch	14
	plotmRECIST	15
	response	16
	selectModelIds	17
	sensitivity	18
	setResponse	18
	show,XevaSet-method	19
	slope	20
	subsetXeva	20
	summarizeMolecularProfiles	21
	summarizeResponse	22
	waterfall	23
Index		25
		_
ABC	compute area between two curves compute area between two time- volume curves	

### Description

compute area between two curves compute area between two time-volume curves

```
ABC(contr.time = NULL, contr.volume = NULL, treat.time = NULL, treat.volume = NULL)
```

addExperimentalDesign

### **Arguments**

```
contr.time time vector for control

contr.volume volume vector for control

treat.time time vector for treatment

treat.volume vector for treatment

degree default TRUE will give angle in Degree and FALSE will return Radians
```

#### Value

returns batch response object

### **Examples**

```
contr.time <- treat.time <- c(0, 3, 7, 11, 18, 22, 26, 30, 32, 35)
contr.volume<- contr.time * tan(60*pi/180)
treat.volume<- treat.time * tan(15*pi/180)
abc <- ABC(contr.time, contr.volume, treat.time, treat.volume)
par(pty="s")
xylimit <- range(c(contr.time, contr.volume, treat.time, treat.volume))
plot(contr.time, contr.volume, type = "b", xlim = xylimit, ylim = xylimit)
lines(treat.time, treat.volume, type = "b")
polygon(c(treat.time, rev(treat.time)), c(contr.volume, rev(treat.volume)), col = "#fa9fb5", border = NA)</pre>
```

addExperimentalDesign Add a new experimental design

#### **Description**

Add a new experimental design in expDesign slot.

### Usage

```
addExperimentalDesign(object, treatment = NULL, control = NULL,
batch.id = NULL, replace = FALSE)
```

#### **Arguments**

object The Xeva dataset

treatment The model.id of treatment

control The model.id of control

batch.id The batch.id for new batch

replace If TRUE will replace the old batch with new values

### Value

returns Xeva dataset with new experimental design added

4 angle

### **Examples**

```
data(brca)
brca <- addExperimentalDesign(object=brca, treatment=c("X.6047.LL71"),
control=c("X.6047.uned"), batch.id="new.batch", replace=FALSE)</pre>
```

angle

compute angle computes angle between two time-volume curves

### **Description**

compute angle computes angle between two time-volume curves

### Usage

```
angle(contr.time = NULL, contr.volume = NULL, treat.time = NULL,
treat.volume = NULL, degree = TRUE)
```

### **Arguments**

```
contr.time time vector for control
contr.volume volume vector for control
treat.time time vector for treatment
treat.volume volume vector for treatment
degree default TRUE will give angle in Degree and FALSE will return Radians
```

### Value

returns batch response object

```
contr.time <- treat.time <- c(0, 3, 7, 11, 18, 22, 26, 30, 32, 35)
contr.volume<- contr.time * tan(60*pi/180)
treat.volume<- treat.time * tan(15*pi/180)
ang <- angle(contr.time, contr.volume, treat.time, treat.volume)
par(pty="s")
xylimit <- range(c(contr.time, contr.volume, treat.time, treat.volume))
plot(contr.time, contr.volume, type = "b", xlim = xylimit, ylim = xylimit)
lines(treat.time, treat.volume, type = "b")
abline(lm(contr.volume~contr.time))
abline(lm(treat.volume~treat.time))</pre>
```

