0.1 Framework of the macroalgae module

Macroalgae, kelp, or seaweed, are macroscopic multicellular marine algae that cover a large range of taxonomic groups. They are of interest from an ecological point of view as they provide habitat for marine animals. They are also interesting from an economic point of view as they can be cultivated and harvested while offering a bio-remediating service to marine waters. There specifically is interest in the use of macroalgae as a bioremediator in integrated multitrophic aquaculture (IMTA) systems, where they can convert excess farm nutrients into biomass and value added products such as alginate.

The macroalgae module (MALG) in DELWAQ models the the dynamics of the kelp *Saccharina latissima* and is based almost entirely on the model described in Ole Jacob Broch (2012). Its applicability to seaweed in general is not known at this time and up to the user to determine. Note that numerous changes have been made to the equations in Ole Jacob Broch (2012) to allow for compatibility with DELWAQ's mass based systems, and also to allow for the inclusion of MALG in 3D models, whereby the size of the seaweed needs to be modelled in addition to the biomass. The primary motivation for this development is for application in the assessment of IMTA systems.

The general model organization will be described here, specifically the implementation in the DELWAQ library. For further details of the derivation of the model equations please refer to Ole Jacob Broch (2012).

Macroalgae can be idealized as 4 distinct state variable components:

- ♦ MacroALgae Structural mass (MALS, gDM m²)
- ♦ MacroALgae Nitrogen storage mass (MALN, gN m²)
- ♦ MacroALgae Phosphorous storage mass (MALP, gP m²)
- ♦ MacroALgae Carbon storage mass (MALC, gC m²)

Collectively these components represent the entire macroalgae "frond", where frond is the term used to describe the macro structure of the algae visible to the naked eye, including the foot (attachment organ). Each component of the frond exhibits its own growth dynamics and interacts with the surrounding water independently. The relationships between the variables is shown in Figure 1.

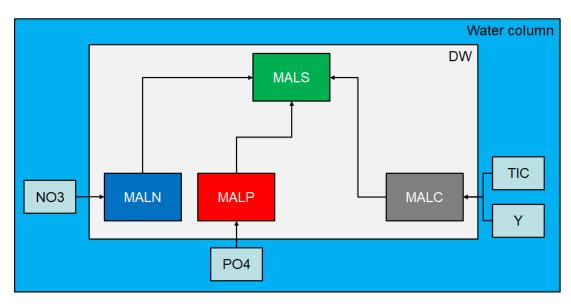


Figure 1: Relationship between the state variables in MALG

For instance, the structural mass represents the part of the macroalgae that increases the area and length of the total front when it grows. It does not take nutrients from the water, does not photosynthesize, and produces detritus when it decays/erodes. It grows by taking nutrients (N/P/C) from its storage and has its own fixed carbon, nitrogen, and phosphorous ratios and thus quotas that it must satisfy to grow. The structural mass has units of dry matter (DM m²) and is analogous to the common notion of 'dry matter'. However, it is lower than the true dry matter of the frond that would be measured by drying the frond because it represents only the dry weight of the plant minus the weight of water, nutrient stores, and carbon stores. The model does also compute dry weight (DW), which is the structural weight including the mass of nutrient stores, and wet weight (WW), which is the structural weight including the mass of nutrient stores with water.

The nitrogen and phosphorous storage (or reserves) uptake nutrients (NO_3^-, PO_4^{3-}) from the ambient water and store them for the structural mass during growth periods. In this model it is assumed the phosphorous storage dynamics are analogous to the nitrogen, but in reality the behaviour of phosphorous storage is not well known.

The carbon storage is the part of the plant responsible for photosynthesizing, producing exudate, and respiring. It produces carbon stores (carbohydrates) by taking up dissolved inorganic carbon and producing oxygen via photosynthesis. It provides carbon to the structural mass during growth periods.

It is assumed that there is constant relationship between volume and area of a macroalgae frond. This means that the density of the macroalgae does not change, even if the ratio of structural mass to stored material changes. Additionally, the C:N and C:P ratios of each component are fixed, but because the ratio of stored mass to structural mass changes, the C:N and C:P ratios of the entire frond will change due to changes in ambient conditions and thus nutrient reserves.

0.1.1 Distribution of macroalgae biomass in the water column

PROCESS: MALDIS

The physiological equations described in Ole Jacob Broch (2012) do not mention any physical description of the seaweed fronds in space aside from their area, which is actually the structural mass state variable in the model. The translation from an area based model to a mass based model is straightforward because Broch uses a constant conversion factor between area and mass. The application of Broch in DELWAQ requires knowledge of the mass per m² and the vertical space occupied by the frond. Thus, the algorithms for distributing the frond mass in space have been developed independently of the physiological formulations in Broch.

