



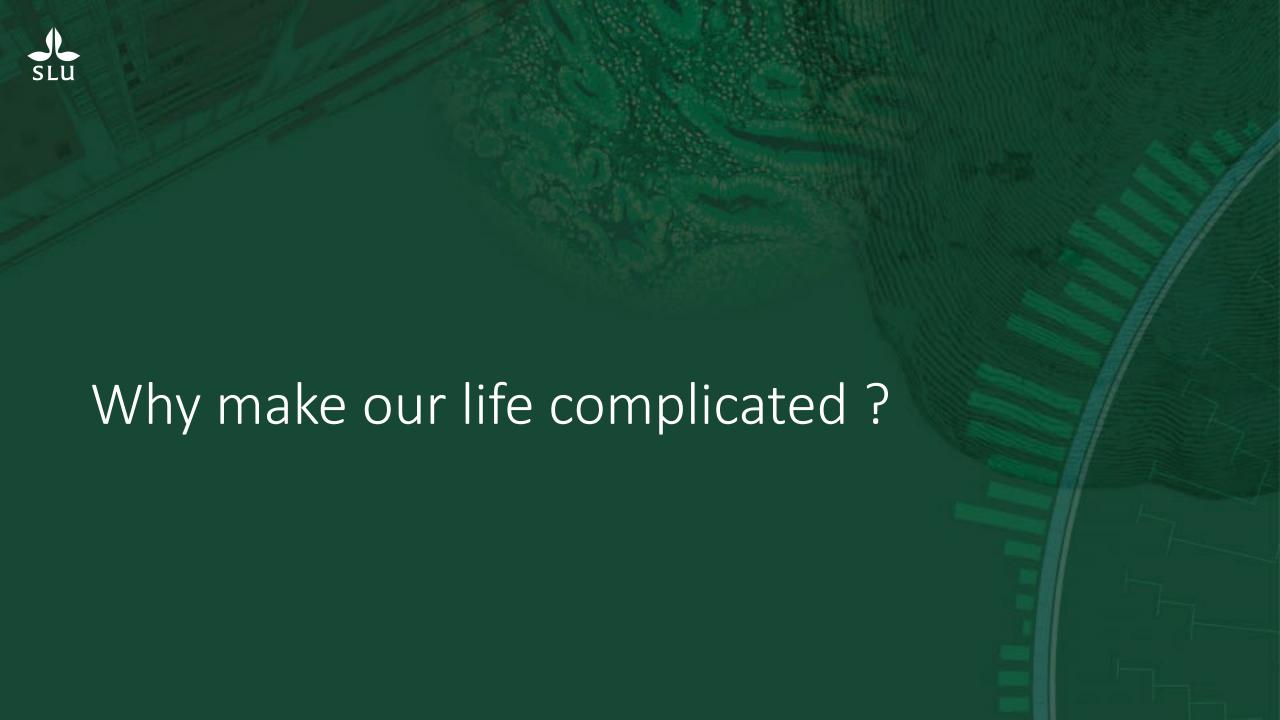
Traditional genomics

One organism to study → One genome



• Many organisms to study together -> A Metagenome



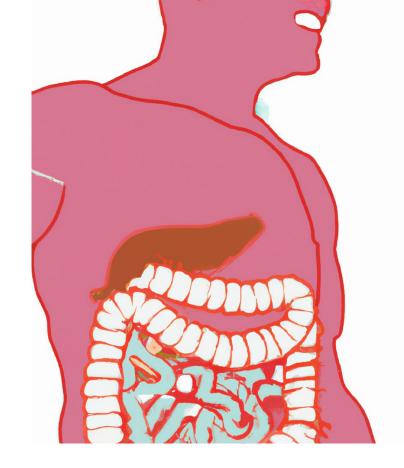


Why make our life complicated?

- It is hard/impossible to separate the organisms:
- Isolating (e.g. making a pure culture) microbes is hard
- We want to study interaction
- We want to study organisms in situ
- It's mainly about the microbes

Human Gut example

- 10*10^13 cells (1.3 x human cells)
- About 0.5% of dry weight
- ~400 species in every gut
- But only ~20% cultivated
- 90% of disease in humans can be related to microbiomes
- Most nutrients that end up in your blood stream have been somewhat metabolized by some microbe



A lake

- 1.35*10^17 cells (a milion per mL)
 - About 135kg of dry weight
 - ~700 species in the water
 - 3.000.000 encoded genes
 - Only a couple of handfull are cultivated.



