- grepq: A Rust application that quickly filters
- ² FASTQ files by matching sequences to a set
- of regular expressions
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- **regular expressions**
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Abstract

Regular expressions (regex) (Kleene 1951) have been an important tool for finding patterns in biological codes for decades (Hodgman 2000 and citations therein), and unlike fuzzy-finding approaches, do not result in approximate matches. The performance of regular expressions can be slow, however, especially when searching for matching patterns in large files. grepq is a Rust application that quickly filters FASTQ files by matching sequences to a set of regular expressions. grepq is designed 23 with a focus on performance and scalability, is easy to install and easy to use, enabling users to quickly filter large FASTQ files, to enumerate named and unnamed variants, to update the order in which patterns are matched against sequences through in-built tune and summarise commands, and optionally, to output a SQLite file for further sequence analysis. grepq is open-source and available on GitHub and Crates.io.

Statement of need

- The ability to quickly filter FASTQ files by matching sequences to a set of regular expressions is an important task in bioinformatics, especially when working with large datasets. The importance and
- challenge of this task will only grow as sequencing technologies

continue to advance and produce ever larger datasets (Katz et al. 2022). The uses cases of *grepq* are diverse, and include pre-processing of FASTQ files before downstream analysis, quality control of sequencing data, and filtering out unwanted sequences.

Where decisions need be made quickly, such as in a clinical settings (Bachurin et al. 2024), biosecurity (Valdivia-Granda 2012), and wastewater-based epidemiology in support of public health measures (Choi et al. 2018; Sims and Kasprzyk-Hordern 2020; Xylogiannopoulos 2021; Merrett et al. 2024), the ability to quickly filter FASTQ files and enumerate named and unnamed variants by matching sequences to a set of regular expressions is attractive as it circumvents the need for more time-consuming bioinformatic workflows.

Regular expressions are a powerful tool for matching sequences,
but they can be slow and inefficient when working with large
datasets. Furthermore, general purpose tools like *grep* (Free
Software Foundation 2023) and *ripgrep* (A. Gallant 2025) are
not optimized for the specific task of filtering FASTQ files, and
ocassionaly yield false positives as they scan the entire FASTQ
record, including the sequence quality field. Tools such *awk* (Aho,
Kernighan, and Weinberger 1988) and *gawk* (Free Software
Foundation 2024) can be used to filter FASTQ files without yielding false positives, but they are significantly slower than *grepq*

and can require the development of more complex scripts to
 achieve the same result.

Implementation

- grepq is implemented in Rust, a systems programming language
 known for its safety features, which help prevent common pro-
- gramming errors such as null pointer dereferences and buffer over-
- 65 flows. These features make Rust an ideal choice for implementing
- a tool like *grepq*, which needs to be fast, efficient, and reliable.
- Furthermore, *grepg* obtains its performance and reliability, in part,
- by using the seq_io (Schlegel and Seyboldt 2025) and regex (Gal-
- lant et al. 2025b) libraries. The seq_io library is a well-tested
- ₇₀ library for parsing FASTQ files, designed to be fast and efficient,
- and which includes a module for parallel processing of FASTQ
- records through multi-threading. The regex library is designed
- to work with regular expressions and sets of regular expressions,
- and is known to be one of the fastest regular expression libraries
- _{rs} currently available (Gallant et al. 2025a). The *regex* library sup-
- ₇₆ ports Perl-like regular expressions without look-around or backref-
- erences (documented at https://docs.rs/regex/1.*/regex/#syntax).
- Further performance gains were obtained by:
 - use of the RegexSet struct from the regex library to match

- multiple regular expressions against a sequence in a single
 pass, rather than matching each regular expression individually (the *RegexSet* is created and compiled once before entering any loop that processes the FASTQ records, avoiding
 the overhead of recompiling the regular expressions for each
 record)
- multi-threading to process the records within an input FASTQ
 file in parallel through use of multiple CPU cores
- use of the *zlib-ng* backend to the *flate2* library to read and
 write gzip-compressed FASTQ files, which is faster than the
 default *miniz_oxide* backend
- use of an optimised global memory allocator (the *mimalloc* library (Mutiple, n.d.)) to reduce memory fragmentation and improve memory allocation and deallocation performance
- buffer reuse to reduce the number of memory allocations and
 deallocations
- use of byte slices to avoid the overhead of converting to and
 from string types
 - in-lining of performance-critical functions
- use of the *write_all* I/O operation that ensures the data is written in one go, rather than writing data in smaller chunks