A simple approach is taken, whereby fronds (typically) only grow in length and retain a constant length: area ratio. The exception to this rule is when a frond is able to reach the end of the water column (i.e. the bed or the water surface) while it still has the capacity to grow. In these situations only the area will increase and not the length. The case for a single frond per computational cell is simple. However, the code must be able to deal with any seeding density, which cannot be uniquely defined by the mass per m² in DELWAQ alone. For example, consider two cases whereby there exists 40g of dry matter per m² as is shown in Figure 2.

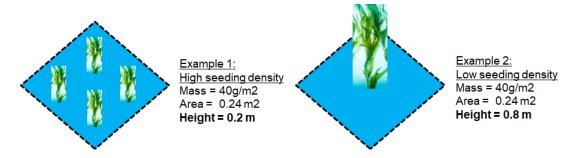


Figure 2: Two situations that have identical mass but non-identical position of the mass in the vertical axis

In the first example there are 4 fronds in a 1 m^2 square. They each weigh 10 g, have a combined area of 0.24 m^2 and are each 0.2 m long. In a second example, there is a single frond in a 1 m^2 square. It weighs 40 g, has an area of 0.24 m^2 , and is 0.8 m long. In a 3D model with grid cells that are <0.5 m thick, the nutrient uptake will take place in different cells in each of the two cases even though the mass per m^2 is identical in DELWAQ. To avoid this ambiguity the user must prescribe a set of parameters to describe the spatial characteristics of the culture. These are as follows:

- ♦ FootDepth The depth below the water surface that the foot of the frond is attached. The frond begins growing from the segment that intersects this depth, and the segment it resides in within a sigma layer model can change with a variable water level as it is assigned each time step.
- $\diamond LmaxMAL$ The maximum length of the frond measured in the vertical (z) axis from the foot to the tip of the frond.
- \diamond SwGroMAL Switch for an upwards (>0) or downwards (<0) growing frond.
- ♦ LinDenMal The linear density of the culture. This describes the amount of mass it takes a m² of culture to grow 1 m. Four small fronds require more mass to increase their collective length by 1 m compared to a single long frond.
- ♦ *ArDenMal* The area density of the culture. This is the ratio between surface area and dry weight.

The model does not explicitly know how many individual plants there are in a given segment, and so the collective mass per segment is generally referred to as a *culture* in this model. However, for most purposes the model can be understood by idealizing the culture as a single frond. Thus, the word 'frond' pertains to the physical structure of the culture, and may represent one or more individual plants.

Based on the parameters described about the culture's position can be completely described in 3 dimensions. To determine the position of the frond in the water column each time step, the process first calculates the length and the area of the culture in this column of segments:

$$LenMAL = MALS/LinDenMAL$$

$$AreMAL = MALS*Surf/ArDenMAL$$

Where:

LenMAL Length of frond in the column (m)

Surf Surface area of the bottom segment (m²)

MALS structural mass (gDM m⁻²)

LinDenMAL The linear density of the culture (g m⁻³) ArDenMAL The area density of the culture (g m⁻²)

This length is then used to check which segments the frond biomass should be present in, and which fraction of the frond exists in each segment. This fraction is sent to each segment for all other computations involving biomass. The calculation of this depends firstly on whether the frond grows upwards or downwards.

IFSwGroDir < 0: Zm = LenMAL + FootDepth

IFSwGroDir > 0: Zm = FootDepth - LenMAL

Where:

Zm Distance from water surface to tip of frond (positive down) (m)

Z1 Distance from water surface to top of segment (positive down) (m)

 Z_2 Distance from water surface to bottom of segment (positive down) (m)

The segment top depth Z1 and segment bottom depth Z2 are then checked against Zm to see if the frond resides entirely within, entirely outside, or partially within the current segment in the segment loop. The fraction of the biomass allocated to the current segment is then the ratio of the segment depth to LenMAL in the column for segments in which the frond completely resides, or the ratio of the difference between Zm and either Z1 or Z2 to LenMAL for segments in which the tip or foot of the frond is found.

0.1.2 Flux of Macroalgae structural biomass

PROCESS: FLMALS

The structural component of the macroalgae is the dry material that gives shape and structure to the macroalgae frond. It does not include N, P, or C stores, but has a fixed N, P, and C component, meaning that these nutrients are required for growth of the structural mass and there is a minimum amount of N,P and C in the structural mass regardless of the reserve nutrient level. As the structural mass grows it increases the length and area of the entire frond. Growth of other components of the frond (nutrient reserves) do not have any effect on the frond length, volume, density, or surface area.

