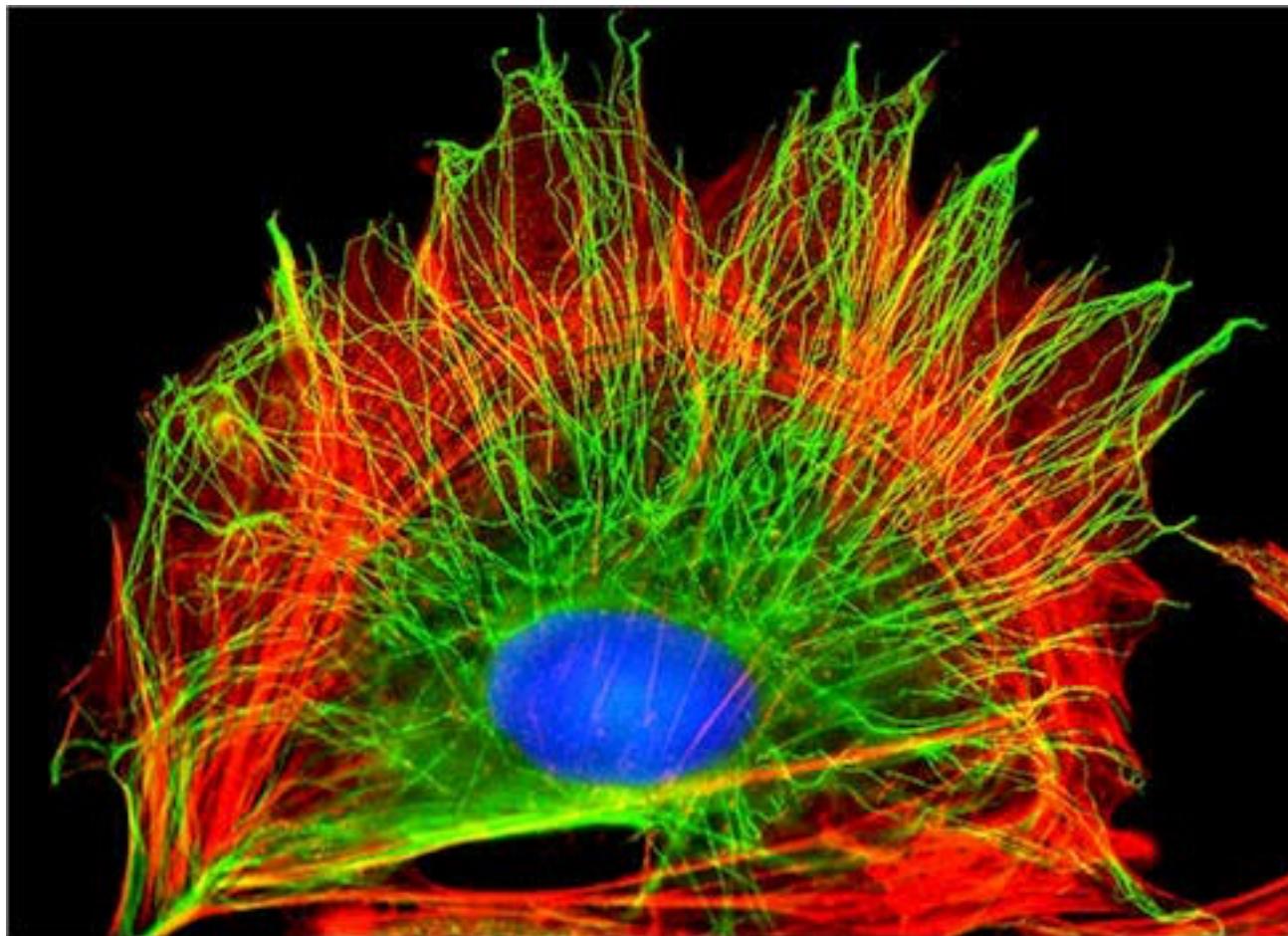
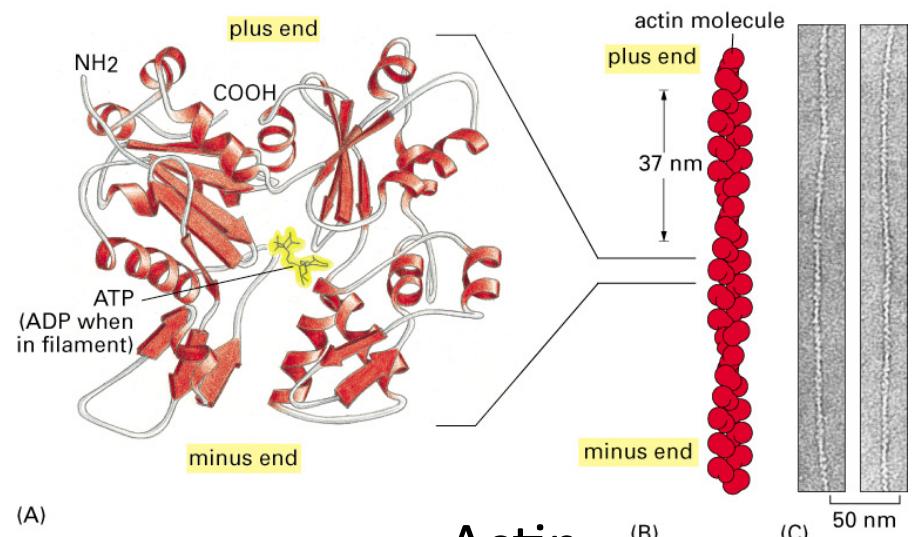
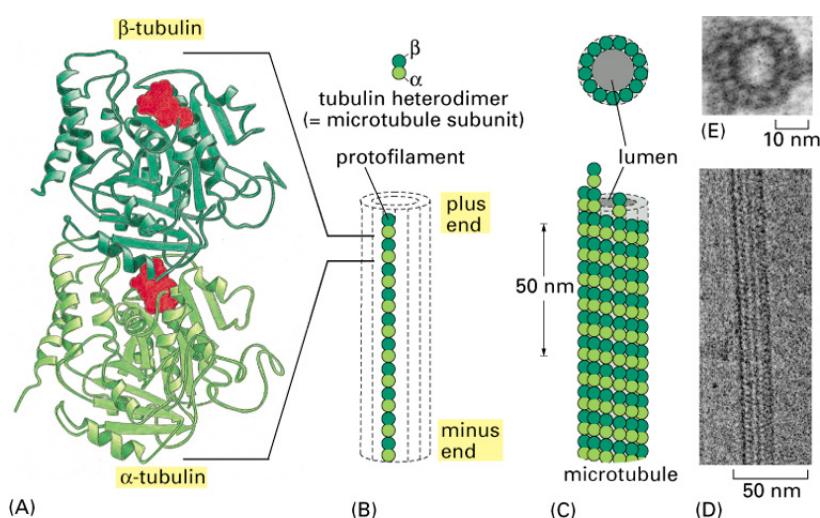