Why does it matter?

They are everywhere They do a lot of stuff:

- 90% of disease in humans can be related to microbiomes
- Most nutrients that end up in your blood stream have been

somewhat metabolized by some microbe

- Microbes do 2/3rds of carbon fixation in aquatic environments
- But also the majority of carbon emissions!



Different Sequencing Strategies give different information

- Who is in the environment? (taxonomy)
- How many of each? (abundance)
- What can they do? (genetic potential)
- What are they doing? (expression analysis)
- What do they eat? (metabolism)
- Where do they come from? (evolution and ecology)



The strategies

- Targeted, a.k.a. amplicon/metabarcoding/edna
- Shotgun sequencing:
 - Short reads (HiSeq, NovaSeq, PacBio Onso)
 - Long reads (PacBio, Nanopore)
- Metatranscriptomic
- Functional metagenomics

Two Main Strategies

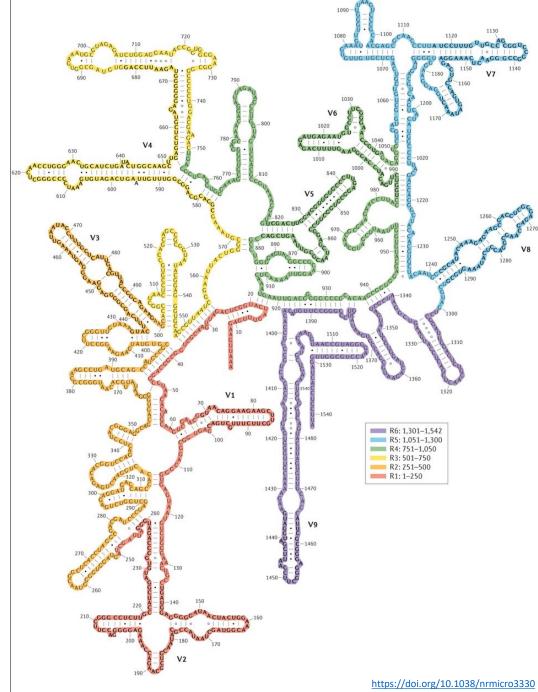
Shotgun	Targeted
Explorative approach	Pick 1 gene to sequence
Sequence everything then rebuild the puzzles	Snapshot of a function or taxonomy
Many different methodologies	Much cheaper and more depth
2 Sub approaches (assembly based and assembly-free)	Data base relient



Targeted: 16s rRNA

The 16S rRNA gene contains conserved and variable regions, allowing for the identification of bacteria and archaea at different taxonomic levels.

- Small genomic region → Cheaper sequencing while maintaining good
- Extensive reference databases → good for finding what we know exist, bad for novel discovery
- Can be easily amplified with universal primers, making it accessible for most sequencing platforms.



Targeted: 16s rRNA

Short read sequencing

- 150-300bp (usually v3-v4 regions)
- High base accuracy
- High number of reads leading to High depth and taxonomic resolution (up to species level)
- Challenging to differentiate closely related species (95% identity same genus, 97% same species and 99% strain)

Long read sequencing

- Entire gene (~1.5 kb)
- Lower accuracy or equal to Illumina (PacBio)
- Less reads → less depth for complex communities
- Full length gene allow for better classification of related species and strain-level differences (can even reach divergent copies within genome*)
- Tools still maturing

DB: https://www.arb-silva.de/

^{*} https://doi.org/10.1038/s41467-019-13036-1

Other genes used for metabarcoding

1. ITS (Internal Transcribed Spacer)

- Fungi
- **Applications**: Widely used in fungal diversity studies, plant-fungal interactions, and environmental microbiology.
- **DB:** https://unite.ut.ee/repository.php

2.18S rRNA

- Eukaryotes
- **Applications**: Commonly used in studies of eukaryotic microbial communities, environmental monitoring, and marine biodiversity.
- **DB:** https://www.arb-silva.de/