The net growth of the structural mass is the resulting rate of structural biomass production and frond erosion (considered to be analogous to mortality used in other DELWAQ processes). This balance can be defined by the following equation:

$$dGrowMALS = MALS \times (\mu - \phi)$$

The growth rate μ of MALS is dependent firstly on the nutrient (MALN, MALP, and MALC) stores available. The growth rate is defined as follows:

$$\mu = f_{density} f_{photoperiod} f_{temperature} \times min \left\{ 1 - \frac{N_{min}}{MALN}, 1 - \frac{P_{min}}{MALP}, 1 - \frac{C_{min}}{MALC} \right\}$$

Where:

```
specific growth rate of macroalgae sturctural mass (d<sup>-1</sup>)
\mu
φ
                 mortality/erosion rate (d^{-1})
f_{density}
                 biomass density limitation function (-)
                 photoperiod limitation function (-)
f_{photoperiod}
                 temperature limitation function (-)
f_{temperature}
                 minimum N storage (gN gDM<sup>-1</sup>)
N_{min}
MALN
                 nitrogen storage (gN/m<sup>2</sup>)
P_{min}
                 minimum phosphorous storage (gP gDM<sup>-1</sup>)
MALP
                 phosphorous storage (gP/m<sup>2</sup>)
                 minimum carbon storage (gC gDM<sup>-1</sup>)
C_{min}
MALC
                 carbon storage (gC/m<sup>2</sup>)
```

Note that in the code the storage terms are temporarily converted to g/gDM by dividing by MALS to comply with the formulations and units outlined in Ole Jacob Broch (2012) The in MALG the storage state variable terms are g m $^{-2}$ as per all non-transportable DELWAQ substances, and this is reflected in the fluxes, which are back calculated to g m $^{-2}d^{-1}$. As the substances are non transportable they reside in the bottom segment. However, depending on the user defined variables, it is possible for the mass and associated fluxes to have no effect in the bottom segment, such as in the case where the algae grow from the water surface downward. This is discussed in MALDIS.

The growth rate of the structural mass is dependent on a density limitation, a photoperiod limitation, and a temperature limitation. These limitations differ from conventional DELWAQ limitations for algae growth in that they do not range strictly between 0 and 1. Instead Broch has tuned them such that their product is equal to the maximum growth rate when at optimal conditions $(0.18 \ d^{-1})$.

The density limitation represents the ability of the frond to only grow so big, and the bigger it gets the lower its growth rate will become. The formulation is as follows:

$$f_{density} = m_1 exp \left\{ -\left(\frac{AreaLoc}{MALS_0}\right)^2 \right\} + m_2$$

Where:

 m_1 growth rate parameter 1 (-) m_2 growth rate parameter 2 (-) AreaLoc locaal frond area in this segment (m²) $MALS_0$ critical biomass area (m²)

This formulation is designed to allow small fronds to grow faster than bigger fronds. This has large implications for the DELWAQ model in contrast to the Broch model which is 'individual based' and thus can integrate this formulation seamlessly. The reason that this complicates the DELWAQ implementation relates back to the MALDIS section. Considering the two setups previously described in Figure 2. It is possible to ascertain that the gross erosion rate within a given segment should be higher for segments containing a single large frond than for those containing a collection of smaller fronds of equivalent mass. This idea is logical if one assumes that the size of the neighbouring frond does not affect the growth of another frond (ignoring light and nutrient competition effects). Thus, decisions made in MALDIS about how to schematize the culture need to also play a role in defining the $MALS_0$ value. The current idea is that:

$$MALS_0 = N_{fronds}/0.06$$

Which corrects for the fact that a given individual plant will grow to a certain length regardless of how many neighbours it has.

The photoperiod limitation is similar to the daylength limitation used in BLOOM and DYNAMO, but instead considers the *normalized change* in daylength compared to the previous day instead of the actual current daylength. In this context 'normalized' means that the change in daylength is relative to the maximum change in daylength (i.e. the daylength change at the equinoxes). This response to normalized daylength changed due to the fact that *Saccharina latissima* is a seasonal anticipator and will grow and store nutrients in accordance with the change in the season as determined by how much longer or shorter the days become. This formulation is given by:

$$f_{photoperiod} = a_1(1 + sin(\tau(n)|\tau(n)|^{\frac{1}{2}})) + a_2$$

Where:

 a_1 photoperiod parameter 1 (-)

 a_2 photoperiod parameter 2 (-)

and τ is a function describing the normalized difference in day length between current day and previous day. This function is calculated by the process DAYLP which essentially identical to the standard DELWAQ process DAYL. The parameters a_1 and a_2 are chosen such that $0.3 > f_{photoperiod} < 2$ at the given latitude. In future implementations it is hoped that the code will calculate this for the user, but it currently requires an iterative approach before the simulation to define the correct values for the given latitude to ensure $0.3 > f_{photoperiod} < 2$.