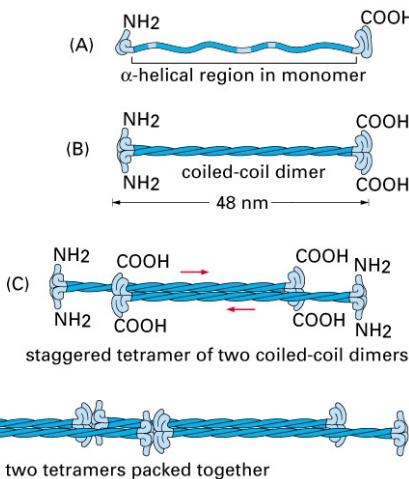
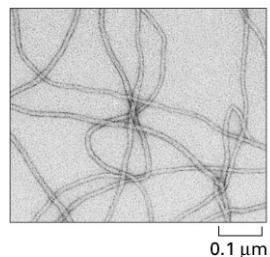
# Streetfighting the cytoskeleton



# Cytoskeletal filament structures

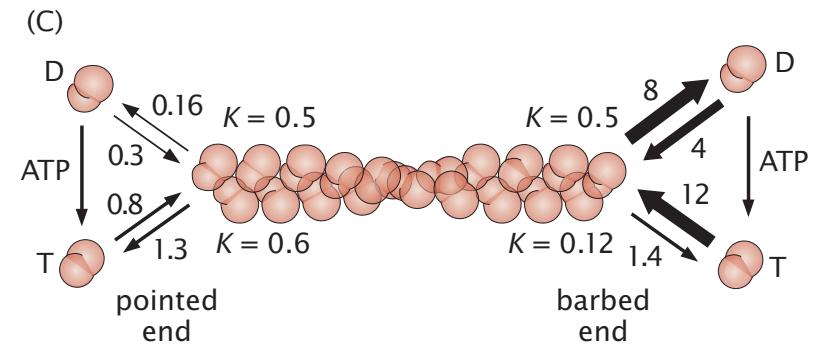
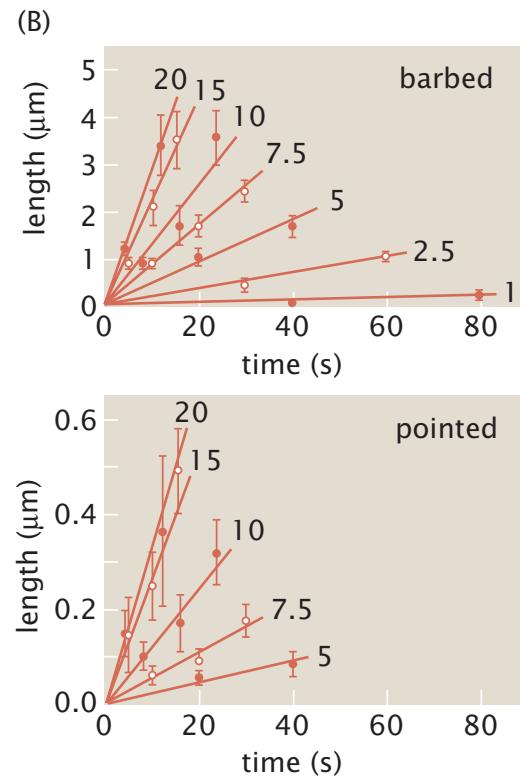
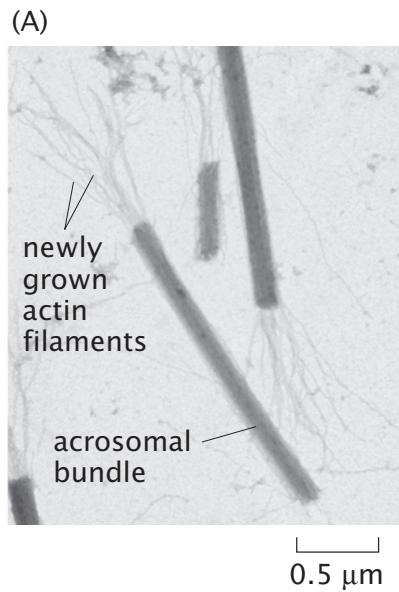


