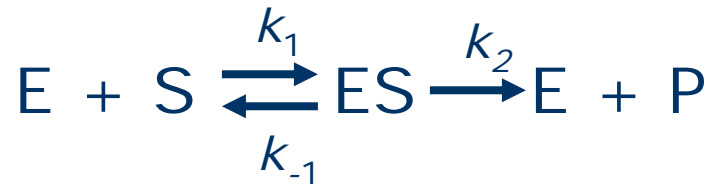


# Enzyme kinetics

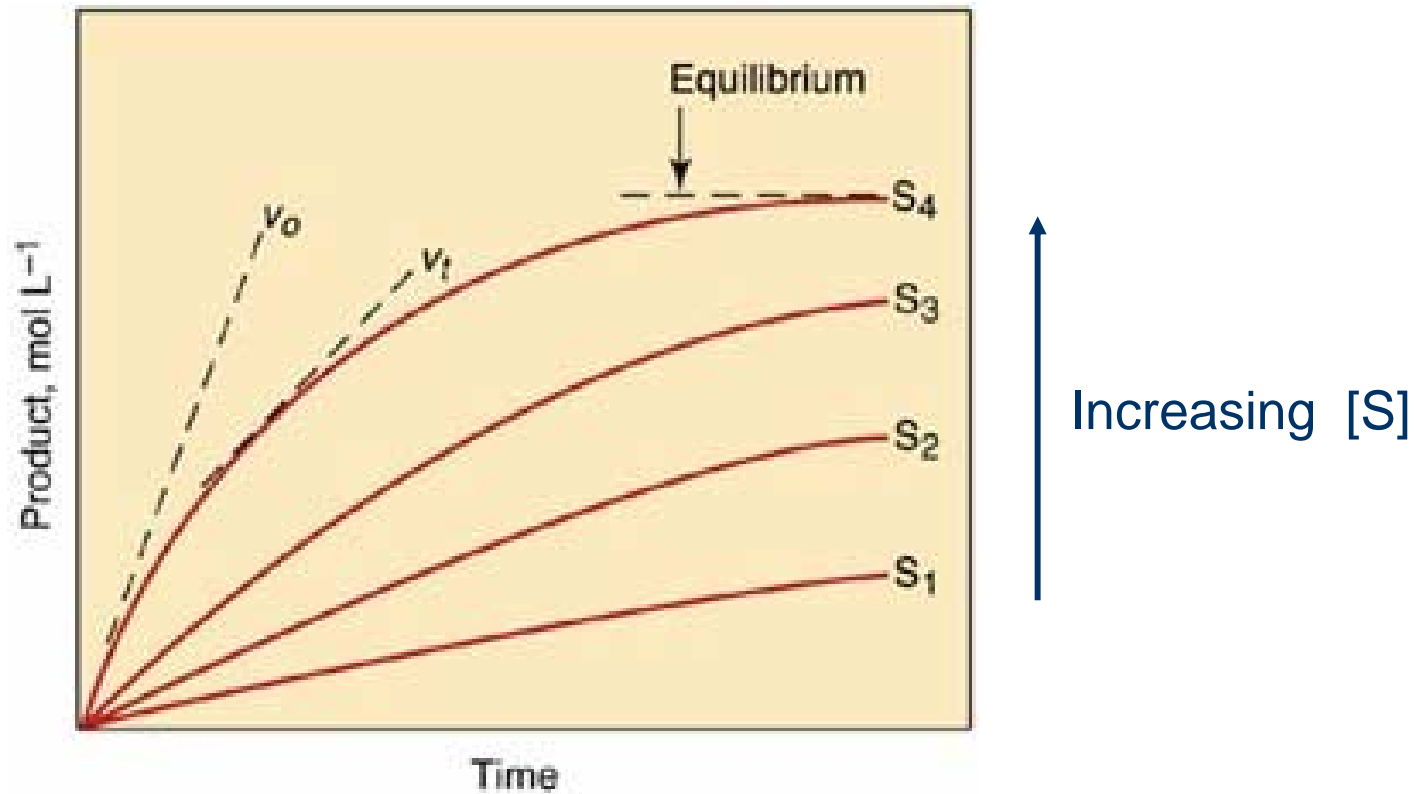
- Study of the rates of enzyme-catalyzed reactions
- Provides information on enzyme specificities and mechanisms

