# Supplement 1: Photosynthesis and fluorescence

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## Contents



*This pdf was generated from an Rmarkdown file, which includes all R code necessary to reproduce the estimations. The Rmarkdown file is available on github (https://github.com/TTRademacher/Exp2019Analysis) and is permanently and publicly archived on the Harvard Forest Data Archive as part of the data set HF???.*

# 1 Photosynthesis

Towards the end of the chilling period (i.e., last ten days of chilling; Fig. 1), we measured instantaneous assimilations rates in leaves towards the top (sub-exposed) and bottom (shade-leaves) of the canopy for all eight trees as well as response curves to $CO_2$ (commonly referred to as A/Ci curves) and light. We conducted those measurement to see if the chilling had affected actual photosynthesis and photosynthetic capacity. All measurements were performed with a LICOR-6400 (Lincoln, Nebraska, USA) from a bucket lift that was parked in between the trees.

## 1.1 Instantaneous photosynthetic rates at top and bottom of the canopy

For instantaneous rates, the best mixed effects model included only interactive fixed effects of time of day and position in the canopy and a random effect accounting for natural between-tree variability (table S1). This model estimated the time of day and position effects at -0.7±0.3 and -2.1±0.5 $\mu mol\, m^{-2}\, s^{-1}$, respectively, and their interactions at 2.6±0.8 $\mu mol\, m^{-2}\, s^{-1}$. This model was only marginally better according to the conditional AIC than several alternative model formulations that included a treatment effect. When the treatment effect was included is was estimated to range from reducing photosynthesis in chilled trees by -0.3±0.7 to -0.7±0.9 $\mu mol\, m^{-2}\, s^{-1}$ compared to control trees. However, any difference between chilled and control trees appears to have been larger at the bottom of the canopy (Fig. S1). Overall, there is little evidence for a feedback effect on instantaneous photosynthesis rates.

Table 1: gives the conditions AIC for various formulations of mixed effect models fitted to the instantaneous photosynthetic rates measured in chilled and control trees towards the end of the chilling period. This table was automatically generated from publicly available code and data (data set ID here) on the Harvard Forest Data Archive.

| Fixed effect | Random effects | conditional AIC |
|---|---|---|
| NA | tree | 466.76 |
| time | tree | 463.52 |
| position | tree | 459.52 |
| treatment | tree | 466.36 |
| time + position | tree | 459.31 |
| time + treatment | tree | 463.22 |
| position + treatment | tree | 459.66 |
| time * position | tree | 448.87 |
| time * treatment | tree | 465.06 |
| position * treatment | tree | 461.71 |
| time + position + treatment | tree | 459.34 |
| time * position + treatment | tree | 448.93 |
| time + position * treatment | tree | 461.42 |
| time * treatment + position | tree | 461.33 |
| time * position * treatment | tree | 452.02 |

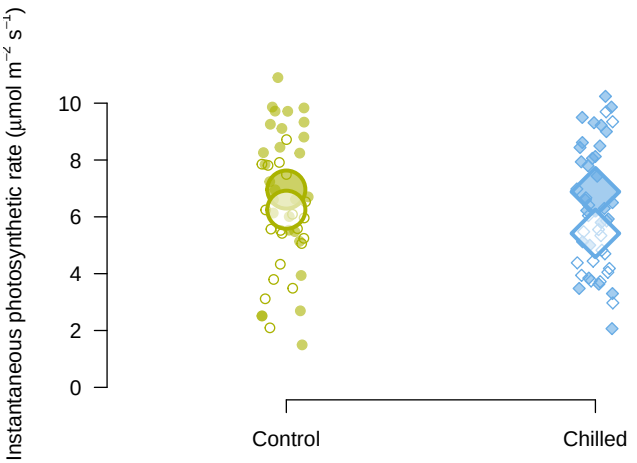


Figure 1: shows instantaneous assimilation rates for chilled (blue diamonds) and control trees (green dots) at the bottom (open symbols) and the top (closed symbols) of the canopy. Treatment group means are displayed with large symbols.