**Microtubule**  
persistence length  $\sim$  1 mm



**Intermediate filament**  
persistence length  $\sim$  2 μm

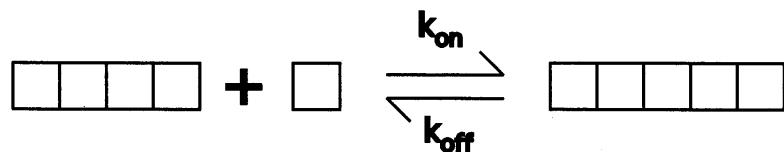
# Cell behaviors that depend on the cytoskeleton usually require polymerization and depolymerization



Kinetic parameters can be inferred from fixed time points

Tom Pollard

# Polymerization can generate force



$$\text{Growth speed } v = \delta * ([m]k_{on} - k_{off})$$

remember:

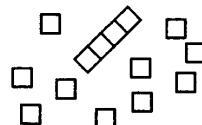
$$\Delta G_0 = -RT * \ln K_{eq}$$

$$\Delta G = \Delta G_0 + RT * \ln Q$$

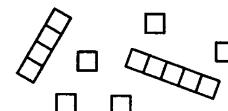
$Q$  = ratio of concentrations of  
products over reactants

$$k_B = R \text{ (gas constant)} / N_A$$

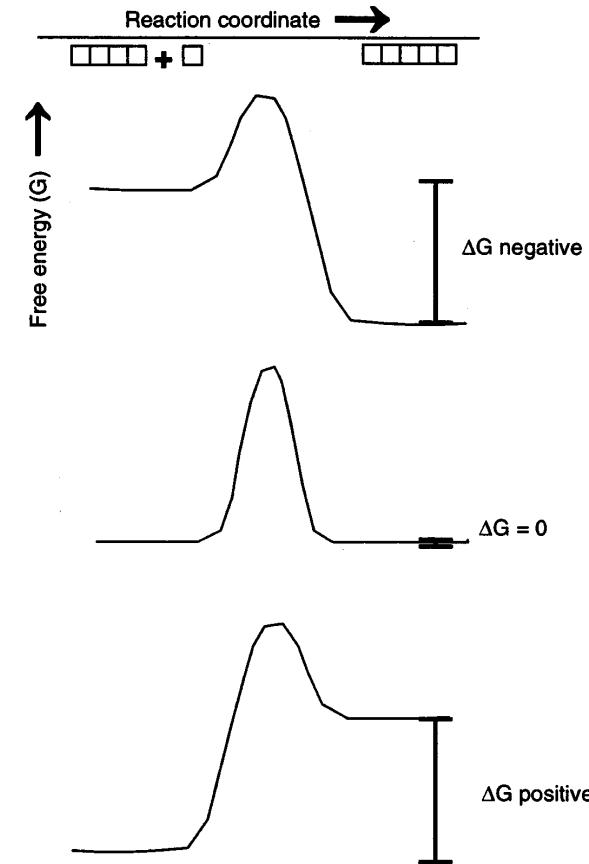
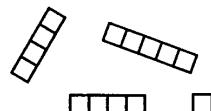
Case I:  
Excess monomer  
Spontaneous elongation