# Formula for a simple enzyme-catalyzed reaction



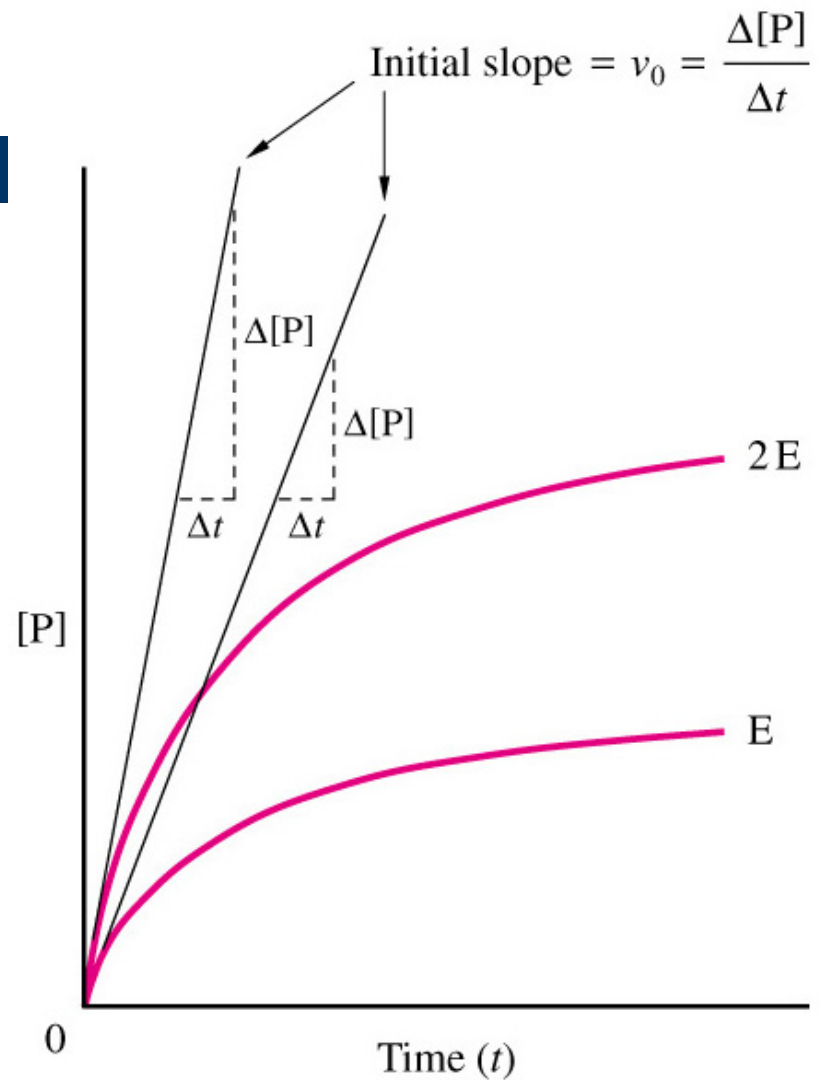
- E = free enzyme
- S = substrate
- ES = enzyme-substrate complex
- P = product

# What are we measuring?



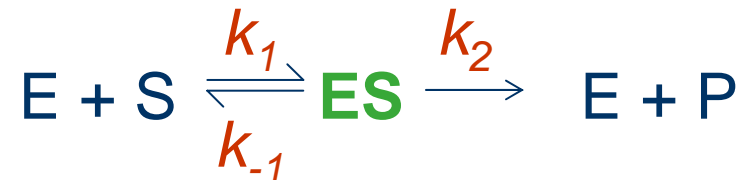
# Initial velocity

- Measured at the very beginning of a reaction when very little P has been made.



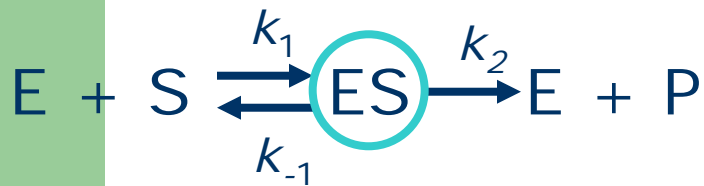
First order equation in respect to substrate concentration