The temperature limitation is a simple piece wise function that identifies an optimal growth between temperatures $10-15^{0}$ C, no growth above 19^{0} C, and linear growth increasing between -1.8 and 10^{0} C. The temperature limitation is therefore:

$$f_{temperature} = \begin{array}{cc} 0.08T + 0.2 & -1.8 < 10 \\ 1 & 10 \le T \le 15 \\ 19/4 - T/4 & 15 \le T \le 19 \\ 0 & T > 19 \end{array}$$

The specific erosion (mortality) rate mrt of the structural biomass is proportional to the total area of the frond and the erosion parameter. The formulation is as follows:

$$\phi = \frac{10^{-6} exp(\epsilon \times LocArea)}{1 + 10^{-6} exp((\epsilon \times MALS) - 1)}$$

Where:

 ϵ erosion parameter (m $^{-2}$)

The structural growth process occurs over all segments which have a biomass fraction >0. This is in spite of the fact that the biomass administratively resides in the bottom segment. During each time step, all segments with a non-zero biomass fraction receive 'ghost' structural and storage mass according to the biomass allocated to it in MALDIS. The local inorganic nutrient, gas, and particulate fluxes are calculated using this mass and the local ambient conditions. The fluxes are then locally applied to ambient state variables exogenous to MALG, but not the MALG parameters (MALS, MALN, MALP, MALC). Instead, the fluxes of MALG state variables are communicated to the bottom segment in a cumulative way, whereby the local fluxes of all segments that share the same bottom segment are summed to compute

the net total change in MALS, MALN, MALP, and MALC resulting from the net growth in the column. Once the bottom segment is reached in the segment loop, the fluxes for each column of segments have been accumulated and the net change in mass of each administrative bottom segment is known. The culture will then become longer or shorter in the next time step to reflect this new state, which again is kept track of by the bottom segment in the column. The consequence of applying this technique is that the local per segment 'ghost reserve masses' adopt the reserve ratio of the whole column. Another way to say this is that all child segments of a given water column have a fixed structural mass to storage mass ratio, and consequently all reserves are equally distributed along the frond.

0.1.3 Flux of Macroalgae nutrient storage

PROCESS: FLMALN

The nutrient storage component(s) of the macroalgae (also referred to as reserves) are those that supply nutrients for growth of the structural mass. The storage is also responsible for taking up dissolved inorganic nutrients from the ambient water. The reserves consist of a nitrogen (MALN) and a phosphorous (MALP) storage component, which are both dealt with in the same nutrient storage subroutine. Note that Ole Jacob Broch (2012) does not include a phosphorous component to the nutrient storage, but it is included in this model for flexibility should new information about phosphorous storage become available. Currently the model is written such that the culture cannot be P limited and no P flux will occur.

The change in a nutrient storage mass is given by the following equations:

$$dUptMALN = J_N - mu(K_C \times k_N \times MALS + MALN)$$

$$dUptMALP = J_P - mu(k_C \times k_P \times MALS + MALP)$$

This represents the balance between uptake J and utilization for growth of the frond's structural mass μ . The uptake is dependent on flow velocity, the amount of stores compared to the minimum and maximum possible, and the ambient nutrient concentration. The formulation for both nitrogen and phosphorus is described as follows:

$$\begin{split} J_N &= f_{velocity} J_{Nmax} (\frac{NO_3^-}{K_{sn} + NO_3^-}) (\frac{MALN_{max} - MALN}{MALN_{max} - MALN_{min}}) \\ J_P &= f_{velocity} J_{Pmax} (\frac{PO_4^{3-}}{K_{sp} + PO_4^{3-}}) (\frac{MALP_{max} - MALP}{MALP_{max} - MALP_{min}}) \\ f_{velocity} &= 1 - exp(-\frac{U}{U_{0.65}}) \end{split}$$

Where:

carbon:dry matter ratio in structural mass (gC gDM⁻¹) k_C nitrogen:carbon ratio in structural mass ($qN qC^{-1}$) k_N phosphorous:carbon ratio in structural mass (gN gC⁻¹) k_P maximum uptake rate nitrogen (gN $m^{-2}d^{-1}$) J_{Nmax} K_{sn} half saturation for inorganic nitrogen uptake $NO_3^$ ambient nitrate concentration (gN m^{-3}) $MALN_{max}$ maximum nitrogen storage (gN gDM⁻¹) $MALN_{min}$ minimum nitrogen storage (gN gDM⁻¹) maximum uptake rate nitrogen gP m $^{-2}$ d $^{-1}$) J_{Pmax} K_{sp} PO_4^{3-} half saturation for inorganic nitrogen uptake (gP m^{-3}) ambient phosphate concentration (gP m⁻³) $MALP_{max}$ maximum phosphorous storage (gP gDM⁻¹) minimum phosphorous storage (gP gDM⁻¹) $MALN_{min}$ water velocity (m s $^{-1}$) water velocity at which uptake rate is 65 of maximum (m s^{-1}) $U_{0.65}$

