
COMPUTATIONAL BIOLOGY

Under Supervisor:

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INTRODUCTION :

Advances in instrumentation and data acquisition modes, such as the recently developed real-time database search (RTS), can reduce interference. However, no diagnostic tool is available currently to rapidly and accurately assess ion interference. To address this need, we developed a multiplexed TMT-based standard, termed the triple knockout (TKO). This standard is comprised of three yeast proteomes in triplicate, each from a strain deficient in a highly abundant protein (Met6, Pfk2, or Ura2). Specifically, we expand the TKO standard to incorporate the TMTpro18-plex reagents (TKO18). We also construct a variant thereof which has been digested only with LysC (TKO18L). We compare proteome coverage and interference levels of TKO18 and TKO18L data that were acquired under different data acquisition modes and analyzed using TVT2.5. Our data illustrate that RTS reduces interference while improving proteome coverage and suggest that digesting with LysC alone only modestly reduces interference, albeit at the expense of proteome depth.

FASTA SEQUENCE:

>sp|P39723|SPC72_YEAST Spindle pole component SPC72 OS=Saccharomyces cerevisiae (strain ATCC 204508 / S288c) OX=559292 GN=SPC72 PE=1 SV=2

MVRRWIPSGRHLRNNDNTGDDDDSEFTNSMDSGMSIPSLRDSMTTRSSHNDPIKPALMND
SNKVKNLEKELTNAKIKIQVLYEYIRRI PNKDGNAPSLGNDTDFRNSIIEGLNLEINKLK
QDLKAKEVEYQDTLQFVQENLENSESIVNTINHLLSFILTHFNEQDENAHLLDKEERETL
EETLELSSDYVLEKMDTLSKFIIQFLQDFLHKSRAESKQDKEEFLSLAQSSPAGSQLES
RDSPSSKEENTDGGYQNDEIHDSNNHIDTENVMANSTSLPISAVESRFEKTLDTQLEIVI
ENLHKEYDQFINSIRLKFEKSQKLEKIIASKLNEQSHLLDSLELEENSSSVIEKQDHLIS
QLKEKIESQSVLINNLEKLKEDIKMKQNEKVLTKETQTKINKLKENNWD SYINDLEK
QINDLQIDKSEEFHVIQNQLDKLDLENYQLKNQLNTLDNQKLILSQYESNFIKFNQNLLL
HLDSIFNILQKILQESSIAQFDRKMKSISV PNALKNLNLIQPKLESLYTFIETALESII
NSYISSLISMETPEQPHQQGNELTATPNKELTLRIEELQRRWISERERRKLDANASEARI
KALEQENESLRSKLFNLSINNP

```
from pyopenms import *  
import matplotlib.pyplot as plt
```

```
def mirror_plot(obs_mz, obs_int, theo_mz, theo_int, title):  
  
    obs_int = [element / max(obs_int) for element in obs_int]  
    theo_int = [element * -1 for element in theo_int]  
    plt.figure(figsize=(12, 8))  
    plt.bar(obs_mz, obs_int, width=3.0)  
    plt.bar(theo_mz, theo_int, width=3.0)  
    plt.title(title)  
    plt.ylabel('intensity')  
    plt.xlabel('m/z')  
    plt.show()
```

```
protein_ids = []  
peptide_ids = []  
SimpleSearchEngineAlgorithm().search("ea07029.mzML", "yeast.fasta", protein_ids, peptide_ids)
```

```
print(len(peptide_ids), len(protein_ids))
```

2 1


```

for peptide_id in peptide_ids:
    print (35*"=")
    print ("Peptide Index m/z:", peptide_id.getMetaValue("scan_index"))
    for hit in peptide_id.getHits():
        index=str(peptide_id.getMetaValue("scan_index"))
        seq=str( hit.getSequence())
        print(" - Peptide hit sequence:", hit.getSequence())
        tsg = TheoreticalSpectrumGenerator()
        theo_spec = MSSpectrum()
        p = Param()
        p.setValue("add_y_ions", "true")
        p.setValue("add_b_ions", "true")
        p.setValue("add_metainfo", "true")
        tsg.setParameters(p)
        peptide = AASequence.fromString(hit.getSequence().toString())
        tsg.getSpectrum(theo_spec, peptide, 1, 2)

    print("Spectrum 1 of", peptide, "has", theo_spec.size(), "peaks.")
    for ion, peak in zip(theo_spec.getStringDataArrays()[0], theo_spec):
        print(ion.decode(), "is generated at m/z", peak.getMZ())
    exp = MSExperiment()
    MzMLFile().load("ea07029.mzML", exp)
    spectra = exp.getSpectrum(peptide_id.getMetaValue("scan_index"))
    alignment = []
    spa = SpectrumAlignment()
    p = spa.getParameters()

    p.setValue("tolerance", 0.5)
    p.setValue("is_relative_tolerance", "false")
    spa.setParameters(p)

    spa.getSpectrumAlignment(alignment, theo_spec, spectra)

```

```
if(len(alignment)!=0):
    print("Number of matched peaks: " + str(len(alignment)))
    print("ion\ttheo. m/z\tobserved m/z")

    for theo_idx, obs_idx in alignment:
        ion_name = theo_spec.getStringDataArrays()[0][theo_idx].decode()
        ion_charge = theo_spec.getIntegerDataArrays()[0][theo_idx]
        print(ion_name + "\t" + str(ion_charge) + "\t"
              + str(theo_spec[theo_idx].getMZ())
              + "\t" + str(spectra[obs_idx].getMZ()))

    theo_mz, theo_int, obs_mz, obs_int = [], [], [], []
    for theo_idx, obs_idx in alignment:
        theo_mz.append(theo_spec[theo_idx].getMZ())
        theo_int.append(theo_spec[theo_idx].getIntensity())
        obs_mz.append(spectra[obs_idx].getMZ())
        obs_int.append(spectra[obs_idx].getIntensity())
    if(max(obs_int)!=0):
        title ="The Seq :"+ seq+" & The index:"+index
        mirror_plot(obs_mz, obs_int, theo_mz, theo_int, title)
```

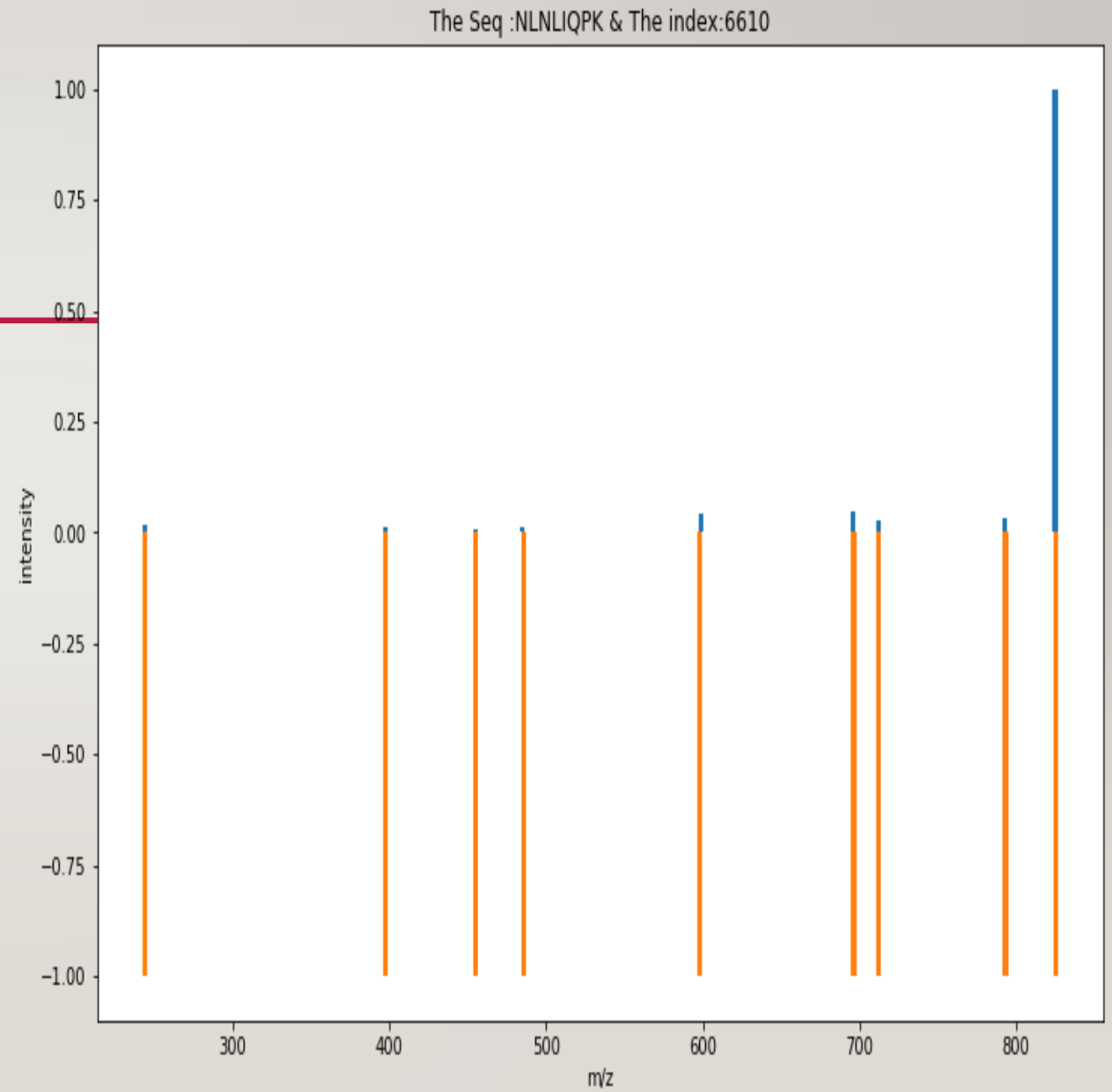
FIRST FILE MZ

Protein

EXP 1:

Peptide

NLNLIQPK



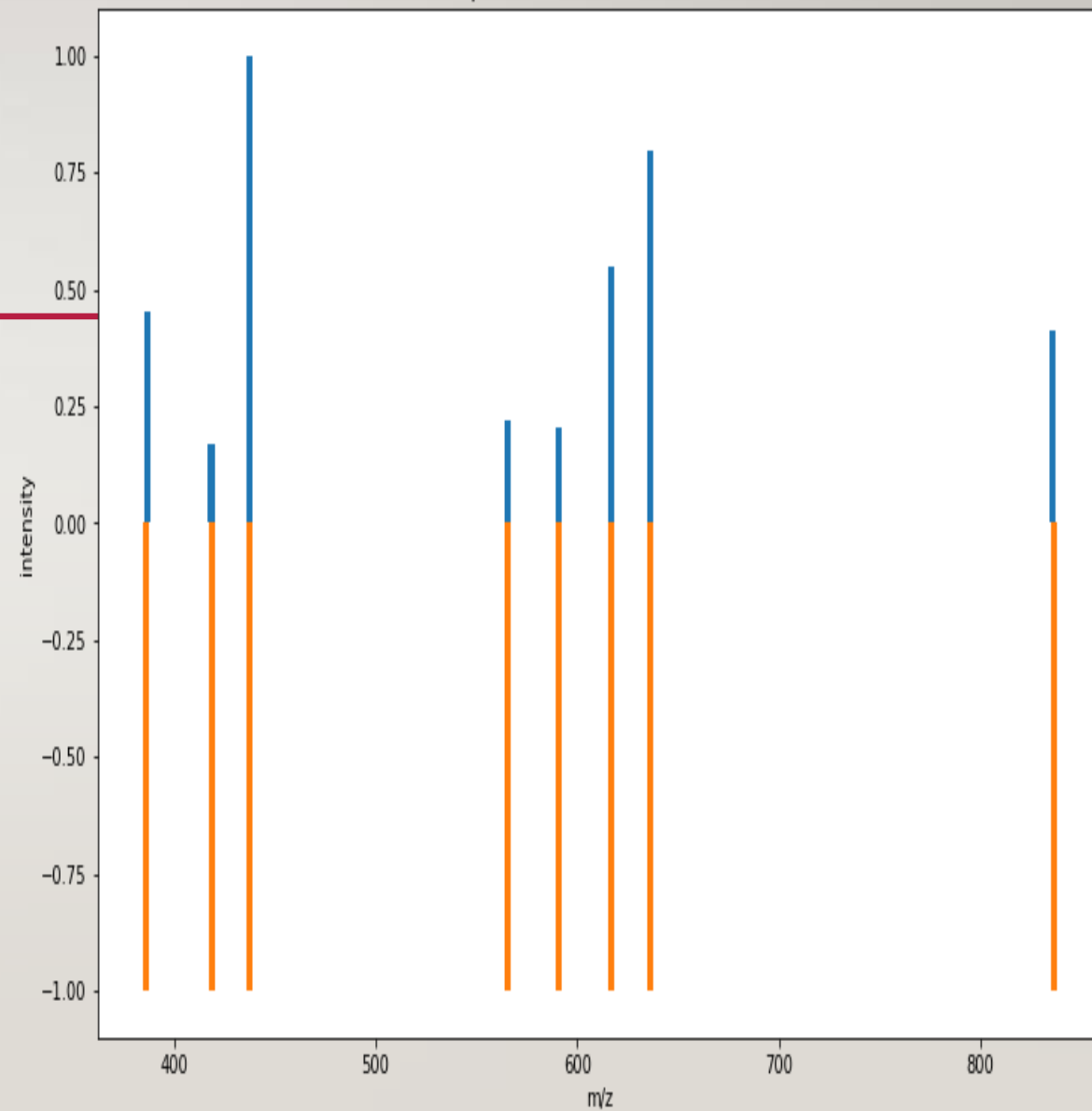
PROTEIN

Exp 2

Peptide

ILQESSIAQFDR

The Seq :ILQESSIAQFDR & The index:10688



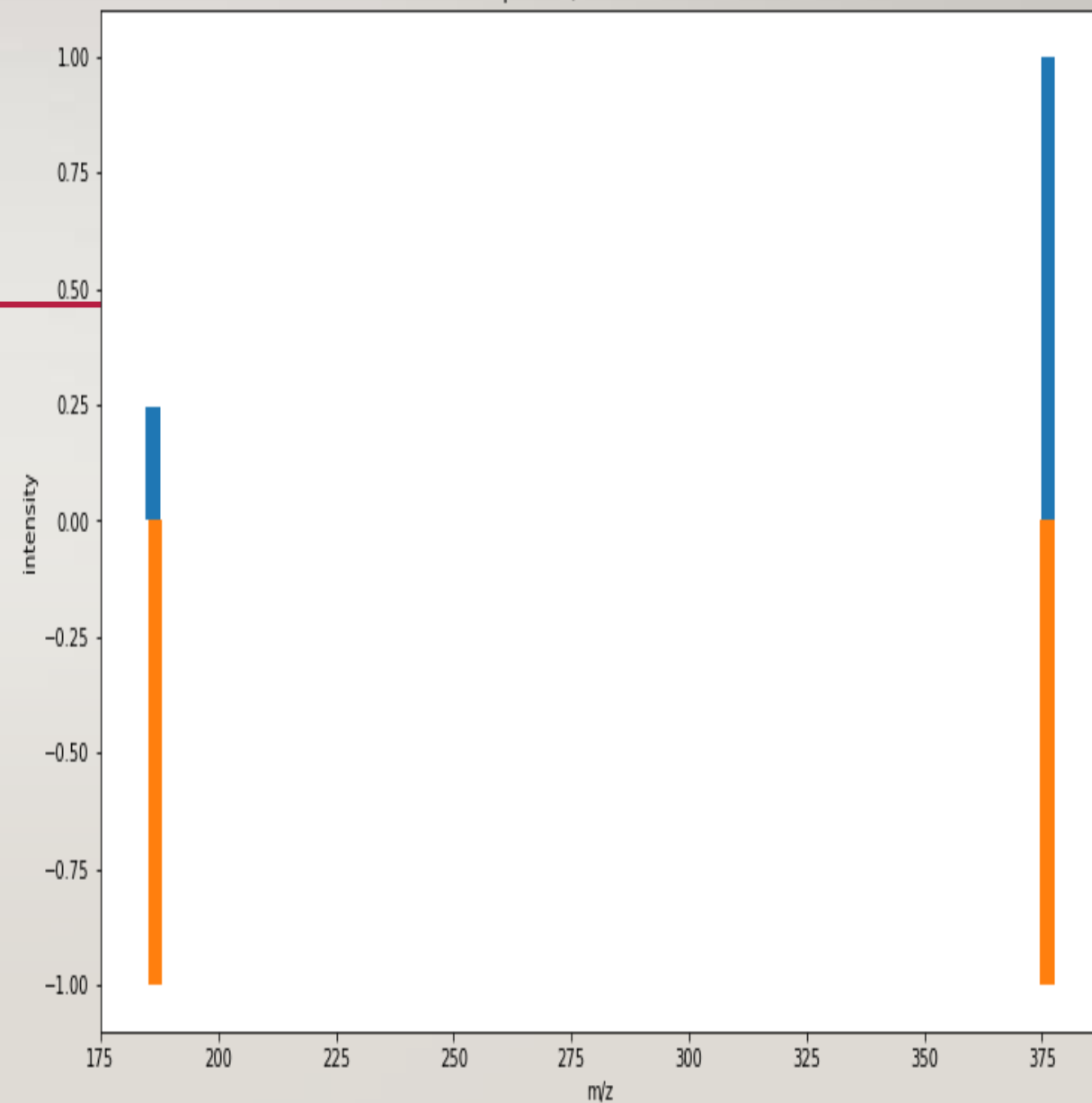
SECOND FILE MZ

Protein

Exp 1

Peptide

ELETQTK



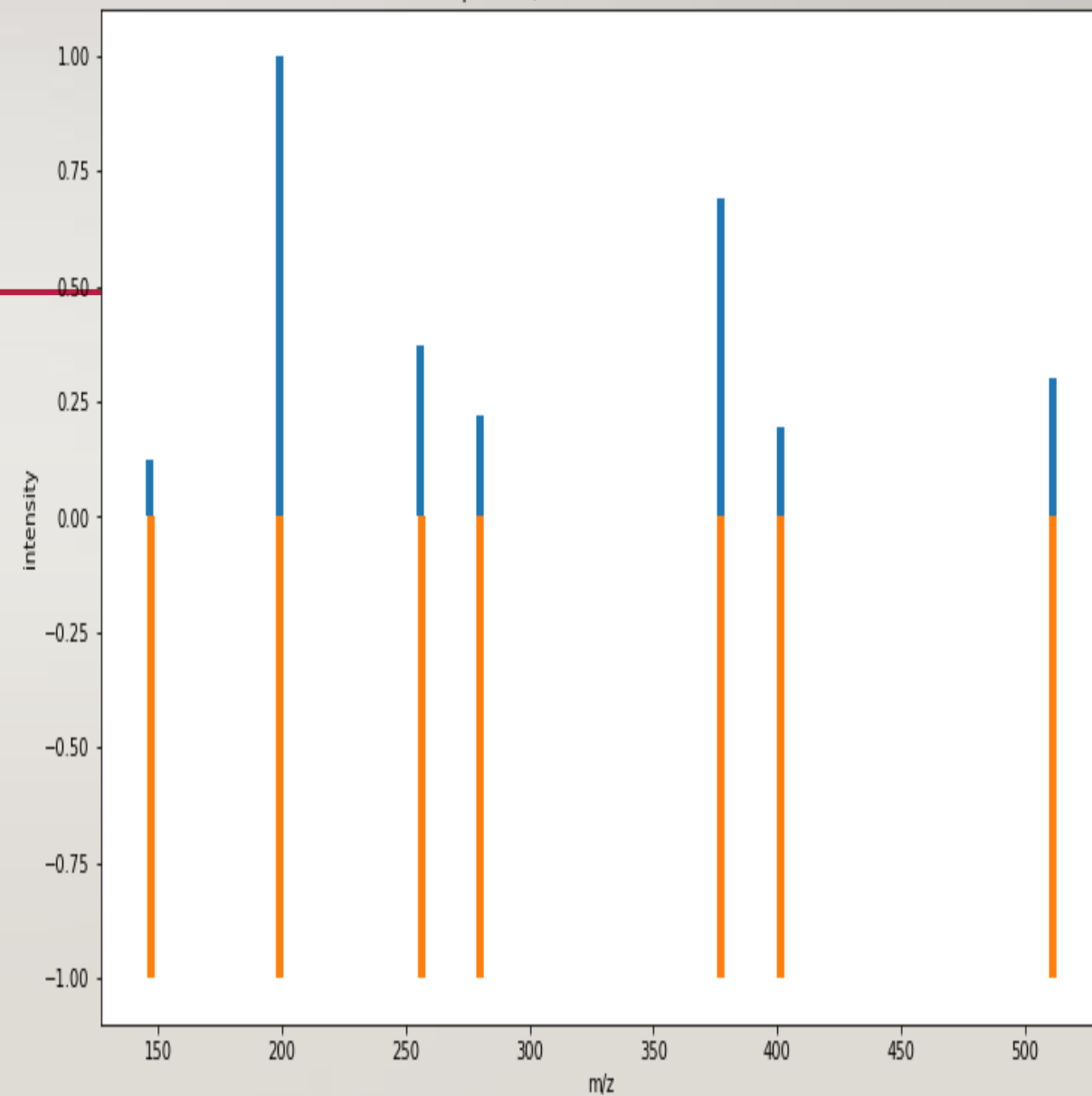
PROTEIN

Exp 2

Peptide

TLDTQLEIVIENLHK