## For enzyme-catalyzed reactions

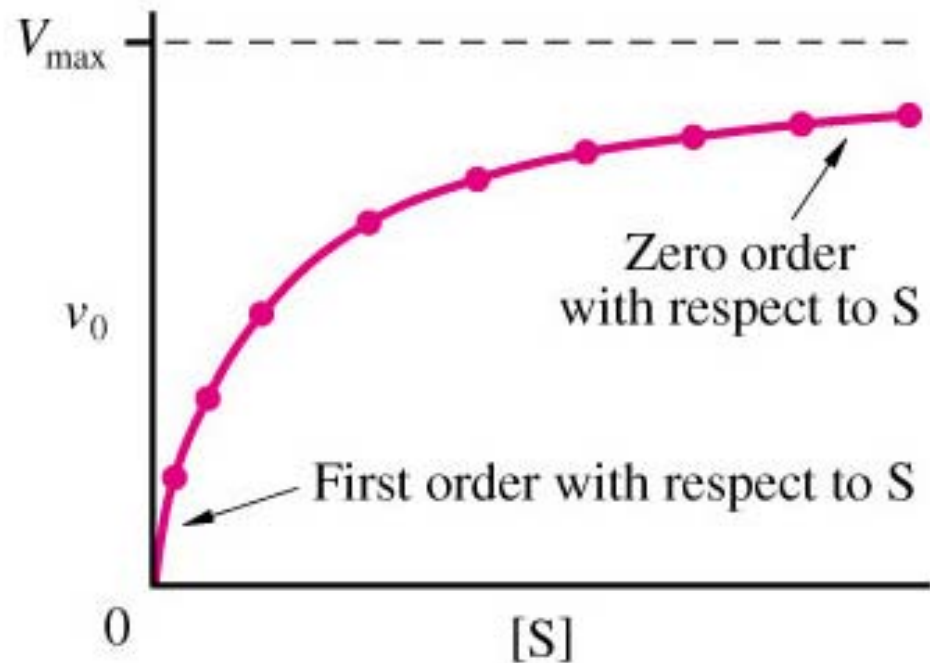


- $k_1$  is rate constant for formation of  $ES$
- $k_{-1}$  is rate constant for conversion of  $ES$  to  $E+S$
- $k_2$  is rate constant for product formation. For this reaction,  $k_2 = k_{cat}$
- Initial velocity assumption: measure activity before appreciable  $P$  accumulates:  $v_0 = k_2 [ES]$

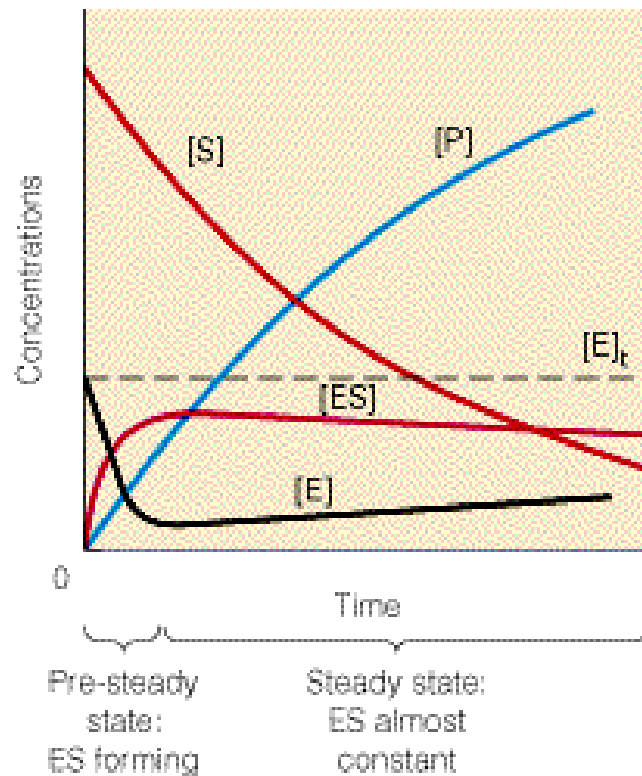
# Enzyme-catalyzed reactions exhibit saturation kinetics



At high [S], the enzyme is said to be **saturated** with respect to substrate



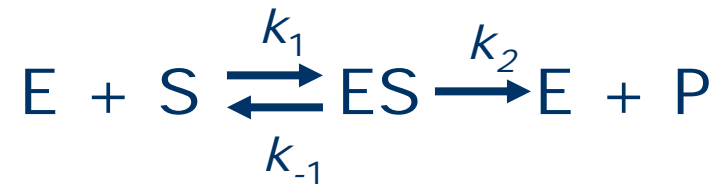
# Steady State



The more ES present, the faster ES will dissociate into  $E + P$  or  $E + S$ . Therefore, when the reaction is started by mixing enzymes and substrates, the  $[ES]$  builds up at first, but quickly reaches a **STEADY STATE**, in which  $[ES]$  remains constant. This steady state will persist until almost all of the substrate has been consumed.

# Michaelis-Menten equation

- If you assume that the formation of ES equals its breakdown, making [ES] constant (steady state), then:



$$k_1 [E][S] = k_{-1} [ES] + k_2 [ES]$$

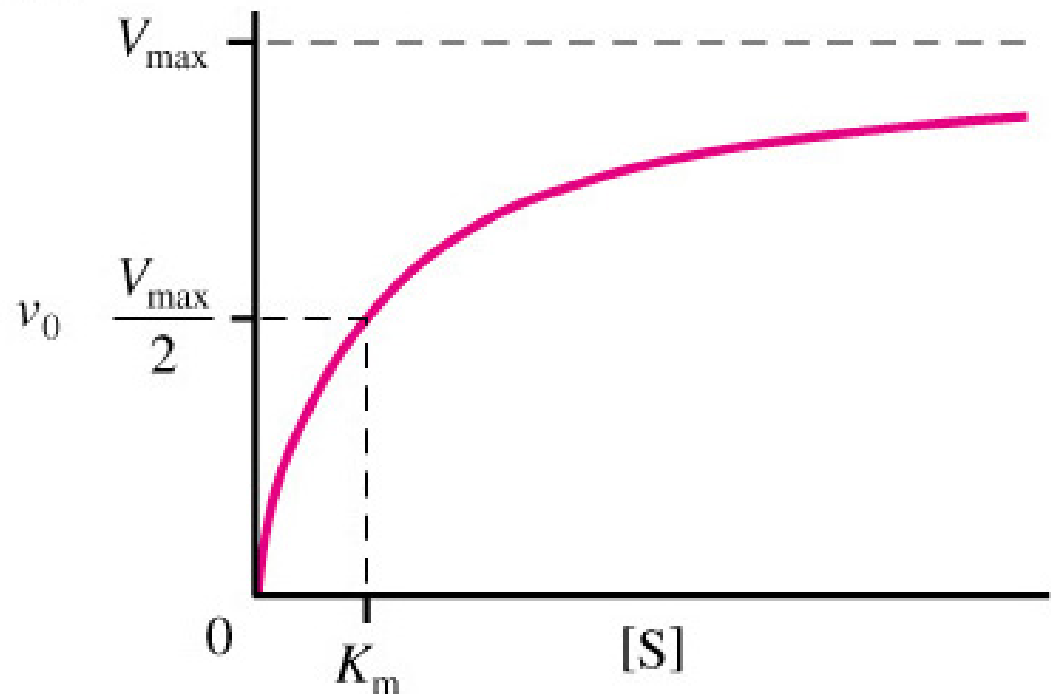
$$K_m = \frac{k_{-1} + k_2}{k_1}$$

$$v = \frac{V_{\max} \cdot [S]}{K_m + [S]}$$



## $K_m$

- $K_m$  is the  $[S]$  at  $1/2 V_{\max}$
- $K_m$  is a constant for a given enzyme
- $K_m$  is an estimate of the equilibrium constant for S binding to E
- Small  $K_m$  means tight binding; high  $K_m$  means weak binding



$K_M$  is a measure of  $[S]$  required for effective catalysis to occur

# Understanding $V_{\max}$

## The theoretical maximal velocity

- $V_{\max}$  is a constant for a given enzyme
- $V_{\max}$  is the theoretical maximal rate of the reaction - but it is NEVER achieved
- To reach  $V_{\max}$  would require that ALL enzyme molecules have tightly bound substrate

# The turnover number

## A measure of catalytic activity

- The  $k_{\text{cat}}$  is a direct measure of the catalytic production of product under saturating substrate conditions.
- $k_{\text{cat}}$ , the turnover number, is the maximum number of substrate molecules converted to product per enzyme molecule per unit of time.
- According to M-M model,  $k_{\text{cat}} = V_{\text{max}}/E_t$
- Values of  $k_{\text{cat}}$  range from less than 1/sec to many millions per sec

# The catalytic efficiency

- It shows what the enzyme can accomplish when abundant enzyme sites are available.
- It is the  $k_{\text{cat}}/K_{\text{M}}$  value that allows direct comparison of the effectiveness of an enzyme toward different substrates.

## Values of $k_{\text{cat}}$ (Turnover Number) for Some Enzymes

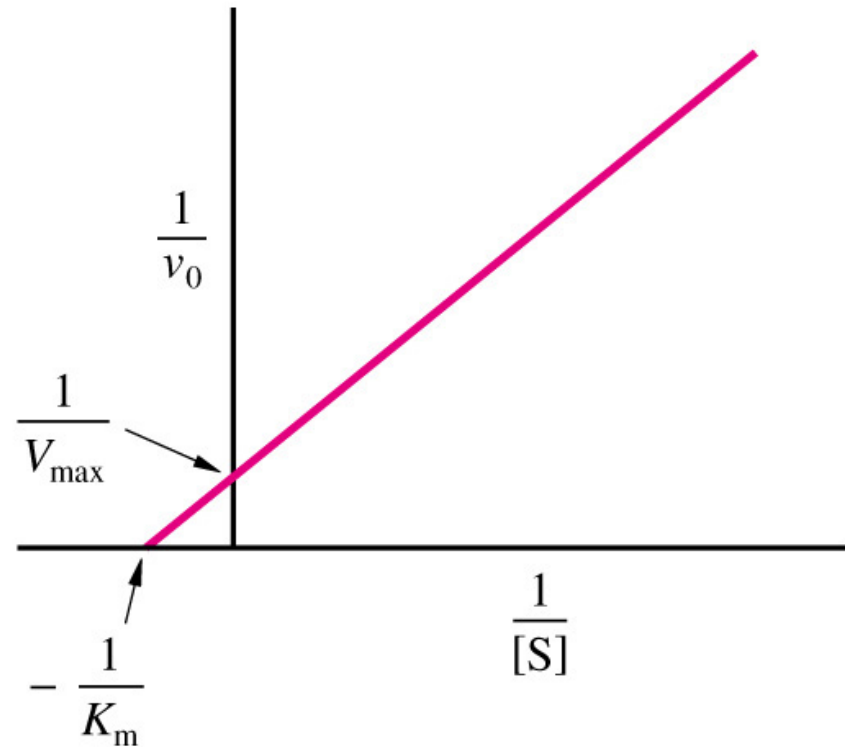
Enzyme	$k_{\text{cat}}$ ( $\text{sec}^{-1}$ )
Catalase	40,000,000
Carbonic anhydrase	1,000,000
Acetylcholinesterase	14,000
Penicillinase	2,000
Lactate dehydrogenase	1,000
Chymotrypsin	100
DNA polymerase I	15
Lysozyme	0.5