### 1.2 A/Ci curves

To estiamte whether chilling affected the photosynthetic apparatus, we derives maximal rates of photosynthetic electron transport ($J_{max}$) and RuBisCO carboxylase activity ($V_{cmax}$), and dark respiration ($R_d$) from measurements of assimilation in response to varying leaf internal $CO_2$ concentrations. We estimated photosynthethic parameters by fitting a model using the 'plantecophys' package (Duursma, 2015), which showed significant difference in photosynthetic parameters between chilled and control trees (Fig. S2). $V_{cmax}$ was 46.6±1.5 $\mu mol\, m^{-2}\, s^{-1}$ for chilled trees instead of 52.1±1.7 $\mu mol\, m^{-2}\, s^{-1}$ for control trees, $J_{max}$ was also lower at 85 $\mu mol\, m^{-2}\, s^{-1}$ for chilled compared to 97 $\mu mol\, m^{-2}\, s^{-1}$ for control trees, whereas dark respiration increased by 5% in chilled trees relative to control.

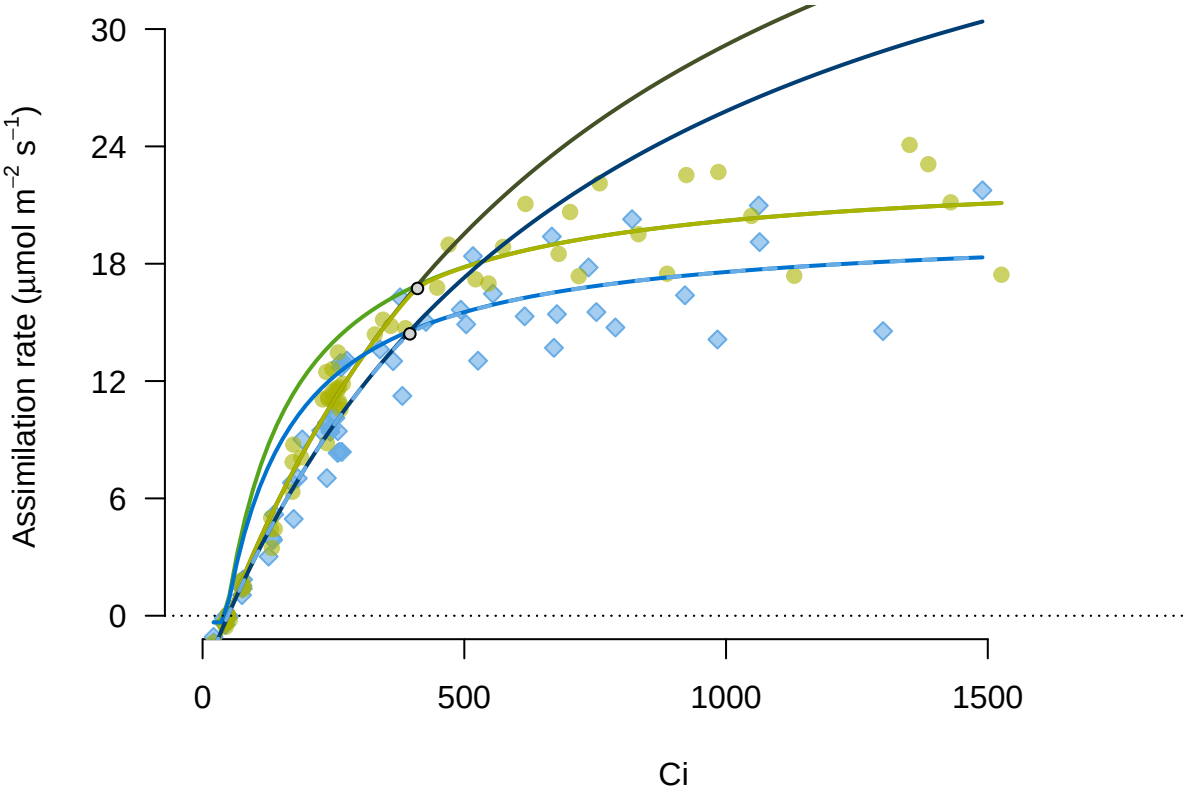