The Seq :TLDTQLEIVIENLHK & The index:8288



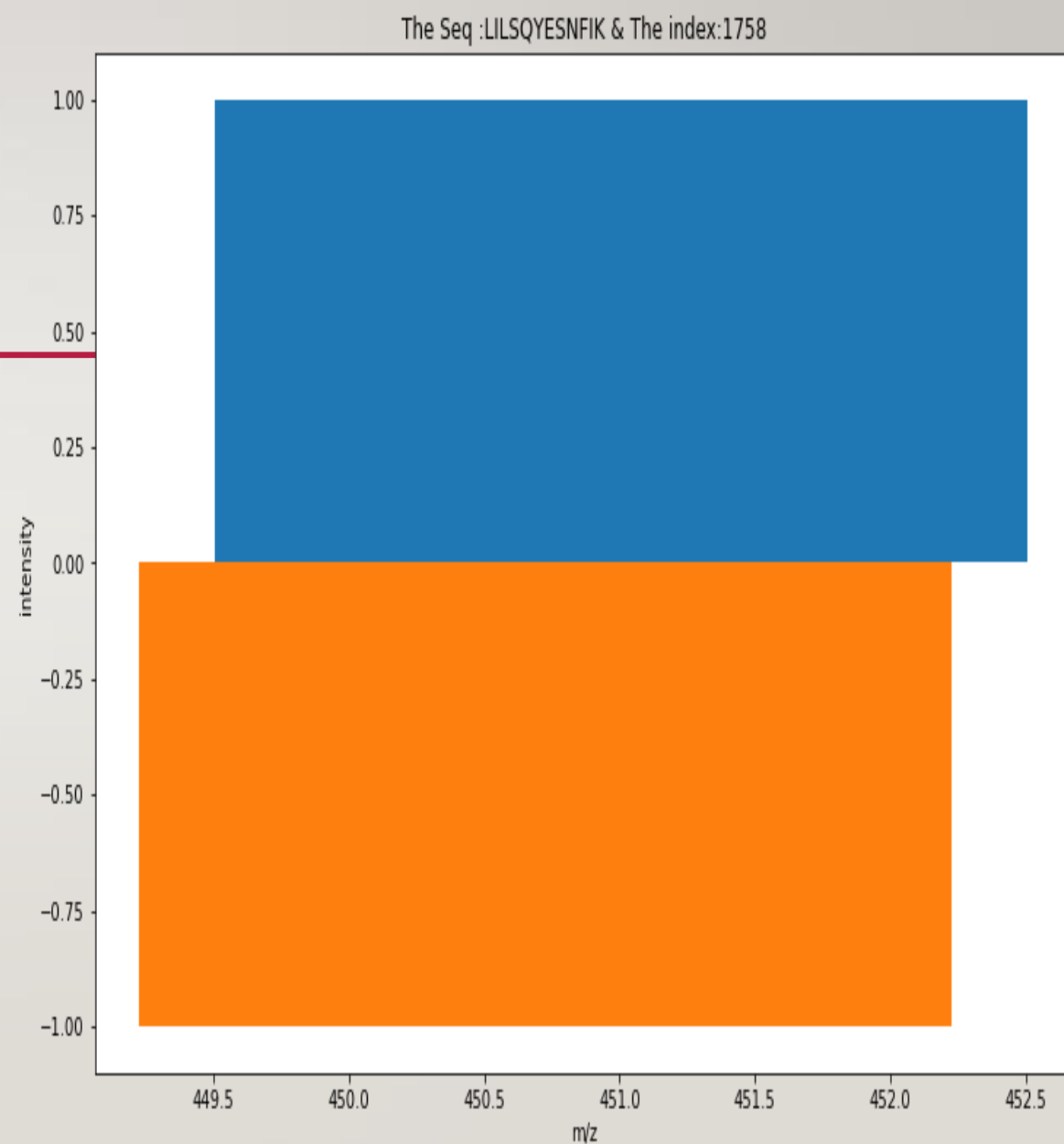
THIRD FILE MZ

Protein

Exp 1

Peptide

LILSQYESNFIK

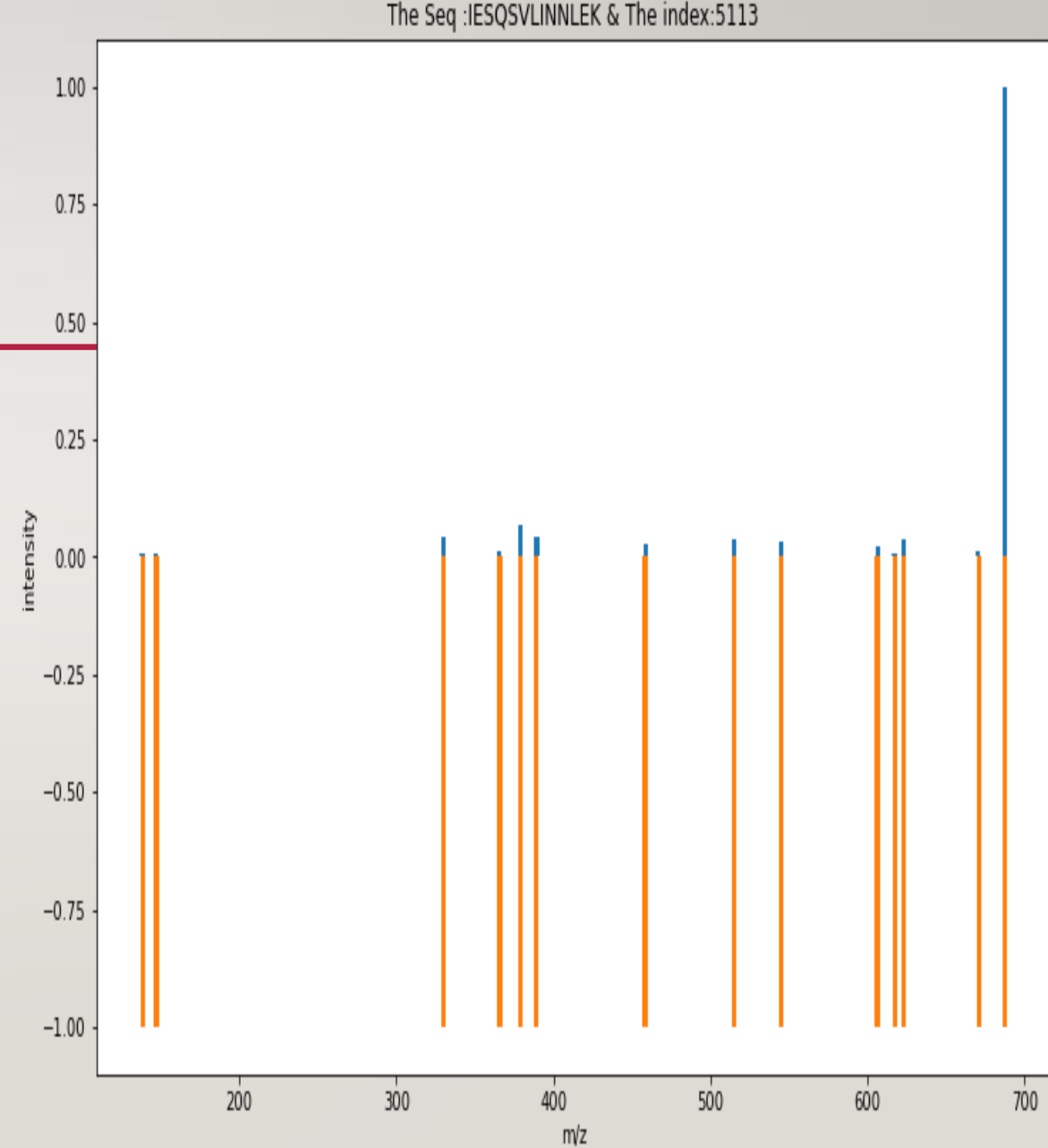


PROTEIN

Exp 2

Peptide

IESQSVLINNLEK



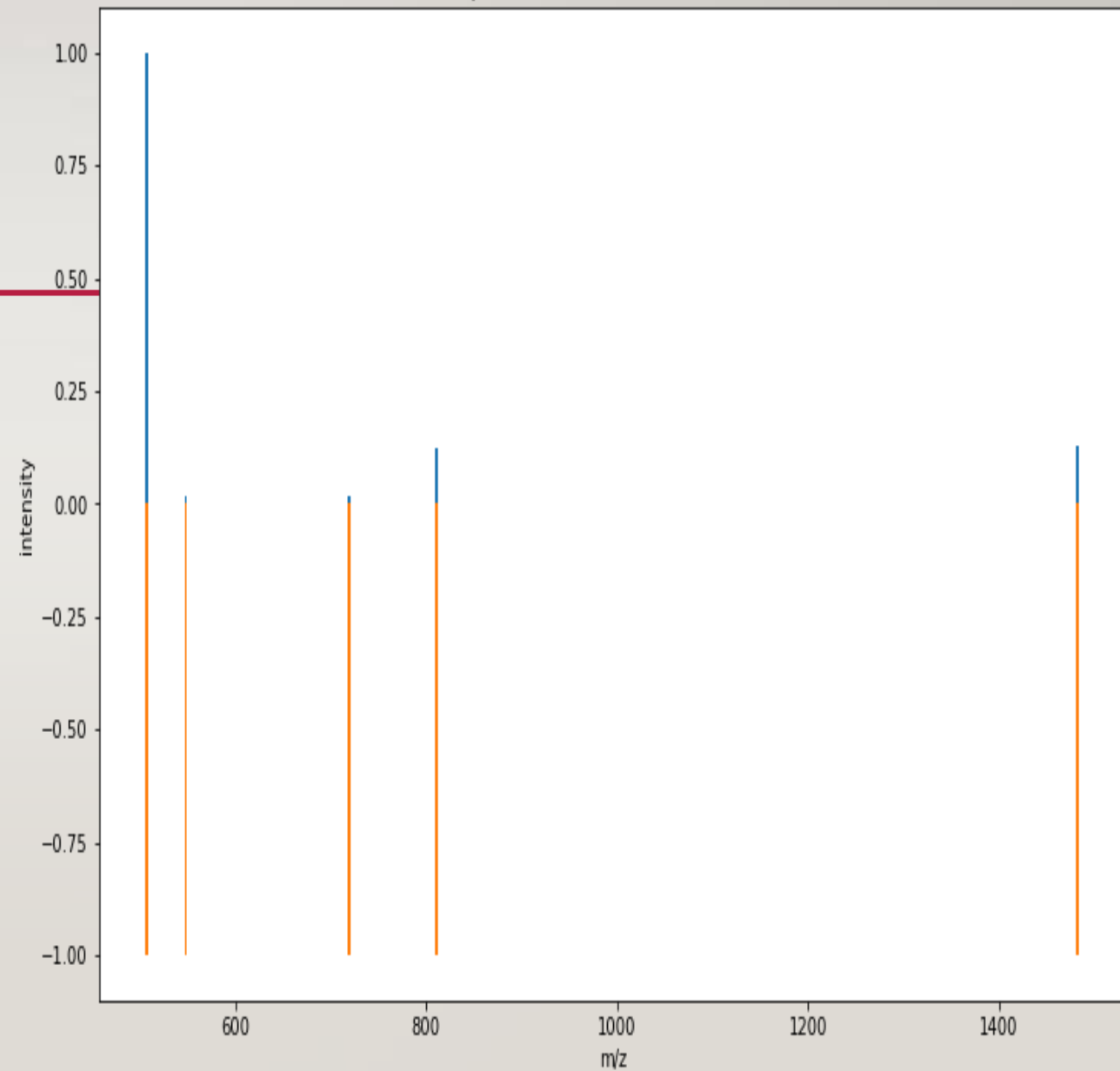
PROTEIN

Exp 3

Peptide

TLDTQLEIVIENLHK

The Seq :TLDTQLEIVIENLHK & The index:5905