## Enzymes Whose $k_{\text{cat}}/K_m$ Approaches the Diffusion-Controlled Rate of Association with Substrate

Enzyme	Substrate	$k_{\text{cat}}$ ( $\text{sec}^{-1}$ )	$K_m$ ( $M$ )	$k_{\text{cat}}/K_m$ ( $\text{sec}^{-1} M^{-1}$ )
Acetylcholinesterase	Acetylcholine	$1.4 \times 10^4$	$9 \times 10^{-5}$	$1.6 \times 10^8$
Carbonic anhydrase	$\text{CO}_2$	$1 \times 10^6$	0.012	$8.3 \times 10^7$
	$\text{HCO}_3^-$	$4 \times 10^5$	0.026	$1.5 \times 10^7$
Catalase	$\text{H}_2\text{O}_2$	$4 \times 10^7$	1.1	$4 \times 10^7$
Crotonase	Crotonyl-CoA	$5.7 \times 10^3$	$2 \times 10^{-5}$	$2.8 \times 10^8$
Fumarase	Fumarate	800	$5 \times 10^{-6}$	$1.6 \times 10^8$
	Malate	900	$2.5 \times 10^{-5}$	$3.6 \times 10^7$
Triosephosphate isomerase	Glyceraldehyde- 3-phosphate*	$4.3 \times 10^3$	$1.8 \times 10^{-5}$	$2.4 \times 10^8$
$\beta$ -Lactamase	Benzylpenicillin	$2 \times 10^3$	$2 \times 10^{-5}$	$1 \times 10^8$

# Measuring $K_m$ and $V_{max}$

- Curve-fitting algorithms can be used to determine  $K_m$  and  $V_{max}$  from  $v$  vs.  $[S]$  plots
- Michaelis-Menton equation can be rearranged to the “double reciprocal” plot and  $K_m$  and  $V_{max}$  can be graphically determined

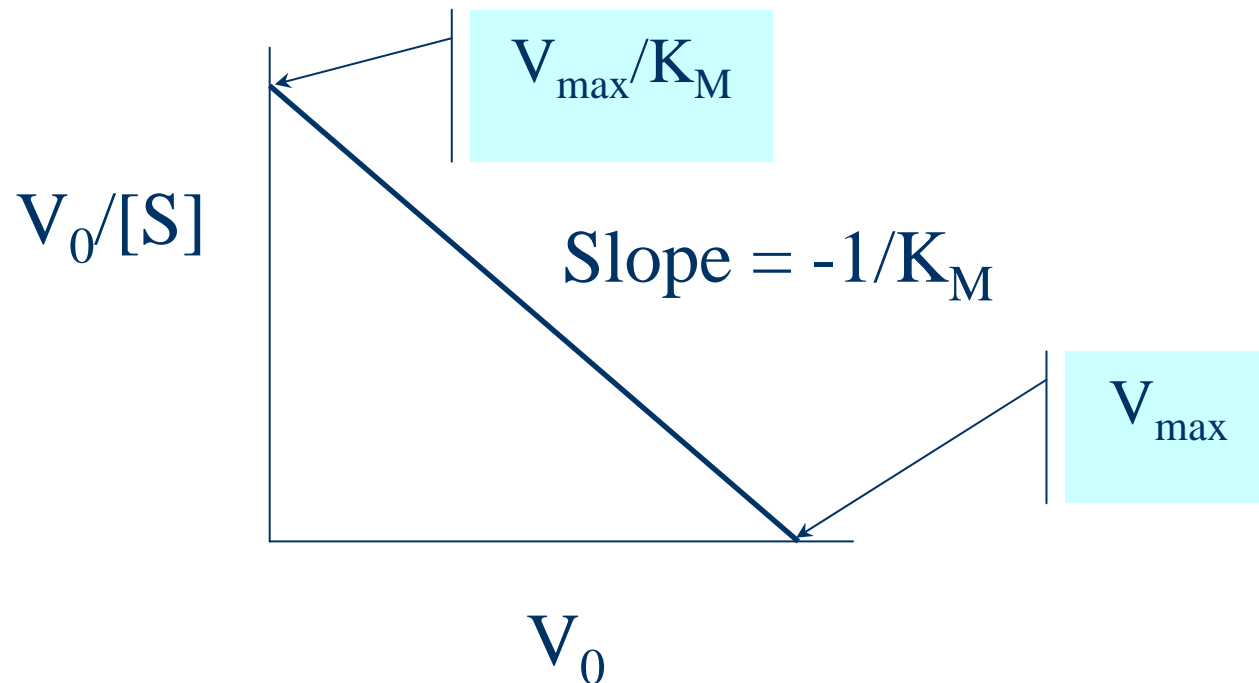


Lineweaver-Burk equation:

$$\frac{1}{v_0} = \left( \frac{K_m}{V_{max}} \right) \frac{1}{[S]} + \frac{1}{V_{max}}$$