Feature set

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102 *grepq* has the following features:

- support for presence and absence (inverted) matching of a set of regular expressions
- IUPAC ambiguity code support (N, R, Y, etc.)
 - support for gzip and zstd compression (reading and writing)
 - JSON support for pattern file input and tune and summarise command output, allowing named regular expression sets and named regular expressions (pattern files can also be in plain text)
 - the ability to:
 - set predicates to filter FASTQ records on the header field (= record ID line) using a regular expression, minimum sequence length, and minimum average quality score (supports Phred+33 and Phred+64)
 - output matched sequences to one of four formats (including FASTQ and FASTA)
 - tune the pattern file and enumerate named and unnamed variants with the *tune* and *summarise* commands: these commands will output a plain text or JSON file with the patterns sorted by their frequency of occurrence in the input FASTQ file or gzip-compressed FASTQ file (or a userspecified number of total matches). This can be useful for

optimizing the pattern file for performance, for example by removing patterns that are rarely matched and reordering nucleotides within the variable regions of the patterns to improve matching efficiency

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- count and summarise the total number of records and the number of matching records (or records that don't match in the case of inverted matching) in the input FASTQ file
- bucket matching sequences to separate files named after each regexName with the -bucket flag, in any of the four output formats

Other than when the **inverted** command is given, output to a 134 SQLite database is supported with the writeSQL option. The 135 SQLite database will contain a table called fastq_data with the following fields: the fastq record (header, sequence and quality 137 fields), length of the sequence field (length), percent GC content (GC), percent GC content as an integer (GC int), number of 139 unique tetranucleotides in the sequence (nTN), percent tetranucleotide frequency within the sequence (TNF), and a JSON array 141 containing the matched regex patterns, the matches and their po-142 sition(s) in the FASTQ sequence (variants). If the pattern file was given in JSON format and contained a non-null qualityEncoding 144 field, then the average quality score for the sequence field (average_quality) will also be written. The -num-tetranucleotides

option can be used to limit the number of tetranucleotides written
to the TNF field of the fastq_data SQLite table, these being the
most or equal most frequent tetranucleotides in the sequence
field of the matched FASTQ records. A summary of the invoked
query (pattern and data files) is written to a second table called
query.

Other than when the *tune* or *summarise* command is run, a FASTQ record is deemed to match (and hence provided in the output) when any of the regular expressions in the pattern file match the sequence field of the FASTQ record. Example output of the *tune* command (when given with the **–json-matches** flag) is shown below:

```
# For each matched pattern in a search of no more than
# 20000 matches of a gzip-compressed FASTQ file, print
# the pattern and the number of matches to a JSON file
# called matches.json, and include the top three most
# frequent variants of each pattern, and their respective
# counts
grepq --read-gzip 16S-no-iupac.json SRX26365298.fastq.gz \
tune -n 20000 -c --names --json-matches --variants 3

Output (abridged) written to matches.json:
{
    "regexSet": {
```

```
"regex": [
            {
                "regexCount": 2,
                "regexName": "Primer contig 06a",
                "regexString": "[AG]AAT[AT]G[AG]CGGGG",
                "variants": [
                    {
                        "count": 1,
                        "variant": "GAATTGGCGGGG",
                        "variantName": "06a-v3"
                    },
                    {
                        "count": 1,
                        "variant": "GAATTGACGGGG",
                        "variantName": "06a-v1"
                    }
                ]
            },
            // matches for other regular expressions...
    ],
    "regexSetName": "conserved 16S rRNA regions"
  }
}
```

To output all variants of each pattern, use the --all argument, for example:

```
# For each matched pattern in a search of no more than
# 20000 matches of a gzip-compressed FASTQ file, print
# the pattern and the number of matches to a JSON file
# called matches.json, and include all variants of each
# pattern, and their respective counts. Note that the
# --variants argument is not given when --all is specified.
grepq --read-gzip 16S-no-iupac.json SRX26365298.fastq.gz \
tune -n 20000 -c --names --json-matches --all
```