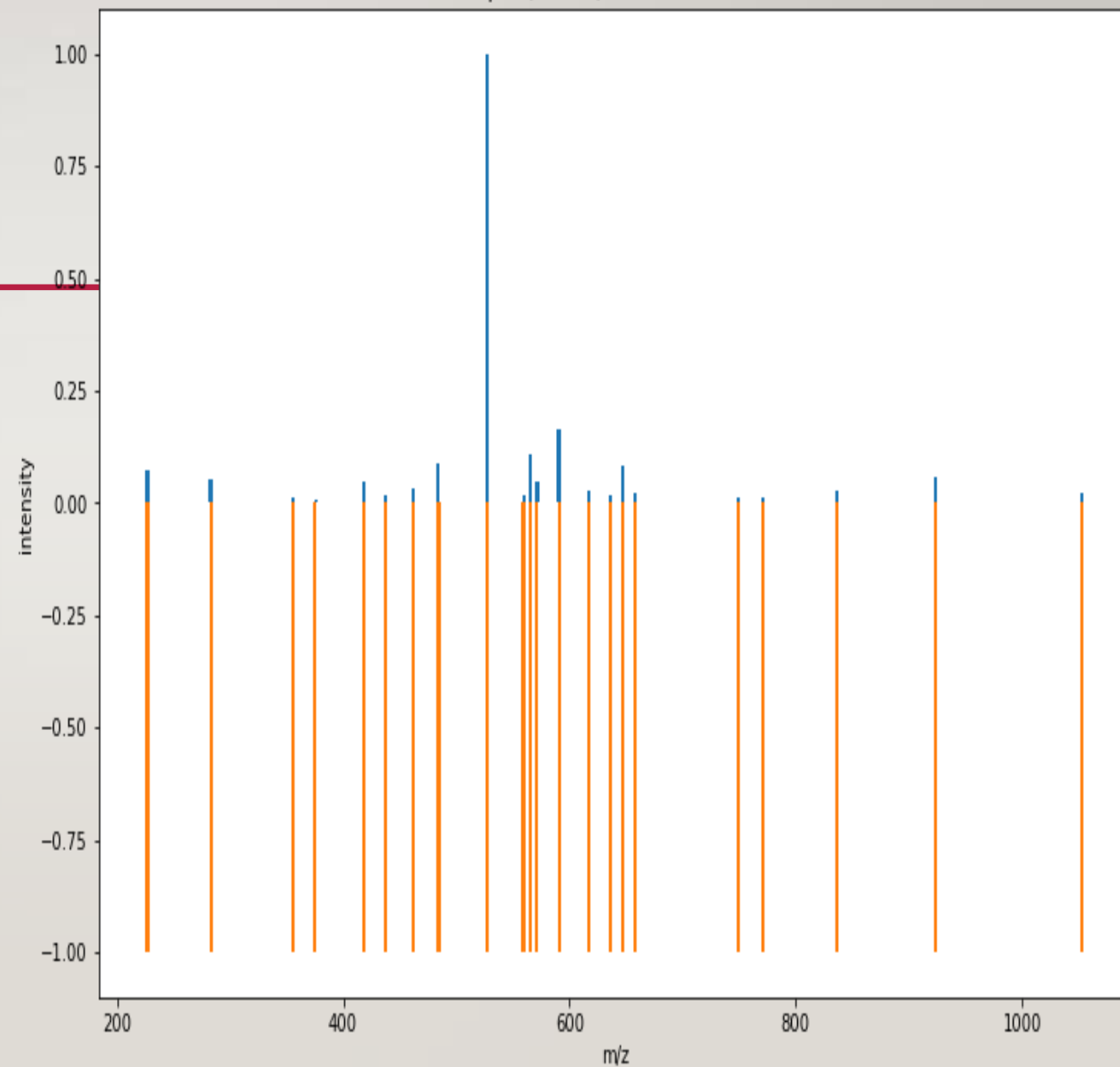
PROTEIN

Exp 4

Peptide

ILQESSIAQFDR

The Seq :ILQESSIAQFDR & The index:7230



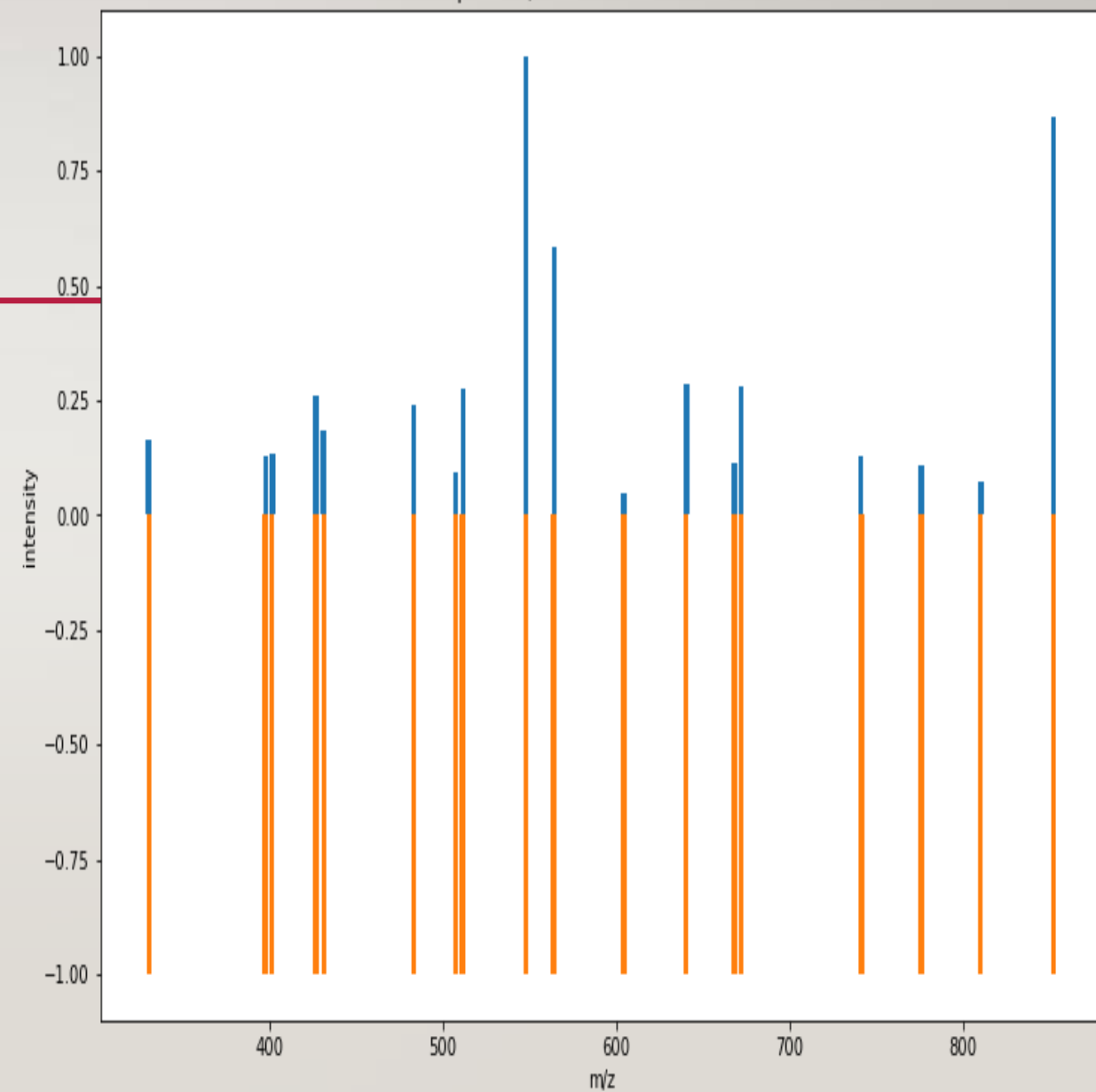
PROTEIN

Exp 5

Peptide

TLDTQRLEIVIENLHK

The Seq :TLDTQLEIVIENLHK & The index:16656



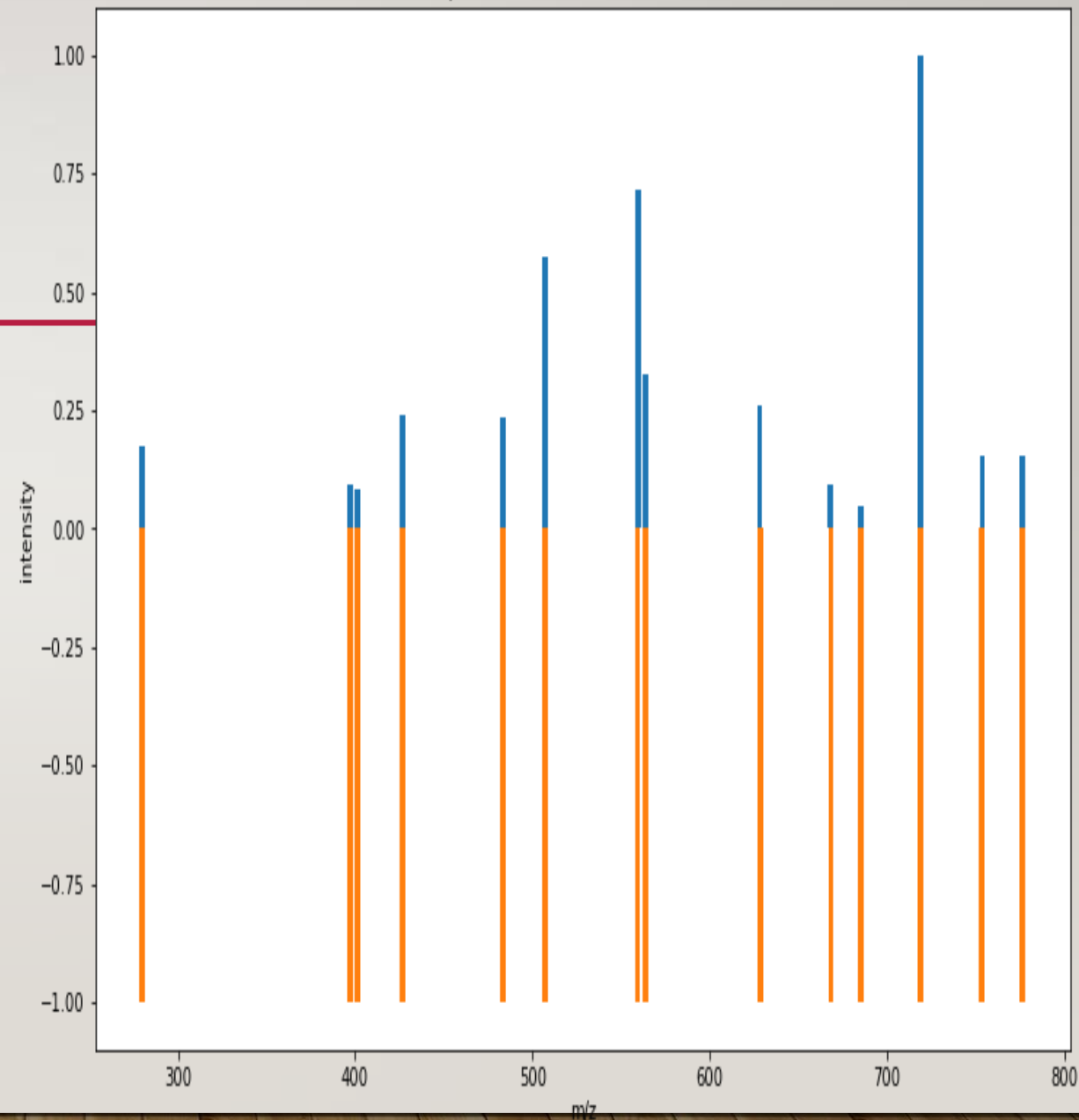
FOURTH FILE MZ

Protein

Exp 1

Peptide

TLDTQRLEIVLHK