# Eadie-Hofstee Plot

$$V_0/[S] = V_{\max}/K_M - V_0/K_M$$





# Enzyme Activity

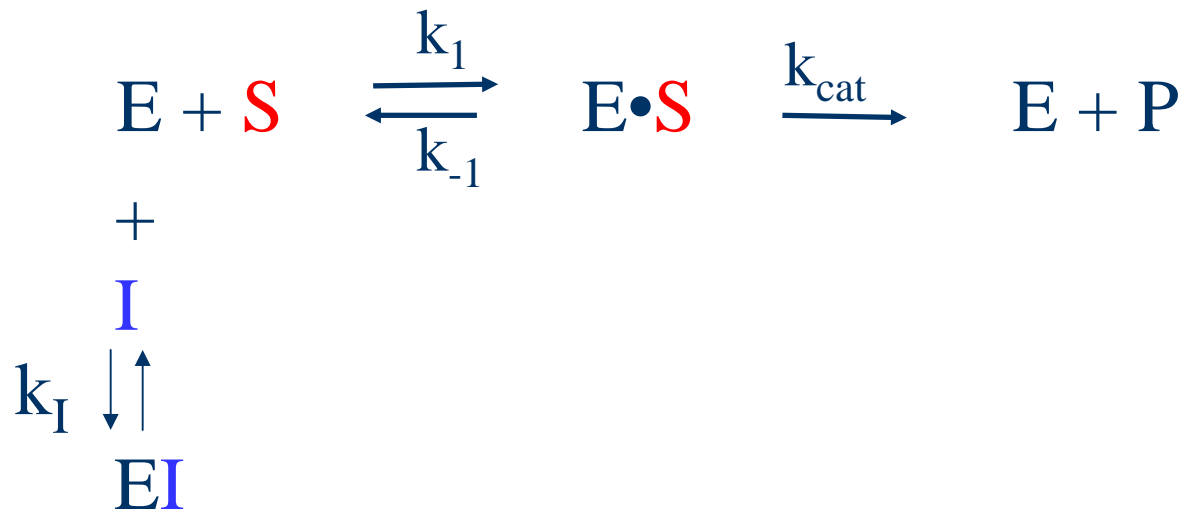
- Amount of reaction that a certain amount of enzyme will produce in a specified period of time
- Activity determined by measuring the amount of product formed or substrate that disappeared
- IU of enzyme activity is
  - The amount of enzyme necessary to produce 1  $\mu$ mole of product (or the loss of 1  $\mu$ mol of substrate) per minute under specified conditions of substrate concentration, pH and Temperature