Case 2:  
At equilibrium  
No change in length

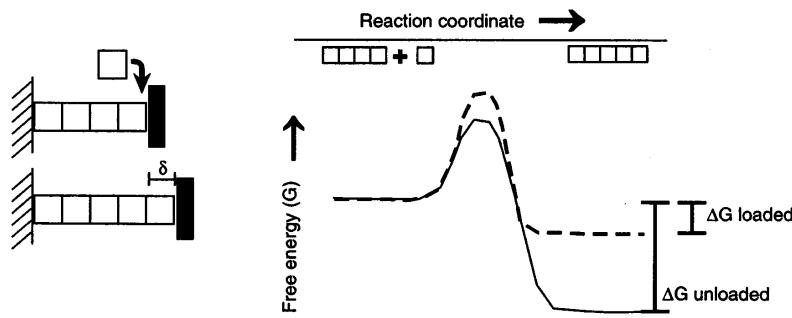


Case 3:  
Excess polymer  
Spontaneous shrinkage

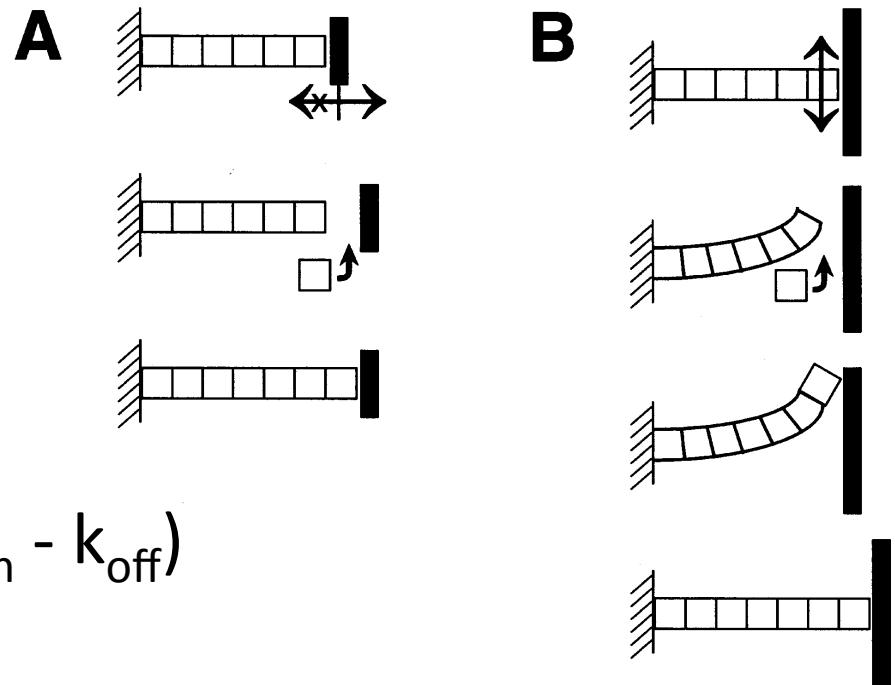


# Force should slow polymerization until stall occurs

$$F_{\max} = (k_B T / \delta) \ln([m]/C_c)$$



Possible kinetic mechanisms:

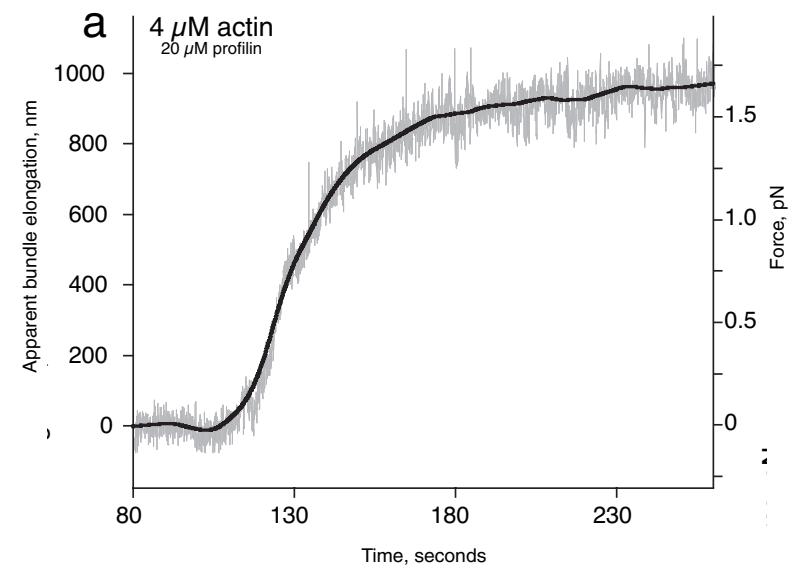
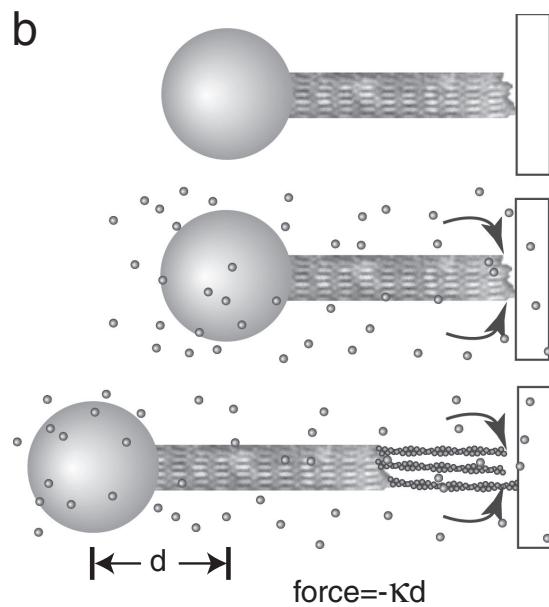
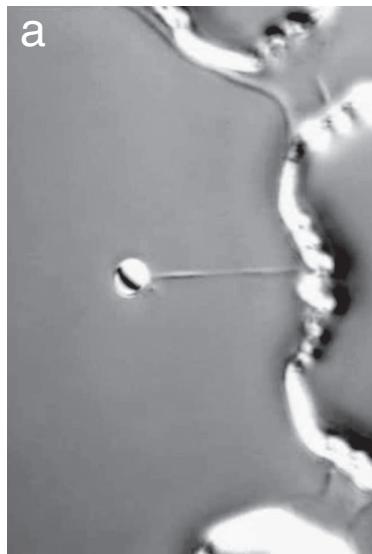