When the count option (-c) is given with the tune or summarise command, grepq will count the number of FASTQ records containing a sequence that is matched, for each matching regular 164 expression in the pattern file. If, however, there are multiple oc-165 currences of a given regular expression within a FASTQ record sequence field, grepq will count this as one match. To ensure all records are processed, the *summarise* command is used instead of the *tune* command. Further, note that counts produced through 169 independently matching regex patterns to the sequence field of a 170 FASTQ record inherently underestimate the true number of those 171 patterns in the biological sample, since a regex pattern may span two reads (i.e., be truncated at either the beginning or end of a read). To illustrate, a regex pattern representing a 12-mer motif has a 5.5% chance of being truncated for a read length of 400 nucleotides (11/400 + 11/400 = 22/400 = 0.055 or 5.5%), assuming a uniform distribution of motif positions and reads are sampled randomly with respect to motifs (this calculation would need to be adjusted to the extent that motifs are not uniformly distributed and reads are not randomly sampled with respect to motifs).

When the count option (**-c**) is not given as part of the *tune* or *sum-marise* command, *grepq* provides the total number of matching
FASTQ records for the set of regular expressions in the pattern
file.

Colorized output for matching regular expressions is not implemented to maximise speed and minimise code complexity, but can be achieved by piping the output to *grep* or *ripgrep* for testing purposes.

189 Performance

The performance of *grepq* was compared to that of *fqgrep*, *seqkit grep*, *ripgrep*, *grep*, *awk*, and *gawk* using the benchmarking tool *hyperfine*. The test conditions and results are shown in **Table 1**, **Table 2** and **Table 3**.

Table 1: Wall times and speedup of various tools for filtering FASTQ records

against a set of regular expressions. Test FASTQ file: SRX26365298.fastq (uncompressed) was 874MB in size, and contained 869,034 records.

tool	wall time (s)		speedup		
	mean	S.D.	× grep	× ripgrep	× awk
grepq	0.192	0.010	1796.76	18.62	863.52
fqgrep	0.338	0.005	1017.61	10.55	489.07
ripgrep	3.568	0.005	96.49	1.00	46.37
seqkit grep	2.885	0.011	119.33	1.24	57.35
grep	344.259	0.545	1.00	0.01	0.48
awk	165.451	1.590	2.08	0.02	1.00
gawk	287.662	1.682	1.20	0.01	0.58

grepq v1.4.0, fqgrep v.1.02, ripgrep v14.1.1, seqkit grep v.2.9.0, grep 2.6.0-FreeBSD, awk v. 20200816, and gawk v.5.3.1. fqgrep and seqkit grep were run with default settings, ripgrep was run with -B 1 -A 2 --colors 'match:none' --no-line-number, and grep was run with -B 1 -A 2 --color=never. awk and gawk scripts were also configured to output matching records in FASTQ format. The pattern file contained 30 regular expression representing the 12-mers (and their reverse compliment) from Table 3 of Martinez-Porchas et al. (2017). The wall times, given in seconds, are the mean of 10 runs, and S.D. is the standard deviation of the wall times, also given in seconds.

Table 2: Wall times and speedup of various tools for filtering gzip-compressed
FASTQ records against a set of regular expressions. Test FASTQ file:
SRX26365298.fastq.gz was 266MB in size, and contained 869,034 records.

tool	wall ti	me (s)	speedup
1001	mean	S.D.	× ripgrep
grepq	1.703	0.002	2.10
fqgrep	1.834	0.005	1.95
ripgrep	3.584	0.013	1.00

Test conditions and tool versions as above, but *grepq* was run with the **-read-gzip** option, *fqgrep*with the **-z** option, and *ripgrep* with the **-z** option. SRX26365298.fastq was gzip-compressed using
the *gzip* v.448.0.3 command (Apple Inc. 2019) using default (level 6) settings. The pattern file

contained 30 regular expression representing the 12-mers (and their reverse compliment) from
Table 3 of Martinez-Porchas et al. (2017). The wall times, given in seconds, are the mean of 10
runs, and S.D. is the standard deviation of the wall times, also given in seconds.