# Enzyme Inhibition

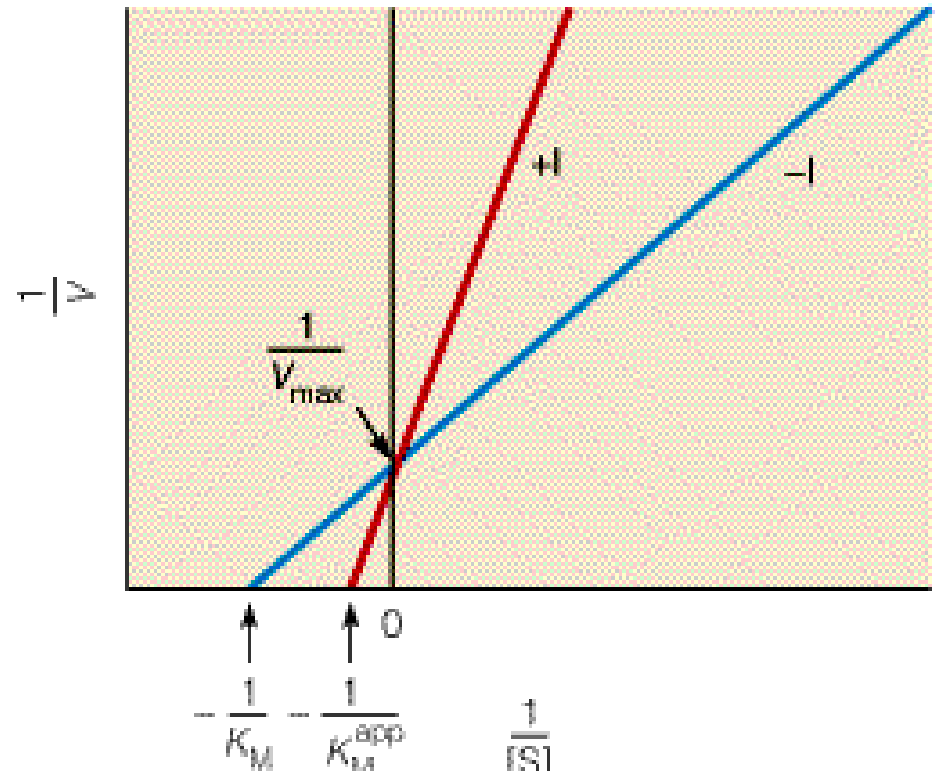
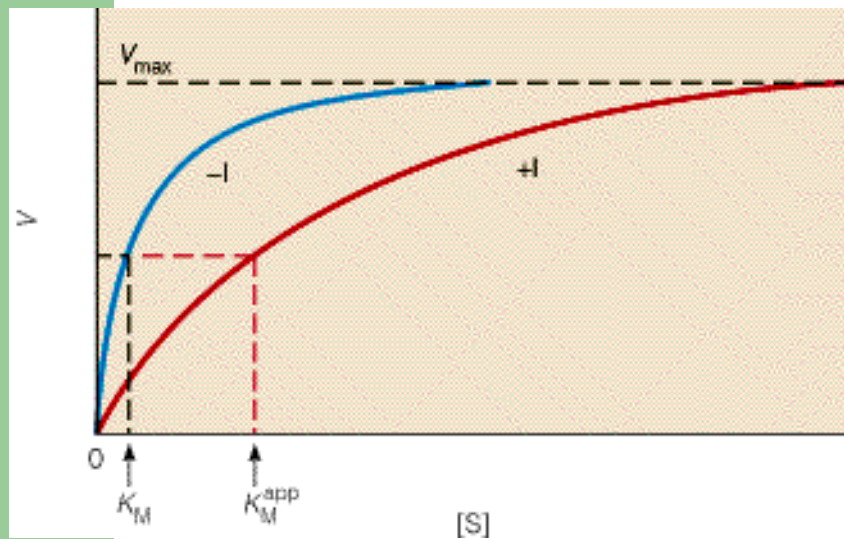
- Many different kinds of molecules inhibit enzymes and act in a variety of ways.
- One major distinction is whether the inhibition is
  - Reversible
    - Competitive with the substrate or
    - Non-competitive with the substrate
  - Irreversible (I is covalently bound, incapacitating the enzyme)

# Reversible Inhibition: Competitive

- Inhibitor resembles substrate but can't undergo the catalytic step, so it wastes the enzyme's time by preventing S binding. i.e. Inhibitor COMPETES with substrate for binding.