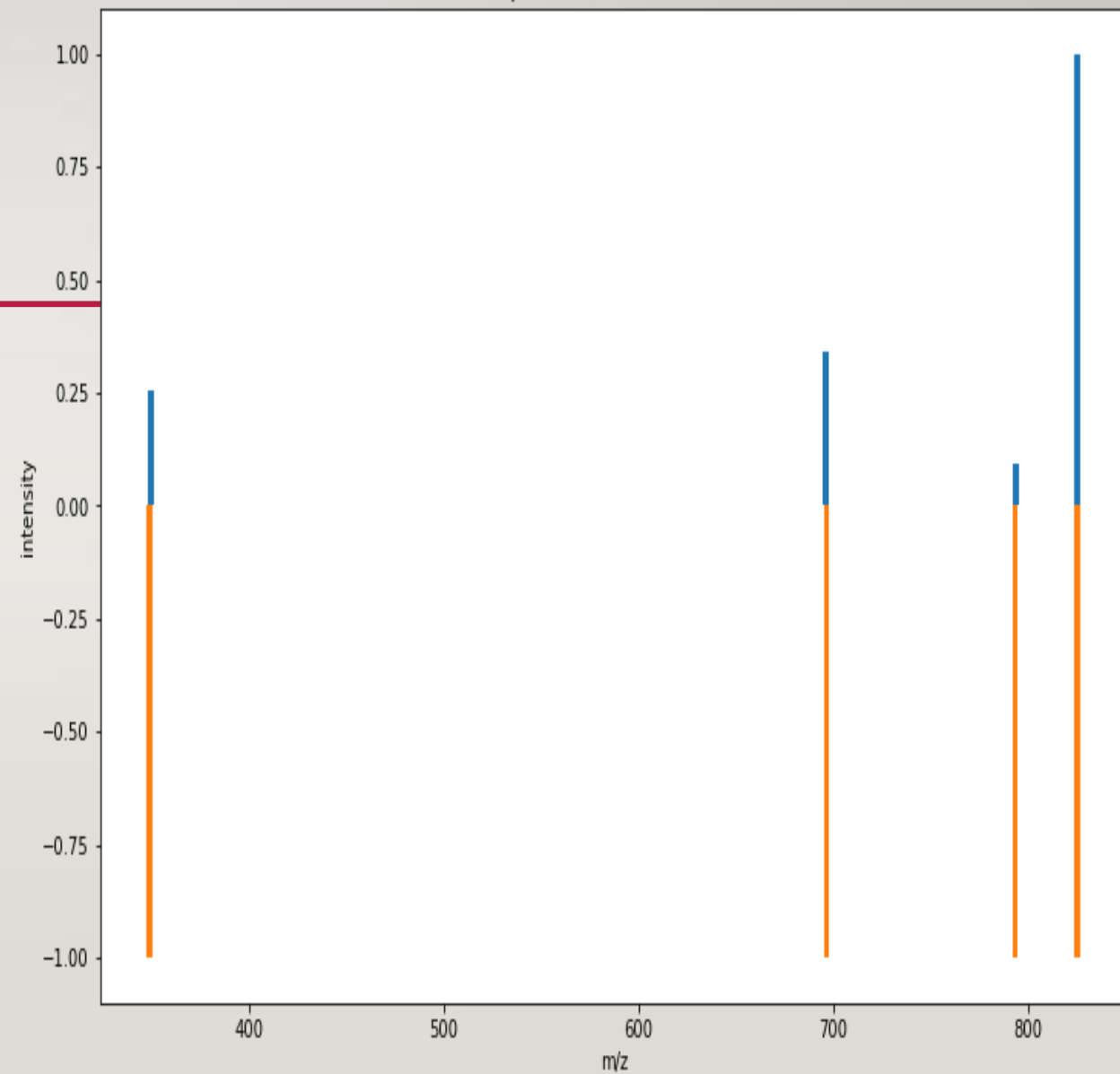
FIFTH FILE MZ

Protein

Exp 1

Peptide

NLNLIQPK



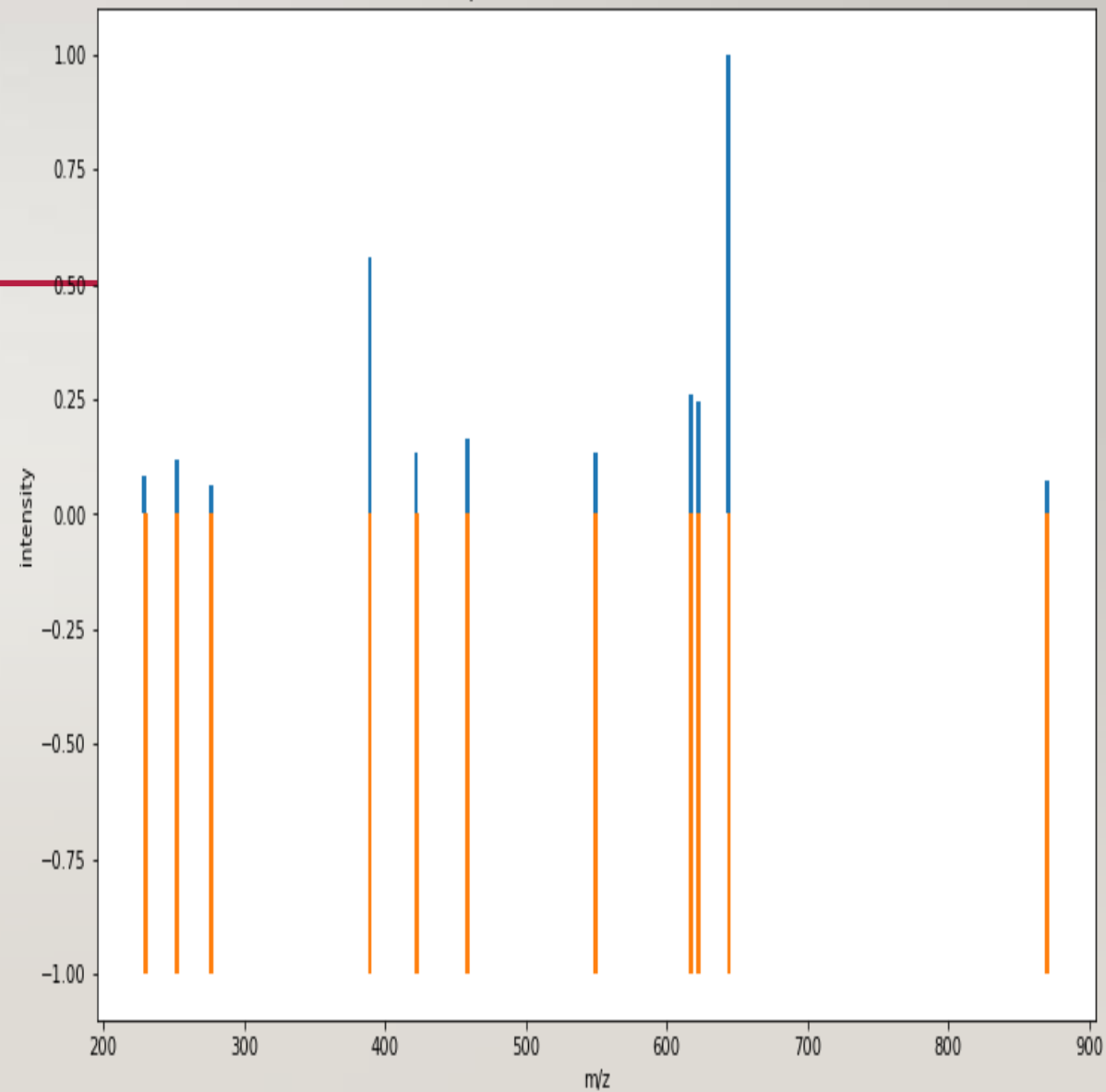
PROTEIN

Exp 2

Peptide

IESQSVLINNLEK

The Seq :IESQSVLINNLEK & The index:4963



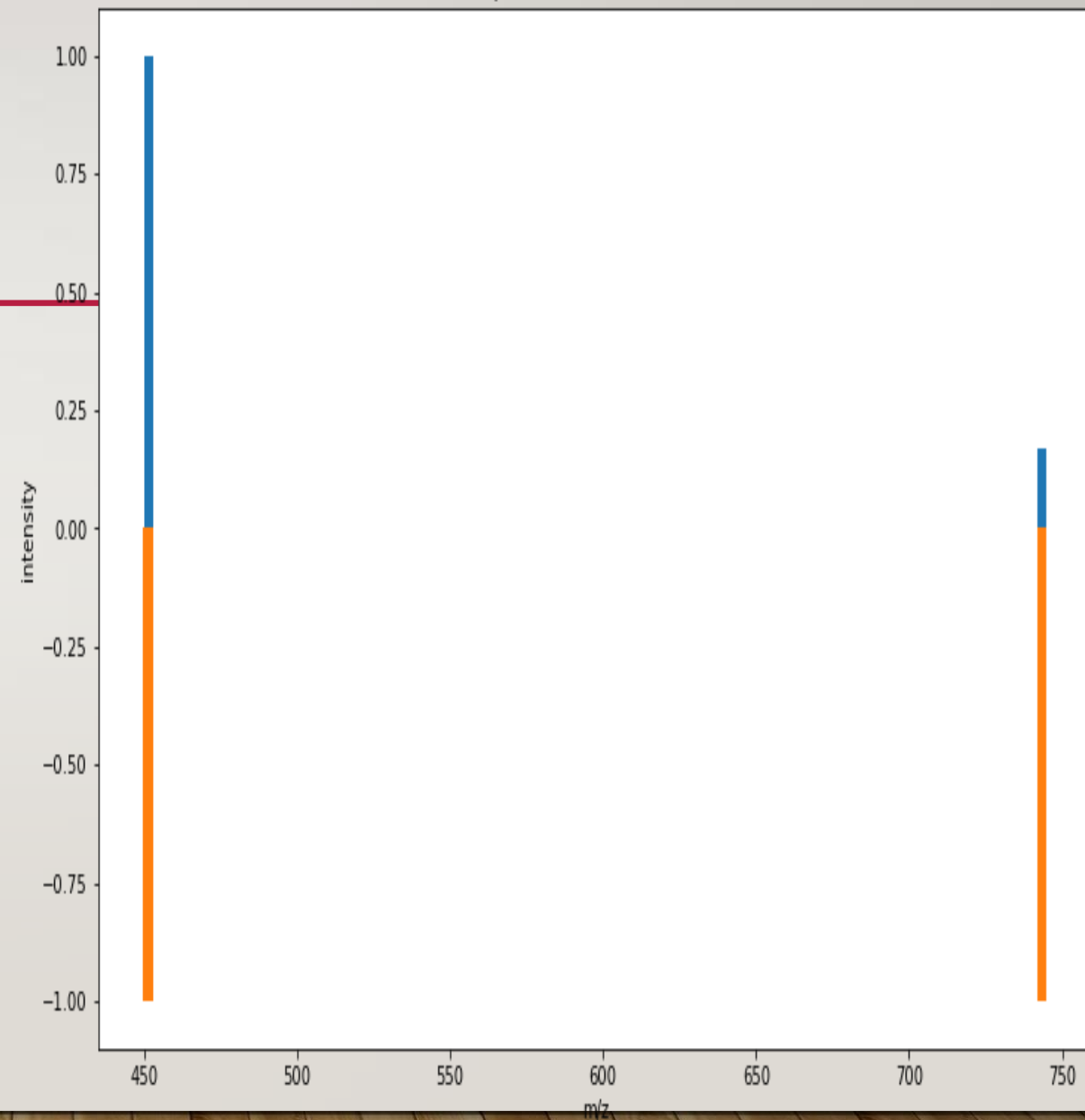
PROTEIN

Exp 3

Peptide

IQVLYEYIR

The Seq :IQVLYEYIR & The index:9567