AUC 5

AUC

AUC returns area under the curve

### **Description**

AUC returns area under the curve

### Usage

```
AUC(time, volume)
```

### **Arguments**

time vector of time

volume first vector of volume

#### Value

returns angle and slope object

### **Examples**

```
time <- c(0, 3, 7, 11, 18, 22, 26, 30, 32, 35)
volume1<- time * tan(30*pi/180)
volume2<- time * tan(45*pi/180)
auc1 <- AUC(time, volume1)
auc2 <- AUC(time, volume2)
par(pty="s")
xylimit <- range(c(time, volume1, volume2))
plot(time, volume1, type = "b", xlim = xylimit, ylim = xylimit)
lines(time, volume2, type = "b")
abline(lm(volume1~time))
abline(lm(volume2~time))</pre>
```

batchNames

Get batch names/ids

### Description

Get batch names/ids from a Xeva dataset. If model.id is specified, will return all batch names conting that model.id

```
batchNames(object, model.id = NULL)
```

6 creatXevaSet

### **Arguments**

object A XevaSet

model.id default NULL. if specified it will return batch names conting that model.id

### Value

A Vector with batch names

### **Examples**

```
data(brca)
batchNames(brca)
batchNames(brca, model.id="X.6047.uned")
```

brca

breast cancer dataset from PDXE

### **Description**

breast cancer dataset from PDXE

### Usage

data(brca)

### **Format**

A Xeva object of PDXE breast cancer dataset

creatXevaSet

Creat Xeva class object creatXevaSet returns Xeva class object

### Description

Creat Xeva class object creatXevaSet returns Xeva class object

```
creatXevaSet(name, model = data.frame(), drug = data.frame(),
  experiment = data.frame(), expDesign = list(),
  modelSensitivity = data.frame(), batchSensitivity = data.frame(),
  molecularProfiles = list(), modToBiobaseMap = data.frame())
```

drugInfo 7

### **Arguments**

name a character string detailing the name of the dataset

model a data. frame containg the annotations for all models used in the experiment drug a data. frame containg the annotations for all the drugs profiled in the data set,

across all data types

experiment a data. frame containg all experiment information

molecularProfiles

a list of ExpressionSet objects containing different molecular profiles

### Value

Returns Xeva object

### **Examples**

\code{NULL}

drugInfo

get drug information get drug information slot

### **Description**

get drug information get drug information slot

### Usage

```
drugInfo(object)
```

### Arguments

object The Xe

The XevaSet to retrieve drug info from

### Value

```
a data.frame with the drug annotations
```

```
data(brca)
drugInfo(brca)
```

8 drugSensitivitySig

drugInfo<-

set drug information set drug information slot

### **Description**

set drug information set drug information slot

### Usage

```
drugInfo(object) <- value</pre>
```

### **Arguments**

object The XevaSet to replace drug info in

value A data. frame with the new drug annotations

#### Value

updated XevaSet

### **Examples**

```
data(brca)
drugInfo(brca)<- drugInfo(brca)</pre>
```

drugSensitivitySig

drugSensitivitySig

### **Description**

Given a Xeva object, and drug name it will return sensitivity value for all the genes/fetures

```
drugSensitivitySig(object, drug, mDataType = NULL, molData = NULL,
  features = NULL, model.ids = NULL, model2bidMap = NULL,
  sensitivity.measure = "slope", fit = c("lm", "maxCor", "gam"),
  standardize = c("SD", "rescale", "none"), nthread = 1, tissue = NULL,
  verbose = TRUE)
```

expDesign 9

### **Arguments**

object	The Xeva dataset		
drug	Name of the drug		
mDataType	molecular data type		
molData	External data matrix. Rows as features and columns as samples		
features	which molecular data fetures to use. Default NULL will use all fetures		
model.ids	which model.id to use from the dataset. Default NULL will use all model.id		
model2bidMap	a datafram with model.id and biobase.id. Default NULL will use internal mapping		
sensitivity.measure			
	Name of the sensitivity measure		
fit	Default 1m. Name of the model to be fitted. Options are "lm", "maxCor", "gam"		
type	Tissue type. Default is NULL which will use 'tissue' from object		

#### **Details**

A matrix of values can be directly passed to molData. fit can be "lm", "maxCor" or "gam". In case where a model.id map to multipal biobase.id the first biobase.id in the datafram will be used.

