Clayton et al. 2018

**Microscopy Methods**

Following standard methodology as described per Gillespie (2006), all samples were screened for gastrointestinal parasites using fecal flotation and sedimentation techniques. A combination of both techniques was necessary to recover the total potential variety of eggs (i.e. lightweight helminthes and protozoa eggs and heavier trematodes) from fecal samples preserved in formalin (Gillespie, 2006).

**Fecal Floatation**

Samples were first examined using the flotation method where one gram of feces was centrifuged with distilled water to rinse out the formalin. Each tube was re-suspended in a sodium nitrate solution, covered with a microscope cover slip, and centrifuged for 10 minutes at 1800 rpm. The coverslip was then placed on a slide and scanned with the 10x objective lens. All parasite eggs, larvae, and cysts were counted and identified. Unknown eggs were also measured and photographed at the 40x objective lens.

**Fecal Sedimentation**

The fecal pellet remaining after flotation examination was suspended in a 40mL solution of diluted soapy water in a 50mL beaker. The fecal solution was filtered through cheesecloth held over the edge of the beaker into a 50mL centrifuge tube. This suspension and filtration technique was repeated, and the sediment was left to settle (~12 hours). Three drops of sediment were transferred onto a slide and were thoroughly mixed with 1 drop of diluted iodine. The sample was then covered with two coverslips placed side-by-side and scanned with the 10x objective lens. All parasite eggs, larvae, and cysts were counted and identified. Unknown eggs were also measured and photographed at the 40x objective lens.