Table 3: Wall times and speedup of various tools for filtering FASTQ records
 against a set of regular expressions. Test FASTQ file: SRX22685872.fastq was
 104GB in size, and contained 139,700,067 records.

tool	wall tin	speedup			
1001	mean	S.D.	× ripgrep		
	Uncompressed				
grepq	26.972	0.244	4.41		
fqgrep	50.525	0.501	2.36		
ripgrep	119.047	1.227	1.00		
	gzip-compressed				
grepq	149.172	1.054	0.98		
fqgrep	169.537	0.934	0.86		
ripgrep	144.333	0.243	1.00		

Test conditions and tool versions as described in the footnote to Table 1. Note that when *grepq* was
run on the gzip-compressed file, a memory resident time for the *grepq* process of 116M as reported
by the *top* command (Apple Inc. 2023c). *fastq-dump* v3.1.1 (Sherry et al. 2012) was used to
download SRX22685872 as a gzip compressed file from the NCBI SRA. The pattern file contained
30 regular expression representing the 12-mers (and their reverse compliment) from Table 3 of
Martinez-Porchas et al. (2017). The wall times, given in seconds, are the mean of 10 runs, and
S.D. is the standard deviation of the wall times, also given in seconds.

Testing

The output of *grepq* was compared against the output of *fqgrep*, *seqkit grep*, ripgrep, grep, awk and gawk, using the stat command (Apple Inc. 2023b),

- 227 and any difference investigated using the diff command (Apple Inc. 2023a).
- Furthermore, a custom utility, spikeq (Crosbie 2024b), was developed to gen-
- erate synthetic FASTQ files with a known number of records and sequences
- 230 with user-specified lengths that were spiked with a set of regular expressions a
- 231 known number of times. This utility was used to test the performance of grepq
- and the aforementioned tools under controlled conditions.
- Finally, a bash test script (see examples/test.sh, available at grepq's Github
- repository) and a simple Rust CLI application, predate (Crosbie 2024a), were
- ²³⁵ developed and utilised to automate system testing, and to monitor for perfor-
- 236 mance regressions.
- 237 grepq has been tested on macOS 15.0.1 (Apple M1 Max) and Linux Ubuntu
- 20.04.6 LTS (AMD EPYC 7763 64-Core Processor). It may work on other plat-
- forms, but this has not been tested.

Availability and documentation

- ²⁴¹ *grepq* is open-source and available at *GitHub* (https://github.com/Rbfinch/gre
- pq) and *Crates.io* (https://crates.io/crates/grepq).
- Documentation and installation instructions for grepg are available at the same
- GitHub repository, and through the -h and -help command-line options, which
- includes a list of all available commands and options, and examples of how to
- use them. Example pattern files in plain text and JSON format are also provided,
- 247 as well as test scripts. *grepq* is distributed under the MIT license.

248 Conclusion

The performance of grepq was compared to that of fqgrep, seqkit grep, ripgrep, 249 grep, awk, and gawk using the benchmarking tool hyperfine. For an uncom-250 pressed FASTQ file 874MB in size, containing 869,034 records, grepq was 251 significantly faster than the other tools tested, with a speedup of 1797 times 252 relative to grep, 864 times relative to awk, and 19 times relative to ripgrep. For 253 a larger uncompressed FASTQ file (104GB in size, and containing 139,700,067 254 records), grepg was 4.4 times faster than ripgrep and marginally slower or of equivalent speed to ripgrep where the same large file was gzip-compressed. 256 When coupled with its exceptional runtime performance, grepq's feature set make it a powerful and flexible tool for filtering large FASTQ files. 258

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266 Conflicts of interest

The author declares no conflicts of interest.