#### Value

A datafram with fetures and values

### **Examples**

expDesign Given a batch.name get batch

### **Description**

Given a batch.name get batch from a Xeva dataset

```
expDesign(object, batch.name = NULL)
```

10 getExperiment

#### **Arguments**

object XevaSet

object batch. name. If NULL will return all batch in the dataset

#### Value

A Vector with all batch.name

### **Examples**

```
data(brca)
expDesign(brca, batch.name = "X-6047.paclitaxel")
```

getExperiment For a given model.id, it will return a data.fram containing all data

stored in experiment slot

#### **Description**

For a given model.id, it will return a data.fram containing all data stored in experiment slot

### Usage

```
getExperiment(object, model.id = NULL, batchName = NULL, expDig = NULL,
  treatment.only = FALSE, max.time = NULL, vol.normal = FALSE,
  return.list = FALSE, impute.value = FALSE, concurrent.time = FALSE)
```

#### **Arguments**

object The XevaSet

model.id The model.id for which data is required, multipal allowed

batchName batch name from the Xeva set

expDig Experiment design

treatment.only Default FALSE. If TRUE give data only for non-zero dose periode (if dose data

avalible)

max.time maximum time for data vol.normal default TRUE will use

return.list default FALSE will return a datafram

impute.value default FALSE. If TRUE will impute the values

concurrent.time

default FALSE. If TRUE will cut the batch data such that control and treatment

will end at same time point

getMolecularProfiles 11

### Value

a data. fram will all the the values stored in experiment slot

### **Examples**

getMolecularProfiles Get Molecular Profiles

### **Description**

Get Molecular Profiles

### Usage

```
getMolecularProfiles(object, data.type)
```

### **Arguments**

object The XevaSet

data.type character, which one of the molecular data types is needed

### Value

a ExpressionSet where sample names are biobase.id of model

```
data(brca)
brca.RNA <- getMolecularProfiles(brca, data.type="RNASeq")</pre>
```

mRECIST

modelInfo

modelInfo Generic Generic for modelInfo method

### **Description**

modelInfo Generic Generic for modelInfo method

### Usage

```
modelInfo(object, mDataType = NULL)
```

### **Arguments**

object

The XevaSet to retrieve drug info from

#### Value

a data. frame with the model annotations

### **Examples**

```
data(brca)
mid <- modelInfo(brca)
head(mid)</pre>
```

mRECIST

Computes the mRECIST

### Description

mRECIST returns the mRECIST for given volume response

### Usage

```
mRECIST(time, volume, min.time = 10, return.detail = FALSE)
```

### Arguments

time Value of best response

volume Value of best average response

min.time minimum time after which tumore volume will be considered return.detail default FALSE. If TRUE will return all intermediate values

### Value

Returns the mRECIST

pdxe 13

### **Examples**

```
time <- c(0, 3, 7, 11, 18, 22, 26, 30, 32, 35)
volume<- c(250.8, 320.4, 402.3, 382.6, 384, 445.9, 460.2, 546.8, 554.3, 617.9)
mRECIST(time, volume, min.time=10, return.detail=FALSE)
```

pdxe

Example dataset with 1x1x1 experiment design

### **Description**

This is PDXE dataset without microarray data.

### Usage

data(pdxe)

#### **Format**

A Xeva object with 1x1x1 experiment design and moleculer data

**PDXMI** 

PDX-MI data

### Description

A dataset containing PDX models minimal information (PDX-MI) standard and corresponding Xeva variable.

### Usage

data(PDXMI)

### **Format**

An object of class data. frame with 45 rows and 4 columns.

#### **Details**

For details about PDX-MI see:

Meehan, Terrence F., et al. "PDX-MI: minimal information for patient-derived tumor xenograft models." Cancer research 77.21 (2017): e62-e66.