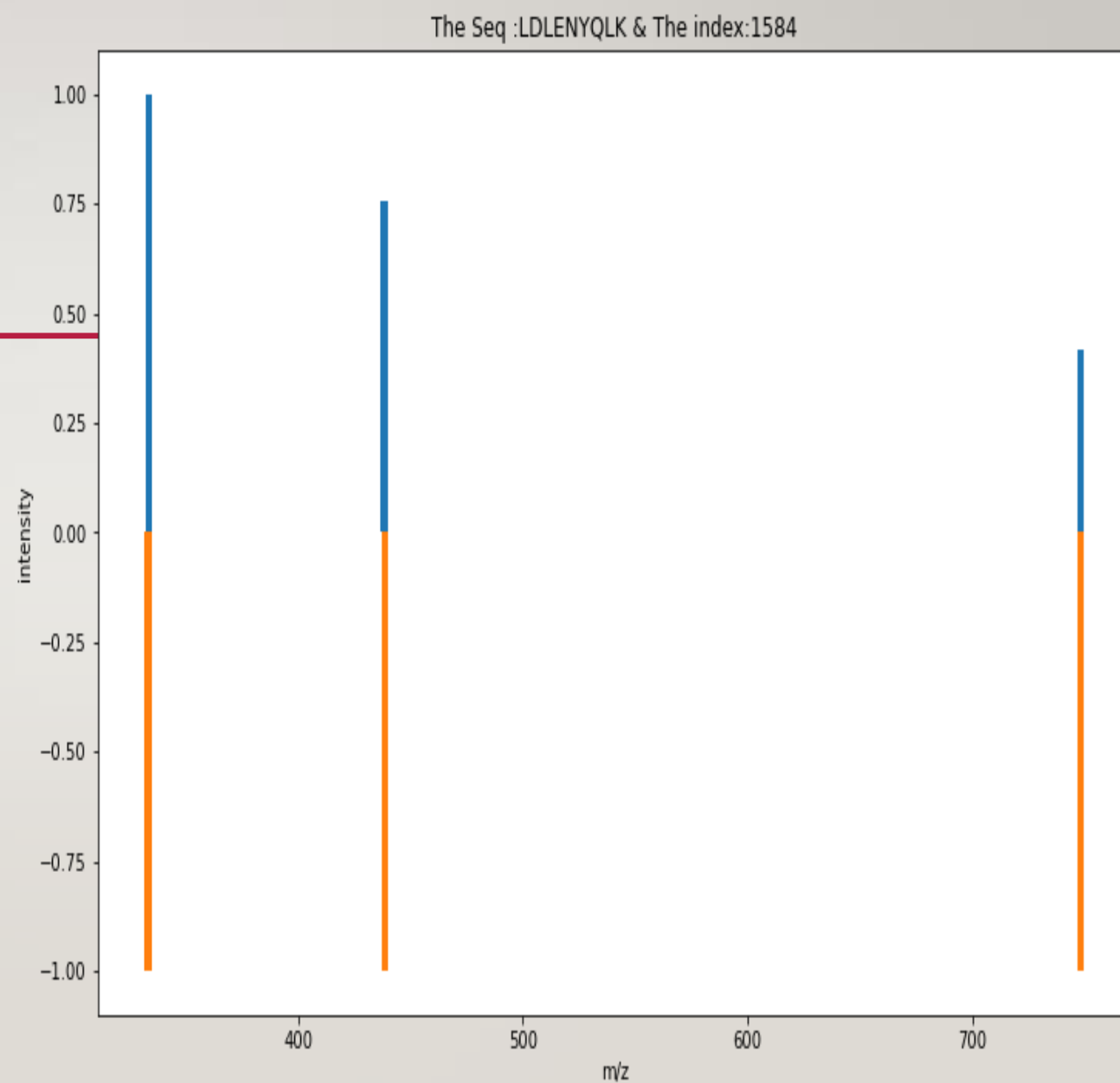
SIXTH FILE MZ

Protein

Exp 1

Peptide

LDLENYQLK

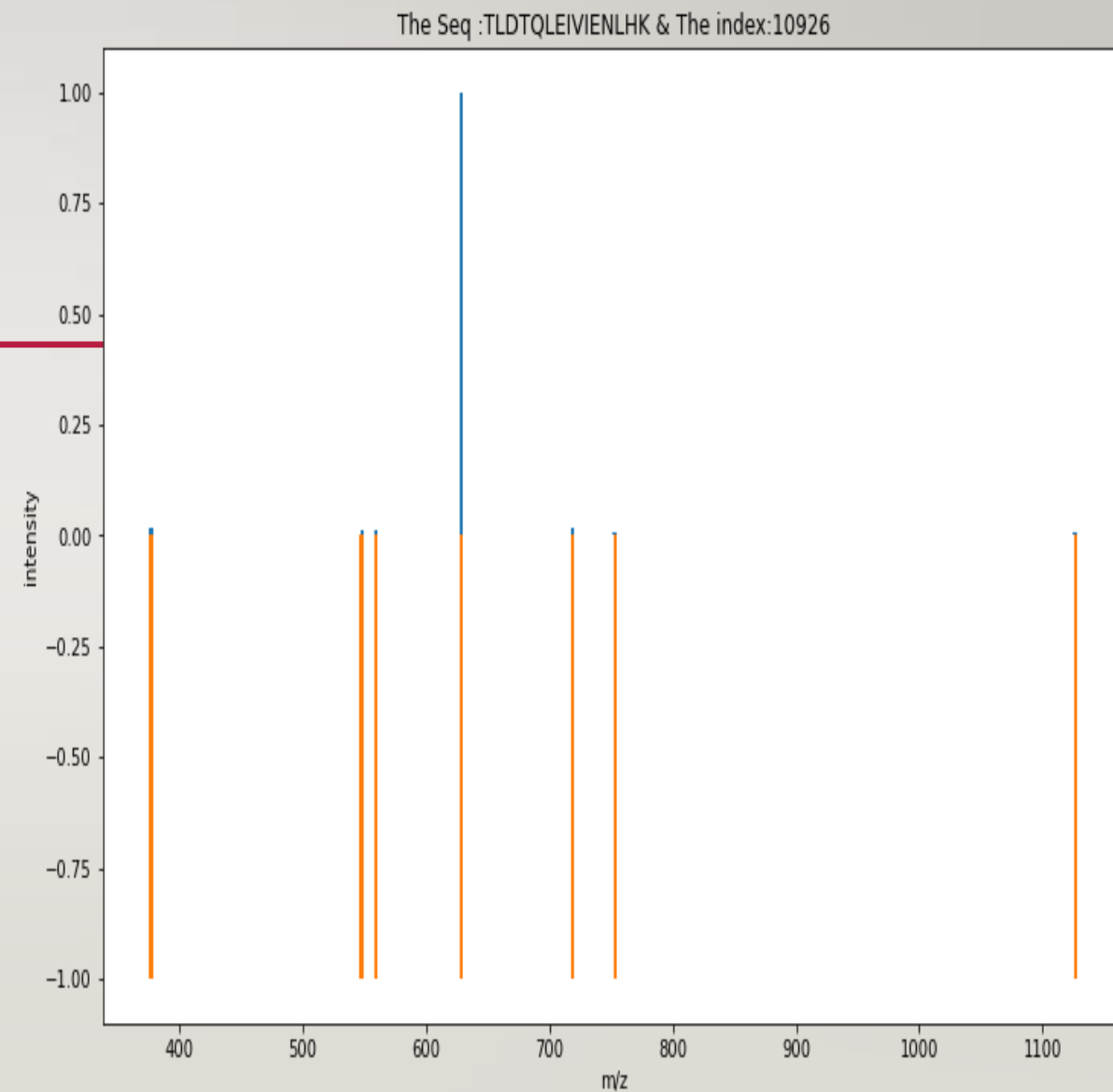


PROTEIN

Exp 2

Peptide

TLDTQLEIVIENLHK



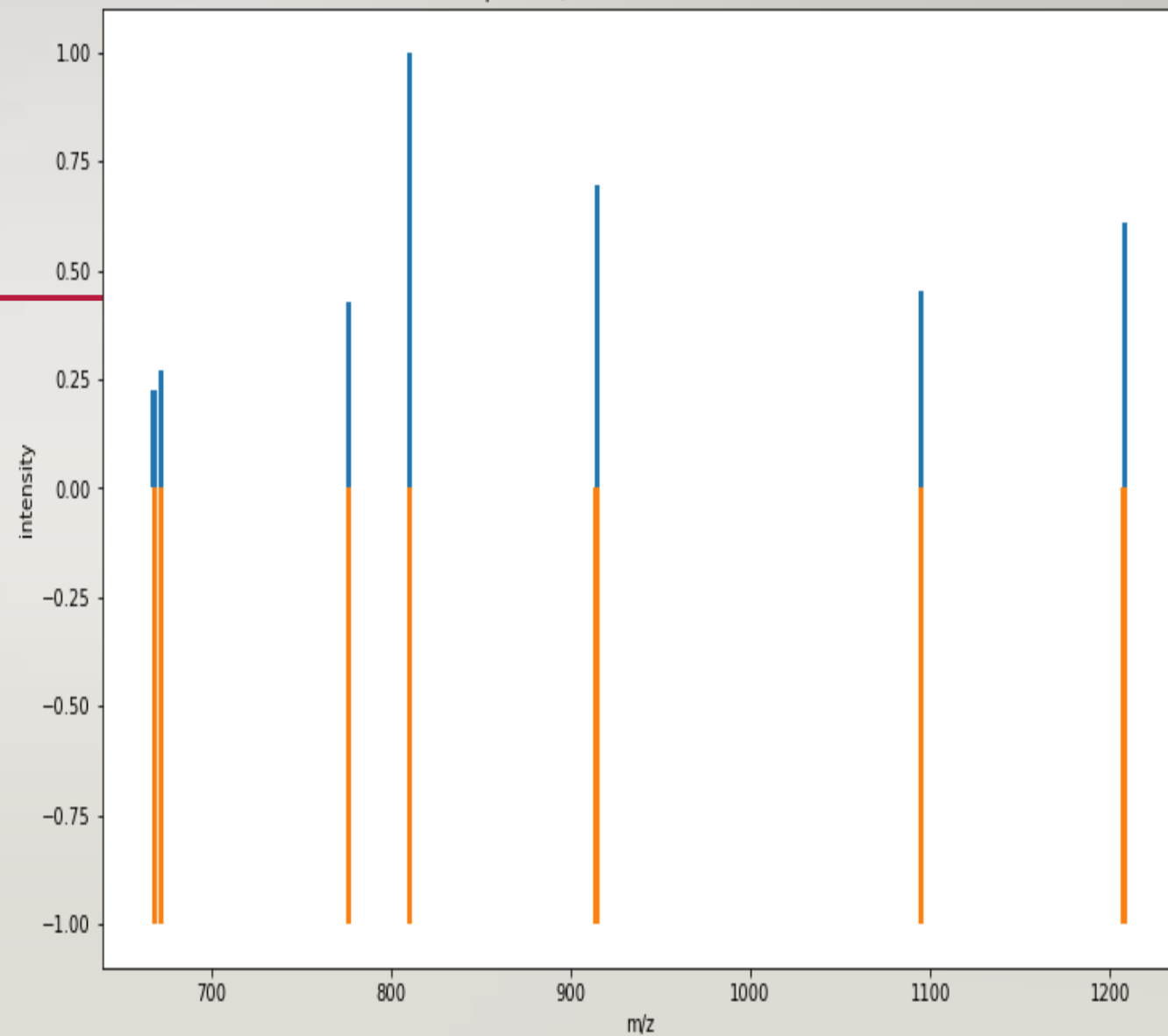
PROTEIN

Exp 3

Peptide

TLDTQLEIVIENLHK

The Seq :TLDTQLEIVIENLHK & The index:17640



SEVENTH FILE MZ

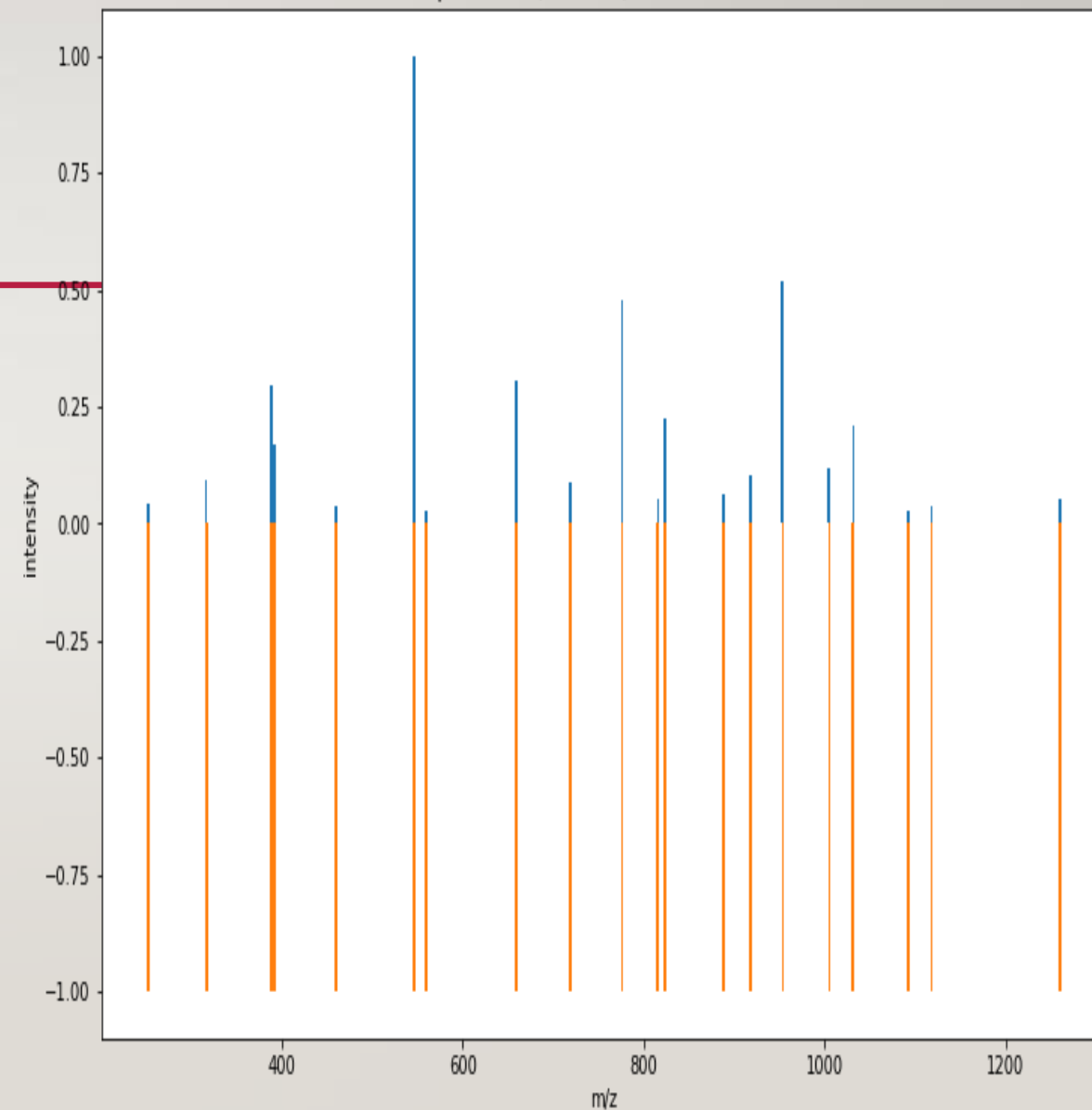