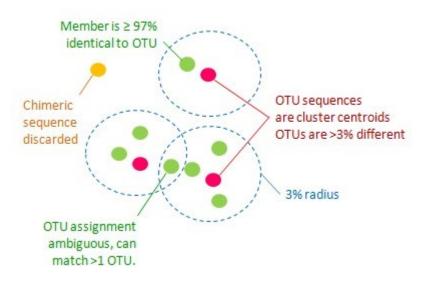
3. COI (Cytochrome c Oxidase I)

- Animals
- **Applications**: Used in biodiversity assessments, conservation studies, and species identification.
- **DB:** https://www.ibol.org/phasel/bold/

Clustering sequence: OTU

Take into account:

- intraspecific variability
- sequencing errors
- 97% similarity: species threshold
- two types: de novo and reference-based



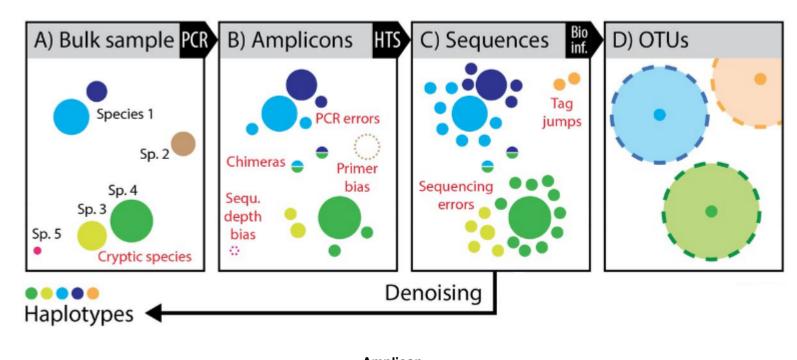
https://drive5.com/usearch/manual/uparseotu_algo.html

Clustering sequence: ASV

• Amplicon sequence variants (today's tutorial)
The "true" biological sequences are inferred from reads
"Biological sequences are discriminated
from errors on the basis of, in part, the
expectation that biological sequences are
more likely to be repeatedly observed than
are error-containing sequences".

• Further reading https://www.nature.com/articles/ismej2017119

OTU vs ASV



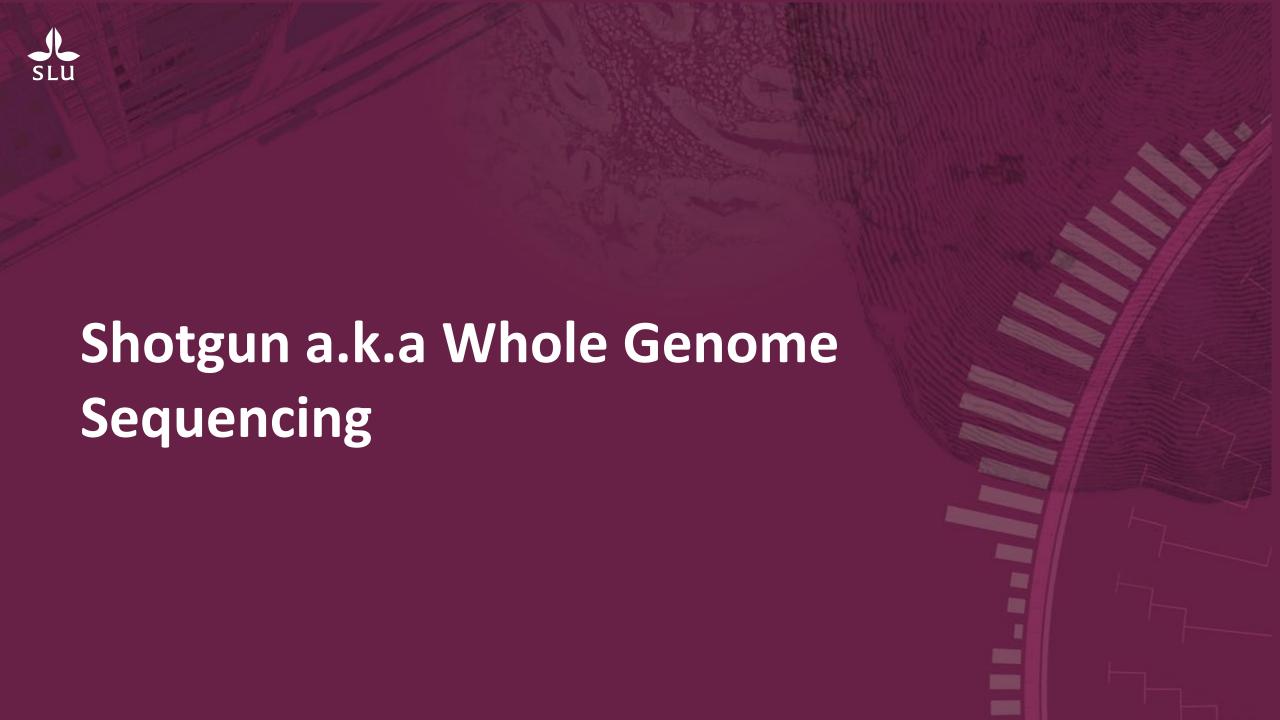
Sample Sequences Amplicon Sequence Variants (ASVs)