#### **Source**

http://cancerres.aacrjournals.org/lookup/doi/10.1158/0008-5472.CAN-17-0582

14 plotBatch

PDX\_MI

PDX-MI: Minimal Information for PDX

### **Description**

PDX-MI: Minimal Information for PDX

### Usage

```
PDX_MI(object)
```

plotBatch

Plot batch data

### **Description**

Plot data for a batch id or experiment design

### Usage

```
plotBatch(object, batchName = NULL, expDig = NULL, max.time = NULL,
  treatment.only = FALSE, vol.normal = FALSE, impute.value = TRUE,
  concurrent.time = FALSE, control.col = "#6baed6",
  treatment.col = "#fc8d59", title = "", xlab = "Time", ylab = "Volume",
  log.y = FALSE, drug.name = NULL, SE.plot = c("all", "none", "errorbar",
  "ribbon"), aspect.ratio = c(1, NULL), minor.line.size = 0.5,
  major.line.size = 0.7)
```

### Arguments

object Xeva object batchName batch name

expDig Experiment design list

max.time maximum time point of the plot, default NULL will plot complete data

treatment.only default FALSE. Given full data treatment.only=TRUE will plot data only during

treatment

vol.normal default FALSE . If TRUE volume will ne normalised impute.value default TRUE, will impute values where missing

control.col color for control plots treatment.col color for treatment plots

title title of the plot xlab title of x axis

plotmRECIST 15

```
ylab title of y axis

log.y default FALSE, if TRUE y axis will be in log

drug.name default NULL will extract drug name from data

SE.plot plot type. Default "all" will plot all plots and average curves. Possible values are "all", "none", "errorbar", "ribbon"

aspect.ratio default 1 will create equeal width and height plot

minor.line.size

line size for minor lines default 0.5

major.line.size

line size for major lines default 0.7
```

### Value

A ggplot2 plot with control and treatment

### **Examples**

plotmRECIST To plot mRECIST values

#### **Description**

plotmRECIST plots the mRECIST matrix obtained from summarizeResponse

### Usage

```
plotmRECIST(mat, control.name = NA, control.col = "#238b45",
  drug.col = "black", colPalette = NULL, name = "Drug & Models",
  sort = TRUE, row_fontsize = 12, col_fontsize = 12, draw_plot = TRUE)
```

### **Arguments**

mat The mRECIST matrix where rows are drugs and columns are patient

control.name name of the control control.col color of the control drug.col color of the drug names

colPalette color palette for mRECIST values

name title of the plot

16 response

sort if matrix should be sorted before ploting

row\_fontsize size of the row name font col\_fontsize size of the column name font

draw\_plot default TRUE will plot the figure. If FALSE will return an object

#### Value

plot

### Examples

```
data(brca)
## select lung cancer pdxe data
brca.mr <- summarizeResponse(brca, response.measure = "mRECIST", group.by="patient.id")
plotmRECIST(brca.mr, control.name = "untreated")</pre>
```

response

compute response

### **Description**

response computes response of a PDX model or batch

#### Usage

```
response(object, model.id = NULL, batchName = NULL, expDig = NULL,
  res.measure = c("angle", "mRECIST", "AUC", "angle", "abc"),
  treatment.only = TRUE, max.time = NULL, impute.value = TRUE,
  min.time = 10, concurrent.time = TRUE, vol.normal = F, verbose = TRUE)
```

### **Arguments**

object Xeva object

model.id model id for which response to be computed batchName batch id for which response to be computed

expDig experiment design for which response to be computed

res.measure response measure

treatment.only Default FALSE. If TRUE give data only for non-zero dose periode (if dose data

avalible)

max.time maximum time for data

impute.value default FALSE. If TRUE will impute the values min.time default **10** days. Used for mRECIST computation