Protein

Exp 1

Peptide

EEFLSAQSSPAGSQLESR

The Seq :EEFLSAQSSPAGSQLESR & The index:8899



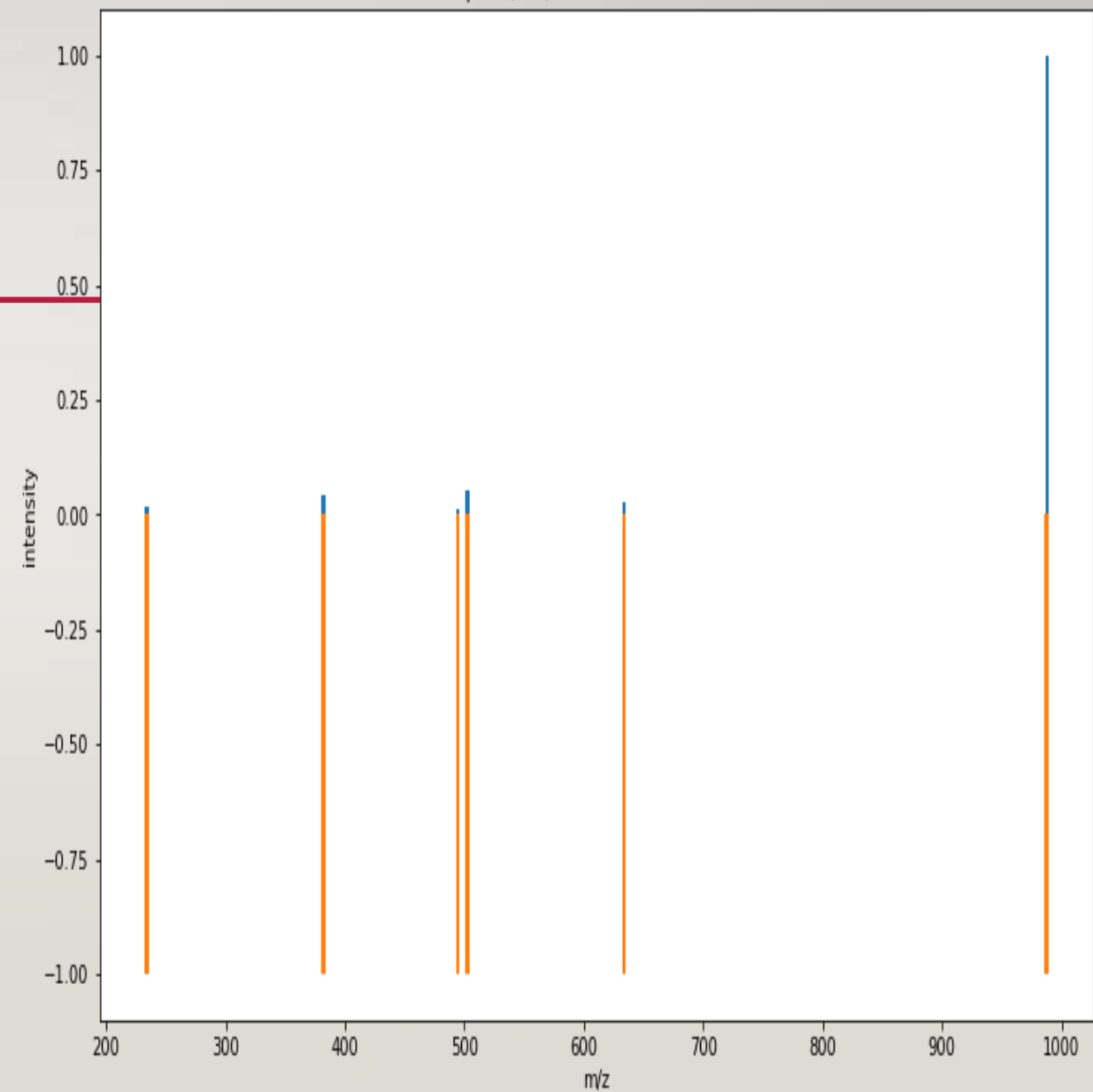
PROTEIN

Exp 2

Peptide

FIIQFLQDFLHSK

The Seq :FIIQFLQDFLHSK & The index:13160



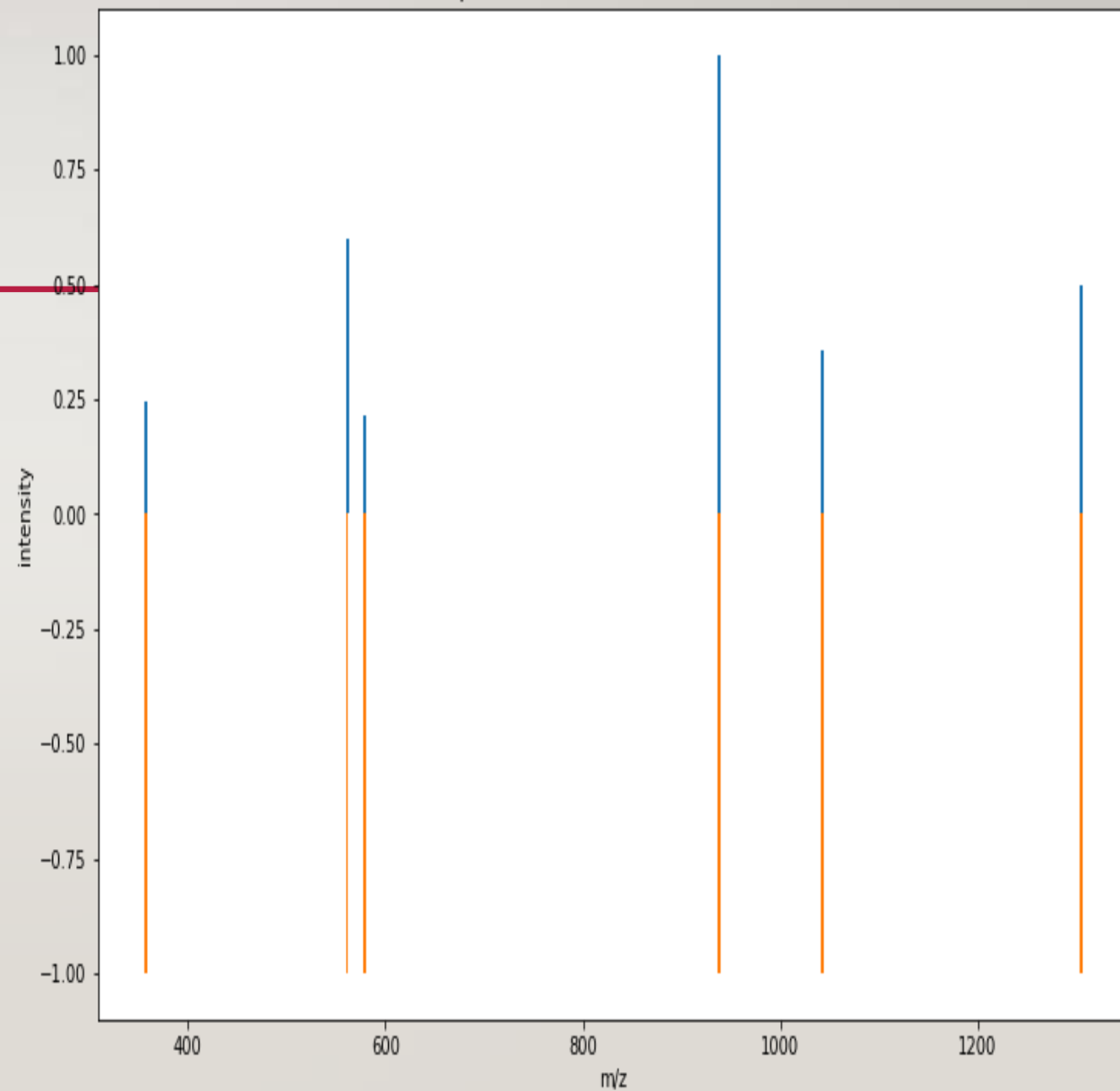
PROTEIN

Exp 3

Peptide

DGNAPSLGNDTDFR