$$\text{Growth speed } v = \delta * ([m]k_{\text{on}} - k_{\text{off}})$$

Expect growth to slow with applied force:

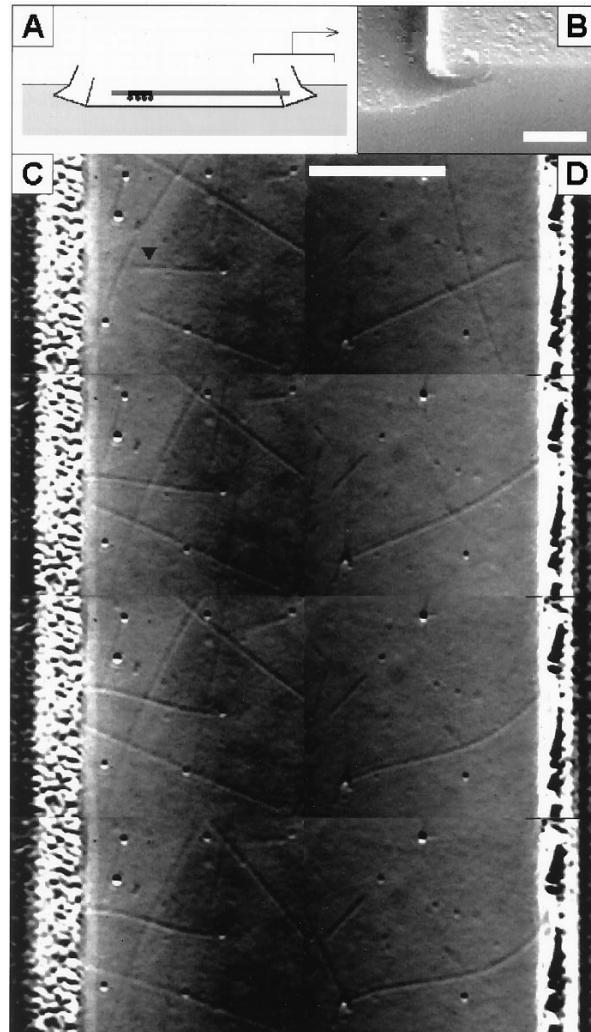
$$v(f) = \delta * ([m]k_{\text{on}} * e^{-(f\delta/k_B T)} - k_{\text{off}})$$