Figure 2: shows photosynthetic assimilation as a function of $CO_2$ concentration for chilled (blue diamands) and control trees (green dots). Blue and green lines display the best fitted photosynthetic model using the 'plantecophys' R package for chilled and control groups respectively.

### 1.3 Light response curves

In contrast to the A/Ci curves, light response curves varied seemingly idiosyncratically between trees and there was no clear association between of chilling and the light response curves in the leaves in the top of the canopy (Fig. S3). One chilled tree showed the lowest light response curve and saturation level, yet the two highest saturation levels were also found in chilled trees.

## 2 Leaf fluorescence

Leaf chlorophyll fluorescence in particular the ratio of variable fluorescence over maximum fluorescence (Fv/Fm) in dark-adapted leaves is know to be a reliable indicator of stress to photosystem II (Kitajima

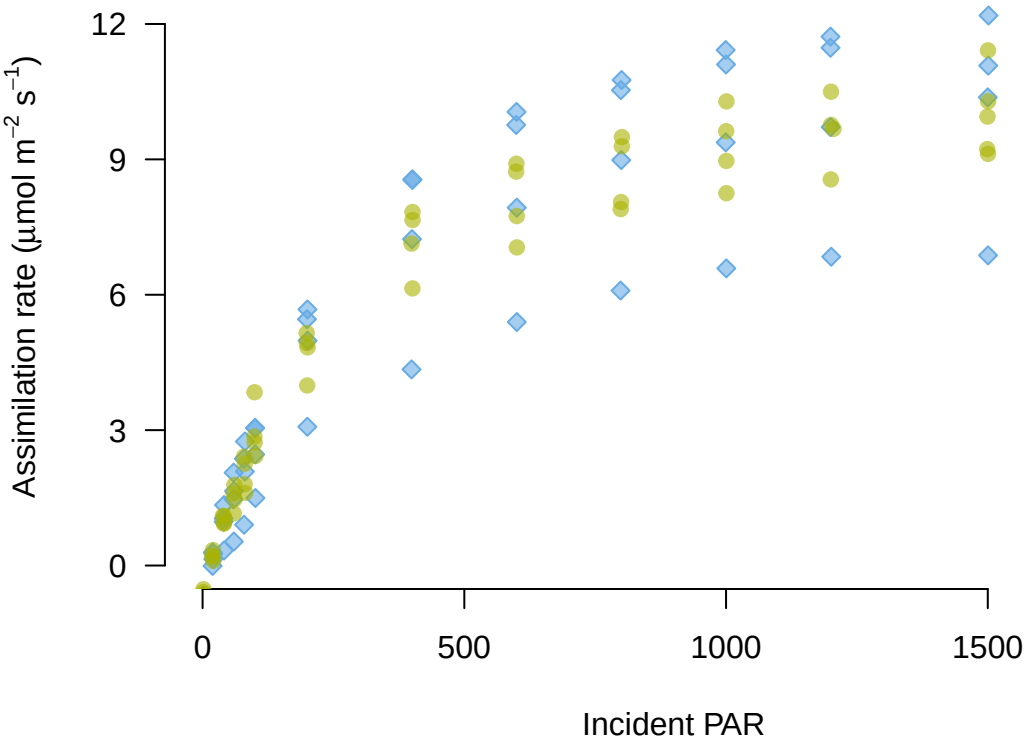


Figure 3: shows the light response curve measurement for chilled (blue diamonds) and control trees (green dots).