concurrent.time

default FALSE. If TRUE will cut the batch data such that control and treatment

will end at same time point

vol.normal default TRUE will use

verbose default TRUE will print infromation

selectModelIds 17

### Value

returns model or batch response object

### **Examples**

selectModelIds

To select model ids based on drug name and/or tissue

### **Description**

To select model ids based on drug name and/or tissue

### Usage

```
selectModelIds(object, drug = NULL, drug.match.exact = TRUE,
   tissue = NULL)
```

### Arguments

object The XevaSet drug Name of the drug drug.match.exact

Default TRUE

tissue Tumor type. Default NULL

### Value

a vector with the matched model.ids

```
data(brca)
selectModelIds(brca, drug="trastuzumab", drug.match.exact=TRUE, tissue="BRCA")
selectModelIds(brca, drug="trastuzumab", drug.match.exact=FALSE)
```

18 setResponse

sensitivity

Get sensitivity for an Xeva object

### **Description**

Given a Xeva object, it will return sensitivity datafram

#### Usage

```
sensitivity(object, type = c("model", "batch"), sensitivity.measure = NULL)
```

### **Arguments**

object The Xeva dataset

type sensitivity type (either model or batch)

sensitivity.measure

Name of the sensitivity.measure. Default NULL, will return all

#### Value

a data. fram with model or batch id and sensitivity values

### **Examples**

```
data(brca)
head(sensitivity(brca, type="batch"))
head(sensitivity(brca, type="model"))
```

setResponse

setResponse sets response of an Xeva object

### Description

setResponse sets response of an Xeva object

```
setResponse(object, res.measure = c("mRECIST", "slope", "AUC", "angle",
   "abc"), min.time = 10, treatment.only = TRUE, max.time = NULL,
   vol.normal = TRUE, impute.value = TRUE, concurrent.time = TRUE,
   verbose = TRUE)
```

show,XevaSet-method 19

### **Arguments**

object Xeva object

res.measure response measure, multipal measure allowed

min.time default **10** days. Used for *mRECIST* computation

treatment.only Default FALSE. If TRUE give data only for non-zero dose periode (if dose data

avalible)

max.time maximum time for data

vol.normal default TRUE will use

impute.value default FALSE. If TRUE will impute the values

concurrent.time

default FALSE. If TRUE will cut the batch data such that control and treatment

will end at same time point

verbose default TRUE will print infromation

#### Value

returns updated Xeva object

### **Examples**

```
data(brca)
brca <- setResponse(brca, res.measure = c("mRECIST"))</pre>
```

show, XevaSet-method

A method to display object for "show" setGeneric is already defined

### **Description**

A method to display object for "show" setGeneric is already defined

```
## S4 method for signature 'XevaSet'
show(object)
```

20 subsetXeva

slope

Computes slope

### **Description**

slope returns the slope for given time and volume data

### Usage

```
slope(time, volume, degree = TRUE)
```

### Arguments

time vector of time volume vector of volume

degree default TRUE will give angle in Degree and FALSE will return Radians

#### Value

returns the slope and a fit object

### **Examples**

```
time <- c(0, 3, 7, 11, 18, 22, 26, 30, 32, 35)
volume<- c(250.8, 320.4, 402.3, 382.6, 384, 445.9, 460.2, 546.8, 554.3, 617.9)
sl <- slope(time, volume)
par(pty="s")
xylimit <- range(c(time, volume))
plot(time, volume, type = "b", xlim = xylimit, ylim = xylimit)
abline(lm(volume~time))</pre>
```

subsetXeva

Subset Xeva object

### **Description**

Subset Xeva object

```
subsetXeva(object, ids, id.name, keep.batch = TRUE)
```

summarizeMolecularProfiles

#### **Arguments**

object the XevaSet

ids ids to be selected for

id.name names of the id

keep.batch Default is TRUE. If FALSE will remove all the other model.ids from the ex-

periemt design that do not belong to selection

#### Value

New Xeva object

### **Examples**

```
data(brca)
print(brca)
df <- subsetXeva(brca, ids = c("X-1004", "X-1008", "X-1286"), id.name="patient.id", keep.batch=TRUE)
print(df)</pre>
```

summarizeMolecularProfiles

summarize Molecular Profiles

#### **Description**

summarizeMolecularProfiles

#### **Usage**

```
summarizeMolecularProfiles(object, drug, mDataType, tissue = NULL,
  sensitivity.measure = NULL, unique.model = TRUE, batchName = NULL,
  expDig = NULL)
```