PCR/sequencing Remove/correct errors

https://trainings.migale.inrae.fr/posts/2022-06-07-module20/content/slides.html#55

For metabarcoding (16s)

- https://benjjneb.github.io/dada2/ (ASV)
- https://mothur.org/ (OTU)
- https://qiime2.org/ (OTU)





Whole Genome sequencing Analysis Methods

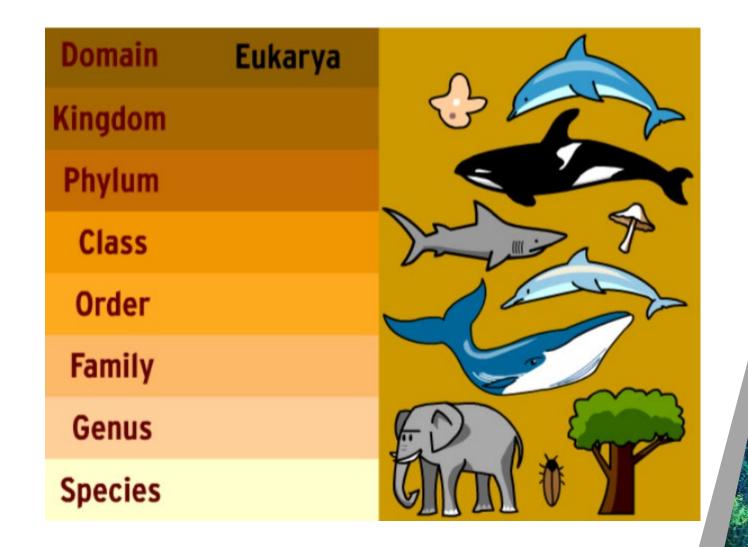
Assembly-free	Assembly based
map/classify reads	Assembling reads into contigs
Direct annotation	Binning the contigs by similarity
Heavily database reliant	Annotation of the bins
Faster, easier	Slow, tricky
More noisy	More specific

Assembly free

- The inventory approach:
 - Annotating the reads directly
 - Heavily relying on databases
 - Good for well known microbiomes (western European poop, agricultural soils)
 - Dangerous for others as methods tend to overclassify

Assembly free

- Taxonomically:
 - Kraken
 - Kaiju
 - Functionally:
 - Blast ...
 - Megan
 - Humann



Tools

- Kraken2/Bracken
 - https://github.com/DerrickWood/kraken2
 - https://github.com/jenniferlu717/Bracken
 - DB: https://benlangmead.github.io/aws-indexes/k2
- Kaiju
 - https://github.com/bioinformatics-centre/kaiju
 - DB: https://bioinformatics-centre.github.io/kaiju/downloads.html
- Krona
 - https://github.com/marbl/Krona/wiki

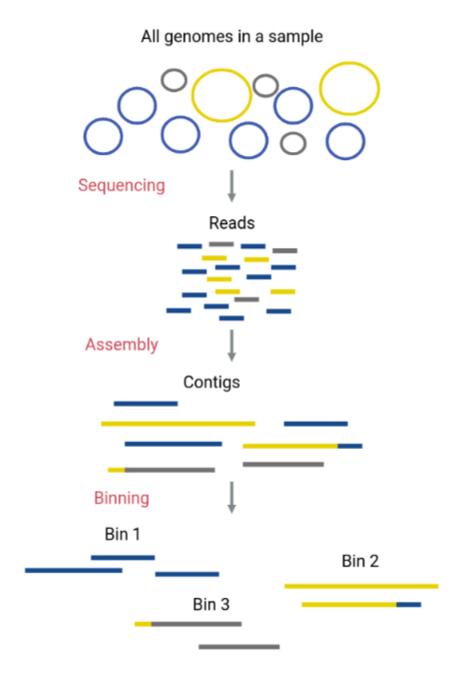
Assembly based

- Turning the reads into genes and more! (aka The Contig):
 - Full length coding sequences
 - Pieces of genomes
 - Or anything else made of DNA
- Increases specificity of previous approaches, but might lose some sensitivity (e.g. not everything assembles