and Butler, 1975) and has also been directly linked to low temperature stress (Baker and Rosenqvist, 2004; Groom and Baker, 1992). To test whether photosystem II was stressed by the chilling in our experiment, we measured minimum fluorescence (Fo), maximum fluorescence (Fm) and the ratio of variable fluorescence over maximum fluorescence (Fv/Fm) in light-adapted and dark-adapted leaves during the last 10 days of chilling (Fig. 1). Directly after each photosynthesis measurement, we removed the leaf from the twig and immediately measured chlorophyll fluorescence with a OS-30P chlorophyll fluorometer (Opti-Sciences, Hudson, New Hampshire, USA). Subsequently, we wrapped the leaves in aluminium foil and stored them in a chest cooler with ice, before re-measuring fluorescense of the dark-adapted leaves in the evening of each sampling day (i.e., after several hours of dark-adaptation). Finally, the leaves were weighted, scanned (Epson Perfection V550 Photo Scanner, Long Beach, California, USA), dried at $60°C$ for 24 hours, and weighted again to obtain leaf area, fresh weight and dry weight for each leaf.

Overall, there were no stark differences in leaf fluorescence between chilled and control trees (Fig. S4 & S5). Dark adapted Fv/Fm were generally close to 0.8, which is close to the assumed optimum for many plant species (Maxwell and Johnson, 2000). However, there was a small difference between Fv/Fm for chilled and control trees (Fig. S4). The estimated treatment effect was -0.14±0.01, when we fit the mixed effects model containing position of the leaf in the canopy, treatment and their interaction. While this model minimised information loss according to the conditional AIC, it only looses marginally more information than a model that does not contain a treatment effect. Although the estimated effect is comparable to declines in wheat and maize leaves due to cold stress (Andrews et al., 1995; Groom and Baker, 1992), it is also of similar magnitude as normal seasonal changes in Fv/Fm of green summer and green autmun leaves in the closely related sugar maple (Junker and Ensminger, 2016). The lack of a clear difference suggests that the maximum potential quantum efficiency of photosystem II was not strongly affected by chilling here.

While there were no differences in minimum and maximum fluorescence between treatments (i.e., chilled versus control), there were differences in the minimum and maximum fluorescence between leaves from the bottom and the top of the canopy, when measured directly after the leaves were removed from the canopy.

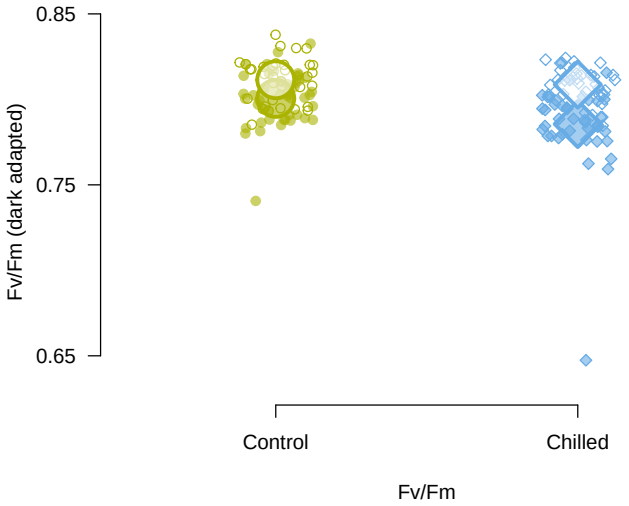


Figure 4: shows Fv/Fm measured immediately after removal and after dark-adapting each leaf for several hours for chilled trees (blue diamonds) and control trees (green dots) for leaves from the bottom (open symbols) and the top (closed symbols) of the canopy.

These differences persist, albeit slightly smaller, once leaves were dark-adapted (Fig. S5). Such differences in fluorescence between sun- and shade-leaves have also been reported for four other temperate deciduous tree species (Lichtenthaler et al., 2007) and are supported by theoretical models (Olioso et al., 1992).