The Seq :DGNAPSLGNDTDFR & The index:21676



EIGHTH FILE MZ

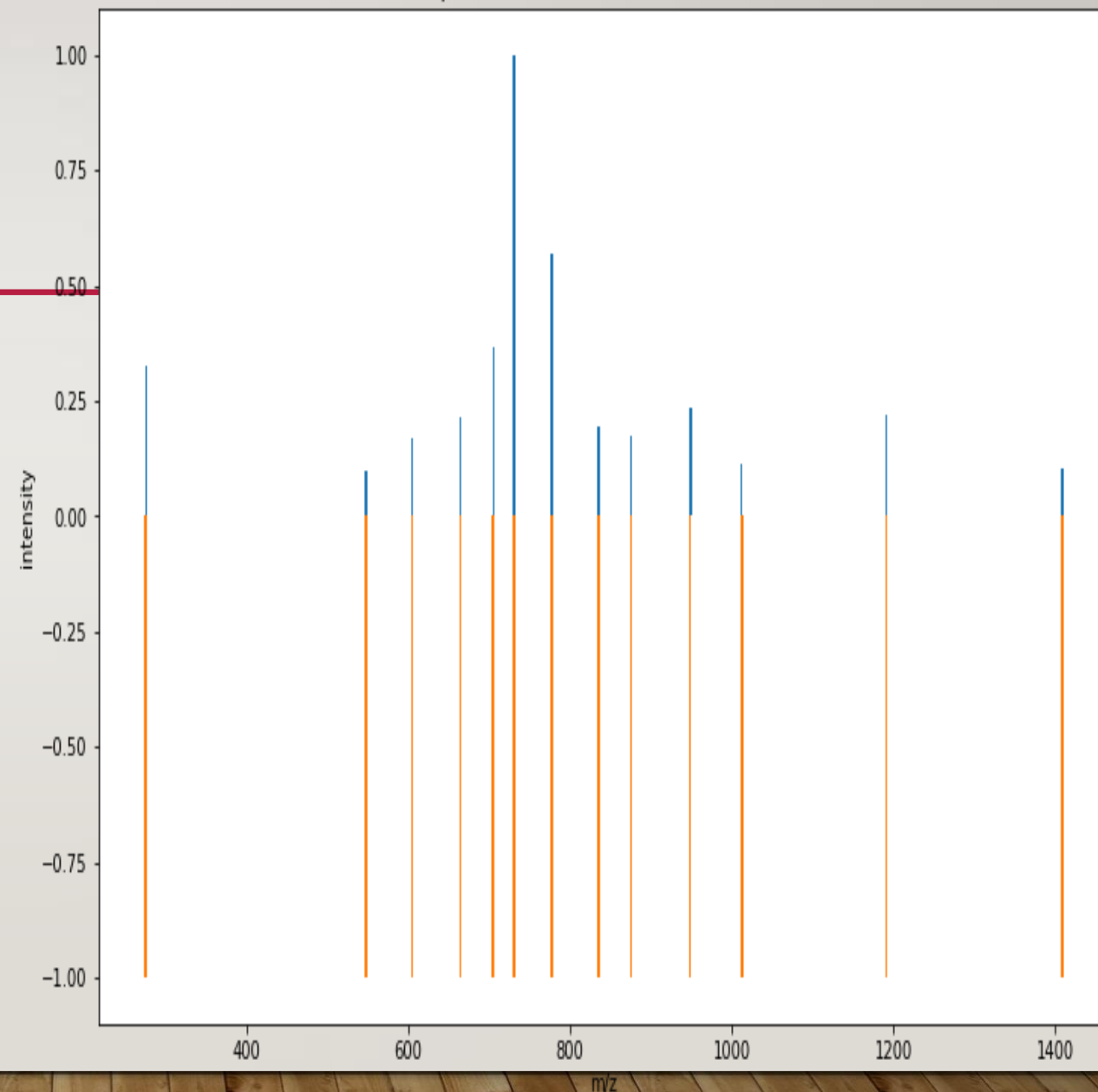
Protein

Exp 1

Peptide

FNQNLLHLDSIFNIL

The Seq :FNQNLLHLDSIFNILQK & The index:22543



LINKS OF RAW AND FASTA FILE:

- RAW FILE LINK:
 - <https://www.ebi.ac.uk/pride/archive/projects/PXD029458>
- FASTA FILE LINK:
 - <https://www.uniprot.org/uniprot/P39723>