The first term in J_N and J_P pertains to the uptake of nutrients and the second term to the utilization of stores by the structural mass during growth. Note how the nutrient requirement (quota) is dependent on the combined quota for structural mass and storage mass. The uptake of nutrients is dependent on the ambient concentration according to Michaelis-Menten kinetics. It is dependent on velocity such that high velocities make it easier for the frond to uptake the nutrients. This is related to an improved mass transfer coefficient. At sufficiently high ambient concentrations and water velocities the uptake rate is J_{max} .

0.1.4 Flux of Macroalgae carbon storage

PROCESS: FLMALC

The carbon storage component of the macroalgae (MALC) supplies carbohydrates for growth of the structural mass. The carbon storage is also responsible for photosynthesis, respiration and exudation. The change in a carbon storage mass is given by the following equation:

$$dUptMALC = P(1 - E) - R - \mu(k_C \times MALS + MALC)$$

Where:

 $P - \text{Gross photosynthetic rate (gCm}^{-3} \text{ d}^{-1})$

E Fraction exudation (-)

R Maintenance respiration rate (gCm $^{-3}$ d $^{-1}$)

Where the first term relates to the net of production, respiration and exudation, and the second term pertains to the utilization of carbon stores by the structural mass.

The carbon production rate (i.e. photosynthesis) is given by the following set of equations:

$$P(T,I) = P_s(1 - exp(-\frac{\alpha \times I}{P_s}))exp(-\frac{\beta \times I}{P_s})$$

$$P_s(T) = \frac{\alpha \times I_{sat}}{ln(1 + \frac{\alpha}{\beta})}$$

$$P_{max}(\beta) = (\frac{\alpha \times I_{sat}}{ln(1 + \frac{\alpha}{\beta})})(\frac{\alpha}{\alpha + \beta})(\frac{\beta}{\alpha + \beta})^{\frac{\beta}{\alpha}}$$

$$P_{max}(T) = \frac{P_1 exp(\frac{T_{AP}}{T_{P1}})}{1 + exp(\frac{T_{APL}}{T} - \frac{T_{APL}}{T_{PL}}) + exp(\frac{T_{APH}}{T_{PH}} - \frac{T_{APH}}{T})}$$

Where:

Photosynthetic rate (gC m $^{-3}$ d $^{-1}$) $P_s(T)$ Saturation photosynthetic rate (gC m $^{-3}$ d $^{-1}$) Incident radiation (W m^{-2}) Saturation radiation (W m^{-2}) I_{sat} Photosynthetic efficiency (gC $d^{-1}W^{-1}$) $\beta(T)$ Photosynthetic light inhibition (gC $d^{-1}W^{-1}$) P_1 P_2 T T_{P1} Reference photosynthetic rate at T₁ Reference photosynthetic rate at T₂ Water temperature (K) temp for reference photosynthetic rate 1 T_{P2} temp for reference photosynthetic rate 2 T_{AP} Arrhenius temperature for photosynthesis (⁰K) T_{APH} Arrhenius temp for photosynthesis high end $({}^{0}K)$ Arrhenius temp for photosynthesis low end (0K) T_{APL}

This set of equations describes a photo inhibitory effect for $I>I_{sat}$. The structural mass is strictly unable to grow above $Temp>19^0$. Although the structural mass cannot grow, photosynthesis production can still occur in the range $>-1^0CTemp<23^0C$. These two temperature controls are fixed for the species and the user cannot flexibly change this unless the code is edited. To adapt the model to tropical species or species that have a different optimal temperature photosynthesis range, both the temperature function and the photosynthetic range parameters have to be edited in addition to the code for the piece-wise temperature growth function for MALS.

The maximum production rate P_{max} can be formulated in two ways. The first is only a function of temperature and describes photosynthesis at $I=I_{sat}$. The second production rate P_{max} is the actual maximum production rate taking into account the effect of temperature on the response of growth to light. This means that growth is non-linear in temperature, as it effects both growth directly and also the way in which growth is affected by light. β is solved for using Newton's method in Broch by differentiating $P_{max}(\beta)$. This involves solving for the value β such that $P_{max}(T)$ equals the $P_{max}(\beta)$ obtained from the temperature relationship at the current temperature and at $I=I_{sat}$. Newton's method is not used in the DELWAQ code and instead β is pre-calculated for all temperatures in the photosynthetic range -2°C to 23°C at an interval of 0.1°C. This is hard coded into the FLMALC subroutine and constitutes a linear approximation of β . Due to this the model should only be used for temperature ranges $-2^{\circ}C < T < 23^{\circ}C$. Currently:

$$\Rightarrow \beta(T) for T < -2^{0}C = \beta(-2^{0}C) \text{ and } \beta(T) for T > 23^{0}C = \beta(23^{0}C).$$