# Direct measurement of actin polymerization force with an optical trap

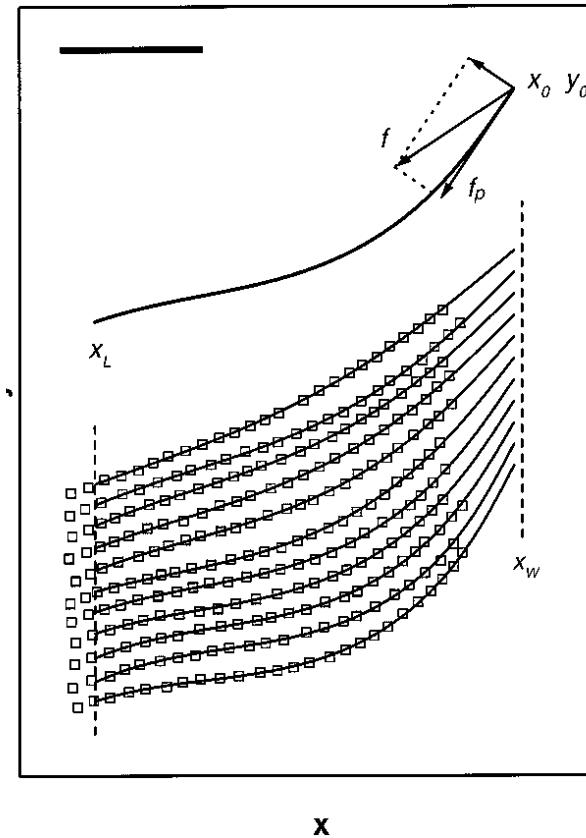


$$F_{\max} = (k_B T / \delta) \ln([m]/C_c)$$

# Measurement of microtubule polymerization force by bending



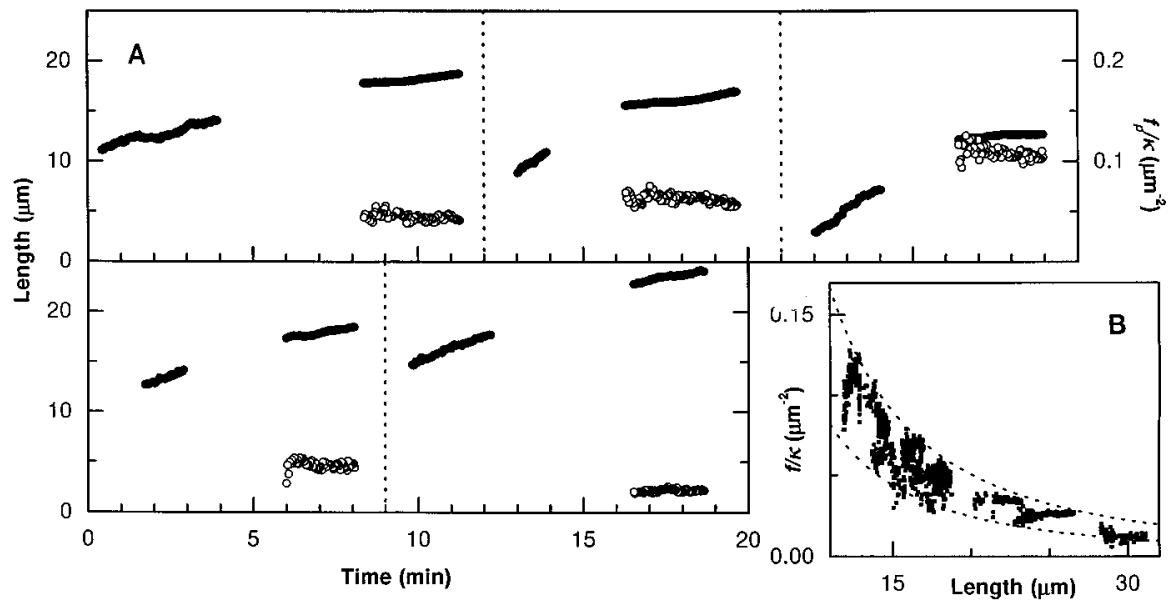
Expect growth to slow with applied force:  
 $v(f) = \delta * [k_{on} * e^{-(f\delta/k_B T)[m]} - k_{off}]$



Dogterom & Yerke, 1997, *Science*, **278**: 856

# Measurement of microtubule polymerization force by bending

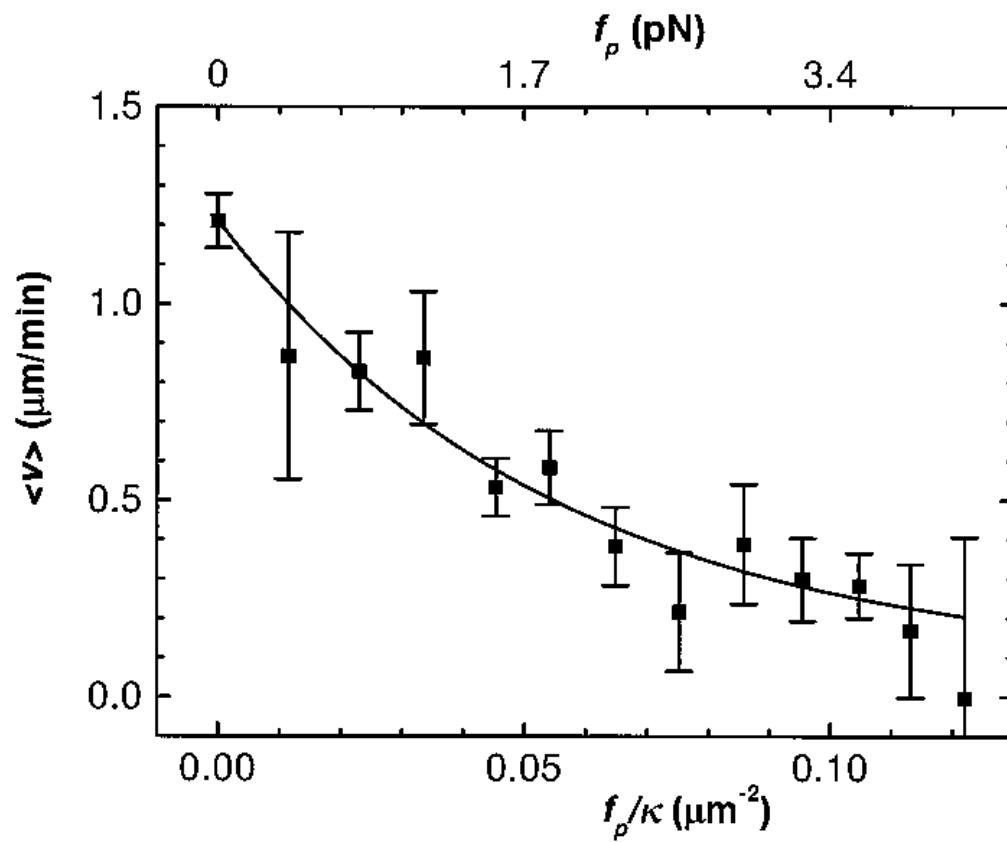
flexural rigidity  
 $\kappa = 34 \pm 7 \text{ pN} \cdot \text{nm}^2$