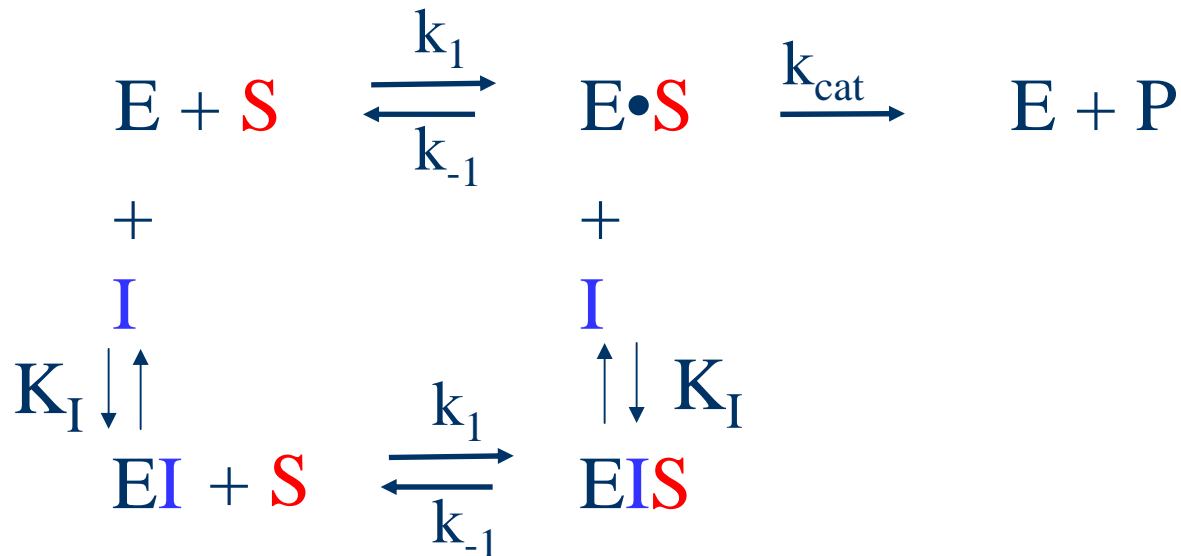


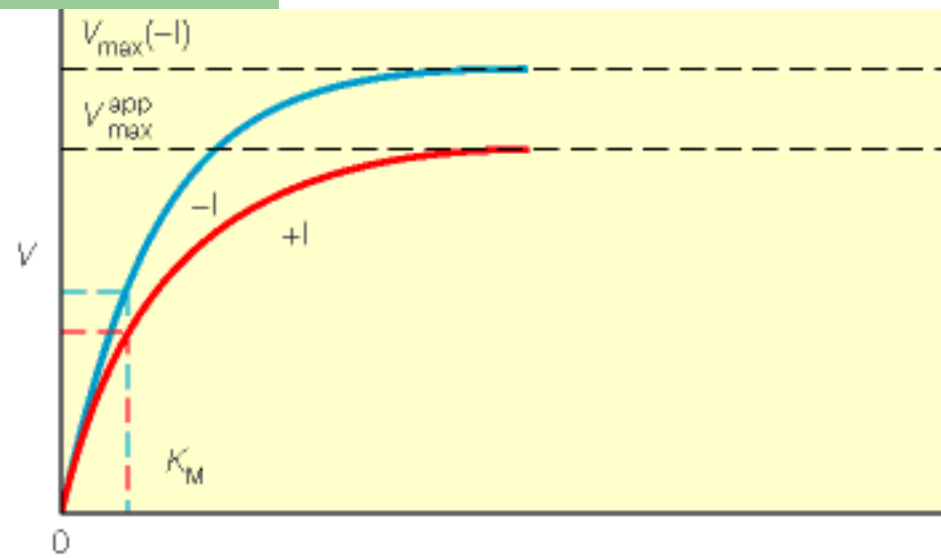
# Reversible Inhibition: Competitive



# Reversible Inhibition: Non-Competitive

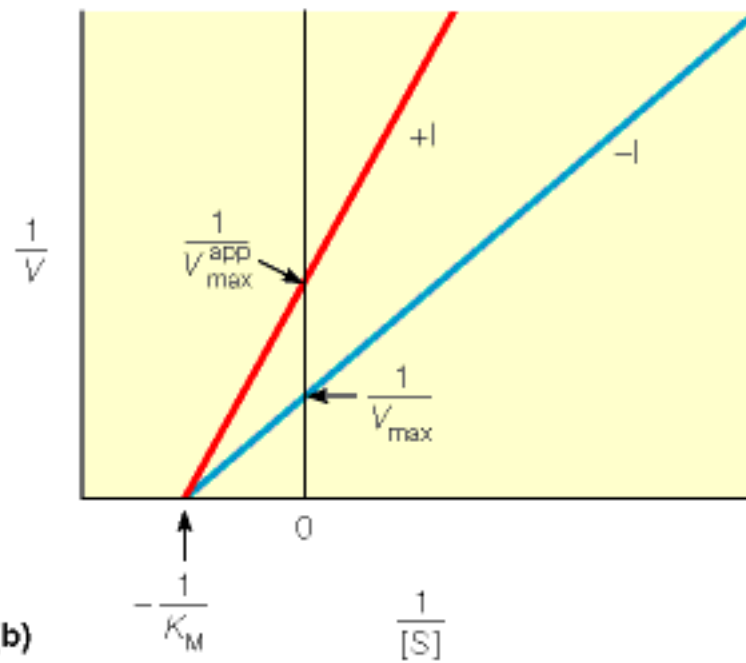
- A molecule or ion binds at a remote site on the enzyme in such a way that it affects  $k_{\text{cat}}$ .



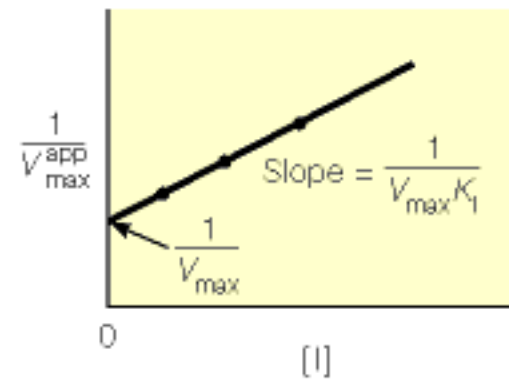


(a)

$[S]$



(b)



(c)

# Enzyme Inhibition

Inhibitor Type	Binding Site on Enzyme	Kinetic effect
Competitive Inhibitor	Specifically at the catalytic site, where it competes with substrate for binding in a dynamic equilibrium- like process. Inhibition is reversible by substrate.	$V_{\max}$ is unchanged; $K_m$ is increased.
Noncompetitive Inhibitor	Binds E or ES complex other than at the catalytic site. Substrate binding unaltered, but ESI complex cannot form products. Inhibition cannot be reversed by substrate.	$K_m$ appears unaltered; $V_{\max}$ is decreased proportionately to inhibitor concentration.
Uncompetitive Inhibitor	Binds only to ES complexes at locations other than the catalytic site. Substrate binding modifies enzyme structure, making inhibitor- binding site available. Inhibition cannot be reversed by substrate.	Apparent $V_{\max}$ decreased; $K_m$ is decreased.