Consequently temperatures above or below the photosynthetic temperature range will result in a production rate equivalent to the rate at the closest temperature in the range. The relationship between temperature, beta, irradiance and gross production is shown in Figure 3

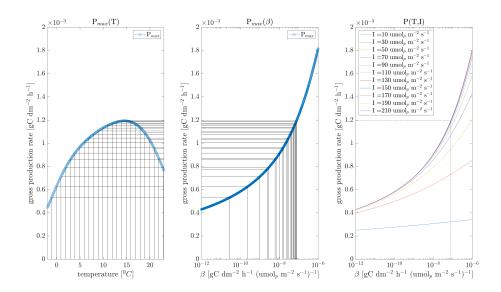


Figure 3: Relationship between temperature, β , irradiance and gross production. Here the black lines demonstrate the process by which for a given temperature $P_{max}(T)$ is calculated, and the corresponding β value required to achieve $P_{max}(T) = P_{max}(\beta)$ is derived, and used to calculate the response of growth rate to irradiance, where $P = P_{max}$ when $I = I_{sat}$

0.1.5 Harvesting of macroalgae

PROCESS: HRVMAL

0.2 Model validation

This section briefly outlines the test cases for the application of the MALG model. Two test cases have been outlined:

- ♦ Broch 1DV model
- ♦ A simple 3D cube model with tidal currents

0.2.1 Testcase: Broch

This testcase involves the reproduction of the model as it was demonstrated in Ole Jacob Broch (2012). The testcase involves a single frond growing off the coast of Nowrway in 250m deep water at approximately 5 m below the surface, although the depth of the frond in Ole Jacob Broch (2012) is never explicitly stated. The kelp grows in a 0D model (a box) with temperature, nitrogen, and light climate forcing derived from measurements and local models. These forcing functions are shown in 4.

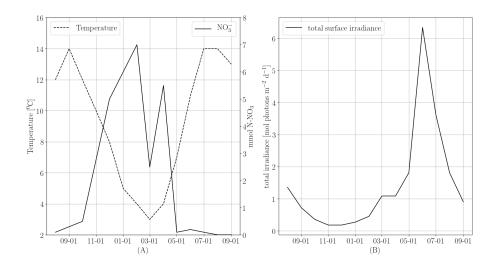


Figure 4: (A) Temperature and NO₃ forcing used in the model. **(B)** 10 m irradiance forcing used in the model.

Only MALG processes and processes relating to reaeration of CO_2 and O_2 and pH are activated in this model. All model constants are set to the default values described in Ole Jacob Broch (2012). There are however a few exceptions:

- $\Leftrightarrow ExtVL$ is set to 0.07 m⁻¹ in Broch, but the depth is not known. In the test case the model is 10 m deep and $ExtVl = 0.18 \text{ m}^{-1}$.
- \diamond Broch states that the maximum growth rate of the structural mass is 0.18 d $^{-1}$. Broch also states that the contribution to the growth rate from MALN=1-=0.65. As $MALN_{max}$ is stated as =0.022, it must be the case that $1-\leq 0.565$. Thus, the μ in MALG is up to 13 percent lower than in Ole Jacob Broch (2012).
- \diamond Broch checks the gross production formulation at I = 10 μ mol m⁻² s⁻¹ and T = 12⁰ and arrives at a value of 2.95×10^{-4} gCdm⁻²h⁻¹. This is lower than the value computed by MALG under the same conditions $(3.21 \times 10^{-4} \text{ gCdm}^{-2}\text{h}^{-1})$. It is at least partially due to the different method of approximating β , but it was also found that $P_{max}(T_{pl}) \neq 3.394 \times 10^{-4} \text{ gCdm}^{-2}\text{h}^{-1}$ in contrast to what was stated in the paper. This discrepancy may be due to discrepancies in T_{apl} and T_{aph} , which were stated to be $27,774^0K$ and $25,924^0K$ respectively in the paper but could not be reproduced using the high and low end temperatures and production rates. Thus the gross production is higher in MALG than in Ole Jacob Broch (2012).