Assembly based

- Genome-resolved metagenomics:
 - Cluster contigs into bins
 - Identify possible genomes
 - Quantify completeness/redundancy
 - Assign taxonomy

Binning



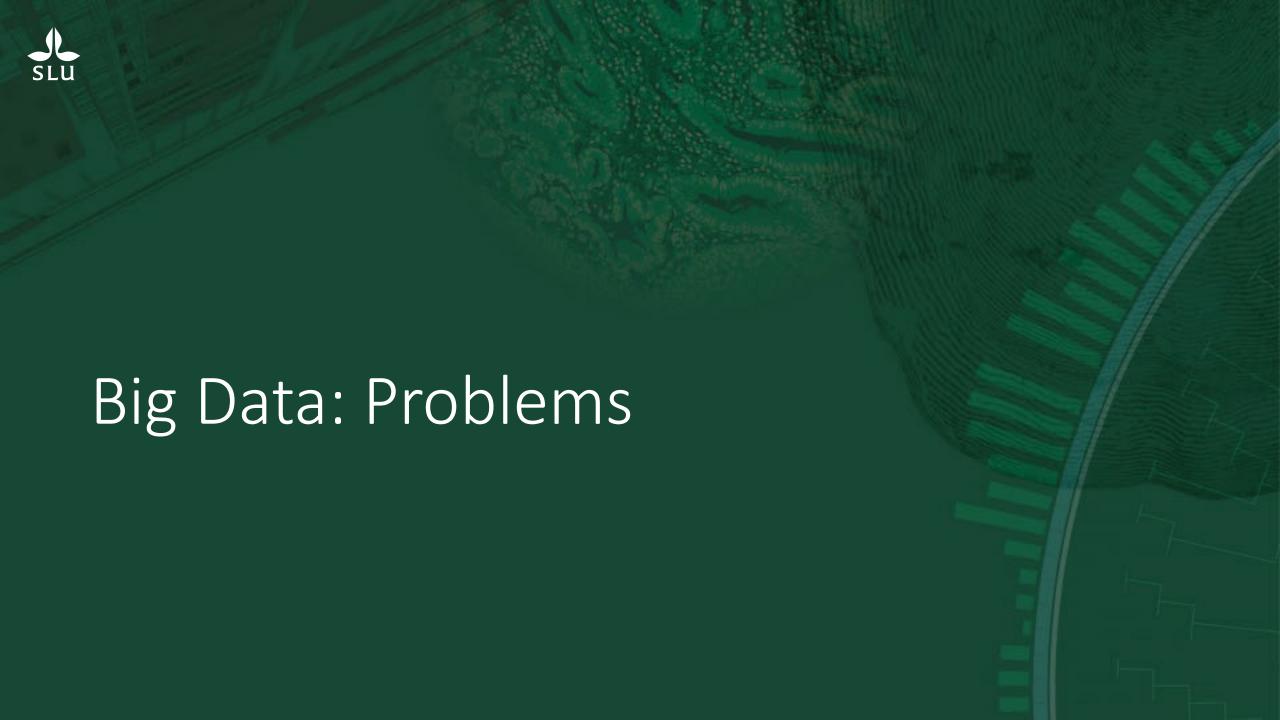


Gene Atlas Approach

If you only care about genes:

- Predict genes and annotate genes:
- Functionally
- Taxonomically
- Gene clustering:
- By similarity
- Into COGs
- Into families
- Quantify and analyse diversity





Storage

- We want to sequence the soil of grapevines with nanopore MinIon (rough output between 100Gb and 300Gb of raw files) once basecalled it becomes 5-15Gb of fastq reads
- To have a good experiment we need at least 5 samples per type of soil and grapevine.
- Let say you want to try 3 different grapevine breed through 2 farms that leave you with the need to store at minimum:
- 3,15 Tb of sequencing data for 1 minimal experiment (100+5)*5*3*2=3150