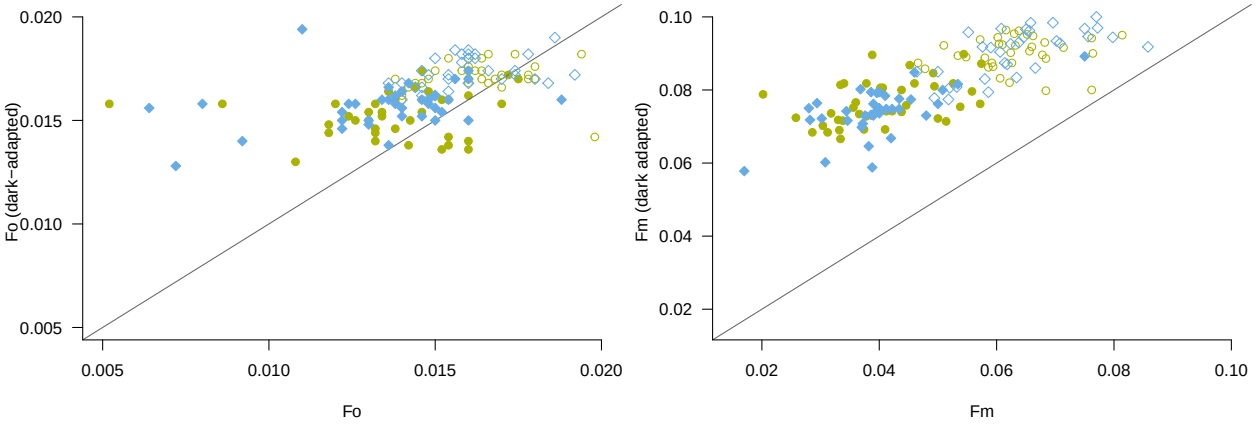


Figure 5: shows minimum (Fo) and maximum fluorescence (Fm) measured immediately after removal and after dark-adapting each leaf for several hours for chilled trees (blue diamonds) and control trees (green dots) for leaves from the bottom (open symbols) and the top (closed symbols) of the canopy.

# References

Andrews, J.R., Fryer, M.J., Baker, N.R., 1995. Characterization of chilling effects on photosynthetic performance of maize crops during early season growth using chlorophyll fluorescence. Journal of Experimental Botany 46, 1195–1203.

Baker, N.R., Rosenqvist, E., 2004. Applications of chlorophyll fluorescence can improve crop production strategies: An examination of future possibilities. Journal of Experimental Botany 55, 1607–1621.

Duursma, R.A., 2015. Plantecophys - An R Package for Analysing and Modelling Leaf Gas Exchange Data. PLoS ONE 10, e0143346.

Groom, Q.J., Baker, N.R., 1992. Analysis of Light-Induced Depressions of Photosynthesis in Leaves of a Wheat Crop during the Winter. Plant Physiology 100, 1217–1223.

Junker, L.V., Ensminger, I., 2016. Relationship between leaf optical properties, chlorophyll fluorescence and pigment changes in senescing Acer saccharum leaves. Tree Physiology 36, 694–711.

Kitajima, M., Butler, W.L., 1975. Quenching of chlorophyll fluorescence and primary photochemistry in chloroplasts by dibromothymoquinone. Biochimica et Biophysica Acta (BBA) - Bioenergetics 376, 105–115.

Lichtenthaler, H.K., Ač, A., Marek, M.V., Kalina, J., Urban, O., 2007. Differences in pigment composition, photosynthetic rates and chlorophyll fluorescence images of sun and shade leaves of four tree species. Plant Physiology and Biochemistry 45, 577–588.

Maxwell, K., Johnson, G.N., 2000. Chlorophyll fluorescence—a practical guide. Journal of Experimental Botany 51, 659–668.

Olioso, A., Méthy, M., Lacaze, B., 1992. Simulation of canopy fluorescence as a function of canopy structure and leaf fluorescence. Remote Sensing of Environment 41, 239–247.