The model settings are listed in Table 1:

parameter	value	unit	description
K0HrvMALS	0.00000	(gDM/m2/d)	zero order harvesting rate Macroalgae
K1HrvMALS	0.00000	(1/d)	first order harvesting rate Macroalgae
MALCmin	0.100000E-01	(gC/gDM)	minimum C in storage
CDRatMALS	0.200000	(gC/gDM)	C to structural dry mass ratio in MALS
ArDenMAL	60.0000	(gDM/m2)	Area density frond (grams/m2 surface area)
R1	0.278500E-03	(gC/dm2/h)	Reference respiration rate at T1
R2	0.542900E-03	(gC/dm2/h)	Reference respiration rate at T2
Tr1	285.000	(degK)	reference temperature 1 for respiration
Tr2	290.000	(degK)	reference temperature 2 for respiration
P1	0.122000E-02	(gC/dm2/h)	Reference photosynthetic rate at T1

P2	0.144000E-02	(gC/dm2/h)	Reference photosynthetic rate at T2
Tp1	285.000	(degK)	temp for reference photosynthetic rate 1
Tp2	288.000	(degK)	temp for reference photosynthetic rate 2
Tap	1694.00	(degK)	Arrhenius temperature for photosynthesis
Taph	25924.0	(degK)	Arrhenius temperature for photosynthesis Arrhenius temp for photosynthesis high end
-	27774.0	, , ,	, , , ,
Tapl	1	(degK)	Arrhenius temp for photosynthesis low end
Tar	11033.0	(degK)	Arrhenius temp for respiration
ExtVI	0.180000	(1/m)	total extinction coefficient visible light
alpha0	0.375000E-04	((gC/dm2/h)/(umol/m2	photosynthetic efficiency MALC
Isat	43.7630	(W/m2)	light intensity where photosynthesis is max
exuMALC	0.500000	(gC/gC)	exudation parameter
MALNmin	0.100000E-01	(-)	minimum N in storage
MALNmax	0.220000E-01	(gN/gDM)	maximum N in MALN
MALPmin	0.100000E-02	(gP/gDM)	minimum P in storage
MALPmax	0.220000E-02	(gP/gDM)	maximum P in MALP
NCRatMALS	0.500000E-01	(gN/gC)	N:C ratio in MALS
PCRatMALS	0.500000E-02	(gP/gC)	P:C ratio in MALS
Ksn	0.560000E-01	(gN/m3)	half saturation MALN N uptake
Ksp	0.126000E-01	(gP/m3)	half saturation MALN P uptake
JNmax	0.336000	(gN/m2/d)	maximum MALN N uptake rate (per area frond)
JPmax	0.336000	(gP/m2/d)	maximum MALP P uptake rate (per area frond)
Vel	0.150000	(m/s)	velocity
Vel65	0.300000E-01	(m/s)	current speed at which J = 0.65Jmax
Latitude	52.0000	(degrees)	latitude of study area
m1	0.108500	(-)	growth rate parameter 1
m2	0.300000E-01	(1/d)	growth rate parameter 2
MALS0	0.600000E-01	(m2)	growth rate parameter 3
a1	1.02000	(-)	photoperiod parameter 1
a2	0.120000	(-)	photoperiod parameter 2
mrtMAL	0.120000	(1/dm2)	epsilon erosion/mortality parameter macro
CDRatMAL	0.200000	, ,	C:DM ratio in MALS
NCRatMAL	0.500000 0.500000E-01	(-)	N:C ratio in MALS
	0.500000E-01	(-)	
PCRatMAL		(-)	P:C ratio in MALS
Kn	2.72000	(gN/gN)	mass of nitrogen reserves per gram nitrogen
Kc	2.12130	(gC/gC)	mass of carbon reserves per gram carbon
Kdw	0.785000E-01	(-)	structural dry weight per unit frond area
FrPO1MAL	0.750000	(-)	fraction of MALS that goes to POC1 in decay
FrPO2MAL	0.250000	(-)	fraction of MALS that goes to POC2 in decay
TotalDepth	10.0000	(m)	total depth water column
FootDepth	-999.999	(m)	location of frond attachment in the water columns
LmaxMAL	10.0000	(m)	Maximum length MALG
SWGroDir	1.00000	(-)	grow direction MALG(1 = up -1 = down)
LinDenMAL	100.000	(g/m3)	linear density of macroalgae
RefDay	0.00000	(d)	daynumber of reference day simulation
AuxSys	86400.0	(scu/d)	ratio between days and system clock
Velocity	0.500000	(m/s)	horizontal flow velocity
VWind	3.00000	(m/s)	wind speed
SWRear	10.0000	(-)	switch for oxygen reaeration formulation (1-13)
KLRear	1.00000	(m/d)	reaeration transfer coefficient
TCRear	1.01600	(-)	temperature coefficient for rearation
Salinity	35.0000	(g/kg)	Salinity
fcover	0.00000	(-)	fraction of water surface covered <0-1>
KLRearMax	1000.00	(m/d)	maximum KLREAR oxygen for temp. correction
KLRearMin	0.200000	(m/d)	minimum rearation transfer coefficient oxygen
INEI ICAIIVIIII	0.20000	(11/G)	minimum rearation transfer coefficient oxygen