Time

To reproduce a typical computational biology paper with minimal expertise it takes 280 Hours.

https://doi.org/10.1371/journal.pone.0080278

That include:

- Understanding the paper and material used
- Installation, set up
- Finding parameters
- Workflow validation
- Computing

Reproducibility

- 1. Access to the tools mentioned in the paper
- 2. Difficulty of installation
- 3. Version issues (machine and software)

https://doi.org/10.1371/journal.pbio.3000333



The issues of Big Data:

- 1. Storage
- 2. Reproducibility
- 3. Computing Power
- 4. Time
- 5. Money
- 6. Consistency
- 7. Usability

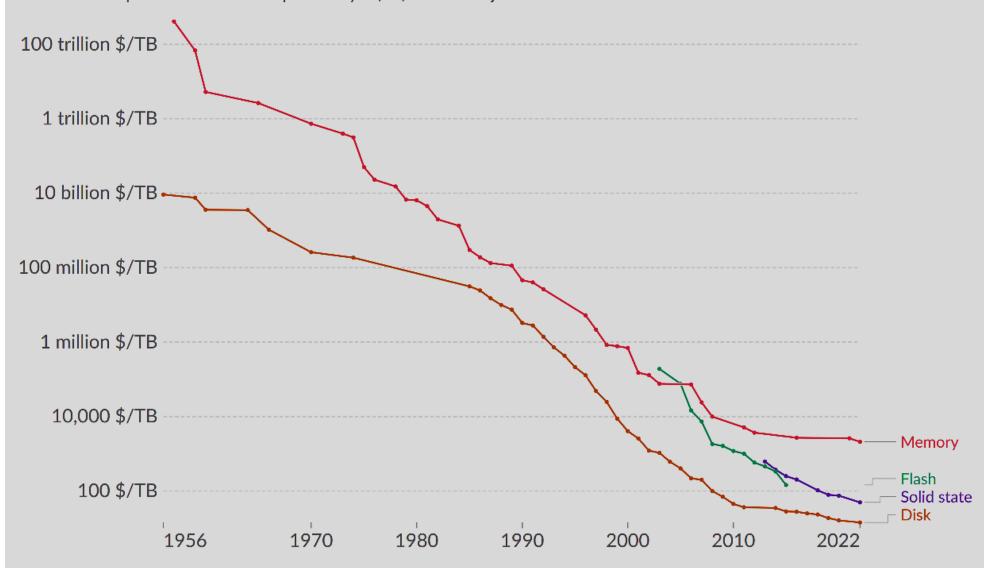




Historical cost of computer memory and storage



This data is expressed in US dollars per terabyte (TB). It is not adjusted for inflation.



Source: John C. McCallum (2022)

OurWorldInData.org/technological-change • CC BY

Note: For each year, the time series shows the cheapest historical price recorded until that year.

Workflow Manager

- One command runs the entire workflow
- Parallelisation is automatic and scheduled based on the available resources
- Allow reuse of code in other scripts
- Continuous checkpoints for resuming and expanding the pipelines