References

Aho, Alfred V., Brian W. Kernighan, and Peter J. Weinberger. 1988. *The AWK*Programming Language. https://www.cs.princeton.edu/~bwk/btl.mirror/.

```
Apple Inc. 2019. The Gzip Command.
      ——. 2023a. The Diff Command.
       —. 2023b. The Stat Command.
273
   ——. 2023c. The Top Command.
274
    Bachurin, Stanislav S, Mikhail V Yurushkin, Ilya A Slynko, Mikhail E Kletskii,
275
       Oleg N Burov, and Dmitriy P Berezovskiy. 2024. "Structural Peculiarities
276
       of Tandem Repeats and Their Clinical Significance." Biochemical and Bio-
277
       physical Research Communications 692: 149349.
278
    Choi, Phil M, Ben J Tscharke, Erica Donner, Jake W O'Brien, Sharon C Grant,
       Sarit L Kaserzon, Rachel Mackie, et al. 2018. "Wastewater-Based Epidemi-
280
       ology Biomarkers: Past, Present and Future." TrAC Trends in Analytical
       Chemistry 105: 453-69.
282
    Crosbie, Nicholas D. 2024a. "predate: Catch bugs and performance regres-
       sions through automated system testing." https://github.com/Rbfinch/preda
284
       te.
             2024b. "spikeg: Generates synthetic FASTQ records free of se-
       quences defined by regex patterns, or containing spiked sequences based
287
       on regex patterns." https://github.com/Rbfinch/spikeq.
288
    Free Software Foundation. 2023. GNU Grep 3.11. Free Software Foundation.
289
       https://www.gnu.org/software/grep/manual/grep.html.
290
       — 2024. GAWK: Effective AWK Programming: A User's Guide for GNU
291
       Awk, for the 5.3.1. Free Software Foundation. https://www.gnu.org/softwa
292
       re/gawk/manual/gawk.html.
293
    Gallant et al. 2025a. "rebar." https://github.com/BurntSushi/rebar.
       — et al. 2025b. "regex." https://github.com/rust-lang/regex.
295
    Gallant, Andrew. 2025. "Ripgrep: Recursively Search the Current Directory for
       Lines Matching a Pattern." https://github.com/BurntSushi/ripgrep.
297
```

- Hodgman, T. Charles. 2000. "A Historical Perspective on Gene/Protein Functional Assignment." *Bioinformatics* 16 (1): 10–15.
- Katz, Kenneth, Oleg Shutov, Richard Lapoint, Michael Kimelman, J Rodney
- Brister, and Christopher O'Sullivan. 2022. "The Sequence Read Archive:
- A Decade More of Explosive Growth." Nucleic Acids Research 50 (D1):
- 303 D387-90.
- Kleene, SC. 1951. "Representationof Events in Nerve Nets and Finite Automata." *CE Shannon and J. McCarthy*.
- Martinez-Porchas, Marcel, Enrique Villalpando-Canchola, Luis Enrique Ortiz
 Suarez, and Francisco Vargas-Albores. 2017. "How Conserved Are the
 Conserved 16S-rRNA Regions?" *PeerJ* 5: e3036.
- Merrett, James E, Monica Nolan, Leon Hartman, Nijoy John, Brianna Flynn,
- Louise Baker, Christelle Schang, et al. 2024. "Highly Sensitive Wastewater
- Surveillance of SARS-CoV-2 Variants by Targeted Next-Generation Ampli-
- con Sequencing Provides Early Warning of Incursion in Victoria, Australia."
- Applied and Environmental Microbiology 90 (8): e01497–23.
- Mutiple. n.d. "Mimalloc: A Rust Wrapper over Microsoft's MiMalloc Memory

 Allocator."
- Schlegel, Markus, and Adrian Seyboldt. 2025. "seq_io: FASTA and FASTQ parsing and writing in Rust." https://github.com/markschl/seq_io.
- Sherry, Stephen, Chunlin Xiao, Kenneth Durbrow, Michael Kimelman, Kurt
 Rodarmer, Martin Shumway, and Eugene Yaschenko. 2012. "NCBI Sra
 Toolkit Technology for Next Generation Sequence Data." In *Plant and*Animal Genome XX Conference (January 14-18, 2012). Plant and Animal
- 322 Genome.
- Sims, Natalie, and Barbara Kasprzyk-Hordern. 2020. "Future Perspectives
 of Wastewater-Based Epidemiology: Monitoring Infectious Disease Spread

- and Resistance to the Community Level." *Environment International* 139: 105689.
- Valdivia-Granda, Willy A. 2012. "Biodefense Oriented Genomic-Based
 Pathogen Classification Systems: Challenges and Opportunities." *Journal* of Bioterrorism & Biodefense 3 (1): 1000113.
- Xylogiannopoulos, Konstantinos F. 2021. "Pattern Detection in Multiple
 Genome Sequences with Applications: The Case of All SARS-CoV-2
 Complete Variants." bioRxiv, 2021–04.