Rain	0.00000	(mm/h)	rainfall rate
coefAOxy	1.66000	(m/d)	gas transfer Oxy coefficient transmission
coefB1Oxy	0.260000	no unit	gas transfer O2 coefficient wind scale 1
coefB2Oxy	1.00000	no unit	gas transfer O2 coefficient wind scale 2
coefC1Oxy	0.660000	no unit	gas transfer O2 coefficient rain scale 1
coefC2Oxy	1.00000	no unit	gas transfer O2 coefficient rain scale 2
coefD1Oxy	1800.06	no unit	fresh water coefficient1 for Schmidt nr Oxy
coefD2Oxy	120.100	no unit	fresh water coefficient2 for Schmidt nr Oxy
coefD3Oxy	3.78180	no unit	fresh water coefficient3 for Schmidt nr Oxy
coefD4Oxy	0.476080E-01	no unit	fresh water coefficient4 for Schmidt nr Oxy
CI	20000.0	(g/m3)	Chloride
SWSatOXY	1.00000	(-)	switch for oxygen saturation formulation (1 2)
SWRearCO2	11.0000	(-)	switch for CO2 reaeration formulation (1113)
KLRearCO2	1.00000	(m/d)	CO2 reaeration transfer coefficient
TCRearCO2	1.01600	(-)	temperature coefficient for rearation CO2
KLRMaxCO2	1000.00	(m/d)	maximum KLREAR CO2 for temperature correction
KLRMinCO2	0.200000	(m/d)	minimum rearation transfer coefficient CO2
coefACO2	1.66000	(m/d)	gas transfer CO2 coefficient transmission
coefB1CO2	0.260000	no unit	gas transfer CO2 coefficient wind scale 1
coefB2CO2	1.00000	no unit	gas transfer CO2 coefficient wind scale 2
coefC1CO2	0.660000	no unit	gas transfer CO2 coefficient rain scale 1
coefC2CO2	1.00000	no unit	gas transfer CO2 coefficient rain scale 2
coefD1CO2	1800.06	no unit	fresh water coefficient1 for Schmidt nr CO2
coefD2CO2	120.100	no unit	fresh water coefficient2 for Schmidt nr CO2
coefD3CO2	3.78180	no unit	fresh water coefficient3 for Schmidt nr CO2
coefD4CO2	0.476080E-01	no unit	fresh water coefficient4 for Schmidt nr CO2
SWSatCO2	1.00000	(-)	switch for CO2 saturation formulation (1 2)
PCO2	0.316000E-03	(atm)	partial atmospheric CO2 pressure
SWTICdummy	0.00000	(-)	dummy option for TIC do not change value
Poros	1.00000	(-)	volumetric porosity
pHmin	1.00000	(-)	minimum allowed calculated pH
pHmax	14.0000	(-)	maximum allowed calculated pH

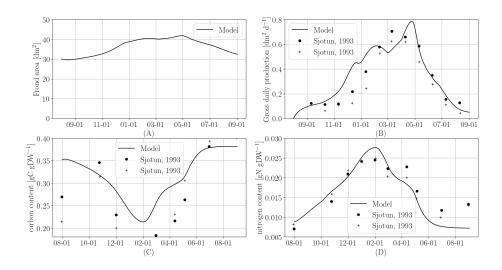


Figure 5: (A) Standing frond area. (B) Gross daily frond area produced. Solid line, model results. Circles, daily area produced estimated from Sjotun (1993), 2-year plants. Crosses, daily area produced estimated fromSjøtun (1993), 3-year plants (C) Carbon content expressed as a fraction of dry weight. Solid line, model results. Circles, Sjøtun (1993) proximal/meristematic tissue. Crosses, apical frond tissue. (D) Nitrogen content expressed as fraction dry weight. Solid line, model results. Circles, Sjøtun (1993) 2-year plants. Crosses, Sjøtun (1993), 3-year plants.

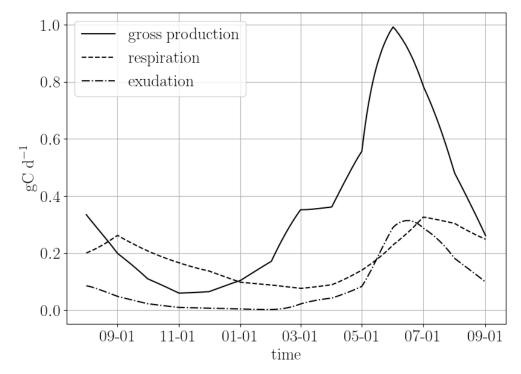


Figure 6: Carbon budget for the kelp plant

0.3 References

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