**Fig. 3.** MT length and applied force obtained from the analysis of MT buckling shapes such as shown in Fig. 2. **(A)** For five different MTs, the length  $L$  as a function of time (at 2-s intervals) is shown both before and after contact with the wall (solid symbols). A segment of time is missing in each case, during which the end of the growing MT was obscured by the presence of the overhang on the wall. Open symbols show the parallel component of the normalized force,  $f_p/\kappa$ . The lower left curve corresponds to the MT shown in Figs. 1D and 2. The upper right curve corresponds to the MT shown in Fig. 1C. **(B)** Total normalized force,  $f/\kappa$ , as a function of MT length for all MT shapes analyzed ( $n = 1316$ ). Each point corresponds to one MT shape. The dashed lines indicate the theoretical length dependence of  $f_c$  in two limiting cases:  $f_c/\kappa = 20.19/L^2$  for a MT with a seed that acts as a perfect clamp (upper curve) and  $f_c/\kappa = \pi^2/L^2$  for a MT with a seed that acts as a perfect hinge (lower curve). In the experiments, the seeds behaved in an intermediate way and, as expected, the forces obtained from the fits fall between these two limiting curves.

Dogterom & Yerke, 1997, *Science*, **278**: 856

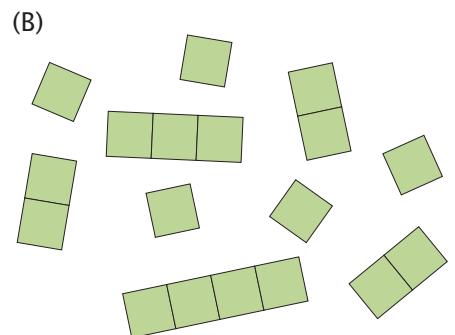
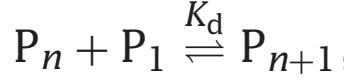
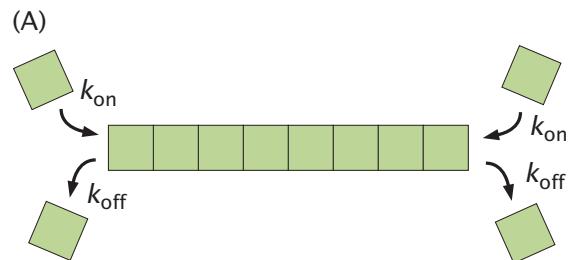
# Measurement of microtubule polymerization force by bending



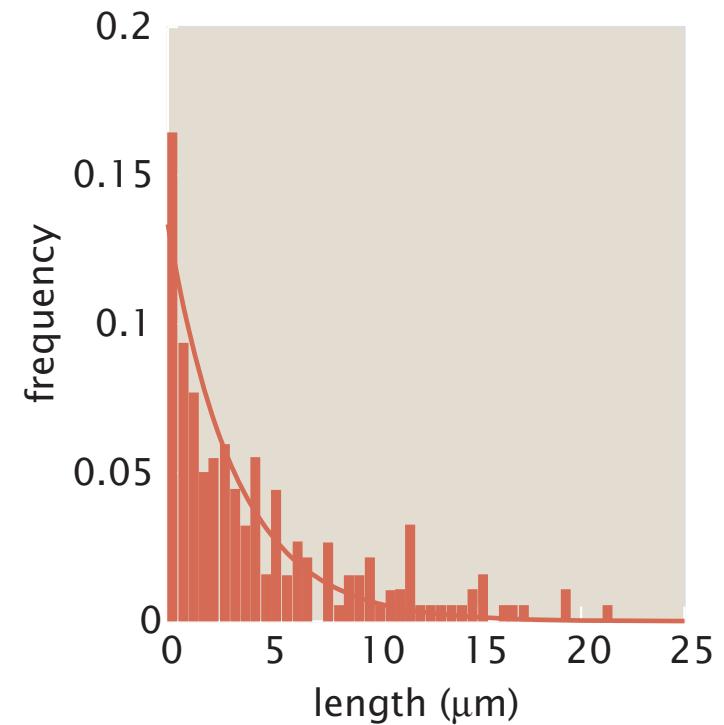
Dogterom & Yerke, 1997, *Science*, **278**: 856

