Analyzing ChIP-Seq data using Bioconductor

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Isoform-specific function of AIB1 during breast cancer progression

- AIB1: Amplified in Breast Cancer 1, transcription coactivator
- High grade tumors, Poor prognosis, Therapy Resistance
- Two isoforms:
- 1) AIB1-FL
- 2) AIB1-D4
- Expression of both isoforms goes up during progression

Experimental Design

- CRISPR engineered breast cancer cell lines that express either one of AIB1 isoforms

- ChIP-Seq: crosslink and pull down AIB1 with DNA, then DNA sequencing to identify difference and similarities.

- Used Bioconductor packages: ChIPseeker and ChIPpeakAnno.

```
```{r setup, include=TRUE}
knitr::opts_chunk$set(echo = FALSE, message=FALSE, warning=FALSE)
Installing packages from Bioconductor, showing an example below
if (!requireNamespace("BiocManager", quietly = TRUE))
install.packages("BiocManager")
BiocManager::install("ChIPseeker")
library(AnnotationDbi)
library(BiocManager)
library(Biostrings)
library(BSgenome)
library(plyr)
library(dplyr)
library(GenomicAlignments)
library(GenomicFeatures)
library(ggplot2)
library(knitr)
library(limma)
library(org.Hs.eg.db)
library(seqRFLP)
library(tibble)
library(wesanderson)
library(ChIPseeker)
library(vennplot)
library(clusterProfiler)
library(TxDb.Hsapiens.UCSC.hg38.knownGene)
library(ChIPpeakAnno)
library(tidyverse)
library(EnsDb.Hsapiens.v86)
save human genome version hg38 with a shorter name
txdb <- TxDb.Hsapiens.UCSC.hq38.knownGene
Converting genes into genomic ranges
columns(EnsDb.Hsapiens.v86)
TxGR<-toGRanges(EnsDb.Hsapiens.v86, feature='gene')</pre>
```

# DNA sequences were aligned to the genome then peaks called using MACS2

```
After algining to the genome, sequencing peaks are shortened to 250bp

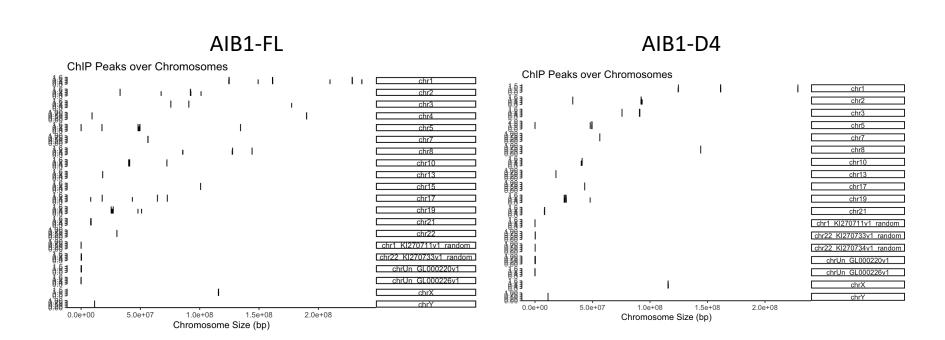
aDCIS_all<-("./aDCIS_all.bed")
aDCIS_all_peak<-readPeakFile(aDCIS_all)
aDCIS_all_peak_250<-resize(aDCIS_all_peak,250,fix="center")
export.bed(aDCIS_all_peak_250,"./aDCIS_peak_250")

aD10_all<-("./aD10_all.bed")
aD10_all_peak<-readPeakFile(aD10_all)
aD10_all_peak_250<-resize(aD10_all_peak,250,fix="center")
export.bed(aD10_all_peak_250,"./aD10_peak_250")
```

# To visualize peaks on chromosomes

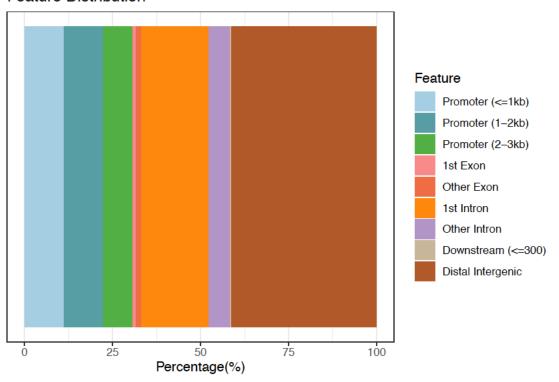
```
ChIPseeker::covplot(aDCIS_all_peak)

ChIPseeker::covplot(aD10_all_peak)
```



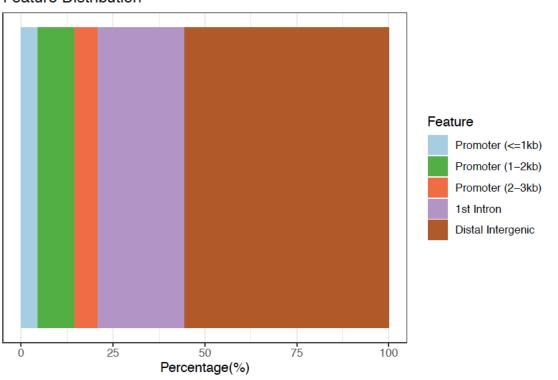
# Distribution of peaks relative to gene features

#### Feature Distribution



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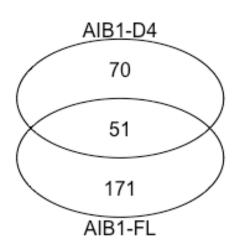


### To determine the overlap of peaks between the two isoforms

```
Determine the overlap of peaks between the two isoforms

aDCIS_short<-("./aDCIS_peak_250")
 aDCIS_short_peak<-readPeakFile(aDCIS_short)
 aD10_short<-("./aD10_peak_250")
 aD10_short_peak<-readPeakFile(aD10_short)
 aDCIS_comp_list<-list(aDCIS_short_peak,aD10_short_peak)
 names(aDCIS_comp_list)<-c("AIB1-FL","AIB1-D4")
ChIPseeker::vennplot(aDCIS_comp_list)

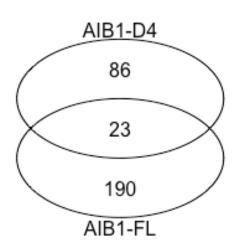
Add statistics to the overlap
enrichPeakOverlap(queryPeak= aDCIS_short,targetPeak= aD10_short, TxDb= txdb,pAdjustMethod = "BH",nShuffle= 1000,chainFile= NULL,verbose= FALSE)</pre>
```



```
qSample tSample qLen tLen N_OL pvalue p.adjust ## 1 aDCIS_peak_250 aD10_peak_250 222 121 51 0.000999001 0.000999001
```

# To annotate identified peaks to genes and check overlap

```
peakAnnoList <- lapply(aDCIS_comp_list, annotatePeak, TxDb=txdb,tssRegion=c(-3000, 3000), verbose=FALSE)
genes= lapply(peakAnnoList, function(i) as.data.frame(i)$geneId)
ChIPseeker::vennplot(genes)</pre>
```



### Converting peak files to Genomic Ranges, overlap and determine distribution from TSS

```
ggplot(data = tibble(dist = pk_anno$distancetoFeature), aes(x = dist))+
 theme_bw()+
 scale_x_continuous(name = 'Distance from TSS (bp)', limits = c(-5e3, 5e3))+
 scale_y_continuous(name = 'Density')+
 geom_vline(xintercept = 0, color = 'red')+
 geom_density(alpha = 0.1, fill = 'blue')
```