### Arguments

object The XevaSet drug Name of the drug

mDataType character, which one of the molecular data types is needed

tissue default NULL will return all across all tissue

sensitivity.measure

default NULL will return all sensitivity measure

unique.model default TRUE will return only one sequncing id, in case where one model id

mapes to several sequencing ids

22 summarizeResponse

#### **Details**

- If a sequencing sample belong to multipal models, summarizeMolecularProfiles will creat saperate column for each model.
- All the models without the moleculer data will be removed from the output expression set.

#### Value

A ExpressionSet where sample names are model.id and sensitivity measure will be present in pData

### **Examples**

summarizeResponse

Summarize Response of PDXs

### **Description**

Summarize Response of PDXs.

### Usage

```
summarizeResponse(object, response.measure = "mRECIST", model.id = NULL,
  batch.id = NULL, group.by = "patient.id", summary.stat = c(";", "mean",
  "median"), tissue = NULL)
```

### **Arguments**

object The XevaSet

response.measure

character . Which response measure to use? Use the responseMeasures function to find out what measures are available for each Xeva set.

group.by default patient.id. How the models should be grouped togather. See details

summary.stat which summary method to use if multipal ids were found

batch.name a vector of batch names. Default NULL will return all batchs

waterfall 23

#### **Details**

There can be two types of response measure

• per model response : One response value for each Model, e.g. mRECIST\_recomputed for each model

• per batch response : One response value for each Batch, e.g. angle between treatment and control groups

In case of per model response output columns will be model.id (or group.by). For per batch response group.by value can be "batch.name".

#### Value

a matrix with rows as drug names, coulmn as group. by and each cell contains response. measure for the pair.

### **Examples**

```
data(brca)
brca.mR <- summarizeResponse(brca, response.measure = "mRECIST", group.by="patient.id")</pre>
```

waterfall

waterfall plot creates waterfall plot for a given drug

#### **Description**

waterfall plot creates waterfall plot for a given drug

### Usage

```
waterfall(object, drug, res.measure, group.by = NULL, tissue = NULL,
model.id = NULL, model.type = NULL, type.color = "#cc4c02",
legend.name = NULL, yname = NULL, title = NULL, sort = TRUE)
```

#### **Arguments**

object the XevaSet drug name of the drug

res.measure PDX model response measure

group.by group response data

tissue tissue

model.id which model.id to plot. Default is NULL will plot all models

model.type type of model such as mutated or wild type

type.color a list with colors used for each type

legend.name name of the legend yname name for y axis title title of the plot

sort default TRUE will sort the data

24 waterfall

## **Index**

```
*Topic datasets
                                                      summarizeMolecularProfiles, 21
    brca, 6
                                                      summarizeResponse, 22
    pdxe, 13
                                                      waterfall, 23
    PDXMI, 13
ABC, 2
{\it add} {\it Experimental Design}, 3
angle, 4
AUC, 5
batchNames, 5
brca, 6
creatXevaSet, 6
drugInfo, 7
drugInfo<-, 8</pre>
drugSensitivitySig, 8
expDesign, 9
getExperiment, 10
getMolecularProfiles, 11
modelInfo, 12
mRECIST, 12
PDX_MI, 14
pdxe, 13
PDXMI, 13
plotBatch, 14
plotmRECIST, 15
response, 16
selectModelIds, 17
\textit{sensitivity}, \textcolor{red}{18}
setResponse, 18
\verb|show,XevaSet-method|, 19|
slope, 20
subsetXeva, 20
```