# Expected length distribution at steady state

Spreading the cytoskeletal butter:



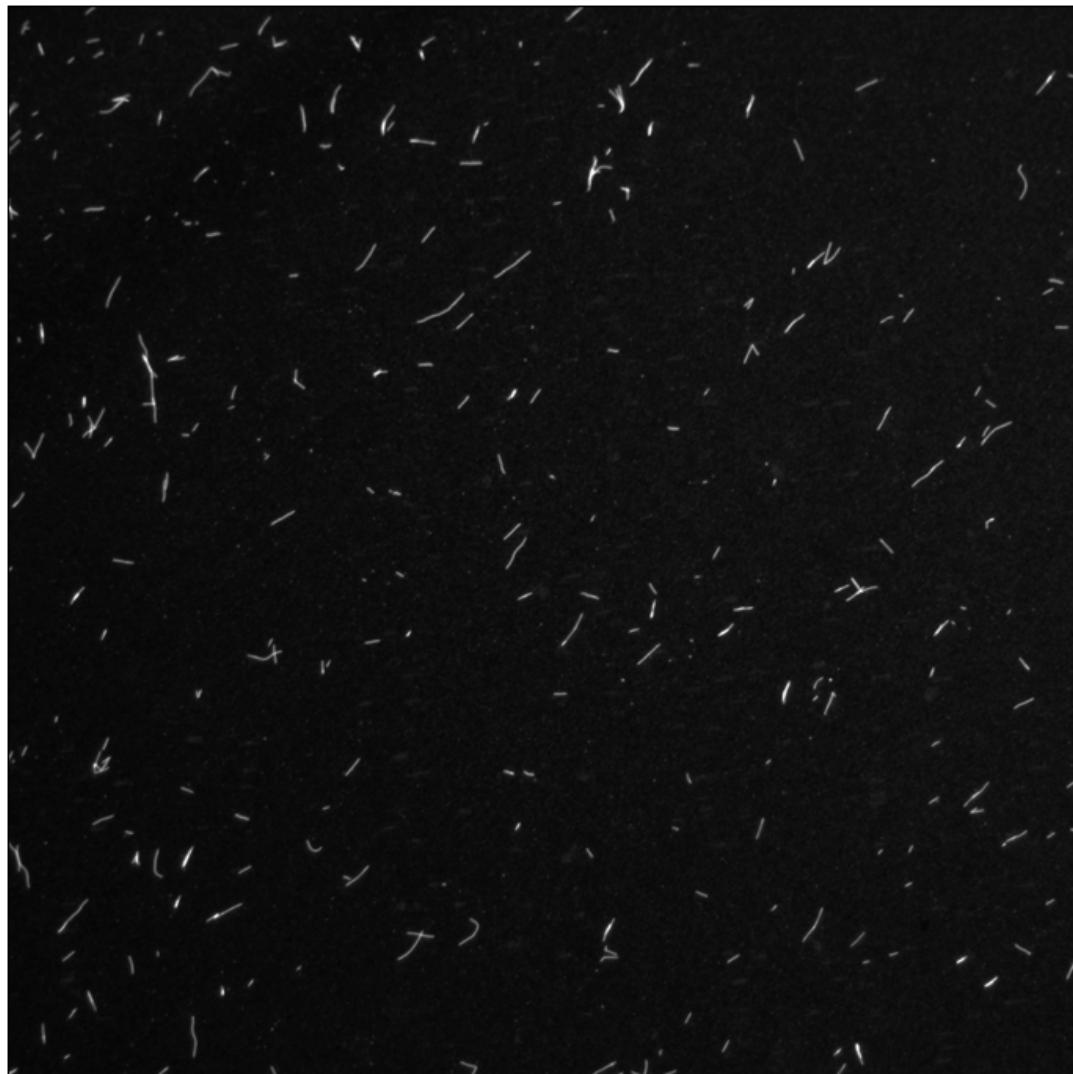
$$[P_n] = K_d e^{n \ln([P_1]/K_d)} = K_d e^{-\alpha n},$$



**Figure 15.23:** Distribution of filament lengths for actin filaments. (Adapted from S. Burlacu et al., *Am. J. Physiol.* 262:C569, 1992.)

Can we simulate this computationally?

# An experimental data point



Tubulin  
20  $\mu\text{M}$   
+DMSO  
80 minutes

Geoff