Usability - Workflow Manager

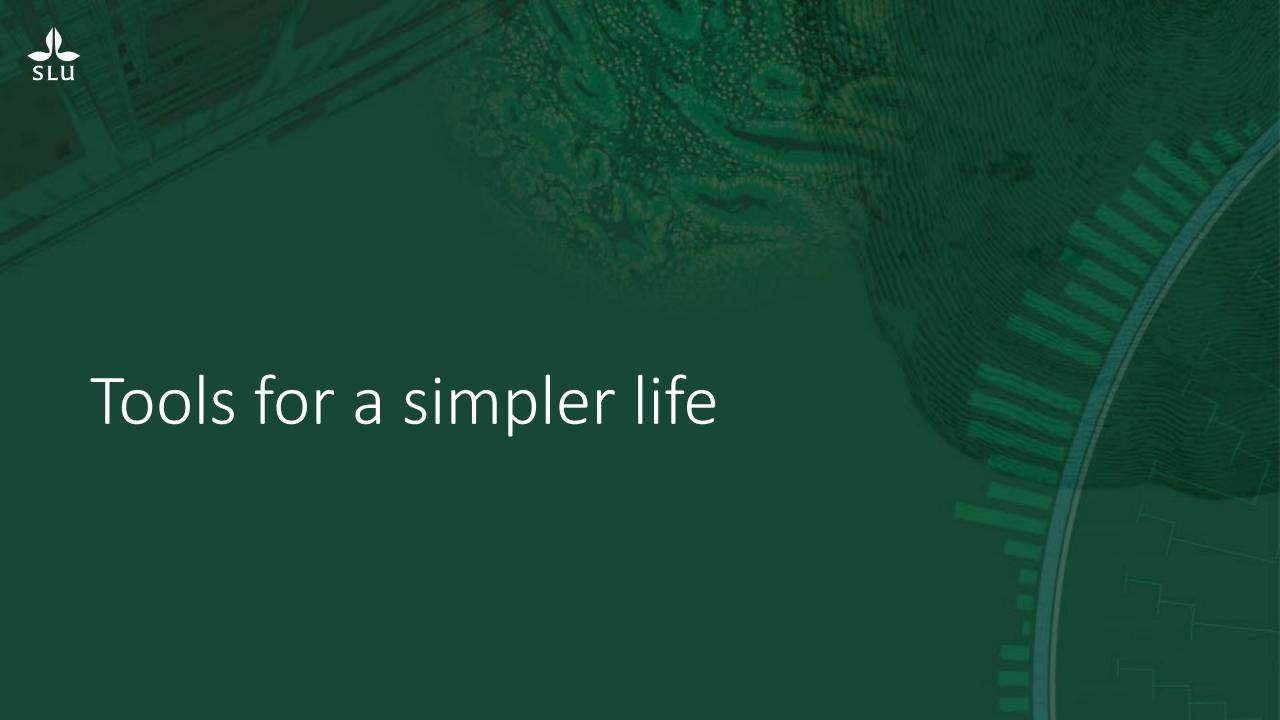
- One command runs the entire workflow
- Parallelisation is automatic and scheduled based on the available resources
- You can write your internal code in any language (bash, R, Python)
- The intermediary files are handled
- Allow reuse of code in other scripts
- Continuous checkpoints for resuming and expanding the pipelines

Reproducibility - Workflows Manager

- Pipeline written in stone and version controlled (Git)
- Support for containers and in some cases conda environment
- The application handling and the configuration/deployment are separated
- The only parameters to change are the resources and environment

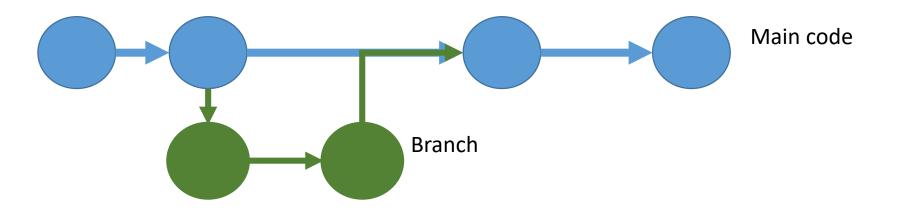
Consistency

- Version control (git, bitbucket, github, gitlab)
- Container (Apptainer, Docker, Podman)
- Open source



Git

- version control system that handles all from small to very large projects
- Simple to use
- Track changes and save specific versions, those version can be changed but are then new versions



Container vs Virtual Machine

- Lighter: MB vs GB
- Faster startup as it virtualises a process and not an OS
- Immutable: does not change over time
- Composable: The output of one can be fed directly in the next
- Transparent









Genomic workflows

- Data analysis apps to retrieve information from datasets
- Huge parallelisation, creating numerous job over a cluster
- Mix of tools and scripts
- Complex configurations and dependencies interactions between said tools

Workflow Manager

SnakeMake	Nextflow
CL oriented tool	CL oriented tool
Rules defined using file name pattern	Can manage any data structure
Built-in support Apptainer	Support all major container runtimes
Custom script for cluster	Built-in support for cluster and cloud services
No support for source code management system	Built-in support for Git/GitHub, manage pipeline revisions
Python based	Groovy/JVM based

For metagenomics (shotgun)



- https://github.com/RVanDamme/MUFFIN
- https://nf-co.re/mag/2.4.0
- https://github.com/metagenome-atlas/